

การผลิตน้ำมันระเหยจากเนื้อเยื่อพะไลงโกธูจพาลำพา

นางสาวศุภวรรณ เจียมทะวงษ์

ศูนย์วิทยทรัพยากร

จุฬาลงกรณ์มหาวิทยาลัย

วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาเภสัชศาสตรมหาบัณฑิต

สาขาวิชาเภสัชเวท ภาควิชาเภสัชเวท

คณะเภสัชศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย

ปีการศึกษา 2544

ISBN 974-17-0229-9

ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

**PRODUCTION OF ESSENTIAL OIL
FROM *ARTEMISIA DUBIA* TISSUE CULTURE**

MISS SUPAWAN CHIAMTAWONGSE

**A Thesis Submitted in Partial Fulfillment of the Requirements
for the Degree of Master of Science in Pharmacy**

**Department of Pharmacognosy
Faculty of Pharmaceutical Sciences**

Chulalongkorn University

Academic Year 2001

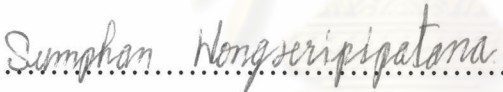
ISBN 947-17-0229-9


Thesis Title PRODUCTION OF ESSENTIAL OIL
FROM *ARTEMISIA DUBIA* TISSUE CULTURE
By Miss Supawan Chiamtawongse
Field of Study Pharmacognosy
Thesis Advisor Associate Professor Nijisiri Ruangrunsi, Ph.D.
Thesis Co-advisor Thanapat Songsak, Ph.D.

Accepted by the Faculty of Pharmaceutical Sciences, Chulalongkorn
University in Partial Fulfillment of the Requirements for the Master's Degree


..... Dean of Faculty of Pharmaceutical Sciences
(Associate Professor Boonyong Tantisira, Ph.D.)

THESIS COMMITTEE


..... Chairman
(Associate Professor Samphan Wongseripipatana, Ph.D.)


..... Thesis Advisor
(Associate Professor Nijisiri Ruangrunsi, Ph.D.)


..... Thesis Co-advisor
(Thanapat Songsak, Ph.D.)


..... Member
(Assistant Professor Sompop Pratharturug, Ph.D.)

ศุภวรรณ เขียมทะวงษ์ : การผลิตน้ำมันระเหยจากเนื้อเยื่อเพาะเลี้ยงโอรุจุพาลำพา (PRODUCTION OF ESSENTIAL OIL FROM *ARTEMISIA DUBIA* TISSUE CULTURE) อาจารย์ที่ปรึกษา : รศ.ดร.นิจศิริ เรืองรังษี, อาจารย์ที่ปรึกษาร่วม : อ.ดร.ธนภัทร ทรงศักดิ์, 101 หน้า. ISBN 974-17-0229-9

การเพาะเลี้ยงเซลล์ของโอรุจุพาลำพา (*Artemisia dubia* Wall. ex Bess.) ซึ่งเป็นพืชในวงศ์ Asteraceae (Compositae) เริ่มต้นจากการนำชิ้นใบที่ผ่านการฆ่าเชื้อแล้ว วางบนอาหารเพาะเลี้ยงกึ่งแข็งชนิด MS ที่ประกอบด้วย 2,4-dichlorophenoxyacetic acid 1 มิลลิกรัมต่อลิตร และ kinetin 0.1 มิลลิกรัมต่อลิตร เพื่อชักนำให้เกิดเป็นแคลลัส หลังจากนั้นนำแคลลัสที่ได้เปลี่ยนถ่ายลงในอาหารเหลวชนิดเดียวกัน เพื่อเพาะเลี้ยงให้เป็นเซลล์เพาะเลี้ยงแขวนลอย เมื่อใช้วิธี Gas Chromatography (GC) และ Gas Chromatography-Mass Spectrometry (GC-MS) วิเคราะห์และเปรียบเทียบองค์ประกอบเคมีของน้ำมันระเหยที่ได้จากต้นจริง และจากการเพาะเลี้ยงเนื้อเยื่อพืช พบว่า น้ำมันระเหยที่ได้จากแคลลัสและเซลล์เพาะเลี้ยงแขวนลอยมีองค์ประกอบเคมีหลักเหมือนกับต้นจริง คือ สารกลุ่ม sesquiterpenes ชื่อ davanone แต่ปริมาณของ davanone ที่วิเคราะห์ได้นั้นน้อยมากเมื่อเปรียบเทียบกับต้นจริง จึงได้นำกลวิธีทางเทคโนโลยีชีวภาพมาใช้เพื่อเพิ่มความสามารถของเซลล์ในการสร้าง davanone ได้แก่ การใช้ตาข่ายในล่อนเพื่อตรึงเซลล์ ร่วมกับการเติม geranyl acetate ความเข้มข้นต่างๆ เพื่อเป็นสารตั้งต้นในกระบวนการเปลี่ยนแปลงทางชีวภาพ และการใช้สารดูดซับเพื่อช่วยดูดซับน้ำมันระเหยที่ถูกขับออกมาจากเซลล์ ซึ่งจะสามารถวัดปริมาณของ davanone ได้สูงขึ้นหลังจากการนำกลวิธีทางเทคโนโลยีชีวภาพดังกล่าวมาใช้

