

การโคลนนิ่งและการศึกษาลำดับเบสซีดีเค็มของตัวไวรัสไซโคโลเอดีซีนและซีดี 163 ที่แยกได้
จากแมคโครไฟจสุกรที่ติดเชื้อไวรัสพีอาร์օาร์ເອສ

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CLONING AND SEQUENCING OF SIALOADHESIN AND CD163 cDNA FROM
PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME VIRUS
INFECTED PORCINE ALVEOLAR MACROPHAGES

Mr. Vo Phong Vu Anh Tuan

A Thesis Submitted in Partial Fulfillment of the Requirements
for the Degree of Master of Science Program in Veterinary Medicine

Department of Veterinary Medicine

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ໄວ ພອງ ວູ້ ຂັນ ທ້ວນ : ກາຣໂຄລນນິ່ງແລກກະຕືກະຊາລຳດັບເປົສີໍດີເຄືນເຂົອງຕັກໄວຮັບໄຊຂອະໄລ ແອດເມື່ອນິນແລກຈີໍດີ 163 ທີ່ແຍກໄດ້ຈາກແມຄໂຄຣຟາຈສຸກຮົກທີ່ຕິດເຫຼືອໄວຮັສພືອເວົ້າຮົກ (CLONING AND SEQUENCING OF SIALOADHESIN AND CD163 cDNA FROM PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME VIRUS INFECTED PORCINE ALVEOLAR MACROPHAGES) ອ.ທີ່ປຶກກະວິທະຍານິພນົມໜັກ: ວ.ສ.ນ.ສ.ພ.ດ.ຮ.ອ. ອົມືຖຸ ນັນທປະເສົາງ, 144 ພໍາ.

ไซโคอะโลแอดไฮซิน และ ชีดี 163 เป็นตัวไวรัสบอย่างน้อย 2 ชนิด ซึ่งพบที่แมคโครฟากสูกรที่ติดเชื้อไวรัสพีอาร์อาร์ເອສ โดยมีความสำคัญต่อการสร้างภูมิคุ้มโรคในสูกรหลังจากได้รับเชื้อไวรัสวัตถุประส่งค์ของงานวิจัยนี้ คือ การโคลนนิ่งและการศึกษาลำดับเบสชีดีเอ็นເຂອງตัวไวรัสไซโคอะโลแอดไฮซิน (ทั้งสายและเฉพาะส่วน N-terminal) และ ชีดี 163 (ทั้งสายและเฉพาะส่วน domain 5) จากแมคโครฟากของสูกรอนุบาลในประเทศไทยที่ติดเชื้อไวรัสพีอาร์อาร์ເອສทั้ง 3 สายพันธุ์ เปรียบเทียบการแสดงออกของทั้งสองยีนกับยีน GAPDH เป็นตัวอ้างอิงภายใน การวิจัยนี้ได้ทำการสกัด RNA ทั้งหมดจากแมคโครฟาก จำนวน 6 ตัวอย่างที่ติดเชื้อไวรัสพีอาร์อาร์ເອສสายพันธุ์จีนสายพันธุ์ยูโรป และสายพันธุ์อเมริกา ต่อมาชีดีเอ็นເຂອງตัวไวรัสไซโคอะโลแอดไฮซิน (ทั้งสายและเฉพาะส่วน N-terminal) และชีดี 163 (ทั้งสายและเฉพาะส่วน domain 5) จะถูกเพิ่มปริมาณโดยใช้เทคนิค PCR เปรียบเทียบการแสดงออกของยีนจากความเข้มของ band ดีเอ็นເกีที่ได้ จำนวนนี้ ชีดีเอ็นເຂອງตัวไวรัสจะถูกโคลนเข้าไปในเวคเตอร์ pCR-XL-TOPO® (ทั้งสาย) และ pCR8®-GW-TOPO® (เฉพาะส่วน N-terminal และ ส่วน domain 5) ทำการวิเคราะห์ และ เปรียบเทียบลำดับนิวคลีโอไทด์ และกรดอะมิโนกรามทั้งส่วนประกอบที่ได้ ผลการวิจัย พบว่า สามารถเพิ่มจำนวนชีดีเอ็นເกี โคลนนิ่งยีนตัวไวรัสทั้งสองเข้าสู่เวคเตอร์ทั้งสองชนิดได้สำเร็จโดยได้ไซโคอะโลแอดไฮซินทั้งสาย จำนวน 10 โคลน ชีดี 163 ทั้งสาย จำนวน 20 โคลน ไซโคอะโลแอดไฮซินเฉพาะส่วน N-terminal จำนวน 24 โคลน และ ชีดี 163 เฉพาะส่วน domain 5 จำนวน 29 โคลน เมื่อเปรียบเทียบส่วนประกอบ ลำดับนิวคลีโอไทด์ และกรดอะมิโนของชีดีเอ็นເຂອງตัวไวรัสทั้งสองแล้ว พบว่า มีความคล้ายคลึงกันมากกว่า 95% นอกจากนี้ การแสดงออกของชีดีเอ็นເຂອງตัวไวรัสทั้งสองชนิดที่แยกได้จากแมคโครฟากสูกรอนุบาลในประเทศไทยที่ติดเชื้อไวรัสพีอาร์อาร์ເອສสายพันธุ์อเมริกามีค่าสูงกว่าสายพันธุ์จีน และสายพันธุ์ยูโรป

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VO PHONG VU ANH TUAN: CLONING AND SEQUENCING OF SIALOADHESIN AND CD163 cDNA FROM PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME VIRUS INFECTED PORCINE ALVEOLAR MACROPHAGES.
ADVISOR: ASSOC. PROF. ATHIPOO NUNTAPRASERT, Ph.D, 144 pp.

Sialoadhesin (Sn) and CD163 receptors were reported as two essential receptors on porcine alveolar macrophages (PAM) for PRRSV infection and importance for the study of the immunity of pigs after infection. The objectives of this study were cloning and sequencing the porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA from 3 strains of PRRSV infected PAM of Thai nursery pigs and comparison of gene expression level with GAPDH gene as internal reference. In this thesis, total RNA was extracted from 6 positive EU, US and HP-PRRSV infected PAM. The porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA were further amplified using PCR technique. Band densitometry was used to quantify expression level. Then, the cDNA were cloned into pCR®-XL-TOPO® (full cDNA) and pCR®8-GW-TOPO® (N-terminal domain and domain 5 cDNA) vectors. The nucleotide and deduced amino acid sequences including compositions were then analyzed and compared. The results showed that 10 clones of Sn (full) and 20 clones of CD163 (full) were constructed in pCR®-XL-TOPO® vectors, while 24 clones of Sn (N-terminal domain) and 29 clones of CD163 (domain 5) were transferred into pCR8®-GW-TOPO® vectors. The sequences and compositions of nucleotide and deduced amino acids of these genes from 3 strains of PRRSV were successfully analyzed and showed more than 95% similarity. Furthermore, the US strain of PRRSV infected PAM collected from Thai nursery pigs showed higher expression level of porcine Sn and CD163 cDNA than that of the HP and EU strains.

Department : Veterinary Medicine Student's Signature

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LIST OF ABBREVIATIONS

ASFV	=	African swine fever virus
aa	=	amino acid
bp	=	base pair
BHK-21	=	Baby hamster kidney
cDNA	=	Complementary DNA
CSF	=	Classical Swine Fever
DCs	=	dendritic cells
DNA	=	deoxyribonucleic acid
dNTP	=	dATP, dGTP, dTTP, dCTP
EAV	=	Equine Arteritis virus
ELISA	=	enzyme-linked immunoassay
ER	=	endoplasmic reticulum
EU	=	European genotype
FMD	=	Foot and Mouth Disease
GAPDH	=	Glyceraldehyde-3-phosphate dehydrogenase
GHR	=	Growth hormone receptors
GHRF	=	Growth hormone releasing factor
GP	=	Glycoprotein
Hb	=	hemoglobin
HP	=	High Pathogenic
Hp	=	haptoglobin
IFN	=	Interferon
IGFBP-5	=	Insulin-like growth factor binding protein-5
IGF-I	=	Insulin-like growth factor-I
IL	=	Interleukin
LB	=	Luria Bertani

LPS	=	Lipopolysaccharides
LV	=	Lelystad Virus
kDa	=	kilodalton
M	=	matrix protein
mRNA	=	Messenger ribonucleic acid
N	=	nucleocapsid protein
NCBI	=	National Center for Biotechnology Information
NSPs	=	non-structural proteins
ORFs	=	open reading frames
PAM	=	porcine alveolar macrophages
PBS	=	Phosphate buffered saline
PCR	=	polymerase chain reaction
PCV2	=	Porcine Circovirus type 2
PED	=	Porcine Epidemic Diarrhea
PK15	=	Porcine kidney 15
PRRS	=	porcine reproductive and respiratory syndrome
PRRSV	=	porcine reproductive and respiratory syndrome virus
RNA	=	Ribonucleic acid
rpm	=	Rounds per minute
RT-PCR	=	Reverse Transcriptase Polymerase chain reaction
SiRNA	=	Small inhibitory RNA
Sn	=	Sialoadhesin
SRCR	=	scavenger receptor cysteine-rich
TLR4	=	Toll-like receptor 4
TNF	=	Tumor necrosis factors
US	=	North American genotype
UV	=	Ultraviolet

CHAPTER I

INTRODUCTION

Many swine viruses can cause the diseases in pigs by replicating inside the target cells. Virus infection usually binds to its specific receptors on the surface of a target cell. The expressed receptor lets the virus to penetrate the cell, spread in the host, and develop the pathogenesis (Norkin, 1995). An important analysis of virus receptors is gained the knowledge for development of clinically effective antiviral agents. The recognition of virus receptor inhibitors of the viral receptor interaction has also useful for the study of viral infection. In addition, the fundamental, biological and clinical importance basis data of viral receptors may have the potential to design the drugs that limit the interaction between virus and its receptor at the initiation of viral binding and may suggest new therapy of viral diseases and the method to blocks viral infection into the target cell of host by using the other related viruses that use the similar receptor. Moreover, the inhibitors of the viral receptor interaction, siRNA functional genomics applications (Behlke, 2006) and the anti-receptor monoclonal antibodies may be clinically useful for inhibition of the replication of major viruses (Norkin, 1995). In swine diseases, there are many reports about viruses and their candidate receptors such as Heparan sulphate (Foot and Mouth Disease (FMD) virus) (Fry et al., 1999); Aminopeptidase N (Porcine Epidemic Diarrhea (PED) virus) (Li et al., 2007); Heparan sulphate and Chondroitin sulphate B (Porcine Circovirus type 2 (PCV2) virus) (Misinzo et al., 2006); Heparan sulphate (Classical Swine Fever (CSF) virus) (Hulst et al., 2000); Heparan sulphate, Sialoadhesin (Sn) and CD163 (porcine reproductive and respiratory syndrome (PRRS) virus) (Duan et al., 1997a; Duan et al., 1997b; Calvert et al., 2007). Among those diseases, PRRS is the most important disease on pig health and welfare worldwide more than 20 years after its emergence (Huang and Meng, 2010). Two receptors on porcine alveolar macrophages (PAM), which are porcine Sn and CD163, have been recognized for PRRS virus (PRRSV) entry and uncoating

(Vanderheijden et al., 2003; Calvert et al., 2007; Van Gorp et al., 2008). The PRRS virus is required these receptors to release viral genome into the cytoplasm of the host cell and spread the infection. Porcine Sn is considered as the attachment and internalization receptor (Vanderheijden et al., 2003). While at an early endosomes stage, the virus can liberate its genome by localization with CD163 (Van Gorp et al., 2009) which is a essential factor for PRRSV genome release (Van Gorp et al., 2008). Furthermore, the co-expression of Sn and CD163 receptors increases virus production and the efficiency of PRRSV infection (Van Gorp et al., 2008). Recently, porcine Sn and CD163 genes were reported to be related to the immunity of pigs after PRRSV infection and may be used to improve PRRS immunity of pigs (Wang et al., 2011). Therefore, the construction of recombinant porcine Sn or CD163 receptors produced by using bacterial system or mammalian system are useful for study of the process of PRRSV into the target cells (Calvert et al., 2007; Pérez et al., 2008; Van Gorp et al., 2008). Furthermore, the understanding of interaction between PRRSV and their receptors may be used to produce peptide mimetic, specific antibodies to block PRRSV entry and inhibit infection of pigs with the PRRSV (Das et al., 2010).

In this study, the porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) receptor cDNA molecules isolated from PAM infected with 3 strains of PRRSV in Thai nursery pigs were cloned, expressed into a bacterial vectors, studied the nucleotide and deduced amino acid sequences. Briefly, the porcine Sn and CD163 receptor cDNA were amplified using polymerase chain reaction (PCR) technique with specific primers. The pCR[®]-XL-TOPO[®] vector containing kanamycin and pCR[®]-8-GW-TOPO[®] vector with spectinomycin resistant genes were used for construction the plasmids with porcine Sn and CD163 cDNA (full, N-terminal domain and domain 5); respectively. Those plasmids were then transformed into *E.coli*. Positive clone of transformants was selected and confirmed correction by using PCR technique. The sequences of study genes were studied and the deduced amino acids and homology were analyzed and compared with the available databases of the reference Sn (refSn) and the reference CD163 (refCD163) cDNA from the

PAM of normal pigs from GenBank. Furthermore, the expression level of porcine Sn and CD163 cDNA from the infected PAM with 3 strains of PRRSV were measured using densitometer and the relative density between these studied genes and housekeeping gene (GAPDH) were analyzed and compared using FusionCapt Advance SL4 machine and Bio 1D software.

The cloning and sequencing of porcine Sn and CD163 cDNA from PRRSV infected PAM in Thai nursery pigs is the first study in Thailand. The knowledge from this study may be useful for understanding these two PRRSV receptors. Their nucleotide and deduced amino acid sequences and compositions may be applied for construction of these recombinant receptors and may helpful to inhibit and control PRRSV infection in pigs in the future.

CHAPTER II

LITERATURE REVIEW

2.1 Genome of Porcine reproductive and respiratory syndrome virus (PRRSV)

PRRSV is enveloped, single stranded and positive sense RNA virus. PRRSV is belonged to the family *Arteriviridae* and the order *Nidovirales* based on the similarity of their genome structure and replication strategy (Snijder and Meulenbergh, 1998). It consists of nine open reading frames (ORFs) as shown in Figure 2.1. All of the ORFs were overlap with each other except between ORF1b and ORF2a. ORF1a and ORF1b (Snijder and Meulenbergh, 1998) together span roughly 75% of the genome from the 5'-end. These ORFs make glycoproteins that are processed by different viral proteases to produce a total thirteen to fourteen non-structural proteins (NSPs), named NSP 1α , NSP 1β , and NSP 2-6, NSP 7α , NSP 7β , and NSP8-12, respectively (Van Aken et al., 2006; Kroese et al., 2008). The ORFs 2a, 2b, 3-7 contain about 25% of the genome at the 3'-end and they encode the structural proteins, namely, the glycoprotein (GP) 2a (GP2a), 2b (or E), GP3, GP4, GP5, the matrix protein (M), and the nucleocapsid protein (N), respectively (Figure 2.2) (Wu et al., 2005). GP2a, GP3, GP4, and GP5 are N-glycosylated and are present on the viral envelope (Dea et al., 2000) as are the non-glycosylated M protein and 2b proteins. GP5, M, and N proteins are known as major structural proteins whereas GP2a, 2b (E), GP3, and GP4 proteins are regarded as minor structural proteins. All of the minor and major surface proteins are requisite for production of PRRSV infection.

