14/

ศึกษาการนำเปลือกหุ้มเมล็ดเทียนเกล็ดหอยมาใช้เป็นสารยึดเกาะ ในการเตรียมยาเม็ด

นาย สรวุฒิ รุจิวิพัฒน์



วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาเภสัชศาสตรมหาบัณฑิต ภาควิชาเภสัชอุตสาหกรรม บัณฑิตวิทยาลัย จุฬาลงกรณ์มหาวิทยาลัย พ.ศ. 2539

> ISBN 974-633-247-3 ลิขสิทธ์ของบัณฑิตวิทยาลัย จุฬาลงกรณ์มหาวิทยาลัย

THE STUDIES OF ISPAGHULA HUSK AS BINDER FOR TABLET PREPARATIONS

Mr. Soravoot Rujivipat

A Thesis Submitted in Partial Fulfillment of the Requirements

for the Degree of Master of Science in Pharmacy

Department of Manufacturing Pharmacy

Graduate School

Chulalongkorn University

1996

ISBN 974-633-247-3

Thesis Title The Studies of Ispaghula Husk as Binder for Tablet Preparations

By Mr. Soravoot Rujivipat

Department Manufacturing Pharmacy

Thesis Advisor Assoc. Prof. Kaisri Umprayn, Ph.D.

Accepted by the Graduate School, Chulalongkorn University in Partial Fulfillment of the Requirements for the Master's Degree

Sand Thorng suran
Dean of Graduate School
(Assoc. Prof. Santi Thoongsuwan, Ph.D.)

Thesis Committee

P. Ialvaniel Chairman (Assoc. Prof. Poj Kulvanich, Ph.D.)

(Assoc. Prof. Kaisri Umprayn, Ph.D.)

Parunce Thanomkist Member
(Assoc. Prof. Parunee Thanomkiat, M. Pharm. St.)

Sucherda Prasentultyahu Member (Assoc. Prof. Suchada Prasentvithyakarn, M. Sc. in Pharm.)

พิมพ์ตันฉบับบทคัดย่อวิทยานิพนธ์ภายในกรอบสีเขียวนี้เพียงแผ่นเดียว

สรวุฒิ รุจิวิพัฒน์ : ศึกษาการนำเปลือกหุ้มเมล็ดเทียนเกล็ดหอยมาใช้เป็นสารยึดเกาะใน การเตรียม ษาเม็ด (THE STUDIES OF ISPAGHULA HUSK AS BINDER FOR TABLET PREPARATIONS) อ. ที่ปรึกษา : รศ. ดร. ไกรสีห์ อัมพรายน์ , 157 หน้า ISBN 974-633-247-3

