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APPENDIX I

REAGENTS AND INSTRUMENTS

A. REAGENTS

Acrylamide (N,N-methylenebisacrylamide) (LKB, Sweden)
Ammonium sulfate (E.Merck, Darmstadt, W. Germany)
Ammonium persulfate (LKB, Sweden)
Azocasein (Sigma,Mo. U.S.A.)
Azocoll (Sigma,Mo. U.S.A.)
3-Aminopropionic acid (Sigma, Mo.,U.S.A.)
Beef extract (Gibco,U.S.A.)
Bis-acrylamide (LKB, Sweden)
Barium chloride (Baker's Analyzed, NY., U.S.A.)
Calcium chloride (Baker's Analyzed, NY.,U.S.A.)
Collagenase Type 1 A (Sigma, Mo., U.S.A.)
Coomasie brilliant blue R 250 (Sigma,Mo.,U.S.A.)
Copper chloride (Baker's Analyzed, NY.,U.S.A.)
DEAE-cellulose (DE-52) (Whatman, U.S.A.)
Dipotassium phosphate (E.Merck, W.Germany)
Dialysis cellulose tubing ,WM cutoff 12,000-14,000
(Spectra/por, Spectrum,CA.U.S.A.)
Elastase Type 1 (Sigma,Mo. U.S.A.)
Elastin congo red (Sigma.Mo. U.S.A.)
Ethylenediaminetetraacetic acid (EDTA) (Sigma.Mo.,U.S.A.)

Glacial acetic acid (E.Merck.,W. Germany)
Glycine (Sigma, Mo.,U.S.A.)
Glycerol (E.Merck.,W.Germany)
Hydrochloric acid (E.Merck.,W.Germany)
p-Hydroxymercuribenzoate (Sigma.Mo.,U.S.A.)
Magnesium chloride (Baker's Analyzed,NY. U.S.A.)
Methanol (E.Merck.,W.Germany)
Mercury chloride (Baker's Analyzed.,NY.,U.S.A.)
Molecular weight markers for gel filtration
(GF-1000)(Sigma.Ma.,U.S.A.)
Molecular weight markers for SDS-PAGE (low MW.)
(Pharmacia,Sweden)
Potassium monophosphate (E.Merck.,W.Germany)
Phenylmethylsulfonylfluoride (Sigma.Mo.,U.S.A.)
1,10 phenanthroline monohydrate (Sigma.Mo.,U.S.A.)
Potassium chloride (E.Merck.,W.Germany)
Sodium dodecyl sulfate (Sigma.Mo.,U.S.A.)
Sodium hydroxide (E.Merck.,W.Germany)
Sephadex G-200 (Pharmacia,Sweden)
Riboflavin (Sigma.Mo.,U.S.A.)
N,N,N,N-tetramethylethylenediamine (TEMED) (LKB.,Sweden)
Trisma-base (Sigma.Mo.,U.S.A.)
Tris-HCl (Sigma.Mo.,U.S.A.)
Trichloreacetic acid (Sigma.Mo.,U.S.A.)
Trypsin (soy bean)inhibitor (Sigma.Mo., U.S.A.)
Zinc chloride (Baker's Analyzed, NY.,U.S.A.)

B. INSTRUMENTS

Fraction collector (2211 superRac, LKB, Bromma, Sweden)

Peristaltic pump (2232, microperpexs., LKB Bromma,
Sweden)

Incubator (Type B-80, Memmert, W. Germany)

Shaker incubator (Lab-Line orbit environ-shaker, U.S.A.)

pH meter (Corning, N.Y., U.S.A.)

Ultrafiltration cell model 8050 (Amicon, Danver, U.S.A.)

Refrigerated superspeed centrifuge (Beckman J2-21, U.S.A.)

Magnetic stirrer (M21/1 Framo-Geratetechnik, W. Germany)

Spectrophotometer (Beckman ACTA III UV-visible, U.S.A.)

Spectrophotometer (HITACHI U-200, Japan)

APPENDIX II

REAGENTS AND PREPARATIONS

1. Media for preparation of protease

1.1 Glycerol beef extract broth (GB)

Beef extract (Gibco)	1 gm.
Glycerol	4 ml.
Phosphate buffer(0.005M) pH 7.0	to 100 ml.