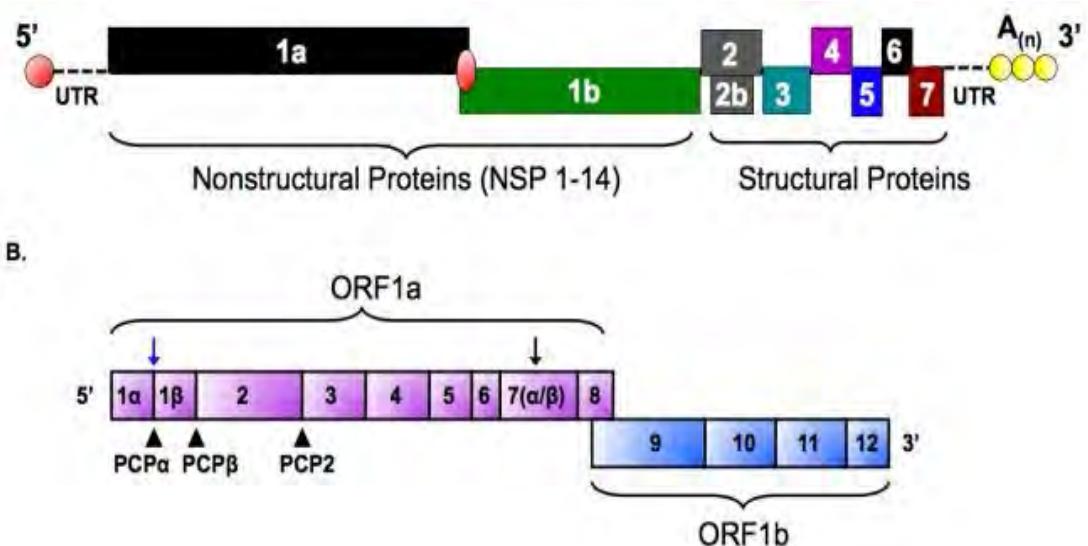


Figure 2.1 Diagram representation of PRRSV genome organization (Das et al., 2010)

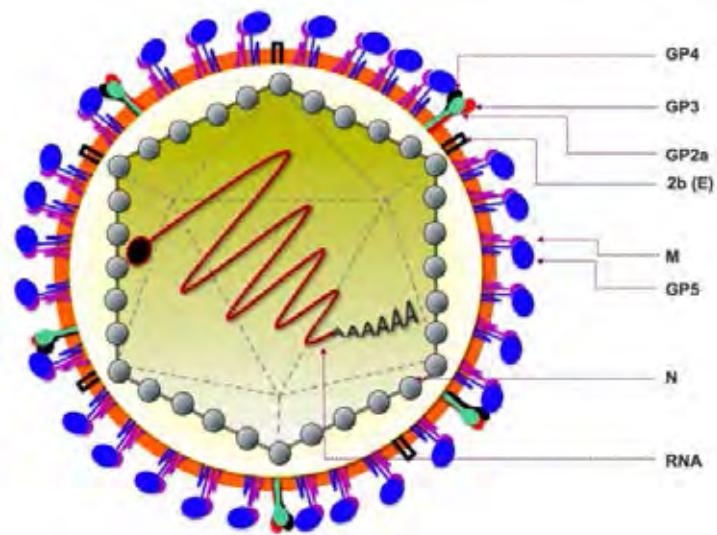


Figure 2.2 Diagram representation of PRRSV particle structure (Das et al., 2010)

2.2. PRRSV strains

Previously, PRRSV can be categorized in genotype based on the same genotypes, common and type specific antigenic determinant present on the structural proteins (Dea et

al., 2000; Forsberg, 2005; Hanada et al., 2005). It has two prototype strains. Genotype I (European genotype) and genotype II (North American genotype) were distinguished based on 40 % dissimilarities of genome sequence homology. These two genotypes were also separate serologically (Kim and Yoon, 2008). The North American strain is VR-2332 (Nelsen et al., 1998) and the European strain is the Lelystad Virus (LV) (Meulenberg et al., 1997). Recently, the new PRRSV strain, as China strain or High Pathogenic (HP) strain (related to the North American PRRSV genotype) is known for rapid and severe spread among Asian countries (Li et al., 2007). The European and North American PRRSV strains cause the same of clinical symptoms, but they are different viral genotypes whose genomes separate by about 37 % (Allende et al., 1999), thus creating a covering of obscurity about the this virus origin. HP-PRRSV strain is greatly pathogenic infectious disease that firstly emerged in pigs in the central region of China (Li et al., 2007; Tian et al., 2009). This HP-PRRS was showed by a prolonged high fever of above 41°C, red discoloration of the ears and body, anorexia, and high mortality in later periods.

2.3 PRRSV cell tropism

The target cells of PRRSV were studied (Duan et al., 1997). However, the porcine alveolar macrophages are primary target cells for the PRRSV infection (Duan et al., 1997; Teifke et al., 2001). PRRSV mostly infects alveolar macrophages of the lung and does not replicate in non-activated monocytes (Thanawongnuwech et al., 2001). Besides primary porcine alveolar macrophages, PRRSV only in vitro infect into the African green monkey kidney cells (MA-104), and the derived cells such as Marc-145 cells (Kim et al., 1993), but those cell lines do not express Sn (Kim et al., 1993; Calvert et al., 2007). From the literature, it is suggesting that porcine alveolar macrophages (PAM) are suitable for studying PRRSV receptors.

2.4. Essential PRRSV receptors

2.4.1. Porcine Sialoadhesin

The Sialoadhesin (Sn), known as CD169 or Siglec-1, is a type I membrane protein (Williams and Barclay, 1988). Sn was initially known as a sialic acid-dependent sheep erythrocyte receptor on resident bone marrow cells of rats and was also characterized in human, mice and swine (Crocker and Gordon, 1985; Vanderheijden et al., 2003). Sn is member of the family of sialic acid binding immunoglobulin-like lectins (siglecs) (Crocker and Gordon, 1985). The siglecs are type I membrane proteins displaying an amino-terminal V-set immunoglobulin domain that binds sialic acid and variable numbers of downstream C2-set immunoglobulin domains (Crocker et al., 2007). The molecular characterization of Sn indicated that it had 17 immunoglobulin domains of extracellular region, a feature that was well conserved in mammals and its cytoplasmic tail was poorly conserved (Crocker et al., 1994). The extracellular region of Sn could be subdivided into a single amino-terminal V-set domain and 16 C2-set domains (Figure 2.3). The 16 C2 repeats ensure that the terminal V-set domain was kept clear of sialic acid residues on the macrophage while active towards sialic acid conjugated on target cells (Munday et al., 1999).

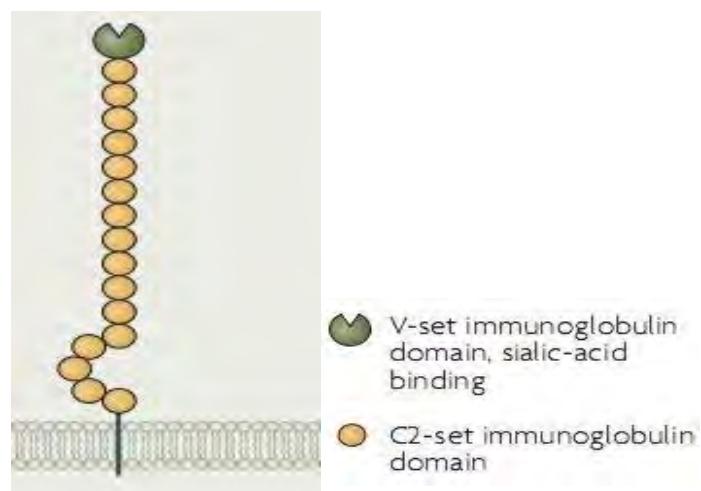


Figure 2.3 Diagram representation of Sn domain (Crocker et al., 1994)

Sn expression was found only on specific subsets of tissue macrophages. Immunocytochemical staining revealed that Sn was constitutively expressed on subpopulations of tissue-resident macrophages, especially those in secondary lymphoid organs (Crocker and Gordon, 1989). Using antibodies raised to the recombinant protein, the expression pattern of human Sn was found to be similar to that of the mouse receptor. Sn was obtained absent from monocytes and other peripheral blood leukocytes, but expressed strongly by tissue macrophages in the spleen, lymph node, bone marrow, liver, colon, and lungs. High expression was also found on inflammatory macrophages presented in affected tissues and was rapidly upregulated by inflammatory macrophages (Hartnell et al., 2001). In contrast, using immunocytochemistry to examine the resident macrophage populations of the nervous system, it showed that Sn was not expressed in the microglia (Perry et al., 1992).

The functions of Sn were suggested as regulators of adhesion, endocytosis and pro-inflammatory (Crocker et al., 2007). It has been reported that the Sn role was as an adhesion molecule for lymphocytes and that it was involved in the modulation of lymphocyte activity (Umansky et al., 1996). Unlike most Sialics, Sn lacks tyrosine-based signaling motifs and its cytoplasmic tail is poorly conserved, which suggests a primary role as a binding partner in cell-cell interactions, rather than in cell signaling (Crocker et al., 1994). It is recently show that under normal conditions, Sn-deficient mice revealed only delicate changes in the haematopoietic and immune systems (Crocker et al, 2007). However, in a model of peptide-induced experimental autoimmune uveoretinitis, the lacking mice showed reduced retinal inflammation and T cells isolated from draining lymph nodes indicated lowered proliferative responses in vitro (Jiang et al., 2006). Furthermore, in two genetically determined models of peripheral and central nervous system demyelination, disease in Sn-deficient mice was improved and numbers of infiltrating CD8+ T cells and macrophages at the sites of inflammation were reduced (Ip et al., 2007). These new findings were consistent with a potentially important role of Sn in modulating T-cell function and activation during

immune responses (Crocker et al., 2007). An additional possibility was that Sn functioned as a phagocytic receptor to clear sialylated pathogens. The cellular and molecular bases for these effects suggested that Sn could mediate both sialic-acid-dependent and sialic-acid-independent interactions with cells of the immune system. Sialic-acid-dependent Sn interactions might be mediated by mucin-like molecules presenting high densities of sialylated O-linked glycans. Sialic-acid-independent Sn interactions could involve the mannose receptor and macrophage galactose-type N-acetylgalactosamine-specific lectin 1 (Kumamoto et al., 2004). The membrane lectins were expressed on dendritic cells (DCs) and have been shown to bind Sn extracted from lymphoid tissues. Although DCs themselves do not normally express Sn, it could be induced on human monocyte-derived DCs following exposure to rhinoviruses *in vitro* (Kirchberger et al., 2005). Interestingly, these DCs were poor stimulators of T cells in mixed lymphocyte reactions, a feature that was partly attributed to the expression of Sn. It is possible that when Sn is expressed by macrophages it is immunostimulatory, whereas on DCs it is immunosuppressive (Kirchberger et al., 2005).

The porcine Sn is imperative for its function as the PRRSV receptor (Crocker et al., 1994; Van Breedam et al., 2010). The porcine Sn gene is 5,193 bp in length and encodes a large protein of 210 kDa (Delputte et al., 2007) and contains 1,730 amino acids (NP_999511, GenBank). The porcine Sn was considered as the attachment and internalization receptor since expression of this protein in non-permissive mammalian cells lead to binding and internalization of PRRSV but not productive viral infection (Vanderheijden et al., 2003). Using monoclonal antibody (41D3) that is specific for porcine Sn, which PRRSV infection is blocked, inhibited this interaction. These findings indicated that Sn is sialic acid binding lectin and that communication between sialic acid on the PRRSV particle and Sn are essential for PRRSV infection of PAM. Similarly, Vanderheijden et al (2003) reported that Sn is concerned in the entry of PRRSV into PAM. The role and co-operation of heparan sulphate and Sn during PRRSV attachment and internalization was

analyzed (Delputte and Nauwynck, 2004). These results revealed that heparan sulphate was not necessary for Sn to function as a PRRSV internalization receptor. In the study, whether the N-terminal domain of Sn is sufficient and/or necessary for PRRSV attachment. An et al. (2010) constructed a chain of truncated segments of Sn and expressed these in the non-permissive PK15 cell line (An et al., 2010). The result shown that the first domain at the N-terminal Sn mediates PRRSV binding to porcine alveolar macrophage cells and contributed to improved understanding the interaction between PRRSV and its target cells. Most recent, the major M/GP5 glycoprotein complex of PRRSV was identified as a ligand for Sn through identifying the viral counterparts for Sn by constructing and validating a soluble form of Sn (Van Breedam et al., 2010). PRRSV has only been obtained to infect and its replication was high levels in MARC-145 cells (Kim et al., 1993), a derived of the African green monkey kidney cell line MA-104. Nevertheless, Sn is absent on MARC-145 cells (Kim et al., 1993; Duan et al., 1998; Wissink et al., 2005) indicating that the molecules other than Sn promote entry of PRRSV in MARC-145 cells. In addition, porcine CD163 obtained from PAM cells was discovered to confer susceptibility to infected PRRSV to non-permissive cells (Calvert et al., 2007).

2.4.2. Porcine CD163

The CD163 is a type I transmembrane protein and is member of scavenger receptor cysteine-rich (SRCR) superfamily (Calvert et al., 2007). The SRCR superfamily are characterized by a cysteine-rich domain (SRCR domain), homologous to the C-terminus of the type I macrophage scavenger receptor (Freeman et al., 1990). Most of the SRCR superfamily proteins are found on cells associated with the immune system (Resnick et al., 1994), but some of the members are also found on other cells, such as hepatocytes (Goldberger et al., 1987) and epithelial cells of the gastrointestinal tract (Li and Snyder, 1995). Functionally, the SRCR domains are thought to mediate protein–protein interactions

and ligand binding (Krieger and Herz, 1994; Pearson, 1996). The SRCR domain has a 100–110 amino acid residues (Resnick et al., 1994). Proteins with SRCR domains are divided into two groups (A and B) based on the localization and number of cysteine residues. Group A all have six cysteine residues per domain but lack cysteine residues at positions 1 and 4. Members of group B have either six or eight cysteine residues per domain, but the cysteine residues at positions 1 and 4 are always present (Resnick et al., 1994). It is identified that CD163 is a group B SRCR protein containing eight cysteine residues per domain, but with only six cysteine resides in domain 8 (Nielsen et al., 2006). Structurally, CD163 has an ectodomain consisting of 9 SRCR domains in tandem. The basic transcript encodes for a protein of 1076 amino acids. The extracellular part contains 1003 amino acids. The transmembrane single segment consists of 24 amino acids. Short cytoplasmic domains consist of 49 amino acids (Ritter et al., 1999; Nielsen et al., 2006). A short linker section then links SRCR domain 9 with a transmembrane domain and an intracellular cytoplasmic tail. Five different isoforms of CD163 have been described so far. They differ in the structure of their cytoplasmic domains and putative phosphorylating sites. Three of these isoforms display different splicing forms of the cytoplasmic domain, which vary from 49 to 84 or 89 amino acids, respectively. The first 42 amino acids after the membrane spanning segment are common for all three isoforms (Nielsen et al., 2006). Two possible alternatives splice sites are observed in an extracellular part of the molecule, one generating a stop codon resulting in a truncated form of protein only consisting of the first three SRCR domains, the other introducing additional 33 amino acids between SRCR 5 and 6 domains (Vilà et al., 2000).

Expression of CD163 is restricted to cells of the monocyte/macrophage lineage. In tissues, CD163 staining is predominantly observed on resident tissue macrophages such as red pulp macrophages in the spleen, Kupffer cells in the liver, and interstitial and alveolar macrophages in the lungs (Fabriek et al., 2005). In addition, the CD163 antigen is present on perifollicular and medullary macrophages in lymph nodes, perifollicular macrophages in

the tonsils, medullary and cortical macrophages in the thymus, and perivascular and meningeal macrophages in the central nervous system (not microglia) (Sánchez et al., 1999; Van den Heuvel et al., 1999). The expression of CD163 is regulated by a variety of factors. Expression of CD163 could correlate with the degree of macrophage activation, because newly infiltrating macrophages are CD163-negative but up-regulate their expression during the healing phase of acute inflammation, in chronic inflammation, and also in wound-healing tissue (Sulahian et al., 2000; Williams et al., 2002). CD163 expression is strongly up-regulated by anti-inflammatory inducers (such as glucocorticoids, IL-6, and IL-10) and down-regulated by proinflammatory agents (such as LPS, TNF- α , and IFN) (Högger et al., 1998; Buechler et al., 2000). However, the mechanisms of CD163 expression are regulated in a complex way: for example, LPS binding to TLR4 induces expression of IL-6 and IL-10, which in turn induce CD163 expression (Weaver et al., 2007). LPS is then able to indirectly induce expression of CD163. Likewise, reduction in CD163 expression mediated by LPS is not a consequence of synthesis inhibition or increased turnover, but rather of induced ectodomain shedding (Weaver et al., 2007).

The role of CD163 has been suggested during erythropoiesis because of its reported ability to bind erythroblasts and promote erythroid expansion in vitro (Fabriek et al., 2007). CD163 was shown to directly interact with erythroblastic cells and a thirteen amino acid motif in the second SRCR domain of CD163 was identified to mediate this binding. Interaction of this CD163 motif with erythroblasts promotes the growth and/or survival of these cells (Fabriek et al., 2007). CD163 was also identified as the endocytic receptor binding hemoglobin (Hb) in complex with the plasma protein haptoglobin (Hp). This specific receptor ligand interaction leading to removal from plasma of the Hp–Hb complex, but not free Hp or Hb, now explains the depletion of circulating Hp in individuals with increased intravascular hemolysis. Besides having a detoxifying effect by removing Hb from plasma, the CD163-mediated endocytosis of the Hp–Hb complex may represent a major pathway for uptake of iron in the tissue macrophages (Graversen et al., 2001). Moreover,

CD163 proposes a role for this molecule as innate immune sensor for bacteria. CD163 has been shown to bind both gram-positive and gram-negative bacteria (Fabriek et al., 2009). Expression of CD163 in monocytic cells promoted bacteria-induced production of pro-inflammatory cytokines, like TNF- α . CD163 is suggested to act as an innate immune sensor for bacteria and inducer of local immunity, rather than as a phagocytic receptor (Fabriek et al., 2009). For other pathogens however, like the porcine viruses ASFV and PRRSV, CD163 serves as a portal that allows infection of their target cells belonging to the monocyte/macrophage lineage (Sánchez-Torres et al., 2003; Van Gorp et al., 2008; Patton et al., 2009).

