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APPENDIX

1. Media and Ingredients

1.1. The defined supplement

The defined supplement was a modification of that described by Kellogg et al. (55) and was composed of 2 mg co-carboxylase, 1 g L-glutamine, 40 g glucose, and 50 mg ferric nitrate per 100 ml of distilled water. The mixture was sterilized by filtration and stored at 4 °C until use.

1.2. Kellogg medium (GCBS)

The Kellogg medium was prepared by dissolving 36 g of GC agar base (BBL) in 1000 ml of distilled water, heated until dissolved completely and sterilized at 15 lbs for 15 min. Ten ml of defined supplement were added when the medium was warm, and poured into each sterile petri dish. All GCBS media were dried at 36-37 °C to remove surface moisture before use.

2. Reagents for Purification of Gonococcal Pili

2.1. Tris-HCl buffer 0.01M, pH 9.5

1.2114 g of Tris (hydroxymethyl) aminomethane was dissolved in about 800 ml of distilled water. Then adjust the pH to 9.5 with 1 N HCl and made up the volume to 1000 ml with distilled water.

2.2. Tris-HCl buffer 0.01 M, pH 7.0, 0.01 M NaN₃

The buffer was prepared by dissolving 0.3028 g of Tris, 0.1625 g of sodium azide in distilled water, then adjusted to pH 7.0 with 1N HCl and was made up the volume to 250 ml.

2.3. Ammonium sulfate, saturated solution

One kilogram of ammonium sulfate (Merk) was dissolved in 1000 ml distilled water and heated until dissolved completely, filtered, allowed to stand overnight at room temperature and pH adjusted to 7.0 with ammonium hydroxide solution.

2.4. Ammonium sulfate, 20% saturated solution

One hundred ml of saturated ammonium sulfate was diluted with 400 ml distilled water.

3. Reagents for Lowry Method (Protein Determination)

3.1. Reagent A

The grams of sodium carbonate (Na₂CO₃) was dissolved in 0.5 N NaOH solution.

3.2. Reagent B

One gram of copper sulfate, hydrated (CuSO₄·5H₂O) was dissolved in 100 ml distilled water.

3.3. Reagent C

The two grams of potassium tartrate was dissolved in 100 ml of distilled water.

3.4. Diluted Folin-Ciocalteu phenol reagent

This solution was freshly prepared by diluting 5 ml of Folin Ciocalteu Phenol reagent (Sigma) with 45 ml of distilled water.

3.5. Bovine serum albumin (CSL) standard (0.3 mg/ml)

4. Reagents for Electron Microscopy

4.1. Fixative solution

4 % glutaraldehyde in 0.1 M phosphate buffer pH 7.4 was prepared by adding 2 ml of 25% glutaraldehyde to 6.25 ml of 0.2 M phosphate buffer pH 7.4 and volume adjusted to 12.5 ml with distilled water.

4.2. Phosphotungstate solution 0.5%, pH 7.0

The staining solution was prepared by dissolving 50 mg of phosphotungstic acid in 10 ml of distilled water, pH adjusted to 7.0 with NaOH solution, filtered through Whatman filter paper No.1 and stored in a dark bottle at 4°C.

5. Reagents for SDS-PAGE

5.1. Stock acrylamide

Thirty grams of acrylamide (LKB) and 0.8 g of N, N'-methylene bisacrylamide (Sigma) were dissolved in 100 ml of distilled water, then filtered and stored in the dark at 4°C.

5.2. Tris-HCl buffer 1.5 M, pH 8.8

18.15 g of Tris was dissolved in 50 ml of distilled water, pH adjusted to 8.8 with HCl and final volume added to 100 ml.

5.3. Tris-HCl buffer 0.5 M, pH 6.8

Three grams of Tris was dissolved in 20 ml distilled water, pH adjusted to 6.8. The final volume was made up to 50 ml with distilled water.

5.4. EDTA 0.2 M

The 2.92 gram of EDTA was dissolved in 50 ml distilled water.

5.5. Sodium Dodecyl Sulfate (SDS), 20%

Ten grams of SDS was dissolved in 50 ml of distilled water.

5.6. N,N,N',N'-tetramethylethylenediamine (TEMED)

(Sigma)

(Sigma)

5.7. Ammonium persulfate solution, 10% (W/V)

The 0.1 g of ammonium persulfate (Sigma) was dissolved in 1 ml distilled water. This reagent was freshly prepared before use.

5.8. Electrode buffer

The buffer was prepared by dissolving 3 g of Tris, 14.4 g of glycine, and 5 ml of 20% SDS in 1000 ml of distilled water, then pH adjusted to 8.3.

5.9. Stock sample buffer (5X)

The stock solution consisted of 2.5 ml of 1.5 M Tris-HCl buffer, pH 6.8, 1 g of SDS, 5 ml of glycerol, 2.5 ml of mercaptoethanol, and 0.5 mg of bromphenol blue. The volume was adjusted to 10 ml with distilled water.

5.10. Staining solution

The staining solution was prepared by dissolving 2.5 g of Coomassie brilliant blue R (Sigma) in 500 ml methanol, 76 ml glacial acetic acid and making up the volume to 1000 ml with distilled water.

5.11. Destaining solution

5.11.1. Destaining I

One litre of destaining solution was prepared as follows : 500 ml methanol, 76 ml glacial acetic acid, and 424 ml of distilled water.

5.11.2. Destaining II

One litre of destaining solution was composed of 50 ml methanol, 75 ml glacial acetic acid, and 875 ml of distilled water.

5.12. SDS molecular weight markers

The standard molecular weight markers purchased from Sigma Chemical Company were composed of egg white lysozyme (14,300), B-lactoglobulin (18,400), Trypsinogen (24,000), Pepsin (34,700), egg albumin (45,000), and bovine plasma albumin (66,000).

6. Reagents for Rabbit Immunization

6.1. Tris-HCl buffer 0.01 M, pH 7.0

0.1211 g of Tris was dissolved in distilled water, pH adjusted to 7.0, sterilized by filtration through a sterile Millipore membrane (0.45 μ m).

6.2. Freund's complete adjuvant (Difco)

6.3. Freund's incomplete adjuvant (Difco)

7. Reagents for Immunodiffusion

7.1. Agarose 1%

0.2 g of agarose was dissolved in 20 ml normal saline solution, boiled until dissolved completely.

7.2. Goat anti rabbit whole serum (Sigma)

7.3. Goat anti rabbit Immunoglobulin G (Sigma)

8. Reagents for Indirect Hemagglutination Test (IHA)

8.1. Formalin treatment of sheep red blood cells.

8.1.1. Sheep red blood cells (SRBC)

Sheep blood was collected in an equal volume of Alsever's solution and stored for 5-10 days at 4 °C before use.

8.1.2. Normal saline solution (NSS)

Nine grams of NaCl dissolved in 1 litre of distilled water.

8.1.3. Formalin solution 7.5%

The 18.7 ml of 37% formaldehyde solution is diluted with 81.3 ml of NSS.

8.2. Tannic acid treatment

8.2.1. Phosphate buffered saline (PBS) pH 7.2

4.672 g $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, 1.53 g KH_2PO_4 , and 6.75 g NaCl dissolved in 1 litre distilled water.

8.2.2. Phosphate buffered saline (PBS) pH 6.4

4.673 g $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, 6.63 g KH_2PO_4 , and 4.5 g NaCl were dissolved in 1 litre distilled water.

8.2.3. Tannin solution (1:40,000)

The solution was freshly prepared by dissolving 2.5 mg Tannic acid (Merk) in 100 ml of PBS pH 7.2

8.3. Normal rabbit serum, 1% (1%PBS-R)

Normal rabbit serum (Gibco) was inactivated at 56 °C for 30 min and absorbed with an equal volume of packed formalin treated SRBC for 10 min at room temperature. After centrifugation, the absorbed normal rabbit serum was then diluted to 1 percent solution in PBS pH 6.4. This diluted serum was kept at -20°C until use.

9. Reagents for Coagglutination Test

9.1. Tryptic Soy Broth (TSB)

Thirty grams of Tryptic soy broth (BBL) was dissolved in 1000 ml distilled water, then swirled until completely dissolved, sterilized by autoclave at 15 lbs for 15 min.

9.2. Phosphate buffer pH 7.2

0.1925 g of KH_2PO_4 and 0.6244 g of K_2HPO_4 were

dissolved in 1 litre of distilled water.

9.3. Formalin solution, 0.5%

0.25 ml of formaldehyde solution was diluted to 20 ml with phosphate buffer pH 7.2.

9.4. PBS pH 7.2, 0.1% NaN₃

Buffer solution prepared by dissolving 1.53 g KH₂PO₄, 3.72 g Na₂HPO₄, 6.75 g NaCl, and 1 g NaN₃ in 1000 ml distilled water. This solution was sterilized by filtration.

10. Reagents for Isolation of IgG from Rabbit Antiserum

10.1. Solution A

1.36 g KH₂PO₄ was dissolved in 100 ml distilled water

10.2. Solution B

17.4 g K₂HPO₄ was dissolved in 1000 ml distilled water.

10.3. Starting buffer

Phosphate buffer, 0.01 M pH 8.0 was prepared by mixing 10 ml of solution A and 190 ml of solution B, and the solution was diluted to 2000 ml with distilled water.

10.4. Solution C

4.08 g of KH_2PO_4 was dissolved in 100 ml of distilled water.

10.5. Solution D

52.2 g of K_2HPO_4 was dissolved in 1000 ml of distilled water.

10.6. Final buffer

Phosphate buffer, 0.3 M pH 8.0 prepared by diluting 50 ml of solution C with 950 ml of solution D.

10.7. DEAE cellulose (DE-52 Whatman)

11. Reagents for Immunoelectrophoresis (IEP)

11.1. Sodium barbital-HCl 0.05 M, pH 8.2

Barbital buffer was composed of 11.3 g of barbital sodium, 0.1 g of NaN_3 , and 16.4 ml of 1N HCl, volume adjusted to 1 litre with distilled water.

11.2. Agarose, 1.5%

0.3 g agarose was dissolved in 20 ml of barbital buffer, boiled until agarose completely dissolved.

12. Reagents for Conjugation of Alkaline Phosphatase to Rabbit IgG

12.1. Alkaline phosphatase enzyme, type VII-S (Sigma), was kept at 4 °C.

12.2. Isophosphate buffer, pH 7.4

3.44 g of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ and 15.2 g of Na_2HPO_4 were dissolved in 1 litre of deionized water.

12.3. MgCl_2 solution, 1 M

2.033 g of $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ was dissolved in 10 ml deionized water.

12.4. PBS- Mg^{++}

Addition of 1 ml of 1 M MgCl_2 to 100 ml of isophosphate buffer and the volume made upto 1 liter with NSS.

12.5. Glutaraldehyde, 10 %

The solution was prepared by diluting 1 ml of 50% glutaraldehyde with 4 ml of isophosphate buffer.

12.6. Lysine 1 M, pH 7.0

0.731 g of Lysine was dissolved in 5 ml of deionized water, pH adjusted to 7.0 with 1 N NaOH.

12.7. Bovine Serum Albumin, 20 mg/ml

0.2 g of BSA was dissolved in 10 ml of deionized water.

12.8. Tris-HCl buffer 0.05 M, pH 8.0

The buffer was prepared by dissolving 0.6055 g of Tris in 100 ml of deionized water, then pH adjusted to 8.0 with 1 N HCl.

13. Reagents for Enzyme-Linked Immunosorbent Assay (ELISA)

13.1. Coating buffer

Carbonate-bicarbonate buffer, pH 9.6 consisted of 0.159 g of Na_2CO_3 , 0.293 g of NaHCO_3 , and 0.02 g of NaN_3 , made up to 100 ml with distilled water, and stored at 4 °C for not more than 2 weeks.

13.2. PBS-T

PBS-Tween pH 7.4 was composed of 8 g NaCl, 0.2 g KH_2PO_4 , 2.9 g $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$, 0.2 g KCl, 0.5 ml Tween 20, and 0.2 g NaN_3 in 1 litre of distilled water and stored at 4°C.

13.3. Sample diluent

2 g of BSA was mixed with 100 ml of PBS-T, and stored at 4°C.

13.4. Triethanolamine 0.25 M, pH 7.6

3.3 ml of triethanolamine was added to distilled water, then adjusted pH to 7.6 Made up the volume to 100 ml

13.5. Mg/Zn solution (100X)

The stock solution prepared by dissolving 10.16 g of $MgCl_2 \cdot 6H_2O$ and 0.017 g of $ZnCl_2$ in 500 ml of distilled water.

13.6. Enzyme diluent

The enzyme diluent for diluting the conjugate contained 4 g of $NaCl$, 25 ml of 0.25 M Triethanolamine pH 7.6, 5 ml of 100X Mg/Zn, 5 ml of 20 mg/ml BSA, and 0.5 g of NaN_3 , made up to 500 ml with distilled water, and stored at 4 °C.

13.7. Substrate diluent

Diethanolamine buffer, 10 % consisted of 9.7 ml of diethanolamine, 80 ml of distilled water, 20 mg of NaN_3 , and 10 mg of $MgCl_2 \cdot 6H_2O$. 1N HCl was added until the pH was 9.8, and distilled water was added to final volume of 100 ml. It was stored at 4°C in the dark.

13.8. Substrate

p-nitrophenyl phosphate (5 mg/tablet, Sigma) tablets were stored at -20°C in the dark until use. Immediately before use, one tablet was dissolved in each 5 ml of substrate diluent at room temperature, and must be used the same day.

13.9. Stopping solution

3 M NaOH was prepared by dissolving 12 g of NaOH in 100 ml of distilled water.

13.9. Stopping

NaOH in 100 ml of distilled

BIOGRAPHY

Mr. Suchart Chanama was born on May 23, 1961 in Bangkok, Thailand. He graduated with the degree of Bachelor of Science in Medical Technology, from the Faculty of Medical Technology, Mahidol University in 1984.

