

CHAPTER I

INTRODUCTION

Surfactants are widely used for various purposes in industry, and most surfactants used are mainly chemically synthesized. In the past few decades, biological surface-active compounds (biosurfactants) have gained prominence and have already taken over for a number of important industrial uses, due to their superior properties biodegradability, production from renewable resources and functionality under extreme conditions; particularly those pertaining during tertiary crude-oil recovery. However, high costs of biosurfactants are a main obstruction to their application. At present, their uses are mainly in the oil and petroleum industries, where they are employed primarily for their emulsification capacity in both tertiary recovery and polluted-sites remediation. Although initial interest and applications are primarily in the area of petroleum engineering and enhanced oil recovery, new applications in medicine, cosmetic and food industries have been recognized. Probably the most important advantage of biosurfactants over chemical surfactants is their ecological acceptability. Many chemically synthesized surfactants cause ecological problems owing to their resistance to degradation, toxicity, and accumulation in natural ecosystems. On the other hand, biosurfactants are biodegradable. However, biosurfactants have several drawbacks:

- (1) poor yields from raw substrate materials
- (2) high capital investment
- (3) need for sterilisation
- (4) problems in the control of the process, for example, foaming
- (5) problems in product recovery and purification
- (6) difficulties in analysing the finished products chemically due to their complex nature

Microorganisms which produce biosurfactants, and the structures of produced biosurfactant, are listed in Table 1.1. Certain microorganisms are likely to be found to be better adapted to particular environments, such as oil reservoirs, soils and deep sea. Rhamnolipids produced by *Pseudomonas* and *Bacillus* were reported to be insensitive to the change of pH, temperature, salinity, calcium or magnesium at concentrations in excess of those found in many oil reservoirs in Venezuela (Rocha *et al.*, 1992). *Bacillus licheniformis* strain JF-2, isolated from oil field injection water, was found to produce biosurfactants under both aerobic and anaerobic conditions (Jenneman *et al.*, 1983; Javaheri *et al.*, 1985). *Bacillus* strain SP018 was found to tolerate anaerobic conditions at 50°C and < 10% NaCl while producing biosurfactants (Pfiffner *et al.*, 1986). Two biosurfactant-producing *Bacillus* strains, AB-2 and Y 12-B, were also isolated from oil sludge-mixed sand and had the ability to grow on hydrocarbon-containing medium at < 50°C (Banat *et al.*, 1993).

Table 1.1 Various biosurfactants produced by microorganisms

Microorganism	Biosurfactant	Reference
<i>Arthrobacter</i> RAG-1	hetropolysaccharides	Rosenberg <i>et al.</i> (1979)
<i>Arthrobacter</i> MIS38	lipopeptide	Morikawa <i>et al.</i> (1993)
<i>Arthrobacter</i> sp.	trehalose, sucrose and fructose lipids	Suzuki <i>et al.</i> (1974)
<i>Bacillus licheniformis</i> JF-2	lipopeptides	Itoh <i>et al.</i> (1974)
<i>Bacillus licheniformis</i> 86	lipopeptides	McInerney <i>et al.</i> (1990)
<i>Bacillus subtilis</i>	surfactin	Horowitz <i>et al.</i> (1990)
<i>Bacillus pumilus</i> A1	surfactin	Arima <i>et al.</i> (1968)
<i>Bacillus</i> sp. AB-2	rhamnolipids	Morikawa <i>et al.</i> (1992)
<i>Bacillus</i> sp. C-14	hydrocarbon-lipid-protein	Banat (1993)
<i>Candida antarctica</i>	mannosylerythritol lipid	Eliseev <i>et al.</i> (1991)
<i>Candida bombicola</i>	sophorose lipids	Kitamoto <i>et al.</i> (1992)
<i>Candida tropicalis</i>	mannan-fatty acid	Gobbett <i>et al.</i> (1984)
<i>Candida lipolytica</i> Y-917	sophoros lipid	Kappell and Fiechter (1976)
<i>Clostridium pasteurianum</i>	neutral lipids	Lesik <i>et al.</i> (1989)
<i>Corynebacterium hydrocarbolastus</i>	protein-lipid-carbohy.	Cooper <i>et al.</i> (1980)
<i>Corynebacterium insidiosum</i>	phospholipids	Zajic <i>et al.</i> (1977)
<i>Corynebacterium lepus</i>	fatty acids	Akit <i>et al.</i> (1981)
Strain MM1	glucose, lipid and hydroxydecanoic acids	Cooper <i>et al.</i> (1979)
<i>Nocardia erythropolis</i>	neutral lipids	Passeri (1992)
<i>Ochrobactrum anthropii</i>	protein	MacDonald <i>et al.</i> (1981)
<i>Penicillium spiculispurum</i>	spiculisporic acid	Wasko and Bratt (1990)
<i>Pseudomonas aeruginosa</i>	rhamnolipid	Ban and Sato (1993)
<i>Pseudomonas fluorescens</i>	lipopeptide	Robert <i>et al.</i> (1989)
<i>Phaffia rhodozyma</i>	carbohydrate-lipid	Neu <i>et al.</i> (1990)
<i>Rhodococcus erythropolis</i>	trehalose dicorynomycolate	Lesik <i>et al.</i> (1991)
<i>Rhodococcus</i> sp. ST-5	glycolipid	Shulga <i>et al.</i> (1990)
<i>Rhodococcus</i> sp. H13-A	glycolipid	Abu-Ruwaida <i>et al.</i> (1991a)
<i>Rhodococcus</i> sp. 33	polysaccharide	Singer and Finnerty (1990)
<i>Torulopsis bombicola</i>	sophorose lipids	Neu <i>et al.</i> (1992)
		Inoue and Ito (1982)

Biosurfactants are mainly classified in to four categories, glycolipid, phospholipid, lipopeptide, 4.polymeric, based on the structure of their hydrophilic part. Among these biosurfactants, glycolipid biosurfactants have been most intensively studied because their production yields are much higher than those of other types of biosurfactants. The production of biosurfactants suitable for specific applications and the development of high added value properties is the central part of the future research development project (Figure 1.1), reinforced by structured screenings and computational monetization.

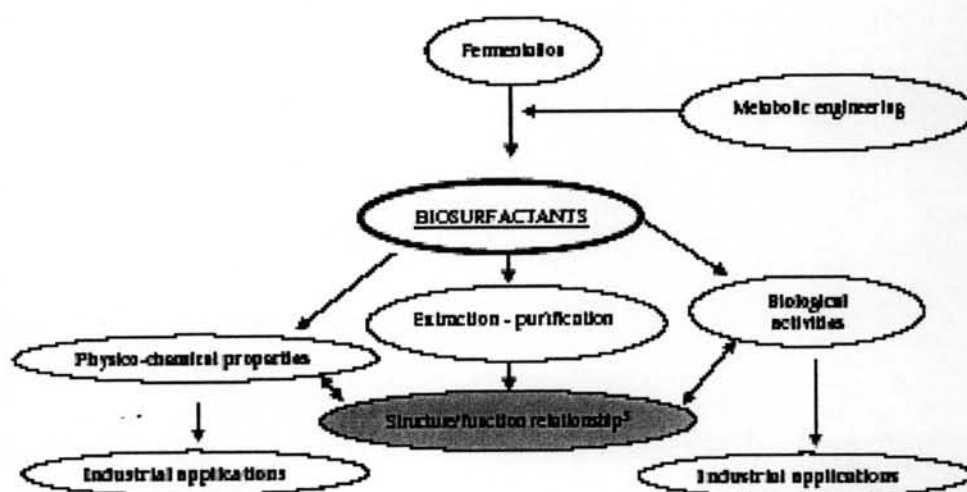


Figure 1.1 Pathways for biosurfactants production and applications

The objective of the present work were to isolate biosurfactant-producing bacteria from oil sludge and to characterize the produced biosurfactants. In addition, the effectiveness of the produced biosurfactants toward oil recovery was investigated.