Porcine CD163 also functions as a cellular receptor for PRRSV (Calvert et al., 2007). The porcine CD163 gene has 3,400 bp in length and encodes a large protein of 150 kDa (Delputte et al., 2007) with 1,133 amino acids (NP_999141, GenBank). Porcine CD163 recognized from PAM cells was obtained to confer susceptibility for PRRSV infection to non-permissive cells (Calvert et al., 2007; Das et al., 2010). Furthermore, porcine CD163 is expressed in MARC-145 cells and cells of monocyte and macrophage lineage that facilitating entry of PRRSV in these cells. Calvert et al. (2007) found that the CD163 expression from PAM, human cells (histiocytic lymphoma), African green monkey kidney cells (MARC-145 and Vero), primary mouse peritoneal macrophages, and canine (histiocytosis) cells involved in encoding functional PRRSV receptors. Moreover, porcine kidney (PK032495), feline kidney (NLFK), or baby hamster kidney (BHK-21) cell lines were recorded as the parental cell lines that were susceptible to PRRSV infection (Calvert et al., 2007). Recombinants of CHO cells stably expressing porcine CD163 have been used for the description of three new monoclonal antibodies against porcine CD163. They used enzyme-linked immunoassay (ELISA) to evaluate levels of soluble CD163 in porcine sera and biological fluids (Pérez et al., 2008). Flow cytometry analysis was used to study the expression of porcine CD163 on PAM (Patton et al., 2009). The findings indicated that the levels of expressed porcine CD163 well correlated with the generally level of PRRSV

copying. These data also showed that the CD163 of expression in dissimilar microenvironments on macrophages (*in vivo*) can ascertain the replication effectiveness and following pathogenesis of PRRSV. The similar study was carried out (Lee et al., 2010) and the outcomes showed that PAM cell lines expressed significant levels of porcine CD163 and were entirely permissive for both EU and US PRRSV strains. The porcine CD163 can enhance virus replication (Lee et al., 2010). The porcine CD163 protein domains concerned in PRRSV infection were identified based on created deletion and chimerical mutants. The infection experiments showed that scavenger receptor cysteine-rich (SRCR) domain 5 (SRCR 5) is essential for PRRSV infection (Van Gorp et al., 2010) and porcine CD163 enhanced virus replication with significant increases in viral protein synthesis and progeny release (Jin Lee and Changhee Lee, 2010). The GP2a and GP4 protein were critical for mediating interglycoprotein interactions (Figure 2.4) (Das et al., 2010) and serves as the viral binding protein that is reliable for intermediate interactions with porcine CD163 for virus infection into susceptible host cell (Das et al., 2010). To identify the porcine CD163 protein domains involved in PRRSV infection, the deletion mutants and chimeric mutants were created. These results showed that scavenger receptor cysteine-rich (SRCR) domain 5 (SRCR 5) is essential for PRRSV infection, while the four N-terminal SRCR domains and the cytoplasmic tail are not required (Van Gorp et al., 2010). The remaining porcine CD163 protein domains need to be present but can be replaced by corresponding SRCR domains from CD163-L1, resulting in reduced (SRCR 6 and interdomain regions) or unchanged (SRCR 7 to SRCR 9) infection efficiency (Van Gorp et al., 2010).

Figure 2.4 Diagram representation of PRRSV glycoproteins and porcine CD163 receptor (Das et al., 2010)

Taken together, the essential PRRSV receptors from review literature are porcine Sn (full and N-terminal domain) and porcine CD163 (full and domain 5). These two receptors were suitable for further studied.

CHAPTER III

MATERIALS AND METHODS

Framework of the study

The framework of this study is shown in Figure 3.1. In this study, a history of a PRRS clinical outbreak farm was used for selecting PRRSV-infected farm. The sera and PAM were collected from nursery pigs with PRRS clinical signs. Then, the molecular technique (RT-PCR) was confirmed the strains of virus from positive PRRSV-infected nursery pigs. The expression and cloning of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA were performed and were compared after their total RNA isolated from PRRSV infected PAM samples. The sequences and compositions of recombinant plasmids and deduced amino acids were studied and compared homology with available database for insight information from GenBank.

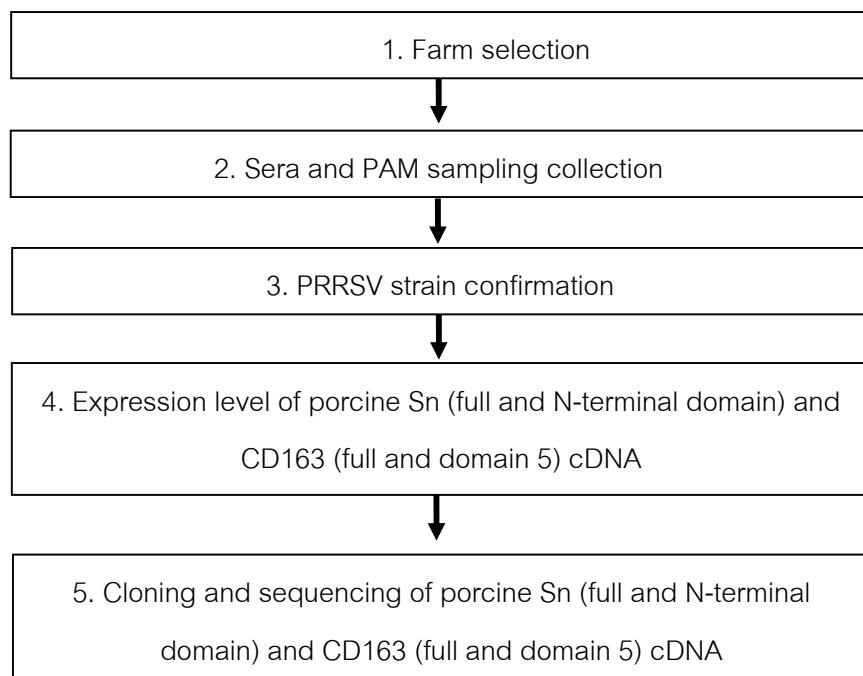


Figure 3.1 Framework of the study

Description of the study

Study farm

Six PRRS outbreak farms (Table 3.1) from the central part of Thailand (Ratchaburi province) were investigated based on clinical signs of PRRSV infection such as abortion and late-term reproductive failure in sows, respiratory disease in nursery and fattening pigs, etc (Li et al., 2007). The candidate farm scale with the sows range from 1,000 to 5,000 was selected.

Table 3.1 Number of Farms, nursery pigs and PAM samples

The positive PRRSV Farms (strain)	Nursery Pigs (3 pigs/farm)	PAM samples
2 (EU strain)	6	2
2 (US strain)	6	2
2 (HP strain)	6	2
Total:	18	6

Sample collection

The eighteen nursery pigs (three nursery pigs/farm) were obtained from those six pig farms as shown in Table 3.1. Five milliliters of sera were collected from each nursery pig with a PRRS clinical sign and stored at -20°C until use. After sera collection, the nursery pigs were euthanized and lungs were collected. The PAM samples were collected by washing broncho-alveolar lung with sterile phosphate buffered saline (PBS) three times with a total volume of approximately 50 ml of PBS. The harvested wash fluid was then centrifuged for 10 minutes at 1,000 rpm. The final cell pellet was re-suspended in 5 ml of

PBS, and the number of PAM was counted to adjust the cell concentration (1×10^6 cells/ml). The PAM was kept fresh and all samples must be processed as soon as possible to maximize the chance of obtaining good quality of RNA. The six PAM samples from the positive PRRSV infected nursery pigs were named as shown in Table 3.2 and used for followed study.

Table 3.2 Sample name of PRRSV infected PAM

Items	PRRSV strain	Sample name
1	HP	Thai-HP-1; Thai-HP-2
2	EU	Thai-EU-1; Thai-EU-2
3	US	Thai-US-1; Thai-US-2

Phase I. Confirmation of positive PRRSV pig farms

The studied Thai nursery pigs were confirmed positive PRRSV infection from sera samples using RT-PCR and ELISA techniques. The PRRSV strains were confirmed by using RT-PCR (Eglia et al., 2001; Vo and Nuntaprasert, 2011) with positive control (PRRSV of ATCC VR2332 for US strain, ATCC LV for EU strain, CH1-a Vaccine for HP strain; respectively) and negative control (DEPC water). The S/P ration of PRRSV immune response was measured by using ELISA kit (IDEXX, USA).

1. Total RNA isolation

Total RNA extractions were performed in a clean and separate area to minimize the chance of cross-contamination. The procedures of total RNA isolation were performed following the protocol of commercial kit's introduction (PureLink™ Viral RNA/DNA Kit,

Invitrogen, USA). According to the protocol, sera samples were added proteinase K and lysis buffer. Next, the mixture was incubated at 56°C for 15 minutes. After that, total RNA were bound onto the Spin Column and then washed with W5 Buffer (contains ethanol). Finally, total RNA was eluted by RNase free water.

2. Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR) two-step protocols

2.1 cDNA synthesis

The RT-PCR kit being used was SuperScript™ RT-PCR System (Invitrogen, San Diego, CA, USA) containing SuperScript II Reverse Transcriptase. Briefly, this step used specific chemicals. The mixture of total RNA, Oligo (dT) and dNTP were incubated at 65°C for 5 minutes and quick chill on ice. The cDNA synthesis mix (RT buffer, MgCl₂, DTT, RNaseOUT and SuperScript) were added to the RNA mixture. Then, the contents of the tube was mixed by pipetting gently up and down and incubated at 50°C for 50 minutes. The final reaction was inactivated by heating at 85°C for 5 minutes. The first-strand cDNA was used as a template for amplification in PCR. The 5 µl of cDNA template were mixed with a reaction mixture containing 1 µl of each specific primer (10 µM), 2 µl of 10X PCR Buffer, 2 µl of 10 mM dNTP Mix, 0.5 µl of Pfu-Taq DNA polymerase (5 U/µl) and 8.5 µl of nuclease-free water (total volume reaction of 20 µl).

2.2 Primers and conditions used for PCR

The reverse-transcribed into cDNA of ORF1 and ORF7 was performed in a one-tube of 20 µl in volume (Eglia et al., 2001; Vo and Nuntaprasert, 2011). The two pairs of primers to amplification of ORF1 and ORF7 PRRSV were shown in Table 3.3 and the sizes of specifically amplified products are 756 bp (US strain), 666 bp (HP strain) and 478 bp (EU strain), respectively.

PCR condition was shown in Table 3.4. The PCR products on agarose gel electrophoresis of 1.5 % were analyzed and readily visible by UV transillumination with an ethidium bromide-stained gel.

Table 3.3 Nucleotide sequences of PCR primers used for RT-PCR

Gene	Sense	Sequence (5' to 3')	GenBank	Predicted product sizes (bp)
1. Sn (full)	+	ATGGACTTCCTGCTCCTGCTC	EU003993	5,193
	-	TCAGACTGTGCTTTACAGA		
2. CD163 (full)	+	ATGGTGCTACTTGAAGACTCT	NM_213976	3,400
	-	TAGTCCAGGTCTTCATCAAGG		
3. Sn (N-terminal)	+	ATGCTCCTGGCTTCATCTGC	EU003993	1,024
	-	TCCACCTCCATGCCCTCATG		
4. CD163 (domain	+	GGACATCCCCTGCTCTGGTC	NM_213976	395
5)	-	CCATGTCCCAGTGAGAGTTG		
5. NSP 2 (full)	+	AAAGACCAGATGGAGGAGGA	GU454850	756
	-	GAGCTGAGTATTTGGCGTG		
6. NSP 2 (deleted)	+	AAAGACCAGATGGAGGAGGA	GU454850	666
	-	GAGCTGAGTATTTGGCGTG		
7. ORF 7	+	ATGCCAATAACAACGGCAAG	GQ330474	478
	-	TCATGCTGAGGGTGATGCTGT		
8. GAPDH	+	TCAATGGAAATCCCATCAC	AF017079	720
	-	TGACAAAGTGGTCGTTGAGG		

Phase II: Expression of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

This study used the PAM samples (as shown in Table 3.1) from positive PRRSV infected Thai nursery pigs for expression of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA. Gene expression level of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA were synthesized using a reverse transcription-polymerase chain reaction (RT-PCR) technique (Lee et al., 2002). The housekeeping gene named glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as a reference control (Foss et al., 1998 ; Suzuki et al., 2000).

1. Total RNA isolation from PAM samples

Total RNA was extracted from total 6 PAM samples of positive 3 strains of PRRSV infected Thai nursery pigs using the protocol of commercial Kit's introduction (Total RNA Minikit, Geneaid, Taiwan). According to the protocol, PAM samples (1×10^6 cells/ml) were efficiently homogenized in a micro-centrifuge tube. The cells were lysed by using RB buffer and β -mercaptoethanol. The total RNA were bound on RB column and then washed with W1 buffer and then were eluted in sterile with RNase free water. The concentration of total RNA was measured by using Nanodrop ND-1000 (Thermo Scientific, Wilming-Ton, DE, USA).

2. Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR)

Five μ g of total RNA from 3 strains of PRRSV infected PAM were reversibly transcribed to cDNA using SuperScriptTM RT-PCR System (Invitrogen, San Diego, CA, USA). This RT-PCR kit contains SuperScript II Reverse Transcriptase. Briefly, this step used specific chemicals. The mixture of RNA, Oligo (dT) and dNTP were incubated at 65°C for 5

minutes and quick chilled on ice. The cDNA synthesis mixture (RT buffer, MgCl₂, DTT, RNaseOUT and SuperScript) were added to RNA mixture. Then, the contents of the tube were mixed by pipetting gently up and down and then incubated at 50°C for 50 minutes. Finally, the reaction was inactivated by heating at 85°C for 5 minutes. The cDNA were used as a template for amplification in PCR step. The concentration of cDNA was measured by using Nanodrop ND-1000 (Thermo Scientific, Wilming-Ton, DE, USA).

3. Primers and conditions used for PCR

The 5 µl of cDNA templates were mixed with a reaction mixture containing 1 µl of each specific primers (10 µM), 2 µl of 10X PCR Buffer, 2 µl of 10 mM dNTP Mix, 0.5 µl of Pfu-Taq DNA polymerase (5 U/µl) and 8.5 µl nuclease-free water (total volume reaction of 20 µl). The specific primers were shown in Table 3.3 and the PCR condition (Vanderheijden et al., 2003; Pérez et al., 2008; Vo and Nuntaprasert, 2012) was shown in Table 3.4. PCR for GAPDH was also done followed the previous studies (Foss et al., 1998; Suzuki et al., 2000).

4. The gene expression level

The PCR products were stained with ethidium bromide in 1.5 % agarose gel electrophoresis and analyzed by using UV transillumination. A FusionCapt Advance SL4 (Vilber Lourmat, Germany) was used to capture the image and using Bio 1D Advance software to analyze density of the expressing bands of porcine Sn and CD163 cDNA. The expression level of porcine Sn and CD163 cDNA were compared with GAPDH housekeeping gene.

Table 3.4 PCR condition program

cDNA	1 cycle	35 cycles			1 cycle	Hold
	Denaturation	Denaturation	Annealing	Extension	Extension	
1. NSP2&ORF 7	94°C (2 minutes)	94°C (30 seconds)	60°C (20 seconds)	72°C (1 minute)	72°C (5 minutes)	4°C
2. Sn (full)	94°C (5 minutes)	94°C (30 seconds)	61°C (30 seconds)	72°C (5 minutes and 20 seconds)	72°C (10 minutes)	4°C
3. Sn (N-terminal domain) & GAPDH	94°C (2 minutes)	94°C (20 seconds)	62°C (20 seconds)	72°C (1 minute)	72°C (5 minutes)	4°C
4. CD163 (full)	94°C (5 minutes)	94°C (30 seconds)	63°C (30 seconds)	72°C (3 minutes and 40 seconds)	72°C (10 minutes)	4°C
5. CD163 (domain 5) & GAPDH	94°C (2 minutes)	94°C (20 seconds)	62°C (20 seconds)	72°C (1 minute)	72°C (5 minutes)	4°C
6. GAPDH	94°C (2 minutes)	94°C (20 seconds)	62°C (20 seconds)	72°C (1 minute)	72°C (5 minutes)	4°C

Phase III. Cloning and sequencing of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

1. Cloning of porcine Sn (full) and CD163 (full) cDNA

The corrected size of PCR products were cut from the agarose gels. These gel bands were then purified by using Gel/PCR DNA Fragments Extraction Kit (Geneaid Company, Taiwan). The gel purified PCR products of porcine Sn (full) and CD163 (full) were measured by using Nanodrop ND-1000 (Thermo Scientific, Wilming-Ton, DE, USA). At the first time, the author cloned these gene receptors directly into the pCR®-8-GW-TOPO® vector using pCR®-8-GW-TOPO® TA Cloning Kit (Invitrogen, USA) according to the manufacturer's instructions. However, no positive colony was detected. Then, the pCR®-XL-TOPO® vector with TOPO®-XL-PCR Cloning Kit (Invitrogen, USA) were chosen for long length (3.0 kb – 10.0 kb) PCR products insertion. Briefly, the TOPO® cloning reaction for eventual transformation into either chemically competent cells were performed in a tube of 6 µl in volume. The mixtures of fresh PCR product and pCR®-XL-TOPO® vector were mixed gently and incubated for 5 minutes at room temperature. The 2 µl of the TOPO® cloning reaction were transformed into a vial of One Shot® TOP10 chemically competent cells, mixed gently and incubated on ice for 30 minutes. Next, the cells were then heat-shock for 30 seconds at 42°C without shaking and transferred into ice. The 250 µl of room temperature S.O.C. medium were added and the tube was shaken horizontally (250 rpm) at 37°C for 1 hour. The 50 µl from each transformant were spread on a Luria Bertani (LB) plate containing 50 µg/ml kanamycin and were further incubated overnight at 37°C. The positive colonies were collected and were cultured overnight in LB medium that containing 50 µg/ml kanamycin. Plasmids DNA were isolated from broth cultures by using the NucleoSpin® Plasmid Kit (Macherey-Nagel, Germany). The positive plasmids were confirmed the presence and correct orientation of the insert by using PCR technique.

