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APPENDIX

I. Reagent for Gram staining

1. Gram crystal violet solution

a. Solution A:

Crystal violet	2.0 g.
Ethanol (95%)	20.0 ml.

Crystal violet was dissolved in ethanol to make a solution.

b. Solution B:

Ammonium oxalate	0.8 g.
Distilled water	80.0 ml.

Ammonium oxalate was dissolved in distilled water. Then, solution A and solution B were mixed to make gram's crystal violet solution.

2. Gram iodine solution

Iodine	1.0 g.
Potassium iodide	2.0 g.
Distilled water	300.0 ml.

Iodine and potassium iodide were mixed in a mortar and distilled water was added to make the solution.

3. Gram safranin solution

Safranin	0.25 g.
Ethanol (95%)	10.00 ml.
Distilled water	100.00 ml.

Safranin was dissolved in ethanol and distilled water was added. The solution was filtrated by filter paper.

Staining procedure

1. The organisms were smeared on a clean slide and allowed to dry.
2. The slide was heated with a flame to fix the smear.
3. Gram crystal violet was dropped on the smear. After 1 minute, the slide was then washed with water and drained.
4. Next, gram iodine solution was dropped on the smear, and washed with water after 1 minute.
5. The smear was decolorized with ethanol (95%) and then washed with water.
6. Gram safranin solution was next dropped on the smear in oder to use as counterstain for 30 second. The smear was allowed to dry.
7. The smear was examined by microscopy under 100x objective len over the entire smear.

II. Catalase test

Several colonies of *S. aureus* was smeared on a clean slide. The 3% hydrogen peroxide was dropped and mixed with the organisms.

The positive result was shown as bubbles formation.

III. Coagulase test

3.1 Slide coagulase test

Several colonies of *S. aureus* was mixed generously with a dropped of human plasma on a clean slide.

The positive result was shown as the white precipitates formation within 5 minutes.

3.2 Tube coagulase test

The 0.5 ml of staphylococcal broth culture or a loopful of staphylococci was mixed with 0.5 ml of human plasma in a clean tube. The tube was incubated at 37°C for 3 to 4 hour.

The positive result was developed as a clot gel.

VI. Media

4.1 Mannitol salt agar

Proteose peptone No.3, Difco	10.0	g.
Bacto beef extract	1.0	g.
Bacto D-mannitol	10.0	g.
Sodium chloride	75.0	g.
Bacto agar	15.0	g.
Phenol red	0.025	g.
Distilled water qs.	1000.0	ml.

4.2 Mueller-Hinton agar medium

Beef, Infusion from	300.0	g.
Bacto casamino acids, Technical	17.5	g.
Starch	1.5	g.
Bacto agar	17.0	g.
Distilled water qs.	1000.0	ml.

For Mueller-Hinton agar containing 5% sodium chloride, 50 grams of sodium chloride was added.

4.3 Mueller-Hinton broth medium

Beef, Infusion from	300.0	g.
Casamino acid, Technical	17.5	g.
Bacto soluble strach	1.5	g.
Distilled water qs.	1000.0	ml.

4.4 Tryptose blood agar base medium

Bacto tryptose	10.0	g.
Bacto beef extract	3.0	g.
Sodium chloride	5.0	g.
Bacto agar	15.0	g.
Distilled water qs.	950.0	ml.

Media Preparation

All of ingredients were dissolved in distilled water and then sterilized by autoclaving at 121°C, 15 pounds /inch² pressure, for 15 minutes. The sterile medium was cooled to 45°C to 50°C, and dispensed into sterile plates or tubes. For sterile tryptose blood agar base medium, 50

ml of sterile defibrinated blood was added aseptically into sterile plates before dispensation.

4.5 0.8% Agarose Gel

The correct amount of powdered agarose electrophoresis medium type was added to a measured quantity of TBE buffer and made 0.8% agarose gel. The slurry was heated in a autoclave at 15 pound/inch² 121°C 15 minutes to dissolve the agarose and was cooled to 50°C. The edge of a clean, dry, plastic plate (10 by 15 cm) was sealed with the autoclave tape so as to form a mold. The edge of the mold was sealed with a small quantity of agarose solution using a pasteur pipette. When the seal was set, the rest of the warm agarose dilution was poured into the mold and was immediately clamp the comb. The sample wells were formed by using the teeth of the comb into position near one end of the gel. The depth of agarose between the bottom of the teeth and the base of the gel was approximately 0.5 to 1.0 mm. After the gel was completely set (30 to 45 minutes at room temperature), the comb and autoclave tape were carefully removed, and the gel was mounted in the electrophoresis tank.

V. Reagents

5.1 Lysis buffer (Treat buffer for plasmid extraction)

Sucrose	8%
Triton X100	5%

EDTA	50 mM
Tris.HCl	50 mM

All of ingredients were dissolved in deionized water and was adjusted the pH to 8.0. (Tris-base was used if it was necessary.)

5.2 Loading buffer (x6)

Bromophenol blue	0.25	g.
Sucrose	40.0	g.
Deionized water qs.	100.0	ml.

Sucrose was dissolved in double distilled water and bromophenol blue was added to make the solution.

5.3 Tris-borate buffer (TBE)

Concentrated stock solution (x5)

Tris base	54.0	g.
Boric acid	27.5	g.
0.5 M EDTA	20.0	ml.

All of ingredients were dissolved in deionized water and adjusted the pH to 8.0. Then deionized water was added to 1000.0 ml.

Working solution

The 100 ml of concentrated stock solution was added into 900 ml. of double distilled water. The working solution consisted of 0.089 M Tris- borate, 0.089 M boric acid and 0.002 M EDTA.

5.4 Ethidium bromide

Ethidium bromide stock solution (5 mg/ml in water) was diluted to 0.5 μ g/ml in TBE.

5.5 10 mM Tris, pH 8.0

121.2 mg of Tris- base was dissolved in 800 ml. of deionized water, and adjusted the pH to 8.0 by adding concentrated HCl. The solution was made up the volume to 1000.0 ml.

5.6 TE buffer

10 mM Tris-base

1 mM EDTA

Tris-base and EDTA were dissolved in deionized water and adjusted the pH to 8.0 by adding concentrated HCl.

5.7 Saturated phenol

Phenol was dissolved in 1 M Tris-base, pH 8.0 to make saturated phenol emulsion. The emulsion was stored at -4°C until used.

5.8 1% Potassium phosphate buffer, pH 6.0

Dibasic potassium phosphate 2.0 g.

Monobasic potassium phosphate 8.0 g.

Distilled water qs to 1000.0 ml.

Dibasic potassium phosphate and monobasic potassium phosphate were dissolved in distilled water, and

adjusted the pH to 6.0 with 18 N phosphoric acid or 10 N potassium hydroxide.

5.9 0.5 Mc Farland standard solution

0.5 ml of 0.048 M Barium chloride (1.175% w/v Barium chloride-2H₂O) was added into 99.5 ml of 0.35 N H₂SO₄ (1% v/v). The suspension was agitated on a vortex mixer just before use. The turbidity of standard should give an optical density of 0.08 to 0.10 at 625 nm when tested in a spectrophotometer with a 1-cm light path.

All of the reagents except ethidium bromide and saturated phenol were sterilized by autoclave at 121°C, 15 pounds pressure, for 15 minutes. Reagents were stored at 4°C until used or 2 weeks.



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