เปลือกหุ้มเมล็คชั้นนอกและชั้นติคกันที่แยกจากเมล็คสุกที่แห้งของเทียนเกล็คหอย (Plantago ovata Forsk.) มีผู้รายงานว่ามีความหนืดและเป็นสารยึดติดที่เป็นเมือก ในการศึกษานี้ได้ประเมินคุณสมบัติการ ยึดเกาะสำหรับการเตรียมยาเม็คเปรียบเทียบกับสารช่วยยึดเกาะอื่นที่ใช้กันทั่วไป เช่น PVP K30, HPC type L, gelatin, corn starch และ Starch 1500° การศึกษาใช้สองวิธี สารช่วยยึดเกาะใช้ระดับ 0.5,1 และ 2% ของ น้ำหนักแท้งสำหรับยาเม็ดพาราเซตามอล และ นิโคตินาไมด์ สำหรับการเตรียมโดยวิธี solution incorporation ส่วนการเตรียบโดยวิธี dry incorporation จะใช้สารช่วยยึดเกาะในปริบาณ 1, 2 และ 4% ของน้ำหนักแห้งของ คำรับ คุณสมบัติช่วยยึดเกาะของสารต่างๆที่ใช้จะประเมินจากคุณสมบัติทางกายภาพของแกรนูล (เช่น ขนาดและการกระจายขนาดของแกรนูล การไหล ความกร่อนของแกรนูล) และเม็ดยา (เช่น ความกร่อน การแตกกระจายตัว การละลายและค่าดัชนีการยึดเกาะ) จากผลการทดลองค่าดัชนีการยึดเกาะที่ได้จากตัวยา ทั้งสองชนิด แสคงให้เห็นว่า เปลือกหุ้มเมล็ดเทียนเกล็ดหอยมีคุณสมบัติช่วยยึดเกาะดีกว่า corn starch และ Starch 1500 แต่ค้อยกว่า PVP K30, HPC type L, gelatin ในกรณีวิธี dry incorporation เปลือกหุ้มเมล็ด เทียนเกล็ดหอยให้คณสมบัติช่วยยึดเกาะดีกว่า Starch 1500° (ยกเว้น นิโคตินาไมด์ ที่ความเข้มข้นของสาร ช่วยยึดเกาะ 4% ของน้ำหนักแห้งของตำรับ ซึ่ง Starch 1500° ดีกว่า) แต่ด้อยกว่า PVP K30 จากการศึกษานี้ ความเข้มข้นของเปลือกหุ้มเมล็คเทียนเกล็คหอยที่เหมาะสมจะใช้เป็นสารช่วยยึคเกาะที่มีประสิทธิภาพสำหรับ ทั้งสองวิธีกับยาทั้งสองชนิค ประมาณ 2% ของน้ำหนักแท้งของตำรับ นอกจากนี้ในกรณีของพาราเซตามอล แกรนูล และยาเบิค พาราเซตามอล พบว่า การเตรียมโดยวิธี solution incorporation จะให้ค่าดัชนีการยึคเกาะ สูงกว่าการเครียมโดย วิธี dry incorporation ยิ่งกว่านั้น นิโคตินาไมด์แกรนูล และยาเม็คนิโคตินาไมด์ ซึ่งเตรียบโดยทั้งสองวิธี จะให้ผลที่ใกล้เคียงกัน

ภาควิชา	ภสัชอุคสาหกรรม	ลายมือชื่อนิสิต ลิวรลิ สิริธรลา
สาขาวิชา	_ 33	ลายมือชื่ออาจารย์ที่ปรึกษา ผู้ผลิง อ้องพบาว
ปีการศึกษา		ลายมือชื่ออาจารย์ที่ปรึกษาร่วม

C 575204 MAJOR MANUFACTURING PHARMACY

KEY WORD: ISPAGHULA HUSK/ TABLET TENSILE SRTENGTH/ BINDER INDEX

SORAVOOT RUJIVIPAT: THE STUDIES OF ISPAGHULA HUSK AS BINDER

FOR TABLET PREPARATIONS. THESIS ADVISOR: ASSOC. PROF.

KAISRI UMPRAYN, Ph.D., 157 PP. ISBN 974-633-247-3

Ispaghula husk (Plantago ovata Forsk.), the epidermis and the collapsed adjacent layers removed from dried ripe seeds, is reported to form viscous and adhesive mucilage. In this study, it was evaluated for binding properties in tablet preparation as comparing with commonly used binders such as PVP K30, HPC type L, gelatin, corn starch and Starch 1500°. Two methods were studies. For solution incorporation, the binders were employed at 0.5, 1 and 2% w/w of paracetamol and nicotinamide tablets. case of dry incorporation method, the amount of binder used were 1, 2 and 4% w/w. The physical properties of granules (such as granule size, size distribution, flowability and granule tablets(such as hardness, friability, and friability) disintegration, dissolution and binder index) were evaluated for According to the results of binder their binding efficacy. index obtained from both drugs illuminated that in solution incorporation method, Ispaghula husk possessed properties superior to corn starch and Starch 1500 but inferior to PVP K30, HPC type L, gelatin. In the case of dry incorportion method, Ispaghula husk gave binding properties superior to Starch 1500 (except for nicotinamide at binder concentration 4% w/w, Starch 1500 was better) but inferior to PVP K30. From this study, suitable concentration of Ispaghula husk used as the effective binding agent for both methods with both drugs was ≅ 2% w/w. In addition, in the case of paracetamol granules and it was found that solution incorporation method tablets, produced binder index higher than dry incorporation method. Furthermore, nicotinamide granules and tablets prepared by both methods gave comparable results

ภาควิชา	ลายมือชื่อนิสิต รื่องลงจน์ โกกู เกุลป.
สาขาวิชา	ลายมือชื่ออาจารย์ที่ปรึกษา <i>X ณารุงา (คำครูงาก</i>
ปีการศึกษา ²⁵³⁸	ลายมือชื่ออาจารย์ที่ปรึกษาร่วม

ACKNOWLEDGMENTS

To my graduate advisor, Associate Professor Dr. Kaisri Umprayn. I would like to express my special, sincere thanks, gratitude and appreciation for his helpful suggestion of problems, meaningful guidance and encouragement throughout my study. His patience, kindness, and understanding are also profoundly appreciated.

I also wish to express my gratitude to all members of my Graduate Committee for their helpful comments, and to Assistant Professor Dr. Poj Kulvanich, Head of the Department of Manufacturing Pharmacy, for his encouragement and providing me with opportunity to study this program.

A special acknowledgement is extended to the Department of Manufacturing Pharmacy, Faculty of Pharmaceutical Science, Chulalongkorn University and the Government Pharmaceutical Organization for the research facilities, including all the people who were concerned with my thesis.

I am indebted to the graduate school, Chulalongkorn University for granting partial financial support to fullfill this thesis.

Above all, I would like to express my infinite thanks and gratitude to dear parents for their love, care and endless encouragement throughout my life.

Finally, my deep appreciation to my fellow and other persons whose name have not been mentioned for their help in anyway all times of my study. Thank you for all.

CONTENTS

	PAGE
ABSTRACT (Thai)	IV
ABSTRACT (English)	V
ACKNOWLEDGMENTS	
CONTENTS	VII
LIST OF TABLES	vIII
LIST OF FIGURES	
LIST OF ABBREVIATIONS	XXI
CHAPTER	
I INTRODUCTION	1
LITERATURE REVIEW	2
PURPOSE OF THE STUDY	15
II EXPERIMENTAL	16
III RESULTS	31
IV DISSCUSSION AND CONCLUSION	91
REFERENCES	99
APPENDICES	104
VITAE	

LIST OF TABLES

Table		Page
1	Characteristics of Plantago seeds from various species	3
2	Summaries some commonly used binders with the usual concentration range of their application, together with the granulating solvent.	6
3	Formulation of paracetamol and nicotinamide tablets prepared by solution incorporation method	19
4	Formulation of paracetamol and nicotinamide tablets prepared by dry incorporation method	19
5	Particle size distribution of paracetamol granules prepared with various binders	41
6	Particle size distribution of nicotinamide granules prepared with various binders	43
7	Physical properties of paracetamol granules prepared with various binders and concentrations.	58
8	Physical properties of nicotinmide granules prepared with various binders and concentrations	60
9	Physical properties of paracetamol tablets prepared with various binders and concentration.	64
10	Physical properties of nicotinamide tablets prepared with various binders and concentration.	66
11	Calibration data between strain and applied forces of strain guages bound on upper punch	108
12	Absorbance of paracetamol standard solution in phosphate buffer pH 5.8	109

Table		Page
13	Absorbance of nicotinamide standard solution in purified water	110
14	Dissolution data of paracetamol tablets prepared with Ispaghula husk by dry incorporation method	123
15	Dissolution data of paracetamol tablets prepared with pregelatinized starch by dry incorporation method	124
16	Dissolution data of paracetamol tablets prepared with polyvinylpyrrolidone by dry incorporation method	125
17	Dissolution data of paracetamol tablets prepared with corn starch by solution incorporation method	126
18	Dissolution data of paracetamol tablets prepared with gelatin by solution incorporation method	127
19	Dissolution data of paracetamol tablets prepared with polyvinylpyrrolidone by solution incorporation method	128
20	Dissolution data of paracetamol tablets prepared with pregelatinized starch by solution incorporation method	129
21	Dissolution data of paracetamol tablets prepared with Ispaghula husk by solution incorporation method	130
22	Dissolution data of paracetamol tablets prepared with hydroxypropyl cellulose by solution incorporation method.	131
23	Dissolution data of nicotinamide blank tablets	132
24	Dissolution data of nicotinamide tablets prepared with Ispaghula husk by dry incorporation method	133
25	Dissolution data of nicotinaamide tablets prepared with pregelatinized starch by dry incorporation method	134
26	Dissolution data of nicotinamide tablets prepared with polyvinylpyrrolidone by dry incorporation method	. 135

Γable		Page
27	Dissolution data of nicotinamide tablets prepared with corn starch by solution incorporation method	136
28	Dissolution data of nicotinamide tablets prepared with polyvinylpyrrolidone by solution incorporation method	137
29	Dissolution data of nicotinamide tablets prepared with gelatin by solution incorporation method	138
30	Dissolution data of nicotinamide tablets prepared with hydroxypropyl cellulose by solution incorporation method.	139
31	Dissolution data of nicotinaamide tablets prepared with pregelatinized starch by solution incorporation method	140
32	Dissolution data of nicotinamide tablets prepared with Ispaghula husk by solution incorporation method	141
33	Analysis of variance and LSD analysis for hardness of paracetamol tablets prepared with various binders at 1 % w/w by dry incorporation method	145
34	Analysis of variance and LSD analysis for hardness of paracetamol tablets prepared with various binders at 2 % w/w by dry incorporation method	146
35	Analysis of variance and LSD analysis for hardness of paracetamol tablets prepared with various binders at 4 % w/w by dry incorporation method	147
36	Analysis of variance and LSD analysis for hardness of paracetamol tablets prepared with various binders at 0.5 % w/w by solution incorporation method	148
37	Analysis of variance and LSD analysis for hardness of paracetamol tablets prepared with various binders at 1 % w/w by solution incorporation method	149
39	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 1 % w/w by dry incorporation method.	. 151

Γable		Page
40	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 2 % w/w by dry incorporation method	152
41	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 4 % w/w by dry incorporation method	153
42	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 0.5 % w/w by solution incorporation method	154
43	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 1 % w/w by solution incorporation method	155
44	Analysis of variance and LSD analysis for hardness of nicotinamide tablets prepared with various binders at 2 % w/w by dry incorporation method	156

LIST OF FIGURES

Figure		Page
1	Stage in the development of moist granules as the proportion of liquid is increased in wet granulation process	8
2	Function block diagram of press and associated measuring system.	23
3	Calibration curve between strain and applied forces of gaugee bound on instrumented upper punch	24
4	Fractured tablets after diameteral compression	26
5	Standard curve of paracetamol in phosphate buffer pH 5.8 at 249 nm	29
6	Standard curve of nicotinamide in purified water at 262 nm.	29
7	Diagram of procedure for assay the quantity of paracetamol tablets	30
8	Diagram of procedure for assay the quantity of nicotinamide tablets	30
9	Photomicrographs of original paracetamol powders	32
10	Photomicrographs of lactose powders	32
11	Photomicrographs of paracetamol granules prepared without binder	32
12	Photomicrographs of paracetamol granules prepared with 2 % Starch 1500 [®] by dry incorporation method	33
13	Photomicrographs of paracetamol granules prepared with 2 % PVP K30 by dry incorporation method	33
14	Photomicrographs of paracetamol granules prepared with 2 % Ispaghula husk by dry incorporation method	33

Figure		Page
15	Photomicrographs of paracetamol granules prepared with 2% gelatin by solution incorporation method	34
16	Photomicrographs of paracetamol granules prepared with 2% HPC type L by solution incorporation method	34
17	Photomicrographs of paracetamol granules prepared with 2% corn starch by solution incorporation method	34
18	Photomicrographs of paracetamol granules prepared with 2% Starch 1500 [®] by solution incorporation method	35
19	Photomicrographs of paracetamol granules prepared with 2% PVP K30 by solution incorporation method	35
20	Photomicrographs of paracetamol granules prepared with 2% Ispaghula husk by solution incorporation method	35
21	Photomicrographs of original nicotinamide powders	36
22	Photomicrographs of nicotinamide granules prepared without binder	36
23	Photomicrographs of nicotinamide granules prepared with 2% Starch 1500 [®] by dry incorporation method	36
24	Photomicrographs of nicotinamide granules prepared with 2% PVP K30 by dry incorporation method	37
25	Photomicrographs of nicotinamide granules prepared with 2% Ispaghula husk by dry incorporation method	37
26	Photomicrographs of nicotinamide granules prepared with 2% gelatin by solution incorporation method	37
27	Photomicrographs of nicotinamide granules prepared with 2% HPC type L by solution incorporation method	38
28	Photomicrographs of nicotinamide granules prepared with 2% corn starch by solution incorporation method	38

Figure		Page
29	Photomicrographs of nicotinamide granules prepared with 2% Starch 1500 [®] by solution incorporation method	38
30	Photomicrographs of nicotinamide granules prepared with 2% PVP K30 by solution incorporation method	39
31	Photomicrographs of nicotinamide granules prepared with 2% Ispaghula husk by solution incorporation method	39
32	Effect of binder types and concentrations on average size of nicotinamide granules prepared by dry incorporation method.	42
33	Effect of binder types and concentrations on average size of nicotinamide granules prepared by solution incorporation method.	42
34	Effect of binder types and concentrations on average size of paracetamol granules prepared by dry incorporation method.	44
35	Effect of binder types and concentrations on average size of paracetamol granules prepared by solution incorporation method	44
36	Compressibility of paracetamol granules prepared by dry incorporation method	46
37	Compressibility of paracetamol granules prepared by solution incorporation method.	46
38	Compressibility of nicotinamide granules prepared by dry incorporation method	47
39	Compressibility of nicotinamide granules prepared by solution incorporation method	47
40	Flowability of paracetamol granules prepared by dry incorporation method	50

Figure		Page
41	Flowability of paracetamol granules prepared by solution incorporation method.	50
42	Flowability of nicotinamide granules prepared by dry incorporation method.	51
43	Flowability of nicotinamide granules prepared by solution incorporation method.	51
44	Percent fine of paracetamol granules prepared by dry incorporation method.	52
45	Percent fine of paracetamol granules prepared by solution incorporation method	52
46	Percent fine of nicotinamide granules prepared by dry incorporation method	53
47	Percent fine of nicotinamide granules prepared by solution incorporation method	53
48	Friability of paracetamol granules prepared by dry incorporation method	56
49	Friability of paracetamol granules prepared by solution incorporation method.	56
50	Friability of nicotinamide granules prepared by dry incorporation method.	57
51	Friability of nicotinamide granules prepared by solution incorporation method	57
52	Effect of various binder and concentrations on hardness of paracetamol tablets	68
53	Effect of various binder and concentrations on hardness of nicotinamide tablets	68
54	Effect of various binder and concentrations on tensile strength of paracetamol tablets	69

Figure		Page
55	Effect of various binder and concentrations on tensile strength of nicotinamide tablets	69
56	Effect of various binder and concentrations on friability of paracetamol tablets.	71
57	Effect of various binder and concentrations on friability of nicotinamide tablets	71
58	Effect of various binder and concentrations on porosity of paracetamol tablets	73
59	Effect of various binder and concentrations on porosity of nicotinamide tablets	73
60	Effect of various binder and concentrations on disintegration time of paracetamol tablets	75
61	Effect of various binder and concentrations on disintegration time of nicotinamide tablets	75
62	Dissolution rate profiles of paracetamol tablets prepared with Ispaghula husk by dry incorporation method	77
63	Dissolution rate profiles of paracetamol tablets prepared with pregelatinized starch by dry incorporation method	77
64	Dissolution rate profiles of paracetamol tablets prepared with PVP K30 by dry incorporation method	78
65	Dissolution rate profiles of paracetamol tablets prepared with corn starch by solution incorporation method	78
66	Dissolution rate profiles of paracetamol tablets prepared with gelatin by solution incorporation method	7 9
67	Dissolution rate profiles of paracetamol tablets prepared with PVP K30 by solution incorporation method	7 9
68	Dissolution rate profiles of paracetamol tablets prepared with pregelatinized starch by solution incorporation method	80

F	igure		Page
	69	Dissolution rate profiles of paracetamol tablets prepared with Ispaghula husk by solution incorporation method	80
	70	Dissolution rate profiles of paracetamol tablets prepared with HPC type L by solution incorporation method	81
	71	Dissolution rate profiles of nicotinamide tablets prepared without binder	81
	72	Dissolution rate profiles of nicotinamide tablets prepared with Ispaghula husk by dry incorporation method	82
	73	Dissolution rate profiles of nicotinamide tablets prepared with pregelatinized starch by dry incorporation method	82
	74	Dissolution rate profiles of nicotinamide tablets prepared with PVP K30 by dry incorporation method	83
	75	Dissolution rate profiles of nicotinamide tablets prepared with corn starch by solution incorporation method	83
	76	Dissolution rate profiles of nicotinamide tablets prepared with PVP K30 by solution incorporation method	84
	77	Dissolution rate profiles of nicotinamide tablets prepared with gelatin by solution incorporation method	84
	78	Dissolution rate profiles of nicotinamide tablets prepared with HPC type L by solution incorporation method	85
	79	Dissolution rate profiles of nicotinamide tablets prepared with pregelatinized starch by solution incorporation method.	85
	80	Dissolution rate profiles of nicotinamide tablets prepared with Ispaghula husk by solution incorporation method	86
	81	Effect of various binder and concentrations on median dissolution time of paracetamol tablets	87
	82	Effect of various binder and concentrations on median dissolution time of nicotinamide tablets	87

Figure		Page
83	Effect of various binder and concentrations on binder index of paracetamol tablets	90
84	Effect of various binder and concentrations on binder index of nicotinamide tablets	90
85	The cumulative percent undersize of paracetamol granules prepared without binder	112
86	Effect of PVP K30 concentration on the cumulative percent undersize of paracetamol granules prepared by dry incorporation method	112
87	Effect of Ispaghula husk concentration on the cumulative percent undersize of paracetamol granules prepared by dry incorporation method	113
88	Effect of pregelatinized starch concentration on the cumulative percent undersize of paracetamol granules prepared by dry incorporation method	113
89	Effect of Ispaghula husk concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method.	114
90	Effect of pregelatinized starch concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method.	114
91	Effect of HPC type L concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method	115
92	Effect of PVP K30 concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method	115
93	Effect of corn starch concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method.	116

Figure		Page
94	Effect of gelatin concentration on the cumulative percent undersize of paracetamol granules prepared by solution incorporation method.	116
95	The cumulative percent undersize of nicotinamide granules prepared without binder	117
96	Effect of PVP K30 concentration on the cumulative percent undersize of nicotinamide granules prepared by dry incorporation method.	117
97	Effect of Ispaghula husk concentration on the cumulative percent undersize of nicotinamide granules prepared by dry incorporation method.	118
98	Effect of pregelatinized starch concentration on the cumulative percent undersize of nicotinamide granules prepared by dry incorporation method	118
99	Effect of corn starch concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method.	119
100	Effect of HPC type L concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method	119
101	Effect of pregelatinized starch concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method	120
102	Effect of Ispaghula husk concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method.	120
103	Effect of PVP K30 concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method	121

Figure		Page
104	Effect of gelatin concentration on the cumulative percent undersize of nicotinamide granules prepared by solution incorporation method.	53

LIST OF ABBREVIATIONS

μm	=	micrometer
avg	=	average
cm	=	centimeter
g	=	gram
kg	=	kilogram
kp	=	kilopound
m	=	meter
mg	= ,	milligram
min	=	minute
ml	=	milliliter
q.s.	=	make to quantity
r.p.m.	=	revolution per minute
S.D.	=	standard deviation
sec	=	second
SEM	_	scanning electron
		photomicroraph
°C	=	degree Celcius