2. Cloning of porcine Sn (N-terminal domain) and CD163 (domain 5) cDNA

The corrected size of PCR products were cut from the agarose gels. These gel bands were then purified by using Gel/PCR DNA Fragments Extraction Kit (Geneaid Company, Taiwan). The gel purified PCR products of porcine Sn (N-terminal domain) and CD163 (domain 5) were measured by using Nanodrop ND-1000 (Thermo Scientific, Wilming-Ton, DE, USA) and further cloned directly into pCR®-8-GW-TOPO® vector with pCR®-8-GW-TOPO® TA Cloning Kit (Invitrogen, USA) according to the manufacturer's instructions. Briefly, the TOPO® cloning reaction for eventual transformation into either chemically competent cells were performed in a tube of 6 µl in volume. The mixtures of fresh PCR product and pCR®-8-GW-TOPO® vector were mixed gently and incubated for 5 minutes at room temperature. The 2 µl of the TOPO® cloning reaction were transformed into a vial of One Shot® TOP10 chemically competent cells, mixed gently and incubated on ice for 30 minutes. Next, the cells were then heat-shock for 30 seconds at 42°C without shaking and transferred into ice. The 250 µl of room temperature S.O.C. medium were added and the tube was shook horizontally (250 rpm) at 37°C for 1 hour. The 50 µl from each transformant were spread on a Luria Bertani (LB) plate that containing 100 µg/ml spectinomycin and were further incubated overnight at 37°C. The positive colonies were collected and were cultured overnight in LB medium that containing 100 µg/ml spectinomycin. DNA plasmids were isolated from broth cultures by using the NucleoSpin® Plasmid Kit (Macherey-Nagel, Germany). The positive plasmids were confirmed the presence and correct orientation of the insert by using PCR technique.

3. Sequencing of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

The positive purified recombinant plasmids were submitted to sequence at AITBIOTECH PTE LTD Company (Singapore). The sequencing primers were designed and

shown in Table 3.5. All sequences were carried out in duplicate and were determined by sequencing with both strands. Nucleotide and deduced amino acids sequences were aligned, studied the composition and compared homology by using the software of Chromas 2.33, Bioedit v7.0.5.3 program (ClustalX 2.0.11) and the MEGA 5.1 program.

Table 3.5 Nucleotide sequence primers used for sequencing

Primer	Sequence (5' to 3')	Predicted product sizes (bp)
Sequence primers of Sn (full)		
M13 reverse	CAGGAAACAGCTATGAC	650
F1 forward	GACGGAGCCGGTCAACCTACA	600
F2 forward	GGAGGGCTCTCACAGCCGCAC	600
F3 forward	CACAGATGCCGGCTCATACCA	600
F4 forward	TGCTGCCCTCTATGCTTGCCG	600
F5 forward	CACCACGGACCTGGCTGCCCC	600
F6 forward	TGTCCTCTACGCACCCCGCAG	600
F7 forward	CACTGTGGACAGCGAGGCCACC	600
F8 forward	CCACCTGGCAGTCGGCTGGT	458
Sequence primers of Sn (N-terminal domain)		
M13 forward	GTAAAACGACGGCCAGT	650
F1 forward	GACGGAGCCGGTCAACCTACA	524
Sequence primers of CD163 (full)		
M13 reverse	CAGGAAACAGCTATGAC	650
F1 forward	AGTCAAATTCAAGAGCGGTG	600
F2 forward	GAGACTTAAAGGTGGAGGCAG	600
F3 forward	CGCGTAGTCTGCTCAAGATA	600
F4 forward	TTTGGGGAAGGAACAGGGCC	600
F5 forward	CTCAGCTTGGAGGCAGGAAA	471
Sequence primers of CD163 (domain 5)		
M13 forward	GTAAAACGACGGCCAGT	541

CHAPTER IV

RESULTS

Phase I. Confirmation of PRRSV strains by using RT-PCR

Table 4.1 Confirmation of positive PRRSV strains from sera samples

Farm	Number of nursery pigs	Number of positive samples			S/P ratio ELISA
		EU	US	HP	
1	3	2	-	-	3.123
2	3	3	-	-	3.100
3	3	-	3	-	3.214
4	3	-	3	-	3.233
5	3	-	-	2	3.115
6	3	-	-	2	3.126
Total	18	5	6	4	

The author firstly selected the PRRSV infected farms with clinical signs for PRRS infection based on historical investigation of those farms. The PRRS suspected nursery pigs were selected and the sera and PAM samples were collected. The total RNA was extracted from sera samples and the ORF 1 and ORF 7 cDNA of PRRSV were amplified using RT-PCR technique. The status of PRRS serology was tested with ELISA kit. The RT-PCR results showed that US, HP and EU strains of PRRSV amplified products were detected about 756 bp, 666 bp and 478 bp, respectively (data not shown). The ELISA results also showed that the average S/P ratio of sera from PRRS suspected nursery pigs were all positive from PRRSV infection (cut-off value is 0.4). In this study, sera samples were PRRSV positive in 15/18 of the cases. Two PAM samples (1 PAM sample/farm) of each PRRSV strains were selected and used for further study.

Phase II: Expression of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

1. The expression level of porcine Sn (full) cDNA

Six PRRSV infected PAM samples were selected from 15 positive sera samples (two infected PAM per PRRSV strain). These purified PCR products for porcine Sn cDNA with two sets of porcine Sn primers were run and stained with ethidium bromide in agarose gel 1.5 % at 100 volts for 45 minutes. The image and relative density were shown in Figure 4.1 and Figure 4.2. The product lengths for each sample were confirmed and showed at 5,193 bp. A FusionCapt Advance SL4 (Vilber Lourmat, Germany) was used to capture the gel image. The bands of Thai-HP-(1 and 2)-Sn; Thai-EU-(1 and 2)-Sn and Thai-US-(1 and 2)-Sn were analyzed the density in each lane using Bio 1D Advance software. The expression level of porcine Sn (full) cDNA was examined by RT-PCR and the density of bands. The result indicated that expression of porcine Sn (full) of PAM from US strain was higher than that of HP and EU strains.

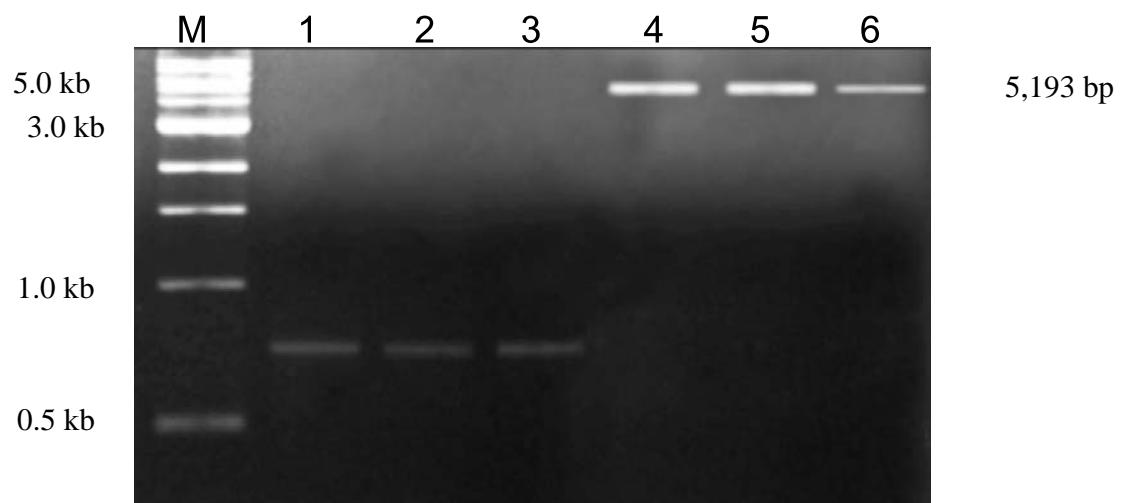


Figure 4.1 Typical 1.5 % agarose gel electrophoresis of amplification of porcine Sn (full) cDNA

- Lane M, 1kb marker (BioLab, USA)
- Lane 1, GAPDH from HP infected PAM
- Lane 2, GAPDH from US infected PAM
- Lane 3, GAPDH from EU infected PAM
- Lane 4, Thai-HP-1-Sn infected PAM
- Lane 5, Thai-US-1-Sn infected PAM
- Lane 6, Thai-EU-1-Sn infected PAM

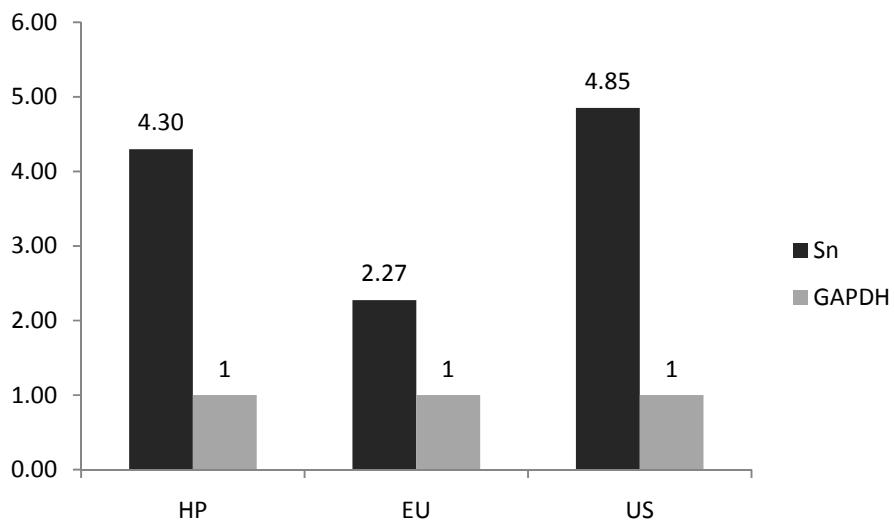


Figure 4.2 Graph shows the relative density of PCR products of porcine Sn (full) and normalized to GAPDH expression

2. The expression level of porcine Sn (N-terminal domain) cDNA

The synthesized N-terminal Sn cDNA from PRRSV infected PAM from each strain were amplified. The expected size of 1,024 bp of these genes were run and stained with ethidium bromide in agarose gel 1.5% at 100 volts for 45 minutes. A FusionCapt Advance SL4 (Vilber Lourmat, Germany) was used to capture the image. The bands of Thai-HP-N-terminal-Sn, Thai-EU-N-terminal-Sn and Thai-US-N-terminal-Sn were measured the density in each lane by using Bio 1D Advance software. The image and relative density were shown in Figure 4.3 and Figure 4.4. The expression of porcine Sn (N-terminal domain) was 6 times higher than GAPDH gene. The band of Thai-US-N-terminal-Sn was showed the highest level at the value of 8.12 followed by 7.46 and 6.35 for Thai-HP-N-terminal-Sn and Thai-EU-N-terminal-Sn, respectively. The result indicated that expression of porcine Sn (N-terminal domain) from US strain was higher than that of HP and EU strains.

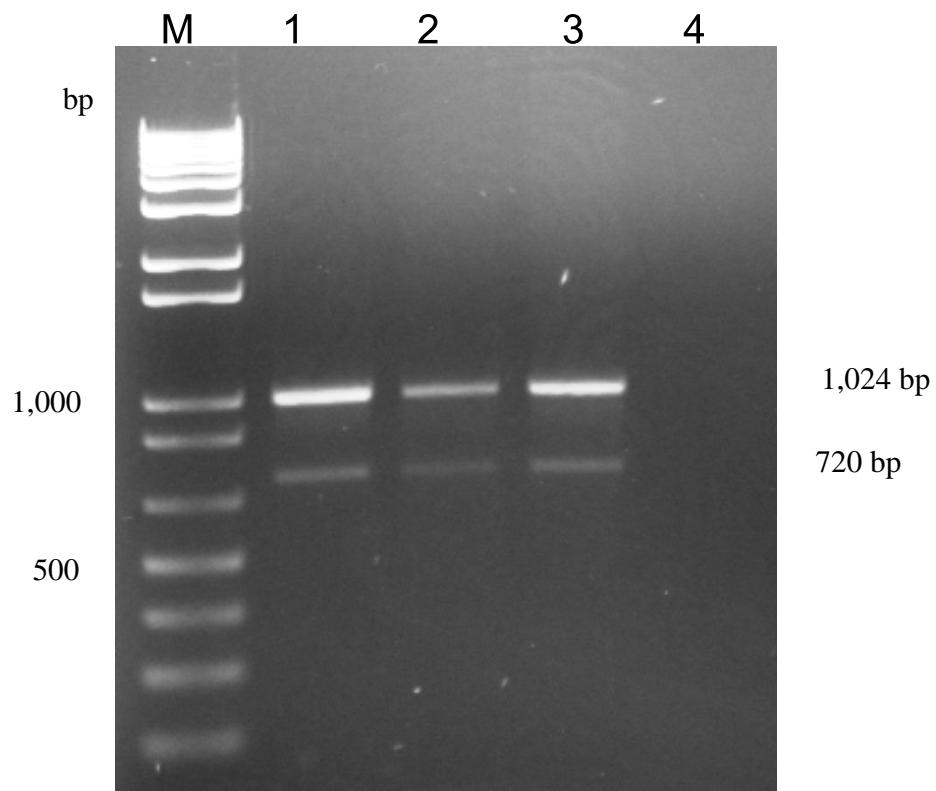


Figure 4.3 Typical 1.5 % agarose gel electrophoresis of amplification of porcine Sn (N-terminal domain) cDNA

Lane M, 1kb marker (Fermantas, Canada)

Lane 1, Thai-HP-1-N-terminal-Sn (1,024 bp); Thai-HP-1-GAPDH (720 bp)

Lane 2, Thai-EU-1-N-terminal-Sn (1,024 bp); Thai-EU-1-GAPDH (720 bp)

Lane 3, Thai-US-1-N-terminal-Sn (1,024 bp); Thai-US-1-GAPDH (720 bp)

Lane 4, negative control

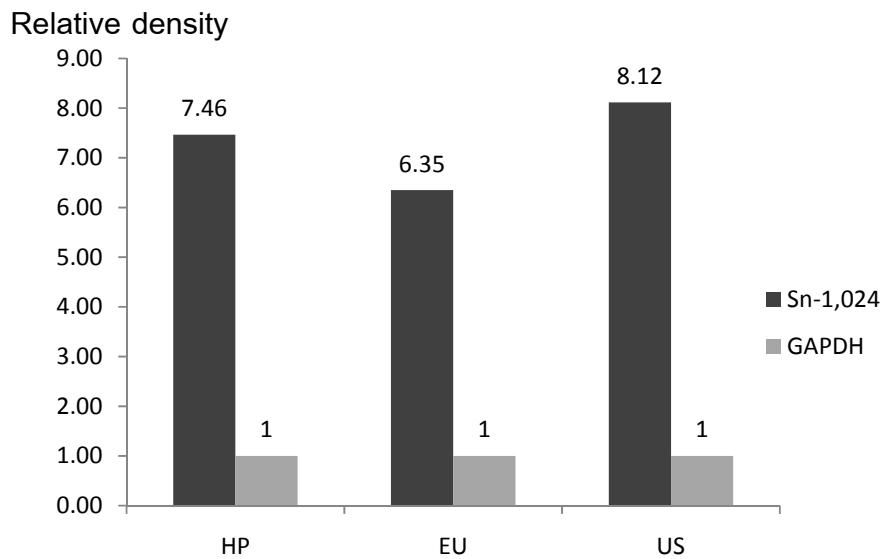


Figure 4.4 Graph shows the relative density of PCR products of porcine Sn (N-terminal domain) and normalized to GAPDH expression

3. The expression level of porcine CD163 (full) cDNA

Six PRRSV infected PAM samples were selected (two infected PAM per PRRSV strain). These purified PCR products for CD163 (full) cDNA with two pairs of porcine CD163 (full) primers were run and stained with ethidium bromide in agarose gel 1.5 % at 100 volts for 45 minutes. The PCR product lengths for each sample were confirmed and shown about 3,400 bp. A FusionCapt Advance SL4 (Vilber Lourmat, Germany) was used to capture the band image as shown in Figure 4.5. The PCR product bands from Thai-HP-CD163, Thai-EU-CD163 and Thai-US-CD163 (as shown in Figure 4.6) were analyzed the density and calculated the relative in each lane by using Bio 1D Advance software. The result indicated that expression of porcine CD163 (full) cDNA from US strain was higher than that of HP and EU strains.