1.2 Glycerol beef extract supplement with calcium chloride (GBC)

Beef extract (Gibco)	1 gm.
Glycerol	4 ml.
CaCl ₂ (0.05 M)	1 ml.
Phosphate buffer (0.005 M) pH 7.0	to 100 ml.

1.3 Beef extract broth (BB)

Beef extract	1 gm.
Phosphate buffer (0.005 M) pH 7.0	to 100 ml.

1.4 Glycerol beef extract broth supplement with 0.5 % glucose (GBG)

Beef extract (Gibco)	1 gm.
Glycerol	4 ml.
Glucose	0.5 gm.
Phosphate buffer (0.005 M) pH 7.0	to 100 ml.

All the medium was mixed well until dissolved completely and sterile by autoclaving at 121° C for 15 min.

2. Reagents for protease assay

2.1 Skim milk plate test

Solution 1	Skim milk	0.9 gm.
	Phosphate buffer pH 7.0	30 ml.
Solution 2	Noble agar	0.6 gm.
	Phosphate buffer pH 7.0	30 ml.

The mixture was heated until dissolved completely. Solution 1, 5 ml was mixed with solution 2, 5 ml, then pour on petridisc which laid on the equilibrium level.

2.2 Azocasein test

2.2.1. 0.5 % azocasein

azocasein (Sigma)	0.5 gm.
Phosphate buffer pH 7.0	100 ml.

2.2.2. 0.5 % TCA

Trichloreacetic acid	5 gm.
D.W.	100 ml.

2.2.3 0.5 N NaOH

NaOH	2 gm.
D.W.	100 ml.

3. Reagents for Protein Estimation

3.1 Na₂CO₃, 2 % in NaOH, 0.1 M (solution A)

Ingradients per 200 ml

Sodium carbonate	4 gm.
0.1 M sodium hydroxide	200 ml.

3.2 $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ 0.5 % in $\text{C}_4\text{H}_4\text{KNaO}_6 \cdot 4\text{H}_2\text{O}$, 1 %
(solution B), Ingredients per 200 ml.

copper sulfate (1 %)	1 gm.
potassium sodium tartrate (2 %)	2 gm.
D.W. to	200 ml.

3.3 Solution C, Ingredients per 51 ml.

Solution A	50 ml.
Solution B	1 ml.

freshly prepared before used.

3.4 Solution D, Ingredients per 100 ml.

Folin Ciocalteus	50 ml.
D.W. to	100 ml.

Stored at 4° C in dark bottle.

4. Buffer solution

1. HCl-glycine 0.1 M pH 3.0

Glycine	7.5 gm.
adjust with 1 N HCl to pH 3.0	
D.W. to	100 ml.

2. HCl-glycine 0.1 M pH 4.0

Glycine	7.5 gm.
adjust with 1 N HCl to pH 4.0	
D.W. to	100 ml.

3. Acetic-acid-sodium acetate 0.1 M pH 5.0

Sodium acetate	0.14 gm.
1 M acetic acid	4.3 ml.
D.W. to	100 ml.



4. Phosphate buffer 0.1 M pH 6.0
- | | |
|--------------------------------|----------|
| 0.5 M KH_2PO_4 | 17.5 ml. |
| 0.5 M K_2HPO_4 | 2.4 ml. |
| D.W. to | 100 ml. |
5. Phosphate buffer 0.1 M pH 7.0
- | | |
|--------------------------------|----------|
| 0.5 M KH_2PO_4 | 7.8 ml |
| 0.5 M KH_2PO_4 | 12.2 ml. |
| D.W. to | 100 ml. |
6. Tris-HCl buffer 0.1 M pH 8.0
- | | |
|-------------------------------|----------|
| Tris base | 1.21 gm. |
| adjust with 1 N HCl to pH 8.0 | |
| D.W. to | 100 ml. |
7. Tris-HCl buffer 0.1 M pH 9.0
- | | |
|-------------------------------|----------|
| Tris base | 1.21 gm. |
| adjust with 1 N HCl to pH 9.0 | |
| D.W. to | 100 ml. |
8. Sodium bicarbonate 0.2 M pH 10.0
- | | |
|------------------|----------|
| NaCO_3 | 2.11 gm. |
| NaHCO_3 | 1.68 gm. |
| D.W. to | 100 ml. |
9. Phosphate buffer 0.01 M pH 7.0
- Solution A
- | | |
|----------------------------------|-----------|
| K_2HPO_4 (0.5 M) | 68.04 gm. |
|----------------------------------|-----------|
- Solution B
- | | |
|----------------------------------|-----------|
| K_2HPO_4 (0.5 M) | 87.09 gm. |
|----------------------------------|-----------|

Solution A ,7.7 ml, Solution 12.3 ml. and add D.W. to 100 ml.

5. Reagents for SDS-PAGE

5.1 Bis-acrylamide solution

Acrylamide	40 gm.
Bis-methyleneacrylamide	1.5 ml.
D.W. to	100 ml.

5.2 TEMED

TEMED	0.5 ml.
D.W. to	100 ml.

5.3 Buffer A (pH 8.8 ,store at 4° C)

Tris base	56.8 gm.
SDS	1.24 gm.
D.W. to	250 ml.

5.4 Buffer B (pH 6.8 ,store at 4° C)

Tris base	37.85 gm.
SDS	2.5 gm.
Adjust pH with HCl, D.W. to	250 ml.

5.5 0.5 % Ammonium persulfate

Ammonium persulfate	0.5 gm.
D.W. to	100 ml.

freshly prepared before used.

5.6 Electrode buffer running (stock solution)

Tris base	15.15 gm.
Glycine	72.0 gm.
SDS	5.0 gm.
D.W. to	100 ml.

Before used the electrode buffer is diluted 1:4.

5.7 Sample buffer

Tris-HCl	9.65 gm.
SDS	4.0 gm.
Glycerol	50.0 ml.
Bromphenol blue	2.0 mg.
D.W. to	100 ml.

5.8 Staining solution

Coomasie brillent blue R 250	0.75 gm.
Methanol	250 ml.
Glacial acetic acid	50 ml.
D.W. to	100 ml.

5.9 Destaining solution

Glacial acetic acid	100 ml.
Methanol	50 ml.
D.W. to	1000 ml.

6. Preparation of separating gel (10 %)

Bis-acrylamide	1.5 ml.
TEMED (0.5 %)	0.3 ml.
Buffer A	1.2 ml.
D.W.	1.5 ml.
0.5 % ammonium persulfate	1.5 ml.

7. Preparation of stacking gel (3 %)

Bis-acrylamide	1.0 ml.
TEMED (0.5 %)	1.0 ml.
Buffer B	1.0 ml.
D.W.	4.5 ml.
0.5 % ammonium persulfate	2.5 ml.

. 8. Polyacrylamide gel cationic system

(Reisfeld and Lewis and Williams)

Solution A	KOH 1 M	48 ml
	Gla. acetic acid	17.2 ml.
	TEMED	4 ml.
	D.W. to	100 ml.
Solution B	KOH 1 M	48 ml.
	Gla. acetic acid	2.9 ml.
	TEMED	0.45 ml.
	D.W. to	100 ml.
Solution C	Acrylamide	29.2 ml.
	Bis-acrylamide	0.8 ml.
	D.W. to	100 ml.
Solution D	Acrylamide	10 ml.
	Bis-acrylamide	2.5 ml.
	D.W. to	100 ml.
Solution E	Riboflavin	0.004 gm.
	D.W. to	100 ml.
Solution F	Ammonium persulfate	0.28 gm.
	D.W. to	100 ml.
Electrode buffer (pH 4.5)		
	3-Aminopropionic acid	31.2 gm.
	Gla. acetic acid	8.0 ml.
	D.W. to	1000 ml.

Before use the electrode buffer is diluted 1:10

9. Preparation of lower gel 7.5 % (pH 4.3)

The above solution are mixed in following proportion:

A:C:H₂O:F = 1:1:1:4

Preparation of upper gel

Solution B:D:E:H₂O = 1:2:1:4

For gel polymerization have to expose light about 1 hour.

10. Staining solution for polyacrylamide gels

1% Amido Black 10B in 7% acetic acid solution

Destaining solution : 7% acetic acid

11. Polyacrylamide gel electrophoresis (anionic system)

Solution A (pH 8.9)

1 N HCl	24.0 ml.
Tris base	18.3 ml.
TEMED	0.115 ml.
D.W. to	50 ml.

Solution B (pH 6.7)

1 N HCl	12 ml.
Tris base	1.495 ml.
TEMED	0.115 ml.
D.W. to	25 ml.

Solution C

Acrylamide	28.0 gm.
Bis-acrylamide	0.736 gm.
D.W. to	100 ml.

Solution D

Acrylamide		5.0 gm.
Bis-acrylamide		1.25 gm.
D.W.	to	100 ml.

Solution E

Riboflavin	2	mg./ 50 ml.
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Solution F

Ammonium persulfate	5	gm /100 ml.
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Preparation of lower gel: The above solution are mixed in the following proportion: A:C:H₂O:F = 1:2:1:4

Preparation of upper gel: B:D:E:H₂O = 1:2:1:4

For the gel polymerization have to expose light about 1 hr.

Sample buffer : 50 % glycerol, bromphenol blue 0.1 %

Electrophoresis buffer (pH 8.3)

Tris base		6.0 gm.
Glycine		28.8 gm.
D.W.	to	1000 ml.

Before use the electrode buffer is diluted 1:10.

12. Reagents for immunoelectrophoresis

12.1 Sodium barbital buffer (0.05 M), pH 8.0

Barbital sodium	47.6 gm.
1 N HCl	69.0 ml.
10 % NaN ₃	4.2 ml.
D.W. to	4200 ml.
adjust pH to 8.2	

12.3 Agar gel (1.5 %)

Special agar Noble	1.5 gm
Sodium barbital buffer.	100 ml.

The gel was heated until dissolved and pipette 2 ml. onto a glass slide, then allowed to cool.

12.4 Dialyzing solution

NaCl	4 gm.
N2B4O7	4 gm.
D.W. to	100 ml.

12.5 Destaining solution

Methanol	75 ml.
Glacial acetic acid	60 ml.
D.W.	30 ml.

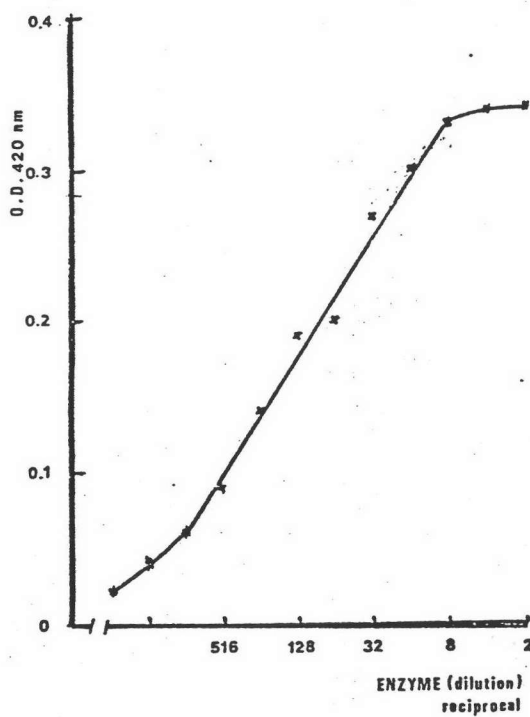


Fig. 18 Proteolytic activity against azocasein ; Plot O.D.420 nm. of azodye release versus reciprocal dilution of enzyme (*P. pseudomallei* crude filtrate).

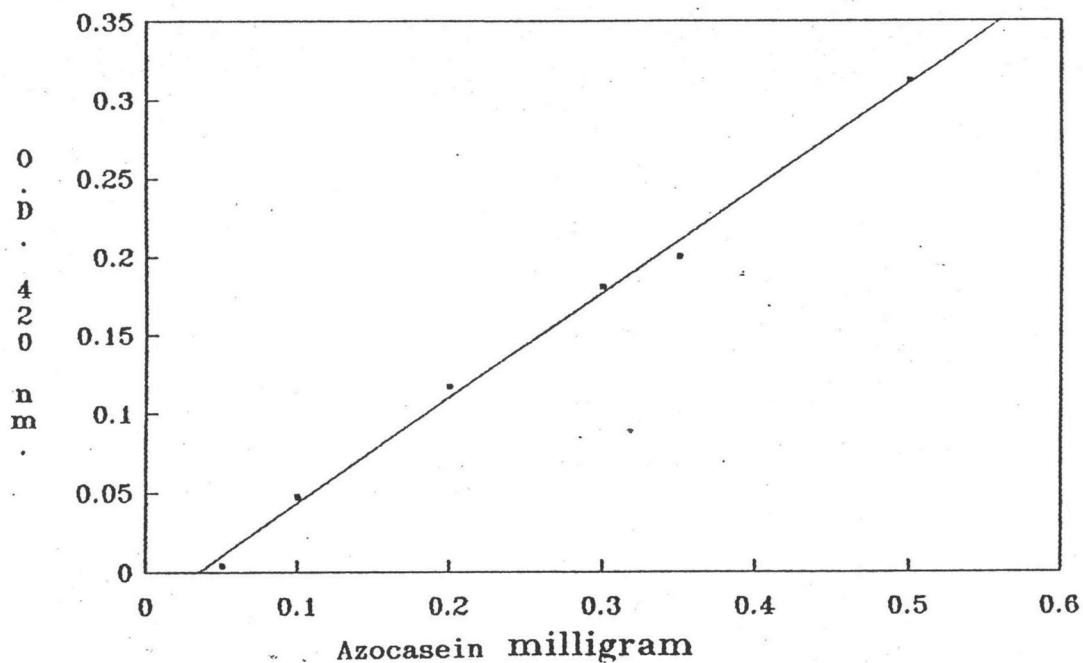


Fig. 19 Linear graph of azodye O.D. 420 nm against dilution of azocasein which was digested by trypsin 1 mg/ml.

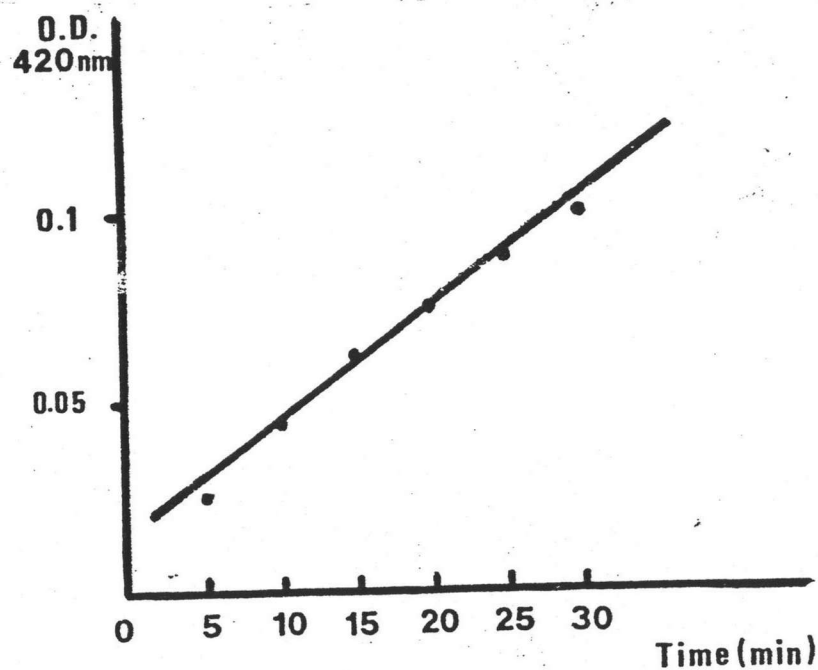


Fig. 20 Kinetic of trypsin 1 PU using azocasein as substrate, showed that 1 PU give 0.1 O.D. 420 nm, 30 min.



CURRICULUM VITAE

Miss Kanchalee Lertpocasombat was born on June 20, 1957, in Bangkok, Thailand. She graduated with the Bachelor of Science (Medical technology) degree from Chulalongkorn University in 1979. She obtained a diploma in Medical Microbiology from Institute for Medical reseach, Kuala lumpur, Malaysia and a certificate in Microbial Disease from the Research Institute for Microbial Diseases, Osaka University, Japan in 1980 and 1989 respectively. She has been working in Department of Microbiology; Bacteriology Unit, Faculty of Medicine, Chulalongkorn University since her graduation.