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ศูนย์วิทยทรัพยากร  
จุฬาลงกรณ์มหาวิทยาลัย



**APPENDICES**

ศูนย์วิทยทรัพยากร  
จุฬาลงกรณ์มหาวิทยาลัย

## APPENDIX A : Preparation for polyacrylamide gel electrophoresis

### 1. Stock reagents

#### 30 % Acrylamide, 0.8% bis-acrylamide, 100 ml

Acrylamide	29.2	g
N,N'-methylene-bis-acrylamide	0.8	g

Adjust volume to 100 ml with distilled water.

#### 1.5 M Tris-HCl pH 8.8

Tris (hydroxymethyl)-aminomethane	18.17	g
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Adjust pH to 8.8 with 1 M HCl and adjust volume to 100 ml with distilled water.

#### 2.0 M Tris-HCl pH 8.8

Tris (hydroxymethyl)-aminomethane	24.2	g
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Adjust pH to 8.8 with 1 M HCl and adjust volume to 100 ml with distilled water.

#### 0.5 M Tris-HCl pH 6.8

Tris (hydroxymethyl)-aminomethane	6.06	g
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Adjust pH to 6.8 with 1 M HCl and adjust volume to 100 ml with distilled water.

#### 1.0 M Tris-HCl pH 6.8

Tris (hydroxymethyl)-aminomethane	12.1	g
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Adjust pH to 6.8 with 1 M HCl and adjust volume to 100 ml with distilled water.

#### Solution B (SDS PAGE)

2.0 M Tris-HCl pH 8.8	75	ml
10% SDS	4	ml
Distilled water	21	ml

#### Solution C (SDS PAGE)

1.0 M Tris-HCl pH 8.8	50	ml
10% SDS	4	ml
Distilled water	46	ml



## 2. SDS-PAGE

### 15 % Separating gel

30 % Acrylamide ml solution	5.0	ml
Solution B	2.5	ml
Distilled water	2.5	ml
10% $(\text{NH}_4)_2\text{S}_2\text{O}_8$	50	$\mu\text{l}$
TEMED	10	$\mu\text{l}$

### 5.0 % Stacking gel

30 % Acrylamide ml solution	0.67	ml
Solution C	1.0	ml
Distilled water	2.3	ml
10 % $(\text{NH}_4)_2\text{S}_2\text{O}_8$	30	$\mu\text{l}$
TEMED	5.0	$\mu\text{l}$

### 5X Sample buffer

1 M Tris-HCl pH 6.8	0.6	ml
50% Glycerol	5.0	ml
10% SDS	2.0	ml
2-Mercaptoethanol	0.5	ml
1 % Bromophenol blue	1.0	ml
Distilled water	0.9	ml

One part of sample buffer was added to four parts of sample. The mixture was heated 5 min. in boiling water before loading to the gel.

### Electrophoresis buffer, 1 litre

(25 mM Tris, 192 mM glycine)

Tris (hydroxymethyl)-aminomethane	3.03	g
Glycine	14.40	g
SDS	1.0	g

Dissolve in distilled water to 1 litre. Do not adjust pH with acid or base (final pH should be 8.3).

## APPENDIX B : Preparation for Tricine-Sodium dodecyl sulfate- Polyacrylamide gel electrophoresis

### 1. Stock reagents

#### 49.5 % Acrylamide, 3% bis-acrylamide, 100 ml

Acrylamide 48 g

N,N'-methylene-bis-acrylamide 1.5 g

Adjust volume to 100 ml with distilled water.

#### Gel buffer, 100 ml

Tris (hydroxymethyl)-aminomethane 36.34 g

SDS 0.3 g

Adjust pH to 8.45 with 1 M HCl and adjust volume to 100 ml with distilled water.

### 2. Tricine-SDS-PAGE

#### 12 % Separating gel

49.5 % Acrylamide solution 2.92 ml

Gel buffer 4 ml

Distilled water 3.88 ml

100% Glycerol 1.2 ml

10%  $(\text{NH}_4)_2\text{S}_2\text{O}_8$  70  $\mu\text{l}$

TEMED 15  $\mu\text{l}$

#### 4.0 % Stacking gel

49.5 % Acrylamide solution 0.25 ml

Gel buffer 0.775 ml

Distilled water 2 ml

10 %  $(\text{NH}_4)_2\text{S}_2\text{O}_8$  30  $\mu\text{l}$

TEMED 5.0  $\mu\text{l}$

**2X Sample buffer**

Gel buffer	3.0	ml
100% Glycerol	2.4	ml
SDS	0.8	g
0.1 % Coomassie Blue G	1.5	ml
0.1 % Phenol red	0.5	ml

Adjust volume to 100 ml with distilled water.

One part of sample buffer was added to one parts of sample. The mixture was heated 5 min. in boiling water before loading to the gel.

**Electrophoresis buffer****Cathode buffer, 100 ml**

Tris (hydroxymethyl)-aminomethane	1.21	g
Tricine	26.3	g
SDS	0.1	g

Dissolve in distilled water to 100 ml. Do not adjust pH with acid or base (final pH should be 8.3).

**Anode buffer, 100 ml**

Tris (hydroxymethyl)-aminomethane	2.42	g
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Adjust pH to 8.9 with 1 M HCl and adjust volume to 100 ml with distilled water.

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## APPENDIX C : Preparation for silver staining

### 1. Stock reagents

#### 0.36% NaOH

NaOH 0.36 g

Adjust volume to 100 ml with distilled water.

#### 1% citric acid

Citric acid 1 g

Adjust volume to 100 ml with distilled water.

#### 50% methanol 10% acetic acid

100 % methanol 50 ml

Glacial acetic acid 10 ml

Adjust volume to 100 ml with distilled water.

#### 1% acetic acid

Glacial acetic acid 1 ml

Adjust volume to 100 ml with distilled water.

### 2. Make fresh reagents

#### Solution A

Silver nitrate 0.8 g

Adjust volume to 4 ml with distilled water.

#### Solution B

0.36% NaOH 21 ml

14.8 M (30%) ammonium hydroxide 1.4 ml

#### Solution C

Add Solution A to Solution B dropwise with constant vigorous stirring, allowing brown precipitate to clear. Adjust volume to 100 ml. Use within 15minutes.

#### Solution D

Mix 0.5 ml 1%citric acid with 50  $\mu$ l 38% formaldehyde, add water to 100 ml.

Solution must be fresh.

### 3. Silver Staining method

1. Wearing gloves, pick up the gel and transfer it to a small container. Soak gel in 50% methanol 10% acetic acid for at least 1 hour with 2 -3 changes of methanol /acetic acid.
2. Rinse 30 minutes with water, with at least 3 changes.
3. Prepare Solutions A, B, then C.
4. Remove gel to a clean container and stain in Solution C for 15 minutes with gentle, constant agitation.
5. Rinse gel twice in deionized water, then soak 2 minutes with gentle agitation.
6. Prepare Solution D.
7. Remove gel to a clean container and develop by washing gel in Solution D. Bands should appear in less than 10 minutes or else change Solution D. If a pale yellow background appears, reaction should be stopped.
8. Stop development by rinsing in 1% acetic acid.
9. Wash gel in water for at least 1 hour with at least three changes of water.
10. Store gel in water or dry gel.



## Biography

Mr. Ekarin Achakulwisut was born on September 15, 1978 in Samutsakorn. He graduated with the degree of Bachelor of Science from the Department of Biochemistry, Chulalongkorn University in 1999. He has further studied in Master's degree of science (Biotechnology), Chulalongkorn University.



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