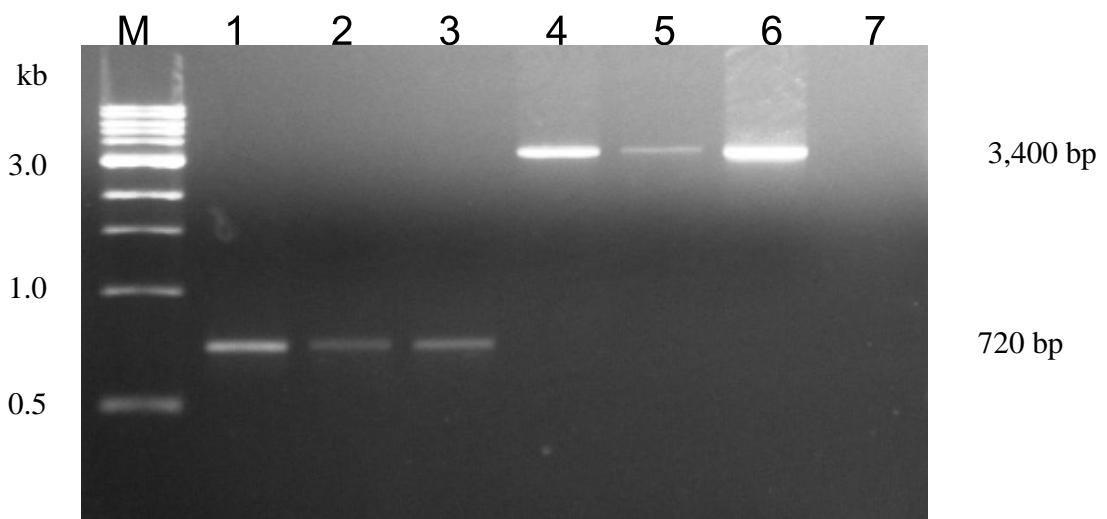


Figure 4.5 Typical 1.5 % agarose gel electrophoresis of amplification of porcine CD163 (full) cDNA

- Lane M, 1kb marker (BioLab, USA)
- Lane 1, GAPDH from EU infected PAM
- Lane 2, GAPDH from HP infected PAM
- Lane 3, GAPDH from US infected PAM
- Lane 4, Thai-HP-1-CD163 infected PAM
- Lane 5, Thai-EU-1-CD163 infected PAM
- Lane 6, Thai-US-1-CD163 infected PAM
- Lane 7, negative control

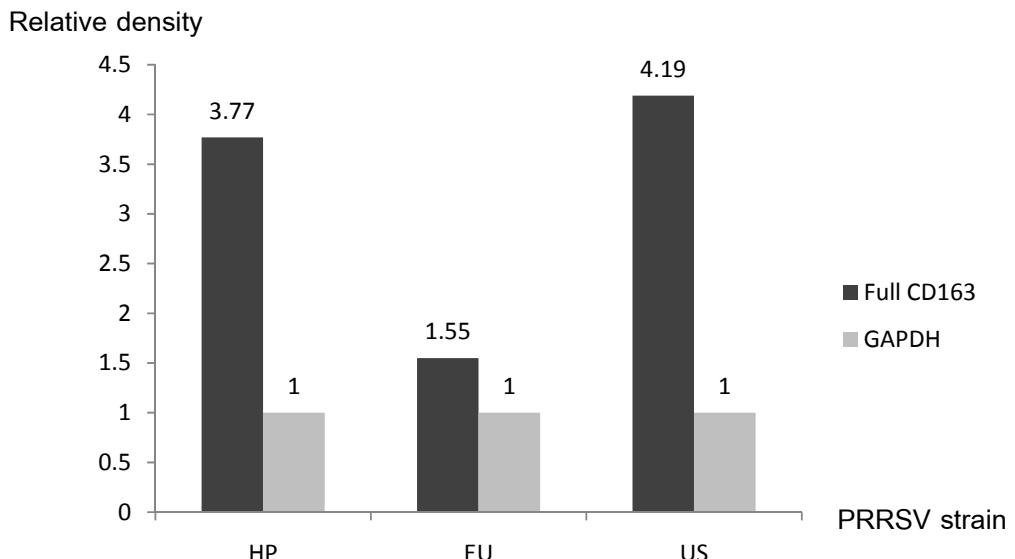


Figure 4.6 Graph shows the relative density of PCR products of porcine CD163 (full) and normalized to GAPDH expression

4. The expression level of porcine CD163 (domain 5) cDNA

The porcine CD163 (domain 5) were amplified from the cDNA of PRRSV infected PAM with specific primers, were run and were stained with ethidium bromide in agarose gel 1.5 % at 100 volts for 45 minutes. A FusionCapt Advance SL4 (Vilber Lourmat, Germany) was also used to capture the image. The expected size of PCR products were about 395 bp and the gel picture was shown in Figure 4.7. The bands of Thai-HP-CD163-DO5, Thai-EUCD163-DO5 and Thai-US-CD163-DO5 were measured the density of the bands in each lane by using Bio 1D Advance software. The relative density was shown in Figure 4.8. The result indicated that expression of porcine CD163 (domain 5) from US strain (4.71) was higher than that of HP (3.70) and EU (3.11) strains.

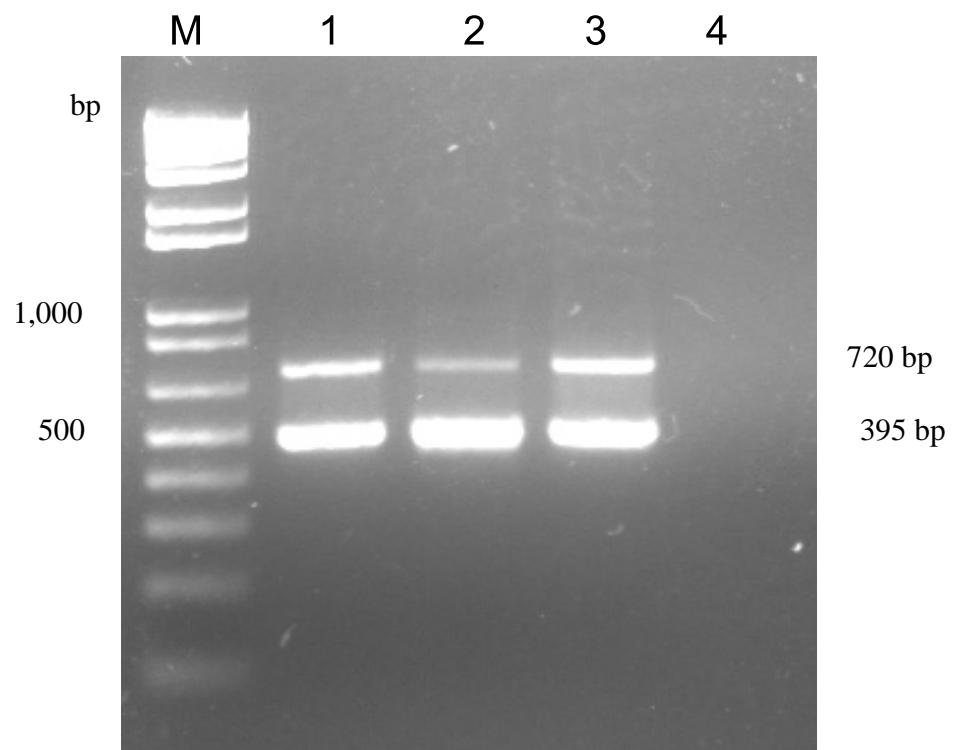


Figure 4.7 Typical 1.5 % agarose gel electrophoresis of amplification of porcine CD163 (domain 5) cDNA

Lane M, 1kb marker (Fermantas, Canada)

Lane 1, Thai-HP-1-CD163-DO5 (395 bp); Thai-HP-1-GAPDH (720 bp)

Lane 2, Thai-EU-1-CD163-DO5 (395 bp); Thai-EU-1-GAPDH (720 bp)

Lane 3, Thai-US-1-CD163-DO5 (395 bp); Thai-US-1-GAPDH (720 bp)

Lane 4, negative control

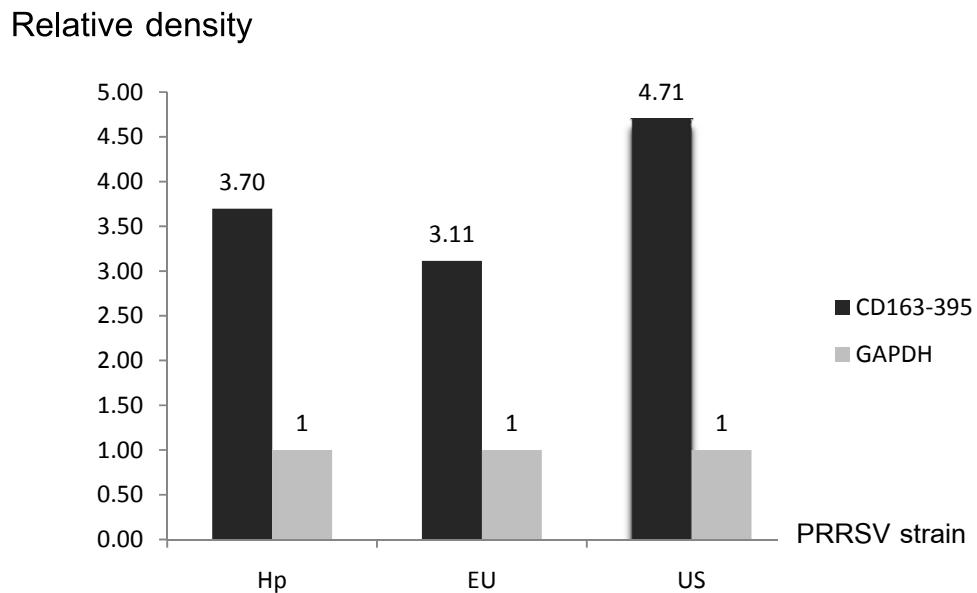


Figure 4.8 Graph shows the relative density of PCR products of porcine CD163 (domain 5) and normalized to GAPDH expression

Phase III: Cloning and sequencing of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

1. Cloning of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA

The corrected sizes of porcine Sn (full) (5,193 bp), porcine Sn (N-terminal domain) (1,024 bp), porcine CD163 (full) (3,400 bp) and porcine CD163 (domain 5) (395 bp) cDNA from 3 strains of PRRSV infected PAM samples were cut and purified from 1.5 % agarose gel. The purified PCR products from 4 genes were first ligated into pCR®8-GW-TOPO® plasmid vectors and transformed into *E.coli* strain One Shot®TOP10 (Invitrogen, USA). Two full receptor genes showed no positive clones. Then, the author used the pCR®-XL-TOPO® plasmid vectors for further long length gene cloning. The corrected insertions of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) were confirmed the corrected

transformants by using PCR technique with specific primers. Ten out of twenty colonies of positive transformants were successfully confirmed for porcine Sn (full) cDNA and named as the clones of pCR®-XL-TOPO®-Sn (4 of Thai-US-1 to 4; 3 of Thai-EU-1 to 3 and 3 of Thai-HP-1 to 3). Twenty-four out of fifty colonies of transformants of porcine Sn (N-terminal domain) cDNA were confirmed and named as clones of pCR®8-GW-TOPO®-Sn (9 of Thai-US-1 to 9; 8 of Thai-EU-1 to 8 and 7 of Thai-HP-1 to 7). Twenty out of forty colonies of porcine CD163 (full) transformants were positive and named as pCR®-XL-TOPO®-CD163 (9 of Thai-US-1 to 9; 5 of Thai-EU-1 to 5 and 6 of Thai-HP-1 to 6) and Twenty-nine out of sixty-three colonies of transformants of porcine CD163 (domain 5) cDNA were confirmed and named as clones of pCR®8-GW-TOPO®-Sn (11 of Thai-US-1 to 11; 8 of Thai-EU-1 to 8 and 10 of Thai-HP-1 to 10). The expected sizes of PCR products for these confirmations were shown at 5,325 bp; 1,170 bp; 3,532 bp and 541 bp for recombinant porcine Sn (full and N-terminal domain) and porcine CD163 (full and domain 5), respectively. In addition, 3 of each corrected recombinant porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) plasmids were further performed the sequencing.

Table 4.2 Positive transformants of recombinant porcine Sn and CD163 plasmids

Sample name	Sn		CD163	
	Full	N-terminal	Full	Domain 5
Thai-HP-1	2	3	4	4
Thai-HP-2	1	4	2	6
Thai-EU-1	1	4	3	4
Thai-EU-2	2	4	2	4
Thai-US-1	2	5	4	5
Thai-US-2	2	4	5	6
Total	10	24	20	29

2. Nucleotide and deduced amino acid sequences of recombinant porcine Sn and CD163 plasmids

Up to now, the author found that the nucleotide and deduced amino acid data of porcine Sn or porcine CD163 cDNA isolated from PRRSV infected PAM or any pig tissues were a few reports. So in this study, the data of porcine Sn and CD163 from PAM of normal pigs from GenBank databases were chosen to analyze alignment, the nucleotide and deduced amino acid compositions and the homology with the data from PAM of PRRSV infected pigs.

2.1 Nucleotide and deduced amino acid sequences of recombinant porcine Sn (full) plasmids

The author selected 3 recombinant plasmids of Thai-HP-1-pCR[®]-XL-TOPO[®]-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR[®]-XL-TOPO[®]-Sn (Thai-EU-1-Sn) and Thai-US-1-pCR[®]-XL-TOPO[®]-Sn (Thai-US-1-Sn) to sequence. The complete sequences of 3 recombinant porcine Sn (full) plasmids were successfully sequenced at the same length of 5,193 bp (Table 4.3) and were analyzed and compared the alignment with the reference Sn (refSn) cDNA (accession number is NM_214346) form the PAM of normal pigs which available databases from GenBank (Figure 4.9). The results indicated that the nucleotide sequences of recombinant porcine Sn (full) plasmids were changed in 10 positions at 130; 762; 1,968; 2,289; 2,718; 2,889; 3,870; 4,295; 4,413 and 4,493, respectively.

The deduced amino acid sequences were summarized in Table 4.4 and the alignment of recombinant porcine Sn (full) plasmids from PRRVS infected PAM (1,730 amino acids) and refSn cDNA (accession number is NP_999511) from normal PAM were shown in Figure 4.10. The results showed that the amino acid sequences of 3 recombinant porcine Sn (full) plasmids were showed the similar pattern and were changed in 2 positions at 44 (Glycine to Arginine) and 1,432 (Leucine to Serine).

Table 4.3 Nucleotide sequences of 3 recombinant porcine Sn (full) plasmids

Transformant's name	Sequence
1. Thai-HP-1- Sn (5,193 bp)	atggacttcctgcctgctccctccctggctcatctgcttagcaggcctggctcggtac ggttccagccccgagaccgtgcagggcatcaaggcctgcctcatcatcccctgcacct tccgctcccgccaacgtggaggtgccccatggcatcacagccatctggtactatgactact caggcaagcgcctggtagtgagccactccaggaacccaaagggtggagaaccactc caaggccggggccctgctgtgggcaggttaacagaggacgtgcagcctgctgaag gacctgcagccccaggactcgggctcataacttccgcttgagatcagcgagggcaacc gctggtcagatgtcaaaggcacagtgtcaccgtacagaggtgcccagcgtgcccaccat tgccttgcagccaagctgcatgagggcatggaggtggacttcaactgctccactccctatgt gtgcccacggagccgtcaacctacagtggcaaggccaggatcccacccgtccgtca cctcccacccatccagaagcttgagccctggcaccagccacatggagaccctgcacatgg ccctgtccctggcaggaccatggccggatcctgagctgccaggtctcagcagccgaacgca ggatgcagaaggagattcacctccaagtgcagtatgcccccaagggtgtggagatcctttc agccactccggacggaacgtcctccctggatctggcaccctcagctgccaggtgaatag cagcaaccctcaggtcagttccgtgcagtgggtcaaggatggacgaagctcaaagacca gaaacgtgtactgcagttgcggccggcagcctgggtgatgtggcgtctacaccctgcaa gccgggaatgccgtggcttcagtctcaccccccgtcagcctccacgtctcatggctga ggtccaggtaaagccctgtggctccatctggagaaccagacggtaacgtggcgtctacaccctgcaat acacctaaggaagcgcccagcgcagctgcgtacagctggtaacaagaaccacacgcccgtct ggagggctctcacagccgcaccctccggctgcactcagttaccaggcggattcgggcttc tacttctgcaggtgcagaacgcccggggcagagagcgtctcccccgtcagcgtgg gtcagccacccaccctcaccccgacctaactgcctccctggagacacaggcggggctg gtgggcacccatgcctcaatgctctggcagcgcagcccccaagctactctggtgttcacacgg ggcctcatctggccttacccctccggggagggtgaccacagcccacgcctcagtgtcgccctc

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Thai-US-1-Sn	GCCGGGAATG CCGTGGGCTC TTCAGTCTCA CCCCCGGTCA GCCTCCACGT

	1110 1120 1130 1140 1150
refSn	CTTCATGGCT GAGGTCCAGG TAAGCCCTGT GGGCTCCATC CTGGAGAAC
Thai-HP-1-Sn	CTTCATGGCT GAGGTCCAGG TAAGCCCTGT GGGCTCCATC CTGGAGAAC
Thai-EU-1-Sn	CTTCATGGCT GAGGTCCAGG TAAGCCCTGT GGGCTCCATC CTGGAGAAC
Thai-US-1-Sn	CTTCATGGCT GAGGTCCAGG TAAGCCCTGT GGGCTCCATC CTGGAGAAC

	1160 1170 1180 1190 1200
refSn	AGACGGTGAC GCTGGCCTGC AATACACCTA AGGAAGCGCC CAGCGAGCTG
Thai-HP-1-Sn	AGACGGTGAC GCTGGCCTGC AATACACCTA AGGAAGCGCC CAGCGAGCTG
Thai-EU-1-Sn	AGACGGTGAC GCTGGCCTGC AATACACCTA AGGAAGCGCC CAGCGAGCTG
Thai-US-1-Sn	AGACGGTGAC GCTGGCCTGC AATACACCTA AGGAAGCGCC CAGCGAGCTG

	1210	1220	1230	1240	1250
refSn	CGCTACAGCT GGTACAAGAA CCACGCCCTG CTGGAGGGCT CTCACAGCCG					
Thai-HP-1-Sn	CGCTACAGCT GGTACAAGAA CCACGCCCTG CTGGAGGGCT CTCACAGCCG					
Thai-EU-1-Sn	CGCTACAGCT GGTACAAGAA CCACGCCCTG CTGGAGGGCT CTCACAGCCG					
Thai-US-1-Sn	CGCTACAGCT GGTACAAGAA CCACGCCCTG CTGGAGGGCT CTCACAGCCG					
	1260	1270	1280	1290	1300
refSn	CACCCCTCCGG CTGCACTCAG TTACCAGGGC GGATTGGGC TTCTACTTCT					
Thai-HP-1-Sn	CACCCCTCCGG CTGCACTCAG TTACCAGGGC GGATTGGGC TTCTACTTCT					
Thai-EU-1-Sn	CACCCCTCCGG CTGCACTCAG TTACCAGGGC GGATTGGGC TTCTACTTCT					
Thai-US-1-Sn	CACCCCTCCGG CTGCACTCAG TTACCAGGGC GGATTGGGC TTCTACTTCT					
	1310	1320	1330	1340	1350
refSn	GCGAGGTGCA GAACGCCCGG GGCAAGAGAGC GCTCTCCCCC TGTCAGCGTG					
Thai-HP-1-Sn	GCGAGGTGCA GAACGCCCGG GGCAAGAGAGC GCTCTCCCCC TGTCAGCGTG					
Thai-EU-1-Sn	GCGAGGTGCA GAACGCCCGG GGCAAGAGAGC GCTCTCCCCC TGTCAGCGTG					
Thai-US-1-Sn	GCGAGGTGCA GAACGCCCGG GGCAAGAGAGC GCTCTCCCCC TGTCAGCGTG					
	1360	1370	1380	1390	1400
refSn	GTGGTCAGCC ACCCACCCCT CACCCGGAC CTAACTGCCT TCCTGGAGAC					
Thai-HP-1-Sn	GTGGTCAGCC ACCCACCCCT CACCCGGAC CTAACTGCCT TCCTGGAGAC					
Thai-EU-1-Sn	GTGGTCAGCC ACCCACCCCT CACCCGGAC CTAACTGCCT TCCTGGAGAC					
Thai-US-1-Sn	GTGGTCAGCC ACCCACCCCT CACCCGGAC CTAACTGCCT TCCTGGAGAC					
	1410	1420	1430	1440	1450
refSn	ACAGGGGGGG CTGGTGGGCA TCCTCCAATG CTCTGTGGTC AGCGAGCCCC					
Thai-HP-1-Sn	ACAGGGGGGG CTGGTGGGCA TCCTCCAATG CTCTGTGGTC AGCGAGCCCC					
Thai-EU-1-Sn	ACAGGGGGGG CTGGTGGGCA TCCTCCAATG CTCTGTGGTC AGCGAGCCCC					
Thai-US-1-Sn	ACAGGGGGGG CTGGTGGGCA TCCTCCAATG CTCTGTGGTC AGCGAGCCCC					
	1460	1470	1480	1490	1500
refSn	CAGCTACTCT GGTGTTGTCA CACGGGGGCC TCATCTTGGC CTCTACCTCC					
Thai-HP-1-Sn	CAGCTACTCT GGTGTTGTCA CACGGGGGCC TCATCTTGGC CTCTACCTCC					
Thai-EU-1-Sn	CAGCTACTCT GGTGTTGTCA CACGGGGGCC TCATCTTGGC CTCTACCTCC					
Thai-US-1-Sn	CAGCTACTCT GGTGTTGTCA CACGGGGGCC TCATCTTGGC CTCTACCTCC					
	1510	1520	1530	1540	1550
refSn	GGGGAGGGTG ACCACAGCCC ACGCTTCAGT GTCGCCTCTG CCCCCAACTC					
Thai-HP-1-Sn	GGGGAGGGTG ACCACAGCCC ACGCTTCAGT GTCGCCTCTG CCCCCAACTC					
Thai-EU-1-Sn	GGGGAGGGTG ACCACAGCCC ACGCTTCAGT GTCGCCTCTG CCCCCAACTC					
Thai-US-1-Sn	GGGGAGGGTG ACCACAGCCC ACGCTTCAGT GTCGCCTCTG CCCCCAACTC					
	1560	1570	1580	1590	1600
refSn	CCTGCGCTG GAGATTCAAG ACCTGGGGCC AACAGACAGT GGGGAATACA					
Thai-HP-1-Sn	CCTGCGCTG GAGATTCAAG ACCTGGGGCC AACAGACAGT GGGGAATACA					
Thai-EU-1-Sn	CCTGCGCTG GAGATTCAAG ACCTGGGGCC AACAGACAGT GGGGAATACA					
Thai-US-1-Sn	CCTGCGCTG GAGATTCAAG ACCTGGGGCC AACAGACAGT GGGGAATACA					

refSn	1610 1620 1630 1640 1650
Thai-HP-1-Sn	TGTGCTCAGC CAGCAGTTCT CTTGGGAATG CGTCCTCCAC CCTGGACTTC
Thai-EU-1-Sn	TGTGCTCAGC CAGCAGTTCT CTTGGGAATG CGTCCTCCAC CCTGGACTTC
Thai-US-1-Sn	TGTGCTCAGC CAGCAGTTCT CTTGGGAATG CGTCCTCCAC CCTGGACTTC

refSn	1660 1670 1680 1690 1700
Thai-HP-1-Sn	CATGCCAATG CAGCCCGCCT CCTCATCAGC CCAGCAGCAG AGGTGGTGGA
Thai-EU-1-Sn	CATGCCAATG CAGCCCGCCT CCTCATCAGC CCAGCAGCAG AGGTGGTGGA
Thai-US-1-Sn	CATGCCAATG CAGCCCGCCT CCTCATCAGC CCAGCAGCAG AGGTGGTGGA

refSn	1710 1720 1730 1740 1750
Thai-HP-1-Sn	AGGGCAGGCG GTGACACTGA GCTGCAGGAG CAGCCTGAGC CTGATGCCTG
Thai-EU-1-Sn	AGGGCAGGCG GTGACACTGA GCTGCAGGAG CAGCCTGAGC CTGATGCCTG
Thai-US-1-Sn	AGGGCAGGCG GTGACACTGA GCTGCAGGAG CAGCCTGAGC CTGATGCCTG

refSn	1760 1770 1780 1790 1800
Thai-HP-1-Sn	ACACCCGTTT TTCTGGTAC CTGAACGGGG CCCTGATTCT CGAGGGGGCCC
Thai-EU-1-Sn	ACACCCGTTT TTCTGGTAC CTGAACGGGG CCCTGATTCT CGAGGGGGCCC
Thai-US-1-Sn	ACACCCGTTT TTCTGGTAC CTGAACGGGG CCCTGATTCT CGAGGGGGCCC

refSn	1810 1820 1830 1840 1850
Thai-HP-1-Sn	AGCAGCAGCC TCCTGCTCCC AGCACGCCTCC AGCACAGATG CCGGCTCATA
Thai-EU-1-Sn	AGCAGCAGCC TCCTGCTCCC AGCACGCCTCC AGCACAGATG CCGGCTCATA
Thai-US-1-Sn	AGCAGCAGCC TCCTGCTCCC AGCACGCCTCC AGCACAGATG CCGGCTCATA

refSn	1860 1870 1880 1890 1900
Thai-HP-1-Sn	CCACTGCCGG GCCCAGAACAA GCCACAGCAC CAGCGGGCCC TCCTCACCTG
Thai-EU-1-Sn	CCACTGCCGG GCCCAGAACAA GCCACAGCAC CAGCGGGCCC TCCTCACCTG
Thai-US-1-Sn	CCACTGCCGG GCCCAGAACAA GCCACAGCAC CAGCGGGCCC TCCTCACCTG

refSn	1910 1920 1930 1940 1950
Thai-HP-1-Sn	CTGTTCTCAC CGTGCTCTAC GCCCACAGGCC AGCCCCGTGTT CACTGCCAG
Thai-EU-1-Sn	CTGTTCTCAC CGTGCTCTAC GCCCACAGGCC AGCCCCGTGTT CACTGCCAG
Thai-US-1-Sn	CTGTTCTCAC CGTGCTCTAC GCCCACAGGCC AGCCCCGTGTT CACTGCCAG

refSn	1960 1970 1980 1990 2000
Thai-HP-1-Sn	CTGGACCCTG ATACTGCAGG AGCTGGGGCC GGACGCCAAG GCCTCCTCTT
Thai-EU-1-Sn	CTGGACCCTG ATACTGCAGG AGCTGGGGCC GGACGCCAAG GCCTCCTCTT
Thai-US-1-Sn	CTGGACCCTG ATACTGCAGG AGCTGGGGCC GGACGCCAAG GCCTCCTCTT

	2410 2420 2430 2440 2450
refSn	ACCGTGACGC TGCTACCTGT :GCCAGAACT GATGCTGCC TCTATGCTTG
Thai-HP-1-Sn	ACCGTGACGC TGCTACCTGT :AGCCAGAACT GATGCTGCC TCTATGCTTG
Thai-EU-1-Sn	ACCGTGACGC TGCTACCTGT :AGCCAGAACT GATGCTGCC TCTATGCTTG
Thai-US-1-Sn	ACCGTGACGC TGCTACCTGT :AGCCAGAACT GATGCTGCC TCTATGCTTG

	2460 2470 2480 2490 2500
refSn	CCGCATCGTC ACCGAGGCTG GTGCTGGCCT CTCCACCCCT GTGGCCCTGA
Thai-HP-1-Sn	CCGCATCGTC ACCGAGGCTG GTGCTGGCCT CTCCACCCCT GTGGCCCTGA
Thai-EU-1-Sn	CCGCATCGTC ACCGAGGCTG GTGCTGGCCT CTCCACCCCT GTGGCCCTGA
Thai-US-1-Sn	CCGCATCGTC ACCGAGGCTG GTGCTGGCCT CTCCACCCCT GTGGCCCTGA

	2510 2520 2530 2540 2550
refSn	ATGTGCTCTA TCCCCCGAT CCTCCAAAGT TGTCAGCCCT CCTGGACGTG
Thai-HP-1-Sn	ATGTGCTCTA TCCCCCGAT CCTCCAAAGT TGTCAGCCCT CCTGGACGTG
Thai-EU-1-Sn	ATGTGCTCTA TCCCCCGAT CCTCCAAAGT TGTCAGCCCT CCTGGACGTG
Thai-US-1-Sn	ATGTGCTCTA TCCCCCGAT CCTCCAAAGT TGTCAGCCCT CCTGGACGTG

	2560 2570 2580 2590 2600
refSn	GACCAGGGCC ACACGGCTGT GTTCGTCTGT ACTGTGGACA GTCGCCCTCT
Thai-HP-1-Sn	GACCAGGGCC ACACGGCTGT GTTCGTCTGT ACTGTGGACA GTCGCCCTCT
Thai-EU-1-Sn	GACCAGGGCC ACACGGCTGT GTTCGTCTGT ACTGTGGACA GTCGCCCTCT
Thai-US-1-Sn	GACCAGGGCC ACACGGCTGT GTTCGTCTGT ACTGTGGACA GTCGCCCTCT

	2610 2620 2630 2640 2650
refSn	TGCCCAGTTG GCCCTGTTCC GTGGGAACA CCTCCTGGCC GCCAGCTCGG
Thai-HP-1-Sn	TGCCCAGTTG GCCCTGTTCC GTGGGAACA CCTCCTGGCC GCCAGCTCGG
Thai-EU-1-Sn	TGCCCAGTTG GCCCTGTTCC GTGGGAACA CCTCCTGGCC GCCAGCTCGG
Thai-US-1-Sn	TGCCCAGTTG GCCCTGTTCC GTGGGAACA CCTCCTGGCC GCCAGCTCGG

	2660 2670 2680 2690 2700
refSn	CACTCCGGCT CCCCCCTCGT GGCGCCTCC AGGCCAAAGC CTCGGCAAC
Thai-HP-1-Sn	CACTCCGGCT CCCCCCTCGT GGCGCCTCC AGGCCAAAGC CTCGGCAAC
Thai-EU-1-Sn	CACTCCGGCT CCCCCCTCGT GGCGCCTCC AGGCCAAAGC CTCGGCAAC
Thai-US-1-Sn	CACTCCGGCT CCCCCCTCGT GGCGCCTCC AGGCCAAAGC CTCGGCAAC

	2710 2720 2730 2740 2750
refSn	TCCTTGCAGC TAGAGGTCCG AGACTTGAGC CTTGGGGACT CTGGCAGCTA
Thai-HP-1-Sn	TCCTTGCAGC TAGAGGTCCG AGACTTGAGC CTTGGGGACT CTGGCAGCTA
Thai-EU-1-Sn	TCCTTGCAGC TAGAGGTCCG AGACTTGAGC CTTGGGGACT CTGGCAGCTA
Thai-US-1-Sn	TCCTTGCAGC TAGAGGTCCG AGACTTGAGC CTTGGGGACT CTGGCAGCTA

	2760 2770 2780 2790 2800
refSn	CCACTGTGAG GCCACCAACA TCCTTGGATC AGCCAACACT TCTCTTACCT
Thai-HP-1-Sn	CCACTGTGAG GCCACCAACA TCCTTGGATC AGCCAACACT TCTCTTACCT
Thai-EU-1-Sn	CCACTGTGAG GCCACCAACA TCCTTGGATC AGCCAACACT TCTCTTACCT
Thai-US-1-Sn	CCACTGTGAG GCCACCAACA TCCTTGGATC AGCCAACACT TCTCTTACCT

					
	3210	3220	3230	3240	3250	
refSn	GGCCTCTACT	CTACAAGGTG	TGGAGGAGCT	TGCAGGCAGC	TCTCCCGCC	
Thai-HP-1-Sn	GGCCTCTACT	CTACAAGGTG	TGGAGGAGCT	TGCAGGCAGC	TCTCCCGCC	
Thai-EU-1-Sn	GGCCTCTACT	CTACAAGGTG	TGGAGGAGCT	TGCAGGCAGC	TCTCCCGCC	
Thai-US-1-Sn	GGCCTCTACT	CTACAAGGTG	TGGAGGAGCT	TGCAGGCAGC	TCTCCCGCC	
					
	3260	3270	3280	3290	3300	
refSn	TACAGGTGGC	CACAGCCCCC	AACACGCTGC	GCCTGGAGAT	CCACAACGCA	
Thai-HP-1-Sn	TACAGGTGGC	CACAGCCCCC	AACACGCTGC	GCCTGGAGAT	CCACAACGCA	
Thai-EU-1-Sn	TACAGGTGGC	CACAGCCCCC	AACACGCTGC	GCCTGGAGAT	CCACAACGCA	
Thai-US-1-Sn	TACAGGTGGC	CACAGCCCCC	AACACGCTGC	GCCTGGAGAT	CCACAACGCA	
					