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

ภาควิชาเภสัชเวท.....
สาขาวิชาเภสัชเวท.....
ปีการศึกษา2544.....

ลายมือชื่อนิสิตศุภวรรณ เขียมทะวงษ์.....
ลายมือชื่ออาจารย์ที่ปรึกษานิจศิริ เรืองรังษี.....
ลายมือชื่ออาจารย์ที่ปรึกษาร่วมธนภัทร ทรงศักดิ์.....

4376628133 : MAJOR PHARMACOGNOSY

KEYWORD : *ARTEMISIA DUBIA* WALL. EX BESS. / ASTERACEAE /
ESSENTIAL OIL / PLANT TISSUE CULTURE / CHEMICAL
CONSTITUENTS

SUPAWAN CHAIMTAWONGSE: THESIS TITLE. PRODUCTION OF
ESSENTIAL OIL FROM *ARTEMISIA DUBIA* TISSUE CULTURE.

THESIS ADVISOR: ASSOC. PROF. NIJSIRI RUANGRUNGSI, Ph.D.

THESIS CO-ADVISOR: THANAPAT SONGSAK, Ph.D., 101 pp.

ISBN 974-17-0229-9

Callus cultures of *Artemisia dubia* Wall. ex Bess., the member of the family Asteraceae (Compositae), were initiated by placing sterilised leaf explants on semi-solid basal MS media, containing 1 mg/l 2,4-dichlorophenoxyacetic acid and 0.1 mg/l kinetin. Suspension cultures were also initiated by using callus cultures on the same media (except without the agar). The chemical constituents of essential oil from callus and suspension cultures were identified and compared to essential oil from intact plant leaves by using Gas Chromatography (GC) and Gas Chromatography-Mass Spectrometry (GC-MS). It was found that callus and suspension cultures produced the same major chemical constituent as the intact plants, sesquiterpenes namely davanone, but it was very low level of davanone recovered from their cultures. Some biotechnological techniques were used for improving the cell capacity to produce davanone. Nylon meshes were used for immobilising cell whilst various concentrations of geranyl acetate were fed for biotransformation process and adsorbent was also used for adsorbing the essential oil excreted from the cells. The level of davanone was increased after using these techniques.

DepartmentPharmacognosy....
Field of study ...Pharmacognosy....
Academic year2001.....

Student's signatureSupawan Chiantawongse.....
Advisor's signatureNisiri Ruangrungsi.....
Co-advisor's signature.....Thanapat Songsak.....

ACKNOWLEDGEMENTS

The author would like to express her sincere gratitude to her thesis advisor, Associate Professor Dr. Nijsiri Ruangrunsi of the Department of Pharmacognosy, Faculty of Pharmaceutical Sciences, Chulalongkorn University, for his helpful suggestion and keen interest.

The author would like to express her deepest appreciation and grateful thanks to her thesis co-advisor, Dr. Thanapat Songsak of the Department of Pharmacognosy, Faculty of Pharmacy, Rangsit University, for his helpful guidance, invaluable advice and continual encouragement throughout the course of this work.

The author would like to thank the tissue culture unit, the Department of Pharmacognosy, Faculty of Pharmacy, Rangsit University, for providing laboratory facilities during this research.

The author would also like to thank Mr. Chattiya Vorkulpinij and Mr. Sittichok Sittichotpong for their helpful guidance and suggestion about Gas Chromatography (GC) and Gas Chromatography-Mass Spectrometry (GC-MS), without them, this research would have not been completed.

The author would like to thank the Graduate School, Chulalongkorn University for granting her partial financial support to conduct this investigation.

Finally, the author would like to express her grateful thanks to her family for their financial support, without whose love, understanding and cheerfulness, this research would have not been succeeded.

CONTENTS

	Page
ABSTRACT (Thai).....	iv
ABSTRACT (English).....	v
ACKNOWLEDGEMENTS.....	vi
CONTENTS.....	vii
LIST OF TABLES.....	xi
LIST OF FIGURES.....	xii
LIST OF ABBREVIATIONS.....	xiii
CHAPTER I INTRODUCTION.....	15
1.1 Plant cell and tissue culture.....	15
1.2 Definition.....	16
1.3 Callus culture.....	16
1.4 Suspension culture.....	16
1.5 Culture medium.....	17
1.5.1 Media compositions.....	17
1.5.2 Plant growth regulators.....	18
1.5.3 Physical conditions.....	19
1.6 Applications.....	19
1.7 Aseptic technique work.....	20
1.8 Secondary metabolites.....	20
1.9 Production of secondary metabolites from plant cell and tissue culture.....	20
1.9.1 Taxol.....	21
1.9.2 Artemisinin.....	22
1.9.3 Ginkgolides.....	23
1.10 Special techniques for improving the secondary metabolite production from plant cell and tissue cultures.....	25
1.10.1 Cell immobilisation.....	25
1.10.2 Feeding of precursors and biotransformation.....	27

CONTENTS (CONT.)

	Page
1.10.3 Permeabilisation.....	29
1.10.4 Elicitation.....	30
1.10.5 Two-phase system.....	32
1.10.6 Cell differentiation.....	33
1.10.7 Used of adsorbent.....	35
1.11 Essential oils.....	35
1.12 Biosynthesis of essential oil constituents.....	36
1.13 Medicinal and commercial uses.....	40
1.14 Production of essential oil from plant cell and tissue cultures..	40
CHAPTER II HISTORICAL.....	44
2.1 Introduction to <i>Artemisia</i> family.....	44
2.2 Example of the <i>Artemisia</i> species that produce essential oils.....	44
2.3 Production of essential oil from <i>Artemisia</i> spp. cell and tissue cultures.....	47
2.4 <i>Artemisia dubia</i> Wall. ex Bess.....	47
2.5 Davanone.....	49
2.6 Biosynthesis of davanone.....	49
CHAPTER III MATERIALS AND METHODS.....	51
3.1 Chemical.....	51
3.2 Plant materials.....	51
3.3 Hydrodistillation of <i>Artemisia dubia</i> Wall. ex Bess. leaves	51
3.4 Identification of essential oil obtained from hydrodistillated of <i>Artemisia dubia</i> Wall. ex Bess. leaves by using Gas Chromatography-Mass spectrometry (GC-MS).....	53
3.5 Preparation of leaf explants.....	53
3.6 General preparation of semi-solid and liquid media.....	54
3.7 Initiation and maintenance of callus cultures	54
3.8 Initiation and maintenance of cell suspension cultures.....	55

CONTENTS (CONT.)

	Page
3.9 Methods for improving the production of essential oil constituents in plant cell culture	55
3.9.1 Cell immobilisation.....	55
3.9.2 Precursor feeding.....	55
3.9.3 Use of adsorbent.....	56
3.9.4 Collected of volatile compounds from <i>Artemisia dubia</i> Wall. ex Bess. immobilised cells which had been fed various concentrations of geranyl acetate.....	56
3.10 Extraction of essential oil.....	58
3.11 Analysis of essential oil from dichloromethane extract by using Gas Chromatography (GC).....	58
3.12 Identification of essential oil from dichloromethane extract by using Gas Chromatography-Mass spectrometry (GC-MS)....	59
3.13 Quantitative analysis of davanone.....	59
CHAPTER IV RESULTS.....	60
4.1 Growth and appearance of callus cultures.....	60
4.2 Growth and appearance of cell suspension cultures.....	62
4.3 Chemical constituents of essential oil from leaves of <i>Artemisia dubia</i> Wall. ex Bess . obtained by hydrodistillation metho.....	64
4.4 GC analysis and GC-MS analysis of dichloromethane extracts from callus and cell suspension cultures.....	67
4.5 GC analysis and GC-MS analysis of dichloromethane extracts of cell suspension cultures after using biotechnological techniques.....	72

CONTENTS (CONT.)

	Page
CHAPTER V DISCUSSIO.....	80
REFERENCES.....	82
APPENDIX.....	94
A. Plant tissue culture terms.....	94
B. Surface sterilising agent formula.....	99
C. MS basal media formul.....	99
D. B5 basal media formula.....	99
VITA.....	101



 ศูนย์วิทยทรัพยากร
 จุฬาลงกรณ์มหาวิทยาลัย

LIST OF TABLES

Table	Page
1. Secondary metabolites produced in high levels by plant cell and tissue cultures.....	24
2. Selected examples of plant species, secondary metabolites and cell immobilisation methods.....	27
3. Selected examples of plant species, precursors feeding and biotransformation products.....	28
4. Selected examples of plant species, permeabilising agents and secondary metabolites.....	30
5. Selected examples of plant species , elicitors and secondary metabolites.....	31
6. Selected example of plant species and two-phase system.....	33
7. Selected examples of plant species and secondary metabolites have been studied in cell differentiation.....	34
8. Selected example of essential oil production in plant cell and tissue cultures.....	43
9. Selected examples of the <i>Artemisia</i> species that produce essential oils.....	45
10. Selected examples of <i>Artemisia</i> spp. which have been studied in essential oil production from cell and tissue cultures.....	47
11. Growth and appearance of callus cultures of <i>A. dubia</i> Wall. ex Bess..	60
12. Growth and appearance of cell suspension cultures of <i>A. dubia</i> Wall. ex Bess.....	62
13. Chemical constituents of essential oil obtained from <i>Artemisia dubia</i> Wall. ex Bess leaves by hydrodistillation.....	66
14. Chemical constituents of essential oil obtained from callus and cell suspension cultures of <i>A. dubia</i> Wall. ex Bess.....	67
15. Davanone contents obtained from dichloromethane extracts.....	76
16. The chemical constituents of plant tissue culture media.....	100

LIST OF FIGURES

Figure	Page
1. Biosynthetic formation of terpenes.....	37
2. Biosynthesis of phenylpropanoid compounds.....	39
3. <i>Artemisia dubia</i> Wall. ex Bess. (<i>A. vulgaris</i> L. var. <i>indica</i> Maxim.)...	48
4. Possible biosynthetic pathway of davanone.....	50
5. Apparatus for essential oil content determination (Cleavenger type apparatus).....	52
6. The apparatus for collecting volatile compounds in immobilised cell which had been fed various concentration of geranyl acetate.....	57
7. The callus cultures of <i>A. dubia</i> Wall. ex Bess.....	61
8. The cell suspension cultures of <i>A. dubia</i> Wall. ex Bess.....	63
9. GC chromatogram of the essential oil from <i>Artemisia dubia</i> Wall. ex Bess. Leaves obtained by hydrodistillation.....	65
10. The GC chromatogram of dichloromethane of callus cultures.....	68
11. The Mass Spectrum of the major constituent, namely davanone.....	69
12. The GC chromatogram of dichloromethane extract of cell suspension cultures.....	70
13. The Mass Spectrum of the major constituent, namely davanone.....	71
14. The GC chromatogram of dichloromethane extracts.....	74
15. A standard curve of davanone contents.....	75
16. Davanone contents ($\mu\text{g/g}$ FW) obtained from dichloromethane extracts of intact plants, callus cultures and cell suspension cultures..	77
17. Davanone contents ($\mu\text{g/g}$ FW) obtained from dichloromethane extractsof immobilised cells after feeding geranyl acetate.....	78
18. Davanone contents ($\mu\text{g/g}$ FW) obtained from dichloromethane extracts, comparison of intact plants, callus cultures, cell suspension cultures and of immobilised cells after feeding various concentrations of geranyl acetate.....	79

LIST OF ABBREVIATIONS

%	=	Percent (part per 100); percentage
μg	=	Microgram
μl	=	Microlitre
/	=	Per
2,4-D	=	2,4-Dichlorophenoxyacetic acid
AOAC	=	Association of Official Analytical Chemists
B5	=	Gamborge B5 medium
$^{\circ}\text{C}$	=	Degree Celsius
cm	=	Centimeter(s)
cont.	=	Continued
DW	=	Dry weight
ed(s)	=	Editor(s)
e.g.	=	For example
EO	=	Essential oil
<i>et al.</i>	=	Et alii
eV	=	Electron volt
FW	=	Fresh weight
FID	=	Flame Ionization Detector
Fig.	=	Figure
g	=	Gram(s)
GC	=	Gas Chromatography
GC-MS	=	Gas Chromatography-Mass Spectrometry
h	=	Hour(s)
IC ₅₀	=	50 % Inhibitory concentration
l	=	Liter(s)
M	=	Molar
mg/l	=	Milligram per liter
min	=	Minute(s)
ml	=	Milliliter(s)

LIST OF ABBREVIATIONS (CONT.)

mm	=	Millimeter(s)
MS	=	Murashige and Skoog's media
NIST	=	National Institute of Standard and Technology
pH	=	The negative logarithm of the concentration of hydrogen ions
ppi	=	Pore per inch
ppm	=	Part per million
rpm	=	Revolution per minute
SD	=	Standard deviation
v/v	=	Volume over volume
w/v	=	Weight over volume



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย