

REFERENCES

1. World Health Organization. Tuberculosis. Fact sheet 104. World Health Organization. 2001: Geneva, Switzerland.
2. Dye, C., et al., Consensus statement. Global burden of tuberculosis: estimated incidence, prevalence, and mortality by country. WHO Global Surveillance and Monitoring Project. *Jama*, 1999. 282(7): p. 677-86.
3. Turenne, C.Y., et al., Necessity of quality-controlled 16S rRNA gene sequence databases: identifying nontuberculous *Mycobacterium* species. *J Clin Microbiol*, 2001. 39(10): p. 3637-48.
4. Falkinham, J.O., 3rd, Epidemiology of infection by nontuberculous mycobacteria. *Clin Microbiol Rev*, 1996. 9(2): p. 177-215.
5. Wayne, L.G. and H.A. Sramek, Agents of newly recognized or infrequently encountered mycobacterial diseases. *Clin Microbiol Rev*, 1992. 5(1): p. 1-25.
6. Wolinsky, E., Mycobacterial diseases other than tuberculosis. *Clin Infect Dis*, 1992. 15(1): p. 1-10.
7. Witebsky, F.G. and P. Kruczak-Filipov, Identification of mycobacteria by conventional methods. *Clin Lab Med*, 1996. 16(3): p. 569-601.
8. Pfyffer, G.E., et al., Comparison of the Mycobacteria Growth Indicator Tube (MGIT) with radiometric and solid culture for recovery of acid-fast bacilli. *J Clin Microbiol*, 1997. 35(2): p. 364-8.
9. Tortoli, E., et al., Use of BACTEC MGIT 960 for recovery of mycobacteria from clinical specimens: multicenter study. *J Clin Microbiol*, 1999. 37(11): p. 3578-82.
10. Somoskovi, A., et al., Comparison of recoveries of mycobacterium tuberculosis using the automated BACTEC MGIT 960 system, the BACTEC 460 TB system, and Lowenstein-Jensen medium. *J Clin Microbiol*, 2000. 38(6): p. 2395-7.
11. Springer, B., et al., Two-laboratory collaborative study on identification of mycobacteria: molecular versus phenotypic methods. *J Clin Microbiol*, 1996. 34(2): p. 296-303.
12. Tortoli, E., et al., Burden of unidentifiable mycobacteria in a reference laboratory. *J Clin Microbiol*, 2001. 39(11): p. 4058-65.
13. Cook, V.J., et al., Conventional methods versus 16S ribosomal DNA sequencing for identification of nontuberculous mycobacteria: cost analysis. *J Clin Microbiol*, 2003. 41(3): p. 1010-5.

14. Butler, W.R., K.C. Jost, Jr., and J.O. Kilburn, Identification of mycobacteria by high-performance liquid chromatography. *J Clin Microbiol*, 1991. 29(11): p. 2468-72.
15. Butler, W.R. and J.O. Kilburn, Identification of major slowly growing pathogenic mycobacteria and *Mycobacterium gordonaiae* by high-performance liquid chromatography of their mycolic acids. *J Clin Microbiol*, 1988. 26(1): p. 50-3.
16. Glickman, S.E., et al., Rapid identification of mycolic acid patterns of mycobacteria by high-performance liquid chromatography using pattern recognition software and a *Mycobacterium* library. *J Clin Microbiol*, 1994. 32(3): p. 740-5.
17. Liebana, E., et al., Assessment of genetic markers for species differentiation within the *Mycobacterium tuberculosis* complex. *J Clin Microbiol*, 1996. 34(4): p. 933-8.
18. Sansila, A., et al., Differentiation between *Mycobacterium tuberculosis* and *Mycobacterium avium* by amplification of the 16S-23S ribosomal DNA spacer. *J Clin Microbiol*, 1998. 36(9): p. 2399-403.
19. Kim, B.J., et al., Differential identification of *Mycobacterium tuberculosis* complex and nontuberculous mycobacteria by duplex PCR assay using the RNA polymerase gene (*rpoB*). *J Clin Microbiol*, 2004. 42(3): p. 1308-12.
20. Telenti, A., et al., Rapid identification of mycobacteria to the species level by polymerase chain reaction and restriction enzyme analysis. *J Clin Microbiol*, 1993. 31(2): p. 175-8.
21. Lappayawichit, P., et al., Differentiation of *Mycobacterium* species by restriction enzyme analysis of amplified 16S-23S ribosomal DNA spacer sequences. *Tuber Lung Dis*, 1996. 77(3): p. 257-63.
22. Hafner, B., et al., Different molecular methods for the identification of rarely isolated non-tuberculous mycobacteria and description of new *hsp65* restriction fragment length polymorphism patterns. *Mol Cell Probes*, 2004. 18(1): p. 59-65.
23. Takewaki, S., et al., Nucleotide sequence comparison of the mycobacterial dnaJ gene and PCR-restriction fragment length polymorphism analysis for identification of mycobacterial species. *Int J Syst Bacteriol*, 1994. 44(1): p. 159-66.
24. Taylor, T.B., et al., Routine use of PCR-restriction fragment length polymorphism analysis for identification of mycobacteria growing in liquid media. *J Clin Microbiol*, 1997. 35(1): p. 79-85.
25. Rossi, M.C., et al., A PCR-colorimetric microwell plate hybridization assay for detection of *Mycobacterium tuberculosis* and *M. avium* from culture samples and Ziehl-Neelsen-positive smears. *J Clin Microbiol*, 2000. 38(5): p. 1772-6.

26. Hong, S.K., et al., Identification of *Mycobacterium tuberculosis* by PCR-linked reverse hybridization using specific *rpoB* oligonucleotide probes. *J Microbiol Methods*, 2004. 59(1): p. 71-9.
27. Stender, H., et al., Fluorescence In situ hybridization assay using peptide nucleic acid probes for differentiation between tuberculous and nontuberculous mycobacterium species in smears of mycobacterium cultures. *J Clin Microbiol*, 1999. 37(9): p. 2760-5.
28. Swanson, D.S., X. Pan, and J.M. Musser, Identification and subspecific differentiation of *Mycobacterium scrofulaceum* by automated sequencing of a region of the gene (*hsp65*) encoding a 65-kilodalton heat shock protein. *J Clin Microbiol*, 1996. 34(12): p. 3151-9.
29. Kurabachew, M., et al., Sequence analysis in the 23S rDNA region of *Mycobacterium tuberculosis* and related species. *J Microbiol Methods*, 2003. 54(3): p. 373-80.
30. Rogall, T., T. Flohr, and E.C. Bottger, Differentiation of *Mycobacterium* species by direct sequencing of amplified DNA. *J Gen Microbiol*, 1990. 136(9): p. 1915-20.
31. Bodinghaus, B., et al., Detection and identification of mycobacteria by amplification of rRNA. *J Clin Microbiol*, 1990. 28(8): p. 1751-9.
32. Kim, B.J., et al., Identification of mycobacterial species by comparative sequence analysis of the RNA polymerase gene (*rpoB*). *J Clin Microbiol*, 1999. 37(6): p. 1714-20.
33. Blackwood, K.S., et al., Evaluation of *recA* sequences for identification of *Mycobacterium* species. *J Clin Microbiol*, 2000. 38(8): p. 2846-52.
34. Peterson, E.M., et al., Direct identification of *Mycobacterium tuberculosis*, *Mycobacterium avium*, and *Mycobacterium intracellulare* from amplified primary cultures in BACTEC media using DNA probes. *J Clin Microbiol*, 1989. 27(7): p. 1543-7.
35. Johansen, I.S., et al., Rapid differentiation between clinically relevant mycobacteria in microscopy positive clinical specimens and mycobacterial isolates by line probe assay. *Diagn Microbiol Infect Dis*, 2002. 43(4): p. 297-302.
36. Mijs, W., et al., Evaluation of a commercial line probe assay for identification of mycobacterium species from liquid and solid culture. *Eur J Clin Microbiol Infect Dis*, 2002. 21(11): p. 794-802.

37. Wilton, S. and D. Cousins, Detection and identification of multiple mycobacterial pathogens by DNA amplification in a single tube. *PCR Methods Appl*, 1992. 1(4): p. 269-73.
38. Kulski, J.K., et al., Use of a multiplex PCR to detect and identify *Mycobacterium avium* and *M. intracellulare* in blood culture fluids of AIDS patients. *J Clin Microbiol*, 1995. 33(3): p. 668-74.
39. Kox, L.F., et al., PCR assay based on DNA coding for 16S rRNA for detection and identification of mycobacteria in clinical samples. *J Clin Microbiol*, 1995. 33(12): p. 3225-33.
40. Pfyffer, G.E., B.A. Brown-Elliott, and J. Wallace, R.J., *Mycobacterium*: General Characteristics, Isolation, and Staining Procedures, in *Manual of Clinical Microbiology*, P.R. Murray, Editor. 2002, ASM Press: Washinton, D.C.
41. Grang, S.M., *Mycobacterium* and Human Disease. 2 ed. 1996, New York: Oxford University Press.
42. Good, R.C. and T.M. Shinnick, *Mycobacterium*, in *Topley&Wilson's Microbiology and Microbial Infections*. 1998, Georgina Bentliff: Great Britain.
43. Tortoli, E., Impact of genotypic studies on mycobacterial taxonomy: the new mycobacteria of the 1990s. *Clin Microbiol Rev*, 2003. 16(2): p. 319-54.
44. Bodmer, T., et al., Improved performance of Gen-Probe Amplified *Mycobacterium* Tuberculosis Direct Test when 500 instead of 50 microliters of decontaminated sediment is used. *J Clin Microbiol*, 1996. 34(1): p. 222-3.
45. Kirschner, P., et al., Genotypic identification of mycobacteria by nucleic acid sequence determination: report of a 2-year experience in a clinical laboratory. *J Clin Microbiol*, 1993. 31(11): p. 2882-9.
46. Mahon, C.R. and G. Manuselis, et.al., *Textbook of Diagnostic Microbiology*. 1995, W.B. Saunder company: the United States of America.
47. Forbes, B.A., D.F. Sahm, and A.S. Weissfeld, *Diagnostic Microbiology*. 1998, Mosby: Texas, The United States of America.
48. Morris, A., *Mycobacteriology: Laboratory Methods and Standards*, in *Guidelines for Tuberculosis Control in New Zealand*. 2003.
49. Ringuet, H., et al., *hsp65* sequencing for identification of rapidly growing mycobacteria. *J Clin Microbiol*, 1999. 37(3): p. 852-7.
50. Vincent, V., et al., *Mycobacterium*: Phenotypic and Genotypic Identification, in

Manual of clinical Microbiology, P.R. Murray, Editor. 2002, ASM Press: Washington, D.C.

51. Hall, G.S. and B.J. Howard, Mycobacteria, in Clinical and Pathogenic Microbiology, B.J. Howard, et al., Editors. 1993, Mosby: Washington, D.C. p. 503-528.
52. Murray, P.R., et al., Manual of Clinical Microbiology, in Manual of Clinical Microbiology, P.R. Murray, et al., Editors. 1999, American Society for microbiology Press: Washington, D.C.
53. Plikaytis, B.B., et al., Differentiation of slowly growing *Mycobacterium* species, including *Mycobacterium tuberculosis*, by gene amplification and restriction fragment length polymorphism analysis. *J Clin Microbiol*, 1992. 30(7): p. 1815-22.
54. Devallois, A., K.S. Goh, and N. Rastogi, Rapid identification of mycobacteria to species level by PCR-restriction fragment length polymorphism analysis of the *hsp65* gene and proposition of an algorithm to differentiate 34 mycobacterial species. *J Clin Microbiol*, 1997. 35(11): p. 2969-73.
55. Steingrube, V.A., et al., PCR amplification and restriction endonuclease analysis of a 65-kilodalton heat shock protein gene sequence for taxonomic separation of rapidly growing mycobacteria. *J Clin Microbiol*, 1995. 33(1): p. 149-53.
56. Brunello, F., et al., Identification of 54 mycobacterial species by PCR-restriction fragment length polymorphism analysis of the *hsp65* gene. *J Clin Microbiol*, 2001. 39(8): p. 2799-806.
57. Kapur, V., et al., Rapid *Mycobacterium* species assignment and unambiguous identification of mutations associated with antimicrobial resistance in *Mycobacterium tuberculosis* by automated DNA sequencing. *Arch Pathol Lab Med*, 1995. 119(2): p. 131-8.
58. Alcaide, F., et al., Heterogeneity and clonality among isolates of *Mycobacterium kansasii*: implications for epidemiological and pathogenicity studies. *J Clin Microbiol*, 1997. 35(8): p. 1959-64.
59. Picardeau, M., et al., Genotypic characterization of five subspecies of *Mycobacterium kansasii*. *J Clin Microbiol*, 1997. 35(1): p. 25-32.
60. Park, H., et al., Detection and identification of mycobacteria by amplification of the internal transcribed spacer regions with genus- and species-specific PCR primers. *J Clin Microbiol*, 2000. 38(11): p. 4080-5.

61. Chapin-Robertson, K., et al., Detection and identification of *Mycobacterium* directly from BACTEC bottles by using a DNA-rRNA probe. *Diagn Microbiol Infect Dis*, 1993. 17(3): p. 203-7.
62. Kaminski, D.A. and D.J. Hardy, Selective utilization of DNA probes for identification of *Mycobacterium* species on the basis of cord formation in primary BACTEC 12B cultures. *J Clin Microbiol*, 1995. 33(6): p. 1548-50.
63. Lebrun, L., et al., Evaluation of nonradioactive DNA probes for identification of mycobacteria. *J Clin Microbiol*, 1992. 30(9): p. 2476-8.
64. Richter, E., et al., Identification of *Mycobacterium kansasii* by using a DNA probe (AccuProbe) and molecular techniques. *J Clin Microbiol*, 1999. 37(4): p. 964-70.
65. Tortoli, E., M.T. Simonetti, and F. Lavinia, Evaluation of reformulated chemiluminescent DNA probe (AccuProbe) for culture identification of *Mycobacterium kansasii*. *J Clin Microbiol*, 1996. 34(11): p. 2838-40.
66. Reisner, B.S., A.M. Gatson, and G.L. Woods, Use of Gen-Probe AccuProbes to identify *Mycobacterium avium* complex, *Mycobacterium tuberculosis* complex, *Mycobacterium kansasii*, and *Mycobacterium gordonaiae* directly from BACTEC TB broth cultures. *J Clin Microbiol*, 1994. 32(12): p. 2995-8.
67. Goto, M., et al., Evaluation of acridinium-ester-labeled DNA probes for identification of *Mycobacterium tuberculosis* and *Mycobacterium avium-Mycobacterium intracellulare* complex in culture. *J Clin Microbiol*, 1991. 29(11): p. 2473-6.
68. Kox, L.F., et al., Multiplex PCR assay for immediate identification of the infecting species in patients with mycobacterial disease. *J Clin Microbiol*, 1997. 35(6): p. 1492-8.
69. Tanaka, H., et al., Comparison of a multiplex-PCR assay with mycolic acids analysis and conventional methods for the identification of mycobacteria. *Microbiol Immunol*, 2003. 47(5): p. 307-12.
70. Fries, J.W., et al., Genus- and species-specific DNA probes to identify mycobacteria using the polymerase chain reaction. *Mol Cell Probes*, 1990. 4(2): p. 87-105.
71. Fries, J.W., et al., Detection of untreated mycobacteria by using polymerase chain reaction and specific DNA probes. *J Clin Microbiol*, 1991. 29(8): p. 1744-7.
72. Soini, H., E.C. Bottger, and M.K. Viljanen, Identification of mycobacteria by PCR-based sequence determination of the 32-kilodalton protein gene. *J Clin Microbiol*, 1994. 32(12): p. 2944-7.

73. Zolg, J.W. and S. Philippi-Schulz, The superoxide dismutase gene, a target for detection and identification of mycobacteria by PCR. *J Clin Microbiol*, 1994. 32(11): p. 2801-12.
74. Kasai, H., T. Ezaki, and S. Harayama, Differentiation of phylogenetically related slowly growing mycobacteria by their *gyrB* sequences. *J Clin Microbiol*, 2000. 38(1): p. 301-8.
75. Roth, A., et al., Differentiation of phylogenetically related slowly growing mycobacteria based on 16S-23S rRNA gene internal transcribed spacer sequences. *J Clin Microbiol*, 1998. 36(1): p. 139-47.
76. Roth, A., et al., *Mycobacterium heckeshornense* sp. nov., A new pathogenic slowly growing *Mycobacterium* sp. Causing cavitary lung disease in an immunocompetent patient. *J Clin Microbiol*, 2000. 38(11): p. 4102-7.
77. El Amin, N.M., et al., Identification of non-tuberculous mycobacteria: 16S rRNA gene sequence analysis vs. conventional methods. *Scand J Infect Dis*, 2000. 32(1): p. 47-50.
78. Hughes, M.S., et al., Identification of mycobacteria from animals by restriction enzyme analysis and direct DNA cycle sequencing of polymerase chain reaction-amplified 16S rRNA gene sequences. *J Clin Microbiol*, 1993. 31(12): p. 3216-22.
79. Anton, A.I., A.J. Martinez-Murcia, and F. Rodriguez-Valera, Intraspecific diversity of the 23S rRNA gene and the spacer region downstream in *Escherichia coli*. *J Bacteriol*, 1999. 181(9): p. 2703-9.
80. Jeng, R.S., et al., The use of 16S and 16S-23S rDNA to easily detect and differentiate common Gram-negative orchard epiphytes. *J Microbiol Methods*, 2001. 44(1): p. 69-77.
81. Frothingham, R., H.G. Hills, and K.H. Wilson, Extensive DNA sequence conservation throughout the *Mycobacterium* tuberculosis complex. *J Clin Microbiol*, 1994. 32(7): p. 1639-43.
82. Ichiyama, S., et al., Evaluation of Gen-Probe Amplified *Mycobacterium* Tuberculosis Direct Test and Roche PCR-microwell plate hybridization method (AMPLICOR MYCOBACTERIUM) for direct detection of mycobacteria. *J Clin Microbiol*, 1996. 34(1): p. 130-3.
83. Tevere, V.J., et al., Detection of *Mycobacterium tuberculosis* by PCR amplification with pan-*Mycobacterium* primers and hybridization to an *M. tuberculosis*-specific probe. *J Clin Microbiol*, 1996. 34(4): p. 918-23.

84. Suffys, P.N., et al., Rapid identification of Mycobacteria to the species level using INNO-LiPA Mycobacteria, a reverse hybridization assay. *J Clin Microbiol*, 2001. 39(12): p. 4477-82.
85. Portaels, F., et al., Direct detection and identification of *Mycobacterium ulcerans* in clinical specimens by PCR and oligonucleotide-specific capture plate hybridization. *J Clin Microbiol*, 1997. 35(5): p. 1097-100.
86. Sanguinetti, M., et al., Routine use of PCR-reverse cross-blot hybridization assay for rapid identification of *Mycobacterium* species growing in liquid media. *J Clin Microbiol*, 1998. 36(6): p. 1530-3.
87. Mcpherson, M.J., P. Quirk, and G.R. Taylor, PCR a Practical Approach. 1991, New York, United States: Oxford University Press.
88. Wolcott, M.J., Advances in nucleic acid-based detection methods. *Clin Microbiol Rev*, 1992. 5(4): p. 370-86.
89. Sanger, F., S. Nicklen, and A.R. Coulson, DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci U S A*, 1977. 74(12): p. 5463-7.
90. Ausubel, F.M., Current Protocols in Molecularbiology. 1993.
91. Hongmanee, P., H. Stender, and O.F. Rasmussen, Evaluation of a fluorescence in situ hybridization assay for differentiation between tuberculous and nontuberculous *Mycobacterium* species in smears of Lowenstein-Jensen and Mycobacteria Growth Indicator Tube cultures using peptide nucleic acid probes. *J Clin Microbiol*, 2001. 39(3): p. 1032-5.
92. Stender, H., et al., Direct detection and identification of *Mycobacterium tuberculosis* in smear-positive sputum samples by fluorescence in situ hybridization (FISH) using peptide nucleic acid (PNA) probes. *Int J Tuberc Lung Dis*, 1999. 3(9): p. 830-7.
93. Thomsen, V.O., A.B. Andersen, and H. Miørner, Incidence and clinical significance of non-tuberculous mycobacteria isolated from clinical specimens during a 2-y nationwide survey. *Scand J Infect Dis*, 2002. 34(9): p. 648-53.
94. Huang, J.Q. and R.H. Hunt, Treatment after failure: the problem of non-responders. Vol. (suppl I): 140-144. 1999: Gut. 45.
95. Corbett, E.L., et al., The growing burden of tuberculosis: global trends and interactions with the HIV epidemic. *Arch Intern Med*, 2003. 163(9): p. 1009-21.
96. World health Organization, Global tuberculosis control: surveillance, planning, financing, WHO Report 2003 (WHO/CDS/TB/2003.316): Geneva: WHO 2003.

97. Wolinsky, E., Nontuberculous mycobacteria and associated diseases. Am Rev Respir Dis, 1979. 119(1): p. 107-59.
98. Horsburgh, C.R., Jr., *Mycobacterium avium* complex infection in the acquired immunodeficiency syndrome. N Engl J Med, 1991. 324(19): p. 1332-8.
99. Horsburgh, C.R., Jr., Epidemiology of mycobacterial diseases in AIDS. Res Microbiol, 1992. 143(4): p. 372-7.
100. Horsburgh, C.R., Jr. and R.M. Selik, The epidemiology of disseminated nontuberculous mycobacterial infection in the acquired immunodeficiency syndrome (AIDS). Am Rev Respir Dis, 1989. 139(1): p. 4-7.
101. Peters, M., et al., Immunosuppression and mycobacteria other than *Mycobacterium tuberculosis*: results from patients with and without HIV infection. Epidemiol Infect, 1989. 103(2): p. 293-300.
102. Selik, R.M., E.T. Starcher, and J.W. Curran, Opportunistic diseases reported in AIDS patients: frequencies, associations, and trends. Aids, 1987. 1(3): p. 175-82.
103. Zakowski, P., et al., Disseminated *Mycobacterium avium-intracellulare* infection in homosexual men dying of acquired immunodeficiency. Jama, 1982. 248(22): p. 2980-2.
104. Nightingale, S.D., et al., Incidence of *Mycobacterium avium-intracellulare* complex bacteremia in human immunodeficiency virus-positive patients. J Infect Dis, 1992. 165(6): p. 1082-5.
105. Kolk, A.H.J., et al., PCR assay for *Mycobacterium tuberculosis* complex and other Mycobacteria. 2000, Department of Biomedical Research, Royal Tropical Institute, Amsterdam, The Netherlands.
106. Hall, L., et al., Evaluation of the MicroSeq system for identification of mycobacteria by 16S ribosomal DNA sequencing and its integration into a routine clinical mycobacteriology laboratory. J Clin Microbiol, 2003. 41(4): p. 1447-53.
107. Kulski, J.K. and T. Pryce, Preparation of mycobacterial DNA from blood culture fluids by simple alkali wash and heat lysis method for PCR detection. J Clin Microbiol, 1996. 34(8): p. 1985-91.
108. Han, X.Y., et al., Rapid and accurate identification of mycobacteria by sequencing hypervariable regions of the 16S ribosomal RNA gene. Am J Clin Pathol, 2002. 118(5): p. 796-801.
109. Kremer K., et al. PCR+Reverse Line Blot Hybridization (PLH) to detect Rifampicin Resistance. 1997.

110. Rogall, T., et al., Towards a phylogeny and definition of species at the molecular level within the genus *Mycobacterium*. Int J Syst Bacteriol, 1990. 40(4): p. 323-30.
111. Masur, H., Recommendations on prophylaxis and therapy for disseminated *Mycobacterium avium* complex disease in patients infected with the human immunodeficiency virus. Public Health Service Task Force on Prophylaxis and Therapy for *Mycobacterium avium* Complex. N Engl J Med, 1993. 329(12): p. 898-904.
112. Hoy, J., et al., Quadruple-drug therapy for *Mycobacterium avium-intracellulare* bacteremia in AIDS patients. J Infect Dis, 1990. 161(4): p. 801-5.

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย



APPENDICES

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

APPENDIX I

REAGENTS AND INSTRUMENTS

A. REAGENT

Absolute ethanol	(Merck, U.S.A)
Agarose (ultrapure)	(Biorad, U.S.A)
Boric acid	(Amresco, U.S.A.)
ECL detection reagent	(Amresco, U.S.A.)
EDAC	(Sigma, U.S.A.)
EDTA	(Amresco, U.S.A)
Ethidium bromide	(Amresco, U.S.A)
Methanol	(Merck, U.S.A)
Mineral oil	(Sigma, U.S.A)
N-acetyl-L-cysteine	(Sigma, U.S.A)
NaCl	(Sigma, U.S.A)
NaHCO ₃	(Sigma, U.S.A.)
Na ₂ HPO ₄	(Sigma, U.S.A)
Sodium citrate	(Sigma, U.S.A)
Sodium dodecyl sulphate	(Amresco, U.S.A)
Sodium hydroxide	(Merck, Germany)
Streptavidin-peroxidase conjugate	(Amresco, U.S.A)
KH ₂ PO ₄	(Sigma, U.S.A)
Tris (ultrapure)	(Amresco, U.S.A)
Tris hydrochloride	(Amresco, U.S.A)

B. MATERIAL

Biodyne C membrane	(Amresco, U.S.A.)
Hyperfilm ECL	(Amresco, U.S.A.)
X-ray film	(Kodak, Japan)

APPENDIX I (CONTINUE)

C. INSTRUMENTS

Hybaid OmniGene thermal cycler	(Hybaid, England)
Camera Gel Doc™ MZL	(Bio-RAD, USA)
Dot botter	(BBL, U.S.A.)
Water bath	(Memmert, U.S.A)
Incubator	(Forma Scientific, U.S.A)
Microcentrifuge	(Eppendorf, U.S.A)
Perkin Elmer GeneAmp PCR system 9600	(Perkin Elmer, U.S.A)
ABI Prism™ 310 Automate sequencer	(Perkin Elmer, U.S.A)
Spectrophotometer	(BIORAD, U.S.A)

APPENDIX II

MEDIA FOR CULTURE

1. Ogawa media

Mineral salt solution

Potassium dihydrogen phosphate anhydrous (KH ₂ PO ₄)	3.0	g
Sodium glutamate	3.0	g
Distilled water	300	ml
Glycerine	18	ml
2%Malachite green solution	18	ml
Homogenised whole eggs	600ml (12-16 eggs)	

Autoclaving mineral salt solution at 121°C for 15 minutes to sterilise. Cool to room temperature. The following ingredients are aseptically pooled in a large, sterile flask and mixed well: glycerine, 2% malachite green solution, homogenised whole eggs. The medium is mixed well and distributed in 6-8 ml volumes in sterile 20x150 mm screw-capped test tubes. Place the bottles in a slanted position in the inspissator and coagulate the medium for 45 minutes at 80-85 °C. Cool and store at 4 °C until used.

APPENDIX III

REAGENTS AND PREPARATIONS

1. 0.5 M Ethylene diamine tetraacetic acid (EDTA), pH 8.0

Dissodium ethylene diamine tetraacetate.2H ₂ O	186.1 g
DDW	800.0 ml
Adjust pH to 8.0	
Adjust volume to 1,000 ml	
Sterililize by autoclaving	

2. 1 M Tris-HCl, pH 8.0

Tris hydrochloride	157.64 g
DDW	800.0 ml
Adjust pH to 8.0	
Adjust volume to 1,000 ml	
Sterililize by autoclaving	

3. 5x Tris-borate buffer (TBE)

Tris base	54 g
Boric acid	27.5 g
0.5 M EDTA (pH 8.0)	20 ml
Adjust volume to 1 liter with distilled water. The solution was mixed and sterilized by autoclaving at 121°C for 15 min.	

4. 10 x TE buffer

Tris	12.11 g
0.5 M EDTA	20 ml
Adjust to pH 8.0 by adding conc. HCl, adjust volume to 1,000 ml. Sterilize by autoclaving	

5. 20xSSPE
- | | | |
|----------------------------------|--------|---|
| Na ₂ HPO ₄ | 28.4 | g |
| NaCl | 210.24 | g |
| EDTA | 7.4 | g |
- Adjust to pH 7.4, Adjust volume to 1,000 ml. Sterilize by autoclaving.
Store at room temperature for no longer than one years.
6. 10% SDS
- | | | |
|-----|-----|----|
| SDS | 10 | g |
| DDW | 100 | ml |
- Dissolve by heating at 65°C for 20 min. Do not autoclaving
7. 0.5 M NaHCO₃, pH 8.4
- | | | |
|--------------------|------|----|
| NaHCO ₃ | 10.5 | g |
| DDW | 250 | ml |
- Adjust to pH 8.4, adjust volume to 250 ml with DDW
Store at room temperature for no longer than one years.
8. 16% (w/v) EDAC (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide)
- | | | |
|------|-----|----|
| EDAC | 1.6 | g |
| DDW | 10 | ml |
- Prepare fresh before use.
9. 2XSSPE
- | | | |
|---------|-----|----|
| 20XSSPE | 20 | ml |
| DDW | 180 | ml |
- Prepare fresh before use.
10. 20 mM EDTA
- | | | |
|---------------------|----|----|
| 0.5 M EDTA (pH 8.0) | 4 | ml |
| DDW | 96 | ml |
- Prepare fresh before use.

11. 2XSSPE/0.1% SDS

20XSSPE	20	ml
10% SDS	2	ml
DDW	178	ml

Prepare fresh before use.

12. 2XSSPE/0.5% SDS

20XSSPE	20	ml
10% SDS	10	ML
DDW	170	ml

Prepare fresh before use.

13. ECL detection reagent

ECL detection reagent 1	5	ml
ECL detection reagent 2	5	ml

Store at 4°C for no longer than six months.

14. 0.1 M NaOH

NaOH	0.4	g
DDW	100	ml

Prepare fresh before use.

15. 0.5 M Na-citrate

Na-citrate	147	g
DDW	1,000	ml

Sterilize by autoclaving.

16. 5 M NaOH

NaOH	200	g
DDW	1,000	ml

Sterilize by autoclaving.

17. Alkaline wash solution (0.05 M Na-citrate and 0.5 M NaOH)
- | | | |
|------------------|----|----|
| 0.5 M Na-citrate | 5 | ml |
| 5 M NaOH | 5 | ml |
| DDW | 40 | ml |
- Sterilize by autoclaving.
18. 0.5 M Tris-HCl, pH 8.0
- | | | |
|--------------------|-------|----|
| Tris hydrochloride | 78.82 | g |
| DDW | 800.0 | ml |
- Adjust pH to 8.0, adjust volume to 1,000 ml
- Sterilize by autoclaving

APPENDIX III (CONTINUE)

REAGENT FOR AGAROSE GEL ELECTROPHORESIS

1. 10 mg/ml Ethidium bromide

Ethidium bromide	1	g
DDW	100	g

Stir on a magnetic stirrer for several hours to ensure that the dye has dissolved.

Wrap the container in aluminum foil or transfer to a dark bottle and stored at 4°C

2. 2% Agarose gel

Agarose (ultrapure)	0.4	g
1 x TBE	20.0	ml
10 mg/ml Ethidium bromide	1.0	μl

Dissolve by heating in microwave oven and occasional mix until no granules of agarose are visible.

BIOGRAPHY

Miss Sanjira Juntarapornchai was born on July 31, 1980 in Bangkok, Thailand. She graduated with bachelor degree of science in Microbiology from the Faculty of Science at Chulalongkorn University in 2002.