	3310	3320	3330	3340	3350	
refSn	GTGCTGGAGG	ATGAAGGC GT	CTACACCTGC	GAGGCCACCA	ACACCTGGG	
Thai-HP-1-Sn	GTGCTGGAGG	ATGAAGGC GT	CTACACCTGC	GAGGCCACCA	ACACCTGGG	
Thai-EU-1-Sn	GTGCTGGAGG	ATGAAGGC GT	CTACACCTGC	GAGGCCACCA	ACACCTGGG	
Thai-US-1-Sn	GTGCTGGAGG	ATGAAGGC GT	CTACACCTGC	GAGGCCACCA	ACACCTGGG	
					
	3360	3370	3380	3390	3400	
refSn	TCAGACCTTG	GCCTCCGCG	CCTTCGATGC	CCAGGCTATG	AGAGTGCAGG	
Thai-HP-1-Sn	TCAGACCTTG	GCCTCCGCG	CCTTCGATGC	CCAGGCTATG	AGAGTGCAGG	
Thai-EU-1-Sn	TCAGACCTTG	GCCTCCGCG	CCTTCGATGC	CCAGGCTATG	AGAGTGCAGG	
Thai-US-1-Sn	TCAGACCTTG	GCCTCCGCG	CCTTCGATGC	CCAGGCTATG	AGAGTGCAGG	
					
	3410	3420	3430	3440	3450	
refSn	TGTGGCCCAA	TGCCACC GTG	CAAGAGGGC	AGCTGGTGAA	CCTGACCTGC	
Thai-HP-1-Sn	TGTGGCCCAA	TGCCACC GTG	CAAGAGGGC	AGCTGGTGAA	CCTGACCTGC	
Thai-EU-1-Sn	TGTGGCCCAA	TGCCACC GTG	CAAGAGGGC	AGCTGGTGAA	CCTGACCTGC	
Thai-US-1-Sn	TGTGGCCCAA	TGCCACC GTG	CAAGAGGGC	AGCTGGTGAA	CCTGACCTGC	
					
	3460	3470	3480	3490	3500	
refSn	CTTGTATGGA	CCACGCACCT	GGCCCAGCTC	ACCTACACGT	GGTACCGAGA	
Thai-HP-1-Sn	CTTGTATGGA	CCACGCACCT	GGCCCAGCTC	ACCTACACGT	GGTACCGAGA	
Thai-EU-1-Sn	CTTGTATGGA	CCACGCACCT	GGCCCAGCTC	ACCTACACGT	GGTACCGAGA	
Thai-US-1-Sn	CTTGTATGGA	CCACGCACCT	GGCCCAGCTC	ACCTACACGT	GGTACCGAGA	
					
	3510	3520	3530	3540	3550	
refSn	CCAGCAGCAG	CTCCCAGGTG	CTGCCCACTC	CATCCTCCTG	CCCAATGTCA	
Thai-HP-1-Sn	CCAGCAGCAG	CTCCCAGGTG	CTGCCCACTC	CATCCTCCTG	CCCAATGTCA	
Thai-EU-1-Sn	CCAGCAGCAG	CTCCCAGGTG	CTGCCCACTC	CATCCTCCTG	CCCAATGTCA	
Thai-US-1-Sn	CCAGCAGCAG	CTCCCAGGTG	CTGCCCACTC	CATCCTCCTG	CCCAATGTCA	
					
	3560	3570	3580	3590	3600	
refSn	CTGTCACAGA	TGCCGCCTCC	TACCGCTGTG	GCATATTGAT	CCCTGCCAG	
Thai-HP-1-Sn	CTGTCACAGA	TGCCGCCTCC	TACCGCTGTG	GCATATTGAT	CCCTGCCAG	
Thai-EU-1-Sn	CTGTCACAGA	TGCCGCCTCC	TACCGCTGTG	GCATATTGAT	CCCTGCCAG	
Thai-US-1-Sn	CTGTCACAGA	TGCCGCCTCC	TACCGCTGTG	GCATATTGAT	CCCTGCCAG	

										
	3610	3620	3630	3640	3650						
refSn	GCACCTCCGCC	TCTCCAGACC	TGTGCCCTG	GATGTCCTCT	ACGCACCCCG						
Thai-HP-1-Sn	GCACCTCCGCC	TCTCCAGACC	TGTGCCCTG	GATGTCCTCT	ACGCACCCCG						
Thai-EU-1-Sn	GCACCTCCGCC	TCTCCAGACC	TGTGCCCTG	GATGTCCTCT	ACGCACCCCG						
Thai-US-1-Sn	GCACCTCCGCC	TCTCCAGACC	TGTGCCCTG	GATGTCCTCT	ACGCACCCCG						
										
	3660	3670	3680	3690	3700						
refSn	CAGACTGCGC	CTGACCCATC	TCTTGGAGAG	CCGTGGTGGG	CAGCTGGCCG						
Thai-HP-1-Sn	CAGACTGCGC	CTGACCCATC	TCTTGGAGAG	CCGTGGTGGG	CAGCTGGCCG						
Thai-EU-1-Sn	CAGACTGCGC	CTGACCCATC	TCTTGGAGAG	CCGTGGTGGG	CAGCTGGCCG						
Thai-US-1-Sn	CAGACTGCGC	CTGACCCATC	TCTTGGAGAG	CCGTGGTGGG	CAGCTGGCCG						
										
	3710	3720	3730	3740	3750						
refSn	TGGTGCTGTG	CACTGTGGAC	AGTCGCCAG	CTGCCAGCT	GACCCTCAGC						
Thai-HP-1-Sn	TGGTGCTGTG	CACTGTGGAC	AGTCGCCAG	CTGCCAGCT	GACCCTCAGC						
Thai-EU-1-Sn	TGGTGCTGTG	CACTGTGGAC	AGTCGCCAG	CTGCCAGCT	GACCCTCAGC						
Thai-US-1-Sn	TGGTGCTGTG	CACTGTGGAC	AGTCGCCAG	CTGCCAGCT	GACCCTCAGC						
										
	3760	3770	3780	3790	3800						
refSn	CATGCTGGCC	GCCTCCTGGC	CTCCTCAACC	GCAGCCTCTG	TCCCCAACAC						
Thai-HP-1-Sn	CATGCTGGCC	GCCTCCTGGC	CTCCTCAACC	GCAGCCTCTG	TCCCCAACAC						
Thai-EU-1-Sn	CATGCTGGCC	GCCTCCTGGC	CTCCTCAACC	GCAGCCTCTG	TCCCCAACAC						
Thai-US-1-Sn	CATGCTGGCC	GCCTCCTGGC	CTCCTCAACC	GCAGCCTCTG	TCCCCAACAC						
										
	3810	3820	3830	3840	3850						
refSn	CCTGCGCTG	GAGCTGTGGG	AGCCCCGGCC	CAGTGATGAG	GGTCTCTACA						
Thai-HP-1-Sn	CCTGCGCTG	GAGCTGTGGG	AGCCCCGGCC	CAGTGATGAG	GGTCTCTACA						
Thai-EU-1-Sn	CCTGCGCTG	GAGCTGTGGG	AGCCCCGGCC	CAGTGATGAG	GGTCTCTACA						
Thai-US-1-Sn	CCTGCGCTG	GAGCTGTGGG	AGCCCCGGCC	CAGTGATGAG	GGTCTCTACA						
										
	3860	3870	3880	3890	3900						
refSn	GCTGCTCGGC	CCGCAGTCCT	CTGGGCCAGG	CCAACACATC	CCTGGAGCTG						
Thai-HP-1-Sn	GCTGCTCGGC	CCGCAGTCCT	CTGGGCCAGG	CCAACACATC	CCTGGAGCTG						
Thai-EU-1-Sn	GCTGCTCGGC	CCGCAGTCCT	CTGGGCCAGG	CCAACACATC	CCTGGAGCTG						
Thai-US-1-Sn	GCTGCTCGGC	CCGCAGTCCT	CTGGGCCAGG	CCAACACATC	CCTGGAGCTG						
										
	3910	3920	3930	3940	3950						
refSn	CGGCTAGAGG	GCGTGCAGGT	GGCACTGGCT	CCATCGGCCA	CTGTGCCGGA						
Thai-HP-1-Sn	CGGCTAGAGG	GCGTGCAGGT	GGCACTGGCT	CCATCGGCCA	CTGTGCCGGA						
Thai-EU-1-Sn	CGGCTAGAGG	GCGTGCAGGT	GGCACTGGCT	CCATCGGCCA	CTGTGCCGGA						
Thai-US-1-Sn	CGGCTAGAGG	GCGTGCAGGT	GGCACTGGCT	CCATCGGCCA	CTGTGCCGGA						
										
	3960	3970	3980	3990	4000						
refSn	GGGGGCCCCCT	GTCACAGTGA	CCTGTGAAGA	CCCTGCTGCC	CGCCCACCCA						
Thai-HP-1-Sn	GGGGGCCCCCT	GTCACAGTGA	CCTGTGAAGA	CCCTGCTGCC	CGCCCACCCA						
Thai-EU-1-Sn	GGGGGCCCCCT	GTCACAGTGA	CCTGTGAAGA	CCCTGCTGCC	CGCCCACCCA						
Thai-US-1-Sn	GGGGGCCCCCT	GTCACAGTGA	CCTGTGAAGA	CCCTGCTGCC	CGCCCACCCA						

	4010	4020	4030	4040	4050
refSn	CTCTCTATGT CTGGTACCAAC AACAGCCGTT GGCTGCAGGA GGGGTCGGCT					
Thai-HP-1-Sn	CCCTCTATGT CTGGTACCAAC AACAGCCGTT GGCTGCAGGA GGGGTCGGCT					
Thai-EU-1-Sn	CCCTCTATGT CTGGTACCAAC AACAGCCGTT GGCTGCAGGA GGGGTCGGCT					
Thai-US-1-Sn	CCCTCTATGT CTGGTACCAAC AACAGCCGTT GGCTGCAGGA GGGGTCGGCT					
	4060	4070	4080	4090	4100
refSn	GCCTCCCTCT CGTTTCCAGC GGCTACACGG GCTCACCGGG GCGCCTATAC					
Thai-HP-1-Sn	GCCTCCCTCT CGTTTCCAGC GGCTACACGG GCTCACCGGG GCGCCTATAC					
Thai-EU-1-Sn	GCCTCCCTCT CGTTTCCAGC GGCTACACGG GCTCACCGGG GCGCCTATAC					
Thai-US-1-Sn	GCCTCCCTCT CGTTTCCAGC GGCTACACGG GCTCACCGGG GCGCCTATAC					
	4110	4120	4130	4140	4150
refSn	CTGCCAGGTC CAGGATGCC AGGGCACACG CATCTCCCAG CCCGCAGCAC					
Thai-HP-1-Sn	CTGCCAGGTC CAGGATGCC AGGGCACACG CATCTCCCAG CCCGCAGCAC					
Thai-EU-1-Sn	CTGCCAGGTC CAGGATGCC AGGGCACACG CATCTCCCAG CCCGCAGCAC					
Thai-US-1-Sn	CTGCCAGGTC CAGGATGCC AGGGCACACG CATCTCCCAG CCCGCAGCAC					
	4160	4170	4180	4190	4200
refSn	TGCACATCCT CTATGCCCT CGGGATGCTG TCCTTTCCCTC CTTCTGGGAC					
Thai-HP-1-Sn	TGCACATCCT CTATGCCCT CGGGATGCTG TCCTTTCCCTC CTTCTGGGAC					
Thai-EU-1-Sn	TGCACATCCT CTATGCCCT CGGGATGCTG TCCTTTCCCTC CTTCTGGGAC					
Thai-US-1-Sn	TGCACATCCT CTATGCCCT CGGGATGCTG TCCTTTCCCTC CTTCTGGGAC					
	4210	4220	4230	4240	4250
refSn	TCAAGGGCCA GCCCTATGGC CGTGGTACAG TGCACTGTGG ACAGCGAGCC					
Thai-HP-1-Sn	TCAAGGGCCA GCCCTATGGC CGTGGTACAG TGCACTGTGG ACAGCGAGCC					
Thai-EU-1-Sn	TCAAGGGCCA GCCCTATGGC CGTGGTACAG TGCACTGTGG ACAGCGAGCC					
Thai-US-1-Sn	TCAAGGGCCA GCCCTATGGC CGTGGTACAG TGCACTGTGG ACAGCGAGCC					
	4260	4270	4280	4290	4300
refSn	ACCTGCCGAG ATGACCCTGT CCCATGATGG CAAGGTGCTG GCCACCAGCC					
Thai-HP-1-Sn	ACCTGCCGAG ATGACCCTGT CCCATGATGG CAAGGTGCTG GCCACCAGCC					
Thai-EU-1-Sn	ACCTGCCGAG ATGACCCTGT CCCATGATGG CAAGGTGCTG GCCACCAGCC					
Thai-US-1-Sn	ACCTGCCGAG ATGACCCTGT CCCATGATGG CAAGGTGCTG GCCACCAGCC					
	4310	4320	4330	4340	4350
refSn	ATGGGGTCCA CGGCTTAGCA GTGGGGACAG GCCATGTCCA GGTGGCCCGC					
Thai-HP-1-Sn	ATGGGGTCCA CGGCTTAGCA GTGGGGACAG GCCATGTCCA GGTGGCCCGC					
Thai-EU-1-Sn	ATGGGGTCCA CGGCTTAGCA GTGGGGACAG GCCATGTCCA GGTGGCCCGC					
Thai-US-1-Sn	ATGGGGTCCA CGGCTTAGCA GTGGGGACAG GCCATGTCCA GGTGGCCCGC					
	4360	4370	4380	4390	4400
refSn	AACGCCCTGC AGCTCGGGGT GCAGAATGTG CCCTCACGTG ACAAGGACAC					
Thai-HP-1-Sn	AACGCCCTGC AGCTCGGGGT GCAGAATGTG CCCTCACGTG ACAAGGACAC					
Thai-EU-1-Sn	AACGCCCTGC AGCTCGGGGT GCAGAATGTG CCCTCACGTG ACAAGGACAC					
Thai-US-1-Sn	AACGCCCTGC AGCTCGGGGT GCAGAATGTG CCCTCACGTG ACAAGGACAC					

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 4410 4420 4430 4440 4450

refSn CTACGTCTGC ATGGACCGCA ACTCCT^TGGG CTCAGTCAGC ACCATGGGC
Thai-HP-1-Sn CTACGTCTGC ATGGACCGCA ACTCCT^TGGG CTCAGTCAGC ACCATGGGC
Thai-EU-1-Sn CTACGTCTGC ATGGACCGCA ACTCCT^TGGG CTCAGTCAGC ACCATGGGC
Thai-US-1-Sn CTACGTCTGC ATGGACCGCA ACTCCT^TGGG CTCAGTCAGC ACCATGGGC

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 4460 4470 4480 4490 4500

refSn AGCTGCAGCC AGAAGGTGTG CACGTGGTAG CTGAGCCAGG GCTGGATGTG
Thai-HP-1-Sn AGCTGCAGCC AGAAGGTGTG CACGTGGTAG CTGAGCCAGG GCTGGATGTG
Thai-EU-1-Sn AGCTGCAGCC AGAAGGTGTG CACGTGGTAG CTGAGCCAGG GCTGGATGTG
Thai-US-1-Sn AGCTGCAGCC AGAAGGTGTG CACGTGGTAG CTGAGCCAGG GCTGGATGTG

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 4510 4520 4530 4540 4550

refSn CCTGAAGGCA CAGCGCTGAA CCTGAGCTGT CGCCTCCCTA GTGGCCCTGG
Thai-HP-1-Sn CCTGAAGGCA CAGCGCTGAA CCTGAGCTGT CGCCTCCCTA GTGGCCCTGG
Thai-EU-1-Sn CCTGAAGGCA CAGCGCTGAA CCTGAGCTGT CGCCTCCCTA GTGGCCCTGG
Thai-US-1-Sn CCTGAAGGCA CAGCGCTGAA CCTGAGCTGT CGCCTCCCTA GTGGCCCTGG

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 4560 4570 4580 4590 4600

refSn GCACATAGGC AACTCCACCT TTGCTTGGTT CCGGAACGGT CGGCAGCTAC
Thai-HP-1-Sn GCACATAGGC AACTCCACCT TTGCTTGGTT CCGGAACGGT CGGCAGCTAC
Thai-EU-1-Sn GCACATAGGC AACTCCACCT TTGCTTGGTT CCGGAACGGT CGGCAGCTAC
Thai-US-1-Sn GCACATAGGC AACTCCACCT TTGCTTGGTT CCGGAACGGT CGGCAGCTAC

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 4610 4620 4630 4640 4650

refSn ACACAGAGTC TGTGCCCA^G CTTACCTTCA CCCATGTGGC CGCGCCCCAA
Thai-HP-1-Sn ACACAGAGTC TGTGCCCA^G CTTACCTTCA CCCATGTGGC CGCGCCCCAA
Thai-EU-1-Sn ACACAGAGTC TGTGCCCA^G CTTACCTTCA CCCATGTGGC CGCGCCCCAA
Thai-US-1-Sn ACACAGAGTC TGTGCCCA^G CTTACCTTCA CCCATGTGGC CGCGCCCCAA

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 4660 4670 4680 4690 4700

refSn GCTGGCTTGT ACCACTGCCA GGCTGAGCTC CCCGCCGGGG CTGCCACCTC
Thai-HP-1-Sn GCTGGCTTGT ACCACTGCCA GGCTGAGCTC CCCGCCGGGG CTGCCACCTC
Thai-EU-1-Sn GCTGGCTTGT ACCACTGCCA GGCTGAGCTC CCCGCCGGGG CTGCCACCTC
Thai-US-1-Sn GCTGGCTTGT ACCACTGCCA GGCTGAGCTC CCCGCCGGGG CTGCCACCTC

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 4710 4720 4730 4740 4750

refSn TGCTCCAGTC TTGCTCCGGG TGCTCTACCC TCCCAAGACG CCCACCATGA
Thai-HP-1-Sn TGCTCCAGTC TTGCTCCGGG TGCTCTACCC TCCCAAGACG CCCACCATGA
Thai-EU-1-Sn TGCTCCAGTC TTGCTCCGGG TGCTCTACCC TCCCAAGACG CCCACCATGA
Thai-US-1-Sn TGCTCCAGTC TTGCTCCGGG TGCTCTACCC TCCCAAGACG CCCACCATGA

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 4760 4770 4780 4790 4800

refSn CTGTTTTGT GGAGCCCGAG GGTGGCATCC AGGGCATTCT GGACTGCCGA
Thai-HP-1-Sn CTGTTTTGT GGAGCCCGAG GGTGGCATCC AGGGCATTCT GGACTGCCGA
Thai-EU-1-Sn CTGTTTTGT GGAGCCCGAG GGTGGCATCC AGGGCATTCT GGACTGCCGA
Thai-US-1-Sn CTGTTTTGT GGAGCCCGAG GGTGGCATCC AGGGCATTCT GGACTGCCGA

	4810 4820 4830 4840 4850
refSn	GTGGACAGTG AGCCCCTAGC CAGCCTGACC CTCCACCTGG GCAGTCGGCT
Thai-HP-1-Sn	GTGGACAGTG AGCCCCTAGC CAGCCTGACC CTCCACCTGG GCAGTCGGCT
Thai-EU-1-Sn	GTGGACAGTG AGCCCCTAGC CAGCCTGACC CTCCACCTGG GCAGTCGGCT
Thai-US-1-Sn	GTGGACAGTG AGCCCCTAGC CAGCCTGACC CTCCACCTGG GCAGTCGGCT

	4860 4870 4880 4890 4900
refSn	GGTGGCCTCC AGCCAGCCTC AGGCTGCCCG TGCCAAGCCG CACATCCGCG
Thai-HP-1-Sn	GGTGGCCTCC AGCCAGCCTC AGGCTGCCCG TGCCAAGCCG CACATCCGCG
Thai-EU-1-Sn	GGTGGCCTCC AGCCAGCCTC AGGCTGCCCG TGCCAAGCCG CACATCCGCG
Thai-US-1-Sn	GGTGGCCTCC AGCCAGCCTC AGGCTGCCCG TGCCAAGCCG CACATCCGCG

	4910 4920 4930 4940 4950
refSn	TCTCAGCCAG TCCCAATGCC TTGCGAGTGG ACATGGAGGA GCTGAAGCCC
Thai-HP-1-Sn	TCTCAGCCAG TCCCAATGCC TTGCGAGTGG ACATGGAGGA GCTGAAGCCC
Thai-EU-1-Sn	TCTCAGCCAG TCCCAATGCC TTGCGAGTGG ACATGGAGGA GCTGAAGCCC
Thai-US-1-Sn	TCTCAGCCAG TCCCAATGCC TTGCGAGTGG ACATGGAGGA GCTGAAGCCC

	4960 4970 4980 4990 5000
refSn	AGTGACCAGG GGGAGTATGT GTGCTCGGCC TCCAATGCCCG TGGGCTCTGC
Thai-HP-1-Sn	AGTGACCAGG GGGAGTATGT GTGCTCGGCC TCCAATGCCCG TGGGCTCTGC
Thai-EU-1-Sn	AGTGACCAGG GGGAGTATGT GTGCTCGGCC TCCAATGCCCG TGGGCTCTGC
Thai-US-1-Sn	AGTGACCAGG GGGAGTATGT GTGCTCGGCC TCCAATGCCCG TGGGCTCTGC

	5010 5020 5030 5040 5050
refSn	CTCTGCTGCC ACCTACTTCG GAACCAGAGC CCTGCATCGC CTGCATCTGT
Thai-HP-1-Sn	CTCTGCTGCC ACCTACTTCG GAACCAGAGC CCTGCATCGC CTGCATCTGT
Thai-EU-1-Sn	CTCTGCTGCC ACCTACTTCG GAACCAGAGC CCTGCATCGC CTGCATCTGT
Thai-US-1-Sn	CTCTGCTGCC ACCTACTTCG GAACCAGAGC CCTGCATCGC CTGCATCTGT

	5060 5070 5080 5090 5100
refSn	TCCAGCACCT TCTCTGGTTC CTGGGGCTGC TGGCGAGCCT CCTCTCCCTA
Thai-HP-1-Sn	TCCAGCACCT TCTCTGGTTC CTGGGGCTGC TGGCGAGCCT CCTCTCCCTA
Thai-EU-1-Sn	TCCAGCACCT TCTCTGGTTC CTGGGGCTGC TGGCGAGCCT CCTCTCCCTA
Thai-US-1-Sn	TCCAGCACCT TCTCTGGTTC CTGGGGCTGC TGGCGAGCCT CCTCTCCCTA

	5110 5120 5130 5140 5150
refSn	CTGTTGGGCC TGGGGGTCTG GTACGCCTGG AGACGGGGAA ATTTTACAA
Thai-HP-1-Sn	CTGTTGGGCC TGGGGGTCTG GTACGCCTGG AGACGGGGAA ATTTTACAA
Thai-EU-1-Sn	CTGTTGGGCC TGGGGGTCTG GTACGCCTGG AGACGGGGAA ATTTTACAA
Thai-US-1-Sn	CTGTTGGGCC TGGGGGTCTG GTACGCCTGG AGACGGGGAA ATTTTACAA

	5160 5170 5180 5190 5200
refSn	GCTGAGAATG GGCGAATATT CAGTAGAGAT GGTATCTCGG AAGGAAACCA
Thai-HP-1-Sn	GCTGAGAATG GGCGAATATT CAGTAGAGAT GGTATCTCGG AAGGAAACCA
Thai-EU-1-Sn	GCTGAGAATG GGCGAATATT CAGTAGAGAT GGTATCTCGG AAGGAAACCA
Thai-US-1-Sn	GCTGAGAATG GGCGAATATT CAGTAGAGAT GGTATCTCGG AAGGAAACCA

	5210 5220 5230 5240 5250
refSn	CGCAGATGTC CACTGACCAG GAAGAAGTTA CTGGAATCGG TGATGATGCG
Thai-HP-1-Sn	CGCAGATGTC CACTGACCAG GAAGAAGTTA CTGGAATCGG TGATGATGCG
Thai-EU-1-Sn	CGCAGATGTC CACTGACCAG GAAGAAGTTA CTGGAATCGG TGATGATGCG
Thai-US-1-Sn	CGCAGATGTC CACTGACCAG GAAGAAGTTA CTGGAATCGG TGATGATGCG

	5260 5270 5280 5290 5300
refSn	GGCTCTGTGA ACCAGGCAGGC ATTTGATCCT GCCCACCTCT GTGAAAACAC
Thai-HP-1-Sn	GGCTCTGTGA ACCAGGCAGGC ATTTGATCCT GCCCACCTCT GTGAAAACAC
Thai-EU-1-Sn	GGCTCTGTGA ACCAGGCAGGC ATTTGATCCT GCCCACCTCT GTGAAAACAC
Thai-US-1-Sn	GGCTCTGTGA ACCAGGCAGGC ATTTGATCCT GCCCACCTCT GTGAAAACAC

	5310 5320
refSn	ACAGTCTGTG AAAAGCACAG TCTGA
Thai-HP-1-Sn	ACAGTCTGTG AAAAGCACAG TCTGA
Thai-EU-1-Sn	ACAGTCTGTG AAAAGCACAG TCTGA
Thai-US-1-Sn	ACAGTCTGTG AAAAGCACAG TCTGA

Figure 4.9 Nucleotide sequence alignments of the reference Sn (refSn) (accession number is NM_214346) with recombinant plasmids of Thai-HP-1-pCR®-XL-TOPO®-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR®-XL-TOPO®-Sn (Thai-EU-1-Sn) and Thai-US-1-pCR®-XL-TOPO®-Sn (Thai-US-1-Sn), respectively. The nucleotide changes are boxed.

Table 4.4 Deduced amino acid sequences of 3 recombinant porcine Sn (full) plasmids

Transformant's name	Sequence
1. Thai-HP-1-Sn (1,730 aa)	MDFLLLLLASSALAGLASWTVSSPETVQGIKGSCLIIPCTFRFPANVEVP HGITAIWYYDYSRKRLVSHSRNPKVVENHFQGRALLGQVEQRTCSLLL KDLQPQDSGSYNFRFEISEGNRSDVKGTWVTTEVPSVPTIALPAKLHE GMEVDFNCSTPYVCPTEPVNLLQWQGQDPTRSVTSHLQKLEPSGTSHME TLHMAWSQDHGRILSCQVSAERRMQKEIHLQVQYAPKGVEILFSHSG RNVLPGDLVTLSCQVNNSNPQVSSVQWVKDGTLKDQKRVLQLRRAAW ADAGVYTCQAGNAVGSSVSPPVSLHVFMAEVQVSPVGSILENQTVTLAC NTPKEAPSELRYSWYKNHALLEGSHSRTLRLHSVTRADSGFYFCEVQNA RGRERSPPVSVVSHPPLTPDLTAFLETQAGLVGILQCSVSEPPATLVL HGLLILASTSGEGDHSPRFSVASAPNSLRLEIQDLGPTDSGEYMCASSS LGNASSTLDFHANAARLLISPAAEVVEGQAVTLSCRSSLMPDTRFSWY LNGALILEGPSSLLPAASSTDAGSYHCRAQNSHSTSGPSSPAVLTVY APRQPVFTAQLDPDTAGAGAGRQGLLCRVSDPPAQLQLLHRGRVVA SSLSWGGGCCTCGCFHRMKVTKAPNLLRVEIRDPVLEDEGVYLCEAS SALGNASASATLDAQATVLVITPSHTLQEGIEANLTCNVSREASGPANFS WFRDGALWAQGPLDTVLLPVARTDAALYACRIVTEAGAGLSTPVALNV LYPPDPPKLSALLDVDQGHTAVFVCTVDSRPLAQLALFRGEHLLAASSAL RLPPRGRQLQAKASANSQLEVRDLSLGDSGYHCEATNILGSANTSLTF QVRGAWVRVSPSPELQEQQAVLSCQVPIGVLEGTSYRWYRDQQQLPGAAHSIL STSATLRFAAITLSQAGAYHCQAQAPGSATTDLAAPVSLHVTYAPRQATL TTLMDSGLGLRGLLLCRVNSDPPAQLRLLHGSRLVASTLQGVEELAGSS PRLQVATAPNTLRLEIHNAVLEDEGVYTCEATNTLGQTLASAADFQAMR VQWPNAUTQEGQLVNLTCLVWTTTHLAQLTYTWYRDQQQLPGAAHSIL LPNVTVTDAASYRCGILIPGQALRLSRPVALDVLYAPRRLRLTHLLESRGG QLAVVLCTVDSRPAAQLTLSHAGRLLASSTAASVPNTRLELWEPRPSDE

GLYSCSARSPLGQANTSLELRLEGVQVALAPSATVPEGAPVTCEDPAA
 RPPTLYVWYHNSRWLQEGLSAASLSFPAATRAHAGAYTCQVQDAQGTRI
 SQPAALHILYAPRDAVLSSFWDSRASPMAVVQCTVDSEPPAEMTLSHDG
 KVLATSHGVHGLAVGTGHVQVARNALQLRVQNVPSPRKDTYVCMDRNS
 SGSVSTMGQLQPEGVHVVAEPGLDVPEGTALNLSCRPLSGPGHIGNSTF
 AWFRNGRQLHTESVPTLTFTHVARAQAGLYHCQAELPAGAATSAPVLLR
 VLYPPKTPTMTVFVEPEGGIQGILDCRVDSEPLASLTHLGSRLVASSQPQ
 AAPAKPHIRVSASNALRVDMEELKPSDQGEYVCSASNALGSASAATYF
 GTRALHRLHLFQHLLWFLGLLASLLFLLLGLGVWYAWRRGNFYKLRMGE
 YSVEVMVRKETTQMSTDQEEVTGIGDDAGSVNQAAFDPAHLCENTQSV
 KSTV

2. Thai-EU-1- MDFLLLLLLASSALAGLASWTVSSPETVQGIKGSCIIPCTFGFPANVEVP
 Sn (1,730 aa) HGITAIWYYDYSRKRLVSHSRNPKVVENHFQGRALLLGQVEQRTCSLLL
 KDLQPQDSGSYNFRFEISEGNRWSDVKGTVVTVTEVPSVPTIALPAKLHE
 GMEVDFNCSTPYVCPTEPVNLQWQGQDPTRSVTSHLQKLEPSGTSHME
 TLHMAISWQDHGRILSCQVSAERRMQKEIHLQVQYAPKGVEILFSHSG
 RNVLPGDLVTLSCQVNSSNPQVSSVQWVKDGTLKDQKRVQLRRAAW
 ADAGVYTCQAGNAVGSSVSPPVSLHVFMAEVQVSPVGSILENQTVTLAC
 NTPKEAPSELRYSWYKNHALLEGSHSRTLRLHSVTRADSGFYFCEVQNA
 RGRERSPPVSVVSHPPLPDLTAFLETQAGLVGILQCSVVSEPPATLVS
 HGGLILASTSGEGDHSPRFSVASAPNSLRLEIQDLGPTDSGEYMCSSSS
 LGNASSTLDFHANAARLLISPAAEVVEGQAVTLSRSSLSLMPDTRFSWY
 LNGALILEGPSSLLPAASSTDAGSYHCRAQNSHSTSGPSSPAVLTVLY
 APRQPVFTAQLDPDTAGAGAGRQGLLLCRVSDPDPQLQLLHRGRVVA
 SSLSWGGGCCTCGGFHRMKVTKAPNLLRVEIRDVLEDEGVYLCEAS
 SALGNASASATLDAQATVLVITPSHTLQEGIEANLTCNVSREASGPANFS
 WFRDGALWAQGPLDTVLLPVARTDAALYACRIVTEAGAGLSTPVALNV
 LYPPDPPKLSALLDVDQGHTAVFVCTVDSRPLAQLALFRGEHLLAASSAL

RLPPRGRQLQAKASANSQLEVRDLSLGDSGSYHCEATNILGSANTSLTF
 QVRGAWRVSPSPELQEGQAVLSCQVPIGVLEGTSYRWYRDGQPLQE
 STSATLRFAAITLSQAGAYHCQAQAPGSATTDLAAPVSLHVTYAPRQATL
 TTLMDSGLGRLLCRVNSDPPAQLRLHGSRLVASTLQGVEELAGSS
 PRLQVATAPNTLRLEIHNAVLEDEGVYTCEATNTLGQTLASAAFDAQAMR
 VQVWPNATVQEGQLVNLTCLVWTTHLAQLTYTWYRDQQQLPGAAHSIL
 LPNVTVTDAASYRCGILIPGQALRLSRPVALDVLYAPRRLRLTHLLESRGG
 QLAWLCTVDSRPAAQLTLSHAGRLLASSTAASVPNTLRELWEPRPSDE
 GLYSCSARSPLGQANTSLELRLEGVQUALAPSATVPEGAPVTCEDPAA
 RPPTLYVWYHNSRWLQEGSAASLSFPAATRAHAGAYTCQVQDAQGTRI
 SQPAALHILYAPRDAVLSSFWDSRASPMAVVQCTVDSEPPAEMTLSHDG
 KVLATSHGVHGLAVGTGHVQVARNALQLRVQNVPQRDKDTYVCMDRNS
 LGSVSTMGQLQPEGVHVVAEPGLDVPEGTALNLSCRPLSGPGHIGNSTF
 AWFRNGRQLHTESVPTLTFTHVARAQAGLYHCQAELPAGAATSAPVLLR
 VLYPPKTPTMTVFVEPEGGIQGILDCRVDSEPLASLTHLGSRLVASSQPQ
 AