

REFERENCES



THAI

- กนกพร กวีพัฒน์, ยุทธนา สมิตะสิริ และ วรรณธนา ขนนไทย 2537 ผลของสารสกัดจากพืชสมุนไพรบางชนิดต่อการสืบพันธุ์ของหนูขาวแต่ละเพศ. เสนอต่อ สถาบันวิจัยและพัฒนาวิทยาศาสตร์และเทคโนโลยี มหาวิทยาลัยเชียงใหม่ ประจำปีงบประมาณ 2536.
- เต็ม สมิตินันท์ 2523 ชื่อพฤษศาสตร์: ชื่อพื้นเมือง “ชื่อพันธุ์ไม้แห่งประเทศไทย” หจก. ฟินนี่ พับลิชชิง. กรุงเทพฯ :183-184.
- ช. นิยมธรรม 2538 ”กวางเครือ” ในอนุกรมวิธานพืช อักษร ก ฉบับ ราชบัณฑิตยสถาน. บริษัท เพื่อนพิมพ์ จำกัด. กรุงเทพฯ.
- วันชัย ดีเอโกนามมูล และ ซาลี ทองเครือ 2544 รายงานการศึกษาเรื่องสถานภาพการวิจัยและพัฒนา กวางเครือในประเทศไทยและสิ่งที่ควรดำเนินการวิจัย 18 มกราคม 2544 ณ อาคารสำนักงานพัฒนาวิทยาศาสตร์และเทคโนโลยีแห่งชาติ : 10-22.
- ทรงพล ชีวะพัฒน์, ปราณีย์ ชวลิตธำรง, สดุดี รัตนจรัสโรจน์, อัญชลี จุฑะพุทธิ และ สมเกียรติ ปัญญา มัง 2543 การศึกษาพิษกึ่งเรื้อรังของกวางเครือขาว. วารสารกรมวิทยาศาสตร์การแพทย์. (42) : 2002-2003.
- พูนศิลป์ ไวทยโชติ, กิตตินันท์ นิวาสะบุตร และ ยุทธนา สมิตะสิริ 2530 การศึกษาเบื้องต้นเกี่ยวกับกวางขาวในสุนัข. การประชุมวิชาการวิทยาศาสตร์และเทคโนโลยีแห่งประเทศไทย ครั้งที่ 13, 20-22 ตุลาคม 2530 ณ มหาวิทยาลัยสงขลานครินทร์ : 498.
- มยุรา อุยะสิทธิรัตน์, ทิพย์อักษร สิ้นชัยศรี, บุญเกตุ ฟองแก้ว และ ยุทธนา สมิตะสิริ 2530 ผลของกวางขาวต่อวงจรการเป็นสัดและรังไข่ของหนูขาว. การประชุมวิชาการมหาวิทยาลัยเกษตรศาสตร์ ครั้งที่ 25 สาขาวิทยาศาสตร์.
- ยุทธนา สมิตะสิริ 2541 ภาพรวมงานวิจัยและพัฒนา กวางเครือขาวตั้งแต่อดีต (พ.ศ.2524) ถึงปัจจุบัน (พ.ศ. 2541). ในเอกสารประกอบการสัมมนาวิชาการเรื่อง กวางเครือ. 1 ธันวาคม 2541 ณ ตึกกรมการแพทย์ กระทรวงสาธารณสุข : 13-27.

- บุษนา สมิตะสิริ, เสรี แปงจิตต์ และ สมบูรณ์ อนันตลาโภชัย 2532 การยับยั้งการให้นม
ในหนูที่กำลังให้นมด้วยกวางขาวเปรียบเทียบกับเอสโตรเจน. วารสารคณะ
วิทยาศาสตร์ มหาวิทยาลัยเชียงใหม่. (16) : 7-11.
- บุษนา สมิตะสิริ และ ศุภชัย โชติพันธุ์วิทยากุล 2540 ผลของกวางขาวต่อห้วนมและ
อวัยวะสืบพันธุ์ของลูกสุกรเพศเมีย. รายงานการวิจัยสำนักวิทยาศาสตร์
มหาวิทยาลัย เทคโนโลยีสุรนารี.
- ยุพดี ลางคลิจันทร์ 2527 การศึกษาผลของกวางขาว (*Pueraria mirifica*) ที่มีต่อ
อวัยวะสืบพันธุ์ ต่อมหมวกไต ตับ พฤติกรรมการสืบพันธุ์ในหนูขาวเพศผู้.
วิทยานิพนธ์วิทยาศาสตรมหาบัณฑิต. คณะวิทยาศาสตร์ มหาวิทยาลัยเชียงใหม่.
- ยุพดี ลางคลิจันทร์ และ บุษนา สมิตะสิริ 2528 ผลของกวางขาวต่อการสืบพันธุ์ในหนู
ขาวเพศผู้. การประชุม วทท. ครั้งที่ 11 มหาวิทยาลัยเกษตรศาสตร์.
- หลวงอนุสารสุนทร 2474 ตำรายาหัวกวางเครือ. โรงพิมพ์อุปติพงษ์. เชียงใหม่. อ้างถึงใน
บุษนา สมิตะสิริ 1998 (2541) ภาพรวมงานวิจัยและพัฒนา กวางเครือขาว
ตั้งแต่อดีต (พ.ศ. 2524) ถึงปัจจุบัน (พ.ศ. 2541). ในเอกสารประกอบการ
สัมมนาวิชาการเรื่องกวางเครือ 1 ธันวาคม 2541 ณ ตึกกรมการแพทย์ กระทรวง
สาธารณสุข : 13-27.
- อำพา เหลืองภิรมย์ และ ชุติมา หาญจวนิช 2541 การศึกษาผลของสารสกัดหัว
กวางเครือต่อน้ำหนักตัว อวัยวะสืบพันธุ์ ต่อมใต้สมอง การหลั่งน้ำนมและการ
ฟักตัวของตัวอ่อนในหนูขาวใหญ่. ภาควิชาชีววิทยา มหาวิทยาลัยขอนแก่น.

ENGLISH

- Amornratanayut W. 2006. Effects of *Curcuma comosa* Roxb. on bone in ovariectomized rats. Thesis: Master of Science Program in Pharmacology. Graduated School. Chulalongkorn University.
- Anderson R., Dart A.M., Star J., Shaw J. and Chin-Dusting J.P. 2004. Plasma C-reactive protein, but not protein S, VICAM-1, von willebrand factor or E-selection, is associated with endothelium dysfunction in coronary artery disease. *Atherosclerosis*. 172: 345-351.
- Anderson T. 1998. Assesment and treatment of endothelial dysfunction in humans. *J Am Coll Cardiol*. 34: 631-638.
- Andrew N.P., Hussian M., Dakak N. and Ouyyumi A.A. 2001. Platelet inhibitory effect of nitric oxide in the human coronary circulation: impact of endothelial dysfunction. *JACC*. 37: 510-516.
- Arnal J.F., Gourdy P., Filipe C., Laurell H., Bayard F. 2004. Characterizing the protective and deleterious vascular effects of estrogens. *Drug Discovery Today: Disease Models*. 1(3) : 213-221.
- Auttapongpaiboon W. 2002. Effects of *Pueraria mirifica* on isolated aorta in high cholesterol-fed rats and ovariectomized rabbits. Thesis: Master of Science in Pharmacology. Gradutaed School. Chulalongkorn University.
- Barchiese F., Jackson E.K. Gillespie D.J. Zacharia L.C., Fingerle J. and Dubbey R.K. 2002. Methoxyestradiol mediate estradiol-induced antimitogenesis in human aortic SMCs. *Hypertension*. 39: 874-879.
- Barnes S., Peterson G. and Coward L. 1995. Rationale for the use of genistein-containing soy matrices in chemoprevention trials for breast and prostate cancer. *J Cell Biochem*. 22: 181-187.

- Bertolini D., Nedwin G., Bringman T., Smith D. and Mundy G. 1986. Stimulation of bone resorptive and inhibition of bone formation in vitro by human necrotic factors. *Nature*. 319: 516-518.
- Biegel L.B., Flaws J.A., Hirshfield A.N., O'Connor J.C., Elliot G.S., Ladics G.S., Silbergeld E.K., Van Pelt C.S., Hurtt M.E. Cook J.C. and Fram S.R. 1998. 90-Day feeding and one-generation reproduction study in Crl:CD BR rats with 17 β -estradiol. *Toxicol Sci*. 44: 116-142.
- Binko J. and Majewski H. 1998. 17 β -estradiol reduces vasoconstriction in endothelial-denuded rat aortas through inducible NOS. *Am J Physiol*. 274: H853-H859.
- Bockman C.S., Gonzalez-Cabrera I. and Abel P.W. 1996. Alpha₂-adrenoceptor subtype causing nitric oxide-mediated vascular relaxation in rats. *J Pharmacol Exp Ther*. 278: 1235-1243.
- Bush T.L., Barret-Connor E., Cowan L.D., Crigui M.H., Wallace R.B. and Suchindran C.M. 1987. Cardiovascular mortality and non-contraceptive use of estrogen in woman: results from the Lipid Research Clinics Program Follow-Up Study. *Circulation*. 75: 1102-1109.
- Caine C.J. 1960. Miroestrol: an oestrogen from the plant *Pueraria mirifica*. *Nature*. 188: 774-777.
- Calif I. 2005. Estrogen's antioxidant power may play key role in cerebral blood vessel health. UCI study.
- Camston J. 2005. How to manage osteoporosis after the menopause. *Best Practice & Research Clinical Rheumatology*. 19(6) : 1007-1019.
- Caramori P.A. and Zago A.J. 2000. Endothelial dysfunction and coronary artery disease. *Arq Bras Cardiol*. 75: 173-181.
- Catania M.A., Crupi A., Firenzuoli F., Parisi A., Sturiale A., Squadrito F., Caputi A.P., and Calapai G. 2002. Oral administration of a soy extract improves endothelial dysfunction in ovariectomized rats. *Planta Med*. 68: 1142-1144.

- Celebrese E.J. 2001. Nitric oxide: Biphasic dose responses. *Crit Rev Toxicol.* 31: 489-501.
- Chansakaow S., Ishikawa T., Seki H., Sekine K., Okada M. and Chaichantipyuth C. 2000a. Identification of deoxymiroestrol as the actual rejuvenating principal of 'Kwao Keur', *Pueraria mirifica*. The known miroestrol may be an artifact. *J. Natural. Products.* (63) : 173-175.
- Chansakaow S., Ishikawa T., Seki H., Sekine K., Okada M., Higuchi Y., Kudo M. and Chaichantipyuth C. 2000b. Isoflavones from *Pueraria mirifica* and their estrogenic activity. *Planta. Med.* (66) : 572-575.
- Chrousos G.P. 2004. *Basic and Clinical Pharmacology.* 9th ed. edited by Katzung B.G. The McGraw-Hill Companies. Singapore : 661-692.
- Clarkson T.B., Morgan T.M. and Anthony M.S. 2001. Inhibition of postmenopausal atherosclerosis progression: a comparison of the effects of conjugated equine estrogens and soy phytoestrogens. *Baillieres Clin Endocrinol Metab.* 86: 41-47.
- Coxman V., Bowman B.M., Mecham M., Roth C.M., Miller M.A. and Miller S.C. 1996. Effects of dihydrotestosterone alone and combined with estrogen on bone mineral density, bone growth, and formation rates in ovariectomized rats. *Bone.* 19(2): 107-114.
- Darignon J. and Ganz P. 2004. Role of endothelial dysfunction in atherosclerosis. *circulation.* 109: III27-III32.
- El-Mas M.M. and Abdel-Rahman A.A. 2004. Differential modulation by estrogen of alpha 2-adrenergic and I₁-imidazoline receptor-mediated hypotension in female rats. *J Appl Physiol.* 97 : 1237-1244.
- Fanti P., Monuier-Faugere M.C., Geng Z., Morris P.E., Cohen D. and Malluche H.H. 1998. The phytoestrogen genistein reduce bone loss in short-term ovariectomized rats. *Osteoporosis International.* 8: 274-281.
- Foegh M.L. and Ramwell P.W. 2004. *Basic and clinical pharmacology.* 9th ed. edited by Katzung B.G. The McGraw-Hill Companies. Singapore : 298-312.

- Fox S.W. and Chow J.W. 1998. Nitric oxide synthase expression in bone cells. *Bone*. 23: 1-6.
- Fulton C.T. and Stallone J.N. 2002. Sexual dimorphism in prostanoid-potentiated vasoconstriction : roles of endothelium and ovarian steroids. *Am J Physiol, Heart Circ Physiol*. 283 : H2062-H2073.
- Furchgott R.F. and Zawadzki J.V. 1980. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature*. 288: 373-376.
- Garnod P., Sornay-Rendn E. and Chapuy M. 1996. Increased bone turnover in late post menopausal women is a major determinant of osteoporosis. *J Bone Miner Res*. 11: 337-349.
- Gass M., Dawson-Hughes B. 2006. Preventing osteoporosis-fractures: an overview. *The American Journal of Medicine* (119-4A) : 3s-11s.
- Gibson J.P., Newbern J.W., Kunh W.L. and Elsea J.R. 1967. Comparative chronic toxicity of three oral estrogens in rats. *Toxicol Appl Pharmacol*. 11: 489-510.
- Gsell S., Eschenhagen T., Kaspereit G., Nose M., Scholz H., Behrens O. and Wieland T. 2000. Apparent up-regulation of stimulatory G-protein α subunit in the pregnant human myometrium is mimicked by elevated smoothelin expression. *FASEB*. 14: 17-26.
- Hamosh M. and Hamosh P. 1975. The effects of estrogen on the lipoprotein lipase activity of rat adipose tissue. *The Journal of Clinical Investigation*. 55:1132-1135.
- Hao Y.J., Tang Y., Chen F.B. and Pei F.X. 2005 Different doses of nitric oxide donor prevent osteoporosis in ovariectomized rats. *Clinical Orthopaedics and Related Research*. 435: 226-231.
- Hart J.E. 1990. Endocrine pathology of estrogen: species differences. *Pharmacol Ther*. 47: 203-218.
- Hermenegildo C., Oviedo P.J. and Cano A. 2006. Cyclooxygenase regulation by estradiol on endothelium. *Curr Pharm Des*. 12: 205-215.

- Heywood R. and Wadsworth P.F. 1980. The experimental toxicology of estrogens. *Pharmacol Ther.* 8: 125-142.
- Holm P., Korsgaard N., Shalmi M., Anderson H.L., Hougaard P., Skouby S.O. and Stender S. 1997. Significant reduction of the antiatherogenic effect of estrogen by long-term inhibition of nitric oxide synthesis in cholesterol-clamped rabbits. *J Clin Invest.* 100: 821-828.
- Hostanska K., Niesslein T., Freudenstein J., Reichling J. and Saller R. 2004. *Cimicifga racemosa* extract inhibits proliferation of estrogen receptor-positive and negative human breast carcinoma cell lines by induction of apoptosis. *Breast Cancer Research Treatment.* 84: 151-160.
- Hukkanen M., Platts L.A. and Lawes T. 2003. Effects of nitric oxide donor nitroglycerin on bone mineral density in a rat model of estrogen deficiency-induced osteopenia. *Bone.* 32: 142-149.
- Hwang J., Wang J., Morazzoni R. Hodis H.N. and Sevanian A. 2003. The phytoestrogens equol increase nitric oxide availability by inhibiting superoxide production: an antioxidant mechanism for cell-mediated LDL modification. *Free Radic Biol Med.* 34(10): 1271-1282.
- Johansen J.S., Riis B.J., Delmas P.O. and Christiansen C. 1988. Plasma BGP: An indicator of spontaneous bone loss and effect of estrogen treatment in post menopausal women. *Eur J Clin Invest.* 18: 191-195.
- Kanaoka K., Kobayashi Y., Hashimoto F. 2000. A common downstream signaling activity of osteoclast survival factors that prevent nitric oxide-promoted osteoclast apoptosis. *Endocrinology.* 141: 2995-3005.
- Kapiotis S., Hermann M. and Held I. 1997. Genistein, the dietary derived angiogenesis inhibitor, prevents LDL oxidation and protects endothelial cells from damage by atherogenic LDL. *Arterioscler Thromb Vasc Biol.* 17: 2868-2874.
- Khalil R.A. 2005. Sex hormones as potential modulators of vascular function in hypertension. *Hypertension.* 46: 249-254.

- Kim H.P., Lee J.Y., Jeong J.K., Bae S.W., Lee H.H. and Jo I. 1999. Non-genomic stimulation of nitric oxide release by estrogen is mediated by estrogen receptor alpha localized in caveolae. *Biochem Biophys Res Commun.* 263: 257-262.
- Kuiper G.G., Lemmen J.G., Carlsson B., Carton J.C., Safe S.H., Saag P.T., Burg B. and Gustafsson J. 1998. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor β . *Endocrinology.* 139: 4252-4263.
- Li M. and Stallone J.N. 2005. Estrogen potentiates vasopressin-induced contraction of female rat aorta by enhancing cyclooxygenase-2 and thromboxane function. *Am J Physiol, Heart Circ Physiol.* 58 : H1542-H1550.
- Lissin L.W. and Cooke J.P. 2000. Phytoestrogens and cardiovascular health. *Journal of the American College of Cardiology* 35(6) : 1403-1410.
- Loose-Mitchell D.S., Stancel G.M. 2001. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 10th ed. edited by Hardman J.G., Limbird L.E., Gilman A.G. The McGraw-Hill Companies : 1597-1629.
- Lotke P.S. 1998. Phytoestrogens: A potential role in hormone replacement therapy. *Prim Care Update Ob/Gyns.* 5(6): 290-295.
- Lutz P.L., Laurindo F.R.M. and Chagas A.C.P. 2003. Endothelium and diseases cardiovascular. Sao Paulo: Atheneu. (2): 17-32.
- Magee A.C. 1963. Biological responses of young rats fed diets containing genistin and genistein. *J Nutr.* 80: 151-156.
- Marcus R. 2001. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 10th ed. edited by Hardman J.G., Limbird L.E., Gilman A.G. The McGraw-Hill Companies : 1715-1739.
- Maturana M.A., Irigoyen M.C. and Spritzer P.M. 2007. Menopause, estrogens, and endothelial dysfunction : current concepts. *clinics.* 62(1) : 77-86.
- McNicol A. 1992. The effects of genistein on platelet function are due to thromboxane receptor antagonism rather than inhibitor of tyrosine kinase. *Prostaglandins, Leukotrienes, Essential Fatty Acids.* 48: 379-384.

- Mercuro G., Longu G., Zoncu S. and Cherchi A. 1999. Impaired forearm blood flow and vasodilator reserve in healthy postmenopausal women. *Am Heart J.* 137: 692-697.
- Mesiano S., Katz S.L., Lee J.Y. and Jaffe R.B. 1999. Phytoestrogens alter adrenocortical function: genistein and daidzein suppress glucocorticoid and stimulate androgen production by cultured adrenal cortical cells. *J Clin. Endocrinol Metab.* 84: 2443-2448.
- Michael E.M. 2002. Genomic and nongenomic effects of estrogen in the vasculatures. *Am. J. Cardiol.* 90 (suppl) : 3F-6F.
- Moien-Afshari F., Kenyon E., Choy J.C., Battistini B., McManus B.M. and Laher I. 2003. Long-term effects of ovariectomy and estrogen replacement treatment on endothelial function in mature rats. *Maturitas.* 49: 213-223.
- Moncada S., Higgs E.A. and Vane J.R. 1977. Human arterial and venous tissues generate prostacyclin (Prostaglandin X), a potent inhibitor of platelet aggregation. *Lancet.* 1: 18-20.
- Murkies A.L., Wilcox G. and Davis S.R. 1998. Clinical review 92: Phytoestrogens. *J Clin Endocrinol Metab.* 83: 297-303.
- Najemnik C., Sinzinger H. and Kritz H. 1999. Endothelial dysfunction, atherosclerosis and diabetes. *Acta Med Austriaca.* 26(5): 148-163.
- Nakashima S., Koike T. and Nozawa Y. 1991. Genistein, a protein tyrosine kinase inhibitor, inhibits thromboxane A₂-mediated human platelet responses. *Mol Pharm.* 39: 475-480.
- Nilsson S. and Gustafsson J.A. 2000. Estrogen receptor transcription and transaction. Basic aspects of estrogen action. *Breast Cancer Res.* 2: 360-366.
- Onoe Y., Miyaura C., Ohta H., Nozawa S. and Suda T. 1997. Expression of estrogen receptor-beta in rat bone. *Endocrinology.* 138: 4509-4512.

- Pare G., Krust A., Karas R., Dupont S., Aronowitz M. and Chambom P. 2000. Estrogen receptor-alpha mediate the protective effects of estrogen against vascular injury. *Circ Res.* 90: 1087-1092.
- Rahimian R., Dube G. and Cornelis B. 1997. Estrogen and selective estrogen receptor modulator LY 117018 enhance release of nitric oxide in rat aorta. *J Pharmacol Exp Ther.* 283: 116-122.
- Ratanachamnong P., Apisariyakul A., Phivthong-ngam L. and Sanrarind Y. 2000. Antioxidative effects of *Pueraria mirifica* in cholesterol-fed rabbits. Mahidol University. 1-90.
- Riggs B.L., Wahner H.W., Seeman E., Offord K.P., Dunn W.L., Mazess R.B., Johnson K.A. and Melton L.J. 1982. Changes in bone mineral density of the proximal femur and spine with aging: Differences between the postmenopausal and senile osteoporosis syndromes. *J Clin Invest.* 70: 716-723.
- Riis B.J. 1991. Biochemical markers of bone turnover in diagnosis and assessment of therapy. *Am J Med.* 91: 64-68S.
- Ruggiero R.J. and Likis F.E. 2002. Estrogen: physiology, pharmacology, and formulations for replacement therapy. *Journal of Midwifery and Women's Health.* 47(3): 130-138.
- Samuels A., Perry M.J. and Gibson R.L. 2001. Role of endothelial nitric oxide synthase in estrogen-induced osteoporosis. *Bone.* 29: 24-29.
- Scheuer J., Malhotra A., Schaible T.F. and Capasso J. 1987. Effects of gonadectomy and hormonal replacement on the rat hearts. *Circ Res.* 61: 12-19.
- Seed M. 2002. The choice of hormone replacement therapy or statin therapy in the treatment of hyperlipidemic postmenopausal woman. *Atherosclerosis supplements* (3) : 53-63.
- Setchell K.D. 1998. Phytoestrogens: the biochemistry, physiology and implications for human health of soy isoflavones. *Am J Clin Nutr.* 68 (suppl): 333S-346S.

- Shimokawa H. 1999. Primary endothelial dysfunction: atherosclerosis. *J Mol Cell Cardiol.* 1(1): 23-37.
- Shirwaiker A., Khan S. and Malini S. 2003. Anti-osteoporotic effect of ethanol extract of *Cissus quadrangularis* Linn. on ovariectomized rat. *Journal of Ethnopharmacology.* 89: 245-250.
- Simchun C. 2006. Effects of *Curcuma comosa* on functions and pathological changes of thoracic aorta in ovariectomized rats. Thesis: Master of Science in Pharmacology. Graduated School. Chulalongkorn University.
- Simionescu M., Simionescu N. and Palade G.E. 1975. Segmental differentiations of cell junctions in the vascular endothelium. The microvasculatures. *J Cell Biol.* 67: 863-885.
- Sirtori C.R., Lovati M.R. and Manzoni C. 1995. Soy and cholesterol reduction: clinical experience. *J Nutr.* 125: 598S-605S.
- Staren E.D. and Omer S. 2004. Hormone replacement therapy in postmenopausal woman. *The American Journal of Surgery* (188) : 136-149.
- Subbiah M.R. 2002. Estrogen replacement therapy and cardioprotection mechanisms and controversies. *Braz J Med Biol Res.* 35: 272-276.
- Suwaidi I.A., Hamasaki S., Higanu S., Nishimura R.A., Holmes D.R. Jr. and Lerman A. 2000. Long-term follow-up of patients with mild coronary artery disease and endothelial dysfunction. *Circulation.* 101: 948-954.
- Taddei S., Virdis A., Ghiadoni L., Mattei P., Sudano I. and Bernini G. 1996. Menopause is associated with endothelial dysfunction in women. *Hypertension.* 28: 575-582.
- Takahashi M., Ikeda U. and Masuyama J.I. 1996. Monocyte-endothelial cell interaction induces expression of adhesion molecules on human umbilical cord endothelial cells. *Cardiovasc Res.* 32: 422-429.
- Thomas G. and Ramwell P.W. 2004. Basic and clinical pharmacology. 9th ed. edited by Katzung B.G. The McGraw-Hill Companies. Singapore : 313-318.

- Tikkanen M.J. and Adlercreutz H. 2000. Dietary soy-derived isoflavones phytoestrogens. Could they have a role in coronary heart disease prevention? *Biochem Pharmacol.* 60: 1-5.
- Tikkanen M.J., Wahala K., Ojala S., Vihma V. and Adlercreutz H. 1998. Effects of soybean phytoestrogens intake on low density lipoprotein oxidation resistance. *Proc Natl Acad Sci.* 95: 3160-3110.
- Tilley L.P. and Smith F.W.K., 2000. *The 5-minute veterinary consult.* 2nd ed. Lippincott Williams and Wilkins. Baltimore, Maryland, USA. : 222.
- Toda T., Uesugi T., Hirai K., Nukaya H., Tsuji K. and Ishida H. 1999. New 6-O-acyl isoflavone glycosides from soybeans fermented with *Bacillus subtilis* (natto). I. 6-O succinylated isoflavone glycosides and their preventive effects on bone loss in ovariectomized rats fed a calcium-deficient diet. *Biol Pharm Bull.* 22:1193-1201.
- Tolbert T. and Oparil S. 2001. Cardiovascular effects of estrogen. *The American Journal of Hypertension* (14) : 1865-1935.
- Tsikas D. 2006. Analysis of nitrite and nitrate in biological fluids by assays based on the Griess reaction: Appraisal of the Griess reaction in the L-arginine/ nitric oxide area of research. *Journal of Chromatography B.* 10(1016) : 1-20.
- Ushiyama Y., Higuchi Y., Takeda S., Masaki T., Shira-Ishi A., Sato K., Kubodera N., Ikeda K. and Ogata E. 2002. ED-71, a vitamin D analog, is a more potent inhibitor of bone resorption than alfacalcidol in an estrogen-deficit rat model of osteoporosis. *Bone.* 30(4): 582-588.
- Usui H., Kurahashi K., Shirahase H., Fukui K. and Fujiwara M. 1987. Endothelium-dependent vasocontraction in response to noradrenaline in the canine cerebral artery. *Jpn J Pharmacol.* 44: 228-231.
- van't Hof R.J., Armour K.J. and Smith L.M. 2000. Requirement of the inducible nitric oxide synthase pathway for IL-1-induced osteoclastic bone resorption. *Proc Natl Acad Sci USA.* 97: 7993-7998.

- Virginia M.M. and Vanhoutte P.M. 1990. 17β -estradiol augments endothelium-dependent contractions to arachidonic acid in rabbit aorta. *Am J Physiol.* 258: R1502-R1507.
- Wakatsuki A., Ikenoue N., Okatani Y. and Fukaya T. 2001. Estrogen-induced small low density lipoprotein particles may be atherogenic in post-menopausal women. *J Am Coll Cardiol.* 37(2): 425-430.
- Wakatsuki A., Ikenoue N. and Sagara Y. 1998. Estrogen-induced small low density lipoprotein particles in post-menopausal women. *Obstet gynecol.* 91(2): 234-240.
- Walsh B.W., Schiff I. and Rosner B. 1991. Effects of postmenopausal estrogen replacement on the concentrations and metabolism of plasma lipoproteins. *N Eng J Med.* 325: 1196-1204.
- Wang A., Nishihashi T., Trandafir C.C., Murakami S., Ji X., Shimizu Y. and Kurahashi K. 2005. Involvement of endothelial cyclo-oxygenase metabolites in noradrenaline-induced contraction of rat coronary artery. *Clinical and Experimental Pharmacology and Physiology.* 32: 628-632.
- Wattanapitayakul S.H., Chularojmontri L. and Srichairat S. 2005. Effects of *Pueraria mirifica* on vascular function of ovariectomized rabbits. *J Med Assoc Thai.* 88 (suppl 1): S21-S29.
- Wimalawansa S.J. 2000a. Restoration of ovariectomy-induced osteoporosis by nitroglycerin. *Calcif Tissue Int.* 66: 56-60.
- Wimalawansa S.J. 2000b. Nitroglycerine therapy is as efficacious as standard estrogen replacement therapy (Premarin) in prevention of oophorectomy-induced bone loss: A human pilot clinical study. *J Bone Miner Res.* 15: 2240-2244.
- Wiseman H. and Duffy R. 2001. New advances in the understanding of the role of steroids and steroid receptors in disease. *Biochem Soc Trans.* 29: 205-208.

- Yang J., Farnell D., Devlin H., Horner K. and Graham J. 2005. The effect of ovariectomy on mandibular cortical thickness in the rat. *Journal of Dentistry*. 33: 123-129.
- Yang Y. and Loscalzo J. 2000. Regulation of tissue factor expression in human microvascular endothelial cells by nitric oxide. *Circulation*. 101: 2144-2148.
- Zhu W., Diwan A.D. and Lin J.H. 2001. Nitric oxide synthase isoforms during fracture healing. *Bone Miner Res*. 16: 535-540.

APPENDICES

Appendix 1

Appendix 1.1 The percentages increase of body weight compared with body weight at the beginning of the study (week 0) in OVX + *P. mirifica* group.

Code	Time period (weeks)						
	0	1	2	3	4	5	6
N1	100.00	100.7692	101.9231	103.0769	103.8462	104.6154	105.7692
N2	100.00	101.1538	103.0769	103.8462	105.00	105.7692	107.6923
N3	100.00	100.7143	101.7857	103.5714	104.2857	105.3571	107.1429
N4	100.00	100.7692	102.6923	103.4615	104.2308	105.00	105.7692
N5	100.00	101.0345	101.7241	102.7586	103.4483	104.4828	105.1724
N6	100.00	100.6897	101.7241	102.7586	104.1379	105.8621	106.8966
N7	100.00	100.8889	102.2222	102.6667	103.1111	103.5556	104.4444
N8	100.00	100.8696	102.1739	103.4783	104.3478	105.6522	106.5217
Mean	100.00	100.8611	102.1653	103.2023	104.051	105.0368	106.1761
S.E.M.	0.00	0.06	0.17	0.16	0.21	0.28	0.38

Appendix 1.2 The percentages increase of body weight compared with body weight at the beginning of the study (week 0) in OVX + Estrogen group.

Code	Time period (weeks)						
	0	1	2	3	4	5	6
N1	100.00	100.6897	101.7241	102.7586	103.4483	104.1379	105.1724
N2	100.00	100.7843	101.9608	102.7451	103.9216	105.098	105.8824
N3	100.00	100.7143	101.7857	102.8571	103.5714	104.6429	105.3571
N4	100.00	101.0638	102.1277	102.8369	103.9007	105.6738	106.383
N5	100.00	101.0204	102.0408	102.7211	103.7415	104.7619	105.4422
N6	100.00	101.0638	102.1277	103.5461	104.6099	105.6738	106.383
N7	100.00	100.4167	101.25	102.0833	102.9167	103.3333	104.1667
N8	100.00	100.40	101.20	102.00	102.80	103.20	104.00
Mean	100.00	100.7691	101.7771	102.6935	103.6138	104.5652	105.3483
S.E.M.	0.00	0.10	0.13	0.17	0.21	0.34	0.32

Appendix 1.3 The percentages increase of body weight compared with body weight at the beginning of the study (week 0) in OVX group.

Code	Time period (weeks)						
	0	1	2	3	4	5	6
N1	100.00	109.6154	111.5385	111.5385	113.4615	113.4615	115.3846
N2	100.00	108.00	111.20	112.00	114.00	115.20	116.00
N3	100.00	103.0769	106.9231	107.6923	108.4615	109.6154	111.5385
N4	100.00	103.5714	107.1429	107.8571	108.9286	110.7143	114.2857
N5	100.00	101.5385	104.6154	106.9231	108.4615	109.6154	111.5385
N6	100.00	101.5385	104.6154	106.9231	108.4615	109.6154	111.5385
N7	100.00	102.0833	106.25	110.4167	114.5833	116.6667	120.8333
N8	100.00	102.6415	103.7736	105.6604	113.2075	116.9811	120.7547
Mean	100.00	104.0082	107.0073	108.6264	111.1957	112.7337	115.2342
S.E.M.	0.00	1.09	1.04	0.84	1.00	1.14	1.36

Appendix 1.4 The percentages increase of body weight compared with body weight at the beginning of the study (week 0) in Sham group.

Code	Time period (weeks)						
	0	1	2	3	4	5	6
N1	100.00	100.7547	101.8868	103.3962	104.9057	106.4151	107.5472
N2	100.00	101.1765	102.7451	103.9216	105.8824	107.8431	109.8039
N3	100.00	101.1765	102.7451	104.7059	107.0588	108.6275	109.8039
N4	100.00	101.9608	103.1373	105.098	105.8824	107.8431	109.8039
N5	100.00	101.7391	102.6087	103.4783	105.2174	106.5217	109.5652
N6	100.00	101.2766	102.1277	104.2553	104.2553	105.5319	106.383
N7	100.00	101.3636	103.1818	104.5455	105.9091	106.8182	109.0909
N8	100.00	100.9756	102.439	103.4146	105.8537	107.3171	109.7561
Mean	100.00	101.3029	102.6089	104.1019	105.6206	107.1147	108.9693
S.E.M.	0.00	0.14	0.16	0.23	0.30	0.35	0.46

Appendix 2

Appendix 2.1 Plasma nitric oxide (NO) level of each group in the experiments.

Group	OVX+ <i>P. mirifica</i>	OVX+Estrogen	OVX	Sham
N1	3.678899	2.669725	2.394495	3.082569
N2	3.266055	2.577982	2.394495	3.036697
N3	3.311927	2.394495	2.211009	2.669725
N4	2.761468	2.944954	2.302752	2.807339
N5	2.669725	2.669725	2.165138	3.220183
N6	2.944954	2.853211	2.394495	2.394495
N7	2.807339	2.899083	2.256881	2.623853
N8	3.633028	2.669725	2.256881	2.990826
Mean	3.134174	2.709862385	2.297018349	2.853211009
S.E.M.	0.14	0.06	0.03	0.10

Appendix 2.2 Plasma alkaline phosphatase (ALP) level of each group in the experiments.

Group	OVX+ <i>P. mirifica</i>	OVX+Estrogen	OVX	Sham
N1	60.8	48.6	145	48.2
N2	10.2	95	108	89.8
N3	84.2	57.6	82.8	64.8
N4	191	180	135	88.8
N5	153	70	182	113
N6	95.3	93.4	118	84.3
N7	102	104	174	89
N8	103	76.6	124	63.1
Mean	99.9375	90.65	133.6	80.125
S.E.M.	19.35356	14.43734	11.7094	7.18348

Appendix 2.3.1 Plasma level of lipid parameters (total cholesterol, HDL-cholesterol, LDL-cholesterol, triglyceride and atherosclerotic index [total cholesterol / HDL-cholesterol ratio]) in OVX + *P. mirifica* group.

Code	Plasma lipid parameters (U)				
	Total-cholesterol	HDL-cholesterol	LDL-cholesterol	Triglyceride	Atherosclerotic index
N1	79	44	22.6	62	0.795455
N2	71	55	24.2	43	0.290909
N3	94	44	18.4	24	1.136364
N4	77	49	33	45	0.571429
N5	72	52	28	30	0.384615
N6	85	44	30	95	0.931818
N7	71	55	28	66	0.290909
N8	81	48	33	69	0.6875
Mean	78.75	48.875	27.15	54.25	0.636125
S.E.M.	2.82	1.67	1.81	8.23	0.1095

Appendix 2.3.2 Plasma level of lipid parameters (total cholesterol, HDL-cholesterol, LDL-cholesterol, triglyceride and atherosclerotic index [total cholesterol / HDL-cholesterol ratio]) in OVX + Estrogen group.

Code	Plasma lipid parameters (U)				
	Total-cholesterol	HDL-cholesterol	LDL-cholesterol	Triglyceride	Atherosclerotic index
N1	86	53	33	104	0.622642
N2	74	50	29	110	0.48
N3	87	51	37	177	0.705882
N4	82	63	31	189	0.301587
N5	97	55	42	112	0.763636
N6	94	58	36	103	0.62069
N7	68	45	23	153	0.511111
N8	91	52	39	136	0.75
Mean	84.88	53.38	33.75	135.50	0.594444
S.E.M.	3.50	1.92	2.14	12.06	0.05551

Appendix 2.3.3 Plasma level of lipid parameters (total cholesterol, HDL-cholesterol, LDL-cholesterol, triglyceride and atherosclerotic index [total cholesterol / HDL-cholesterol ratio]) in OVX group.

Code	Plasma lipid parameters (U)				
	Total-cholesterol	HDL-cholesterol	LDL-cholesterol	Triglyceride	Atherosclerotic index
N1	84	48	28.6	37	0.75
N2	86	44	29	65	0.954545
N3	67	42	6.2	69	0.595238
N4	61	41	18.2	69	0.487805
N5	90	49	24.4	83	0.836735
N6	92	52	23.2	84	0.769231
N7	64	42	34.8	51	0.52381
N8	79	47	18.2	83	0.680851
Mean	77.88	45.63	22.83	67.63	0.699777
S.E.M.	4.32	1.40	3.10	5.92	0.05641

Appendix 2.3.4 Plasma level of lipid parameters (total cholesterol, HDL-cholesterol, LDL-cholesterol, triglyceride and atherosclerotic index [total cholesterol / HDL-cholesterol ratio]) in Sham group.

Code	Plasma lipid parameters (U)				
	Total-cholesterol	HDL-cholesterol	LDL-cholesterol	Triglyceride	Atherosclerotic index
N1	56	34	15.2	34	0.647059
N2	56	38	9.4	78	0.473684
N3	63	40	8.2	75	0.575
N4	65	43	10	67	0.511628
N5	74	48	17.6	70	0.541667
N6	72	58	10	46	0.241379
N7	72	45	14.2	64	0.6
N8	67	41	21.2	24	0.634146
Mean	65.63	43.38	13.23	57.25	0.52807
S.E.M.	2.49	2.58	1.63	7.10	0.04598

Appendix 3

Appendix 3.1

Appendix 3.1.1 The percentages of aortic contraction due to noradrenaline (NA)

induction in isolated rat thoracic aortas in OVX + *P. mirifica* group.

Code	Percentages of aortic contraction					
	NA 10 ⁻⁹	NA 10 ⁻⁸	NA 10 ⁻⁷	NA 10 ⁻⁶	NA 10 ⁻⁵	NA 10 ⁻⁴
N1	10.19	15.47	20.98	48.45	100	91.92
N2	1.23	7.58	26.37	62.3	100	97.39
N3	4.22	7.54	26.65	58.7	100	92.6
N4	3.8	4.05	31.64	71.04	100	87.2
N5	0.48	5.43	33.69	93.41	100	93.41
N6	4.63	5.21	25.35	77.08	100	93.4
N7	12.82	14.9	47.39	85.54	100	70.34
N8	2.79	5.13	46.31	91.11	100	92.99
Mean	5.02	8.16	32.30	73.45	100.00	89.91
S.E.M.	1.52	1.59	3.46	5.74	0.00	2.96

Appendix 3.1.2 The percentages of aortic contraction due to noradrenaline (NA)

induction in isolated rat thoracic aortas in OVX + Estrogen group.

Code	Percentages of aortic contraction					
	NA 10 ⁻⁹	NA 10 ⁻⁸	NA 10 ⁻⁷	NA 10 ⁻⁶	NA 10 ⁻⁵	NA 10 ⁻⁴
N1	8.58	12.13	25.73	46.99	100	86.94
N2	15.34	17.24	51.02	87.27	100	98.14
N3	5.37	9.22	45.92	81.29	100	94.82
N4	2.66	3.42	30.08	79.13	100	99.81
N5	16.87	23.16	32.61	65.8	100	94.56
N6	3.7	11.1	22.83	57.98	100	73.84
N7	2.29	12.13	24.03	55.38	100	82.15
N8	3.57	5.46	21.7	68.2	100	89.95
Mean	7.30	11.73	31.74	67.75	100.00	90.03
S.E.M.	2.05	2.22	3.90	4.96	0.00	3.10

Appendix 3.1.3 The percentages of aortic contraction due to noradrenaline (NA) induction in isolated rat thoracic aortas in OVX group.

Code	Percentages of aortic contraction					
	NA 10 ⁻⁹	NA 10 ⁻⁸	NA 10 ⁻⁷	NA 10 ⁻⁶	NA 10 ⁻⁵	NA 10 ⁻⁴
N1	3.16	10.74	39.49	73.7	100	79.46
N2	3.22	6.51	50.12	70.2	100	59.35
N3	9.78	35.61	52.54	66.08	100	85.8
N4	0.31	4.97	50.33	86.49	100	80.61
N5	1.33	5.29	37.13	80.47	100	86.82
N6	2.36	4.46	11.55	52.23	100	77.56
N7	1.07	5.16	11.58	46.01	100	72.67
N8	7.28	10.71	19.33	45.79	100	76.29
Mean	3.56	10.43	34.01	65.12	100.00	77.32
S.E.M.	1.16	3.71	6.17	5.51	0.00	3.06

Appendix 3.1.4 The percentages of aortic contraction due to noradrenaline (NA) induction in isolated rat thoracic aortas in Sham group.

Code	Percentages of aortic contraction					
	NA 10 ⁻⁹	NA 10 ⁻⁸	NA 10 ⁻⁷	NA 10 ⁻⁶	NA 10 ⁻⁵	NA 10 ⁻⁴
N1	21.94	49.85	66.19	84.87	100	94.48
N2	3.51	7.63	51.29	87.87	100	81.71
N3	3.99	10.1	33.68	80.14	100	90.12
N4	5.56	9.1	32.43	73.04	100	88.8
N5	6.77	14.41	23.61	66.32	100	88.54
N6	3.87	7.65	37.99	80.74	100	96.39
N7	5.26	12.8	41.78	76.95	100	96.23
N8	5.37	9.22	45.92	81.29	100	94.82
Mean	7.03	15.10	41.61	78.90	100.00	91.39
S.E.M.	2.16	5.04	4.63	2.40	0.00	1.79

Appendix 3.2

Appendix 3.2.1 The percentages of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX + *P. mirifica* group.

Code	Percentages of aortic relaxation					
	Ach 10 ⁻⁹	Ach 10 ⁻⁸	Ach 10 ⁻⁷	Ach 10 ⁻⁶	Ach 10 ⁻⁵	Ach 10 ⁻⁴
N1	6.61	18.53	30.75	54.74	70.98	78.59
N2	15.94	37.05	44.02	68.13	115.54	145.22
N3	42.57	76.15	123.85	170.74	185.68	191.02
N4	31.95	80.78	109.09	184.94	204.16	234.03
N5	2.06	17.53	93.81	202.27	257.84	311.34
N6	9.36	20.62	34.06	52.34	64.12	67.24
N7	28.08	66.25	76.97	140.38	150.16	186.75
N8	7.82	37.43	61.45	86.59	118.99	151.96
Mean	18.05	44.29	71.75	120.02	145.93	170.77
S.E.M.	5.12	9.32	12.39	21.80	23.71	28.24

Appendix 3.2.2 The percentages of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX + Estrogen group.

Code	Percentages of aortic relaxation					
	Ach 10 ⁻⁹	Ach 10 ⁻⁸	Ach 10 ⁻⁷	Ach 10 ⁻⁶	Ach 10 ⁻⁵	Ach 10 ⁻⁴
N1	2.21	7.46	19.26	40.49	56.48	61.89
N2	39.75	56.78	117.98	161.2	176.66	218.61
N3	32.28	62.17	83.33	135.45	187.57	220.11
N4	19.52	33.92	65.08	85.86	112.52	128.66
N5	14.32	38.01	53.73	89.33	103.14	130.51
N6	28.1	40.51	75.39	93.65	138.14	158.04
N7	11.12	17.71	34.8	68.35	104.59	120.03
N8	18.57	57.03	131.3	257.29	303.71	372.81
Mean	20.73	39.20	72.61	116.45	147.85	176.33
S.E.M.	4.29	6.89	13.57	24.08	26.81	33.62

Appendix 3.2.3 The percentages of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX group.

Code	Percentages of aortic relaxation					
	Ach 10^{-9}	Ach 10^{-8}	Ach 10^{-7}	Ach 10^{-6}	Ach 10^{-5}	Ach 10^{-4}
N1	28.03	55.3	60.61	89.39	102.27	116.67
N2	2.65	15.42	24.14	42.21	59.5	78.5
N3	2.25	37.08	55.06	94.38	135.96	125.84
N4	7.46	26.87	29.35	39.8	49.75	62.19
N5	10.26	32.48	61.82	83.76	94.59	112.54
N6	2.31	8.33	21.53	35.65	49.77	77.31
N7	13.01	21.11	29	65.88	74.41	117.27
N8	20.51	31.74	62.61	83.98	105.35	134.07
Mean	10.81	28.54	43.01	66.88	83.95	103.05
S.E.M.	3.32	5.09	6.54	8.62	10.89	9.35

Appendix 3.2.4 The percentages of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in Sham group.

Code	Percentages of aortic relaxation					
	Ach 10^{-9}	Ach 10^{-8}	Ach 10^{-7}	Ach 10^{-6}	Ach 10^{-5}	Ach 10^{-4}
N1	86.58	159.73	214.77	362.42	496.64	671.14
N2	11	21.36	53.07	100.65	164.53	172.69
N3	15.34	26.69	40.18	66.26	97.24	167.79
N4	23.86	44.57	73.14	97.14	103.57	130.29
N5	24.24	56.95	94.24	132.02	165.07	192.61
N6	10.6	27.22	53.01	97.71	118.34	139.54
N7	15.11	27.46	56.68	82.12	98.99	113.6
N8	32.28	62.17	83.33	135.45	187.57	220.11
Mean	27.38	53.27	83.55	134.22	178.99	225.97
S.E.M.	8.86	16.13	19.78	33.61	47.02	64.75

Appendix 3.3

Appendix 3.3.1 The percentages of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX + *P. mirifica* group.

Code	Percentages of aortic relaxation					
	SNP 10 ⁻⁹	SNP 10 ⁻⁸	SNP 10 ⁻⁷	SNP 10 ⁻⁶	SNP 10 ⁻⁵	SNP 10 ⁻⁴
N1	17.52	76.62	108.91	122.67	132.9	148.31
N2	33.9	108.83	152.65	165.03	170.65	174.87
N3	19.21	92.44	185.22	225.96	229.77	232.89
N4	16.89	37.87	111.31	207.89	267.48	270.93
N5	13.96	77.58	129.5	135.59	139.43	141.88
N6	4.89	47.94	152.66	199.42	205.49	208.69
N7	10.16	57.71	107.02	108.21	123.22	127.24
N8	27.1	63.87	157.42	243.87	276.13	298.71
Mean	17.95	70.36	138.09	176.08	193.13	200.44
S.E.M.	3.24	8.26	10.02	17.86	21.47	22.32

Appendix 3.3.2 The percentages of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX + Estrogen group.

Code	Percentages of aortic relaxation					
	SNP 10 ⁻⁹	SNP 10 ⁻⁸	SNP 10 ⁻⁷	SNP 10 ⁻⁶	SNP 10 ⁻⁵	SNP 10 ⁻⁴
N1	29.27	98.13	173.24	205.81	211.43	220.73
N2	11.87	33.52	78.68	102.93	106.5	112.89
N3	28.21	44.6	136.97	175.4	183.03	207.45
N4	24.88	75.47	98.52	125.27	132.33	135.81
N5	27.87	85.15	119.65	147.05	153.62	157.85
N6	34.01	68.02	211.02	331.74	360.58	368.63
N7	27.72	62.62	175.67	221.85	231.98	247.18
N8	55.92	100.32	223.73	267.57	271.29	285.09
Mean	29.97	70.98	152.19	197.20	206.34	216.96
S.E.M.	4.34	8.45	18.49	26.99	29.11	29.75

Appendix 3.3.3 The percentages of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX group.

Code	Percentages of aortic relaxation					
	SNP 10 ⁻⁹	SNP 10 ⁻⁸	SNP 10 ⁻⁷	SNP 10 ⁻⁶	SNP 10 ⁻⁵	SNP 10 ⁻⁴
N1	35.45	49.09	70.91	89.09	93.64	103.73
N2	37.53	105.3	161.74	177.75	184.57	187.75
N3	45.1	70.59	172.55	211.76	243.14	282.35
N4	3.22	39.48	53.18	73.33	76.55	93.47
N5	26.79	119.67	197.5	233.1	245.89	255.19
N6	18.01	49.84	57.23	133.7	163.76	177.07
N7	18.97	36.32	88.82	183.68	199.12	216.47
N8	36.03	54.71	123.25	154.83	160.52	166.52
Mean	27.64	65.63	115.65	157.15	170.90	185.32
S.E.M.	4.82	10.94	19.89	19.86	21.90	23.45

Appendix 3.3.4 The percentages of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in Sham group.

Code	Percentages of aortic relaxation					
	SNP 10 ⁻⁹	SNP 10 ⁻⁸	SNP 10 ⁻⁷	SNP 10 ⁻⁶	SNP 10 ⁻⁵	SNP 10 ⁻⁴
N1	38.46	61.54	71.15	98.08	105.77	135.77
N2	37.62	156.81	212.29	223.9	228.24	232.19
N3	36.5	156.39	213.26	224.59	230.6	233.77
N4	34.9	110.47	247.31	300.17	310.47	310.62
N5	32.22	63.01	108.59	179.76	195.99	214.49
N6	33.26	63.39	85.56	138.08	148.74	173.64
N7	19.47	41.53	76.02	117.55	149.69	168.47
N8	28.21	44.6	136.97	175.4	183.03	207.45
Mean	32.58	87.22	143.90	182.19	194.07	209.55
S.E.M.	2.20	16.84	24.92	23.42	22.36	18.79

Appendix 4

Appendix 4.1 The relative bone weight of each group in the experiments.

Group	OVX+ <i>P. mirifica</i>	OVX+Estrogen	OVX	Sham
N1	0.230461538	0.217018868	0.205283599	0.236343576
N2	0.221296296	0.232936803	0.193109004	0.230269316
N3	0.191254237	0.23125	0.19655143	0.232527307
N4	0.212785714	0.213357143	0.215636286	0.186241091
N5	0.209864407	0.201035714	0.199549672	0.248654869
N6	0.21052459	0.210357143	0.194161165	0.230796784
N7	0.232979592	0.222533333	0.177484652	0.24404874
N8	0.227041667	0.202137661	0.209352258	0.219276596
Mean	0.217026005	0.216328333	0.198891008	0.228519785
S.E.M.	0.00487	0.00426	0.00412	0.00682

Appendix 4.2 The relative bone ash of each group in the experiments.

Group	OVX+ <i>P. mirifica</i>	OVX+Estrogen	OVX	Sham
N1	0.142076923	0.138566038	0.127762712	0.147535714
N2	0.131777778	0.146951673	0.120701754	0.144333333
N3	0.114237288	0.13475	0.123649123	0.146290909
N4	0.129333333	0.132571429	0.123508772	0.115186441
N5	0.130857143	0.137678571	0.120034483	0.155176471
N6	0.135762712	0.141333333	0.121533333	0.140074074
N7	0.139114754	0.122035088	0.110258065	0.151773585
N8	0.134204082	0.127703704	0.1301	0.134893617
Mean	0.132170502	0.135198729	0.12219353	0.141908018
S.E.M.	0.00297	0.00277	0.0021	0.00443

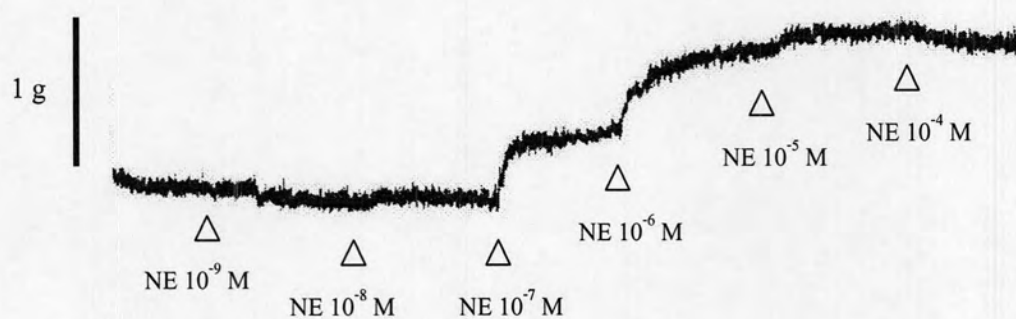
Appendix 4.3 The bone calcium level (mg/g Ash) of each group in the experiments.

Group	OVX+<i>P. mirifica</i>	OVX+Estrogen	OVX	Sham
N1	299.0660531	295.0571895	305.5187052	297.107238
N2	323.608769	283.7718189	311.1918605	297.4980754
N3	332.7299703	311.5247742	310.4426788	295.7742978
N4	322.0504009	354.7309833	291.1184211	320.9093584
N5	291.8941048	298.6530172	307.4573864	318.0439727
N6	303.9409617	299.1699092	315.6133295	288.6171338
N7	291.6354557	290.3301887	294.5693911	294.2317255
N8	283.5493754	317.7544566	301.9748391	343.8643533
Mean	306.0593864	306.3740422	304.7358264	307.0057694
S.E.M.	6.32994	7.90828	2.9791	6.66755

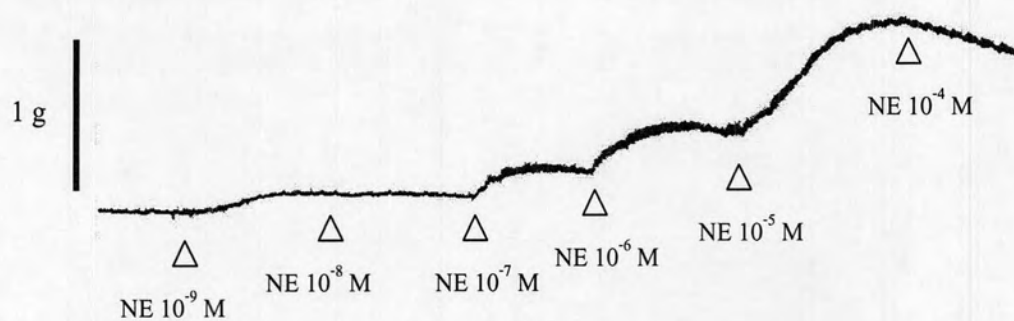
Appendix 5

Appendix 5.1

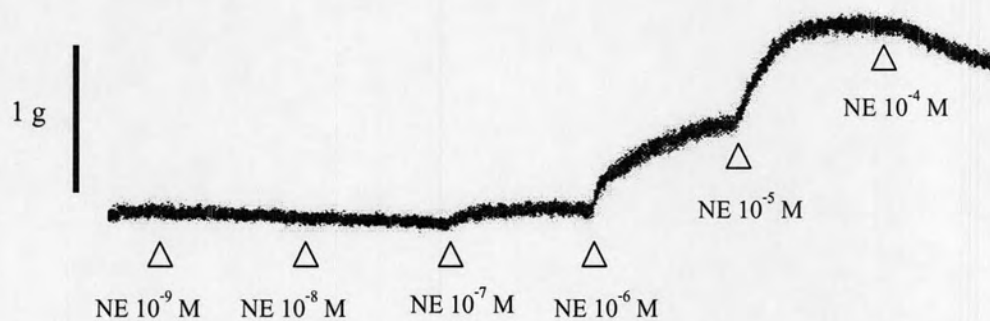
Appendix 5.1.1 The isometric tension changes due to noradrenaline (NA) induction in isolated rat thoracic aortas in OVX + *P. mirifica* group.



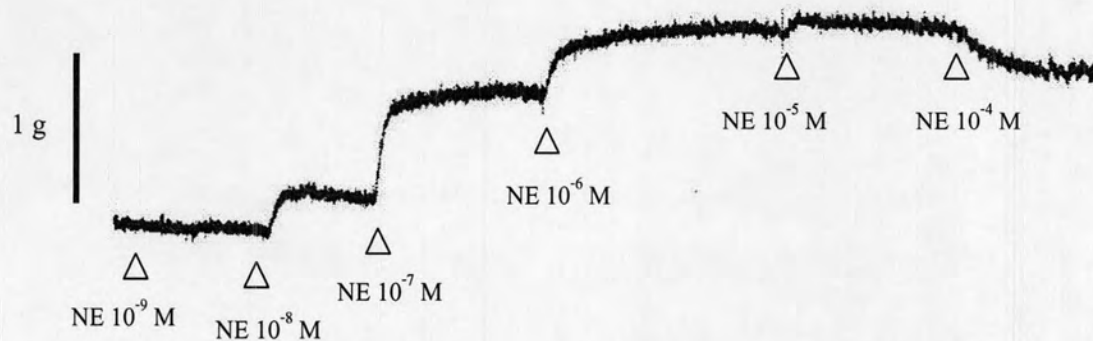
Appendix 5.1.2 The isometric tension changes due to noradrenaline (NA) induction in isolated rat thoracic aortas in OVX + Estrogen group.



Appendix 5.1.3 The isometric tension changes due to noradrenaline (NA) induction in isolated rat thoracic aortas in OVX group.

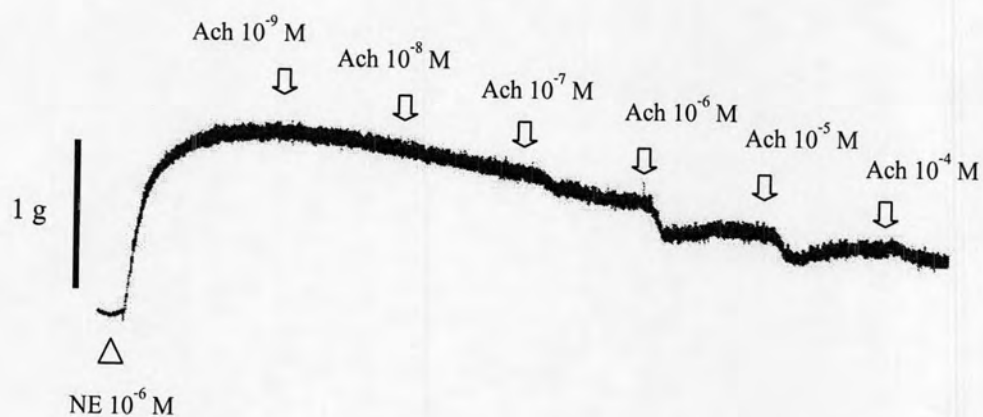


Appendix 5.1.4 The isometric tension changes due to noradrenaline (NA) induction in isolated rat thoracic aortas in Sham group.

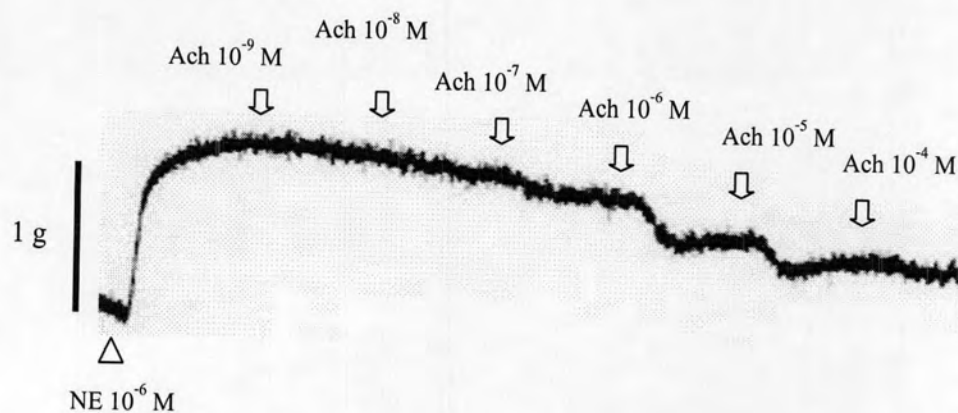


Appendix 5.2

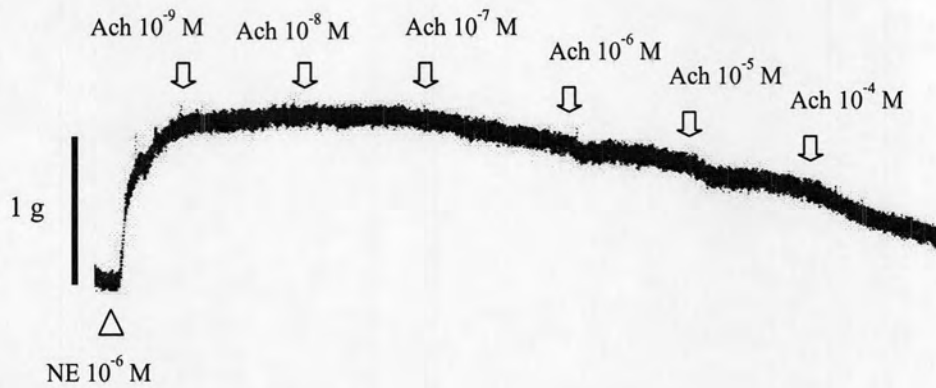
Appendix 5.2.1 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX + *P. mirifica* group.



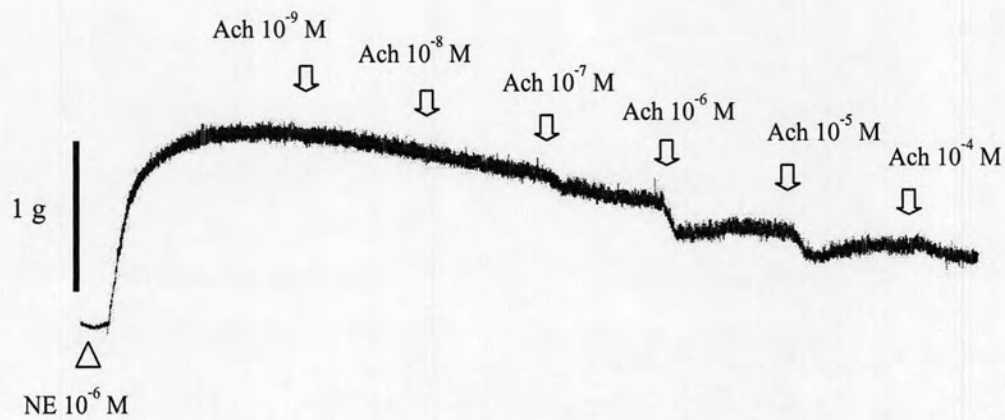
Appendix 5.2.2 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX + Estrogen group.



Appendix 5.2.3 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in OVX group.

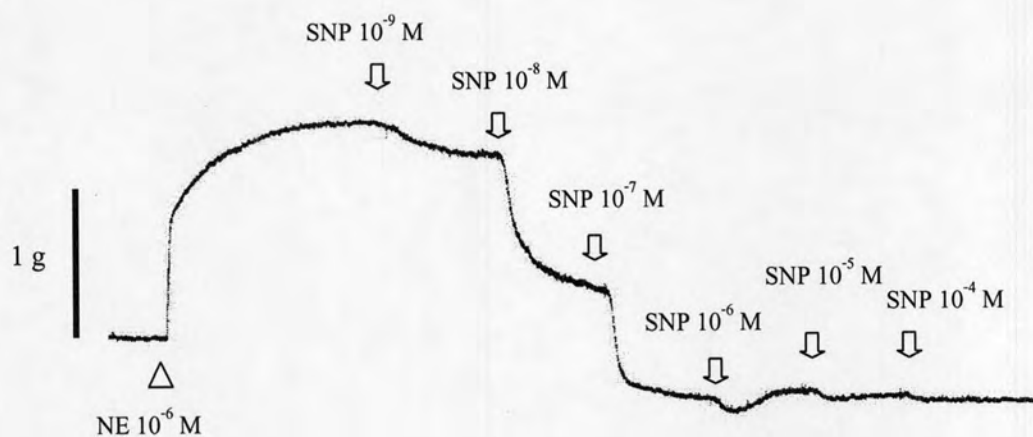


Appendix 5.2.4 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to acetylcholine (Ach) stimulation in isolated rat thoracic aortas in Sham group.

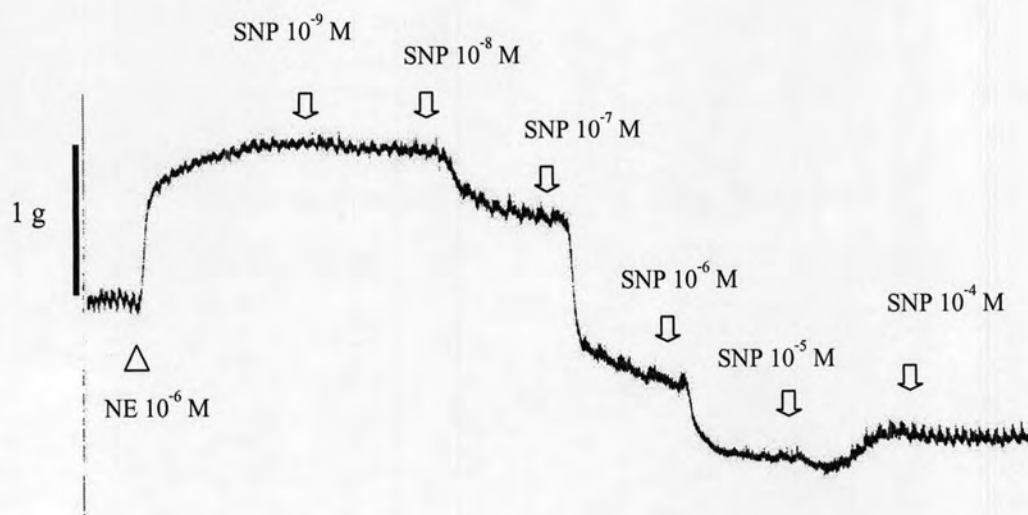


Appendix 5.3

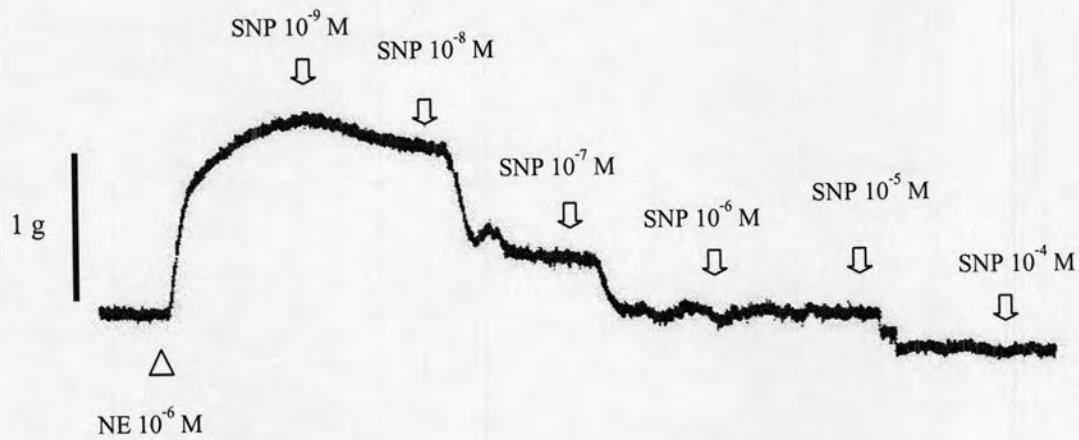
Appendix 5.3.1 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX + *P. mirifica* group.



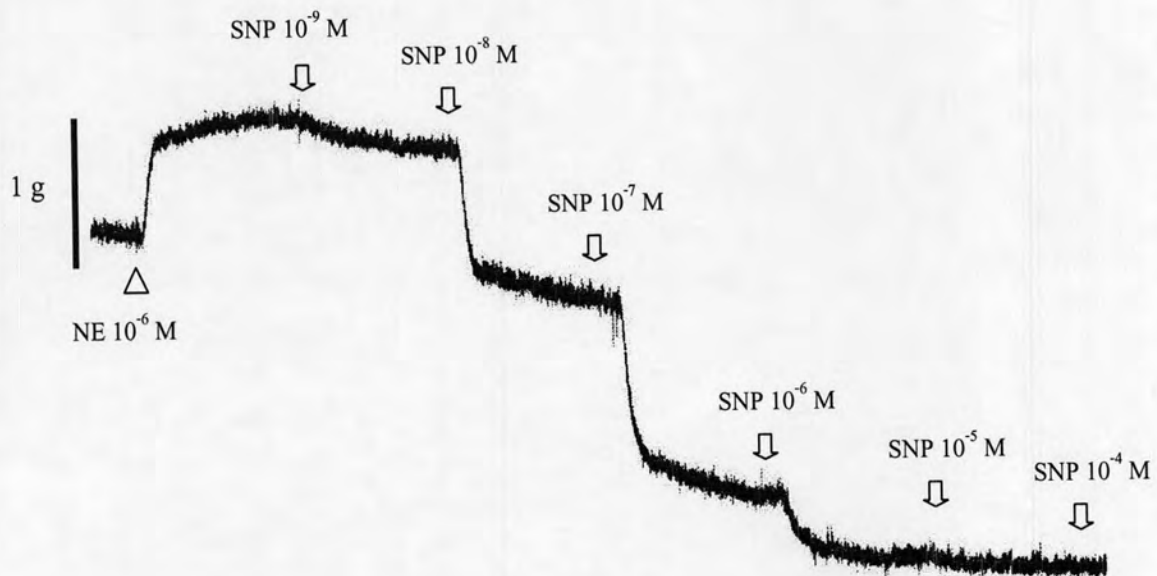
Appendix 5.3.2 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX + Estrogen group.



Appendix 5.3.3 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in OVX group.



Appendix 5.3.4 The isometric tension changes of noradrenaline-precontracted aortic relaxation due to sodium nitroprusside (SNP) stimulation in isolated rat thoracic aortas in Sham group.



Appendix 6

Preparation of reagents

Modified Krebs-Henseleit (KHS) buffer solution

Compositions of Modified Krebs-Henseleit (KHS) buffer solution were consisted of 119 mM NaCl, 4.7 mM KCl, 2.5 mM $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, 1 mM $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 25 mM NaHCO_3 , 1.2 mM KH_2PO_4 and 11.2 mM Glucose.

The solution was continuously bubbled with carbogen gas (95% O_2 + 5% CO_2) and maintained temperature at 37 C°, acid-base status at pH 7.4 throughout the experiments.

Vascular functions testing solutions

Noradrenaline (NA)

Noradrenaline working solutions (10^{-6} M to 10^{-1} M) were prepared to obtain the final concentration of 25 μl noradrenaline (10^{-9} M to 10^{-4} M) in 25 ml organ bath. Ascorbic acid (10^{-4} M) was used as an antioxidant in these working solutions.

Acetylcholine (Ach)

Acetylcholine (10^{-6} M to 10^{-1} M) were used as working solutions to obtain the final concentration of 25 μl acetylcholine (10^{-9} M to 10^{-4} M) in 25 ml organ chamber.

Sodium nitroprusside (SNP)

Sodium nitroprusside working solutions (10^{-6} M to 10^{-1} M) were prepared to obtain the desired concentration of 25 μl sodium nitroprusside (10^{-9} M to 10^{-4} M) in 25 ml organ bath.

Decalcifying agents

Solution A (20% sodium citrate) and solution B (45% formic acid) were used as bone decalcifying agents. The equal volumes of both solutions were mixed freshly and used in bone decalcification process. Mixed solution was changed every 1-3 days until the bones were softened.

Griess reagents

Color reagent 1 (1% sulfanilamide [p-Aminobenzenesulfonamide] in 3N HCl) and color reagent 2 (0.1% NNED [N-(1-Naphthyl) ethylenediamine dihydrochloride]) were used as Griess reagents to determine plasma nitric oxide level in colorimetric non-enzymatic assay.

Appendix 7

เลขที่60/ 2549.....

คณะสัตวแพทยศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย
 ใบอนุญาตให้ใช้สัตว์ใน
 งานวิจัย งานทดสอบ งานผลิตชีววัตถุ งานสอน และงานอื่นๆ

ใบอนุญาตนี้ให้ไว้เพื่อแสดงว่าคณะกรรมการควบคุมดูแลการเลี้ยงและการใช้สัตว์เพื่อ
 งานทางวิทยาศาสตร์ คณะสัตวแพทยศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย ได้พิจารณาโครงการวิจัย เรื่อง
 “ผลของกาวเครือขาว (*Pueraria mirifica*) ต่อการทำงานและการเปลี่ยนแปลงทางพยาธิสภาพของหลอดเลือด
 แดงใหญ่และกระดูกในหนูขาวที่ผ่าตัดเอารังไข่ออก” ซึ่งมีรองศาสตราจารย์ ภญ.ดร.สุหัตรา ศรีไชยรัตน์
 เป็นหัวหน้าหรือเจ้าของโครงการแล้วเห็นสมควรอนุญาตให้ดำเนินการตามโครงการนี้ได้ โดยมีเงื่อนไข
 ว่าผู้ให้สัตว์ในความรับผิดชอบของโครงการต้องปฏิบัติตามข้อมูลที่กรอกในแบบฟอร์มขออนุญาตใช้
 สัตว์ที่ คณะสัตวแพทยศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย สำหรับการวิจัยอย่างเคร่งครัด กรณีที่มีการปฏิบัติ
 อย่างหนึ่งอย่างใด นอกเหนือจากที่ระบุในแบบฟอร์มขออนุญาตและเสนอในโครงการ คณะกรรมการ
 ควบคุมดูแลการเลี้ยงและการใช้สัตว์เพื่องานทางวิทยาศาสตร์จะดำเนินการงดใบอนุญาตฯ นี้ และแจ้ง
 หน่วยงานที่เกี่ยวข้องทราบ

ลงนาม.....
 (ผู้ช่วยศาสตราจารย์ น.สพ.ดร.สุวรรณเกียรติ สว่างคุณ)
 ประธานคณะกรรมการควบคุมดูแลการเลี้ยงและ
 การใช้สัตว์เพื่องานทางวิทยาศาสตร์

ลงนาม.....
 (รองศาสตราจารย์ สพ.ญ.ดร.เจนนุช ว่องธวัชชัย)
 รองคณบดีฝ่ายวิจัยและบริการวิชาการ

วันที่ออกใบอนุญาต.....27.....ธันวาคม 2549.....

วันที่หมดอายุ.....26.....ธันวาคม 2550.....



BIOGRAPHY

Mr. Chenphop Sawangmake was born in November 30, 1979 in Nan province, Thailand. He graduated with Doctor of Veterinary Medicine (D.V.M.) in 2004 from the Faculty of Veterinary Medicine, Chiang Mai University, Chiang Mai, Thailand. After graduation, he worked as a clinical practitioner in Yindee small animal hospital, Samparn, Nakornprathom.

He studied in Master of Science Program in Veterinary Pharmacology, Department of Veterinary Pharmacology, Faculty of Veterinary Science, Chulalongkorn University in academic year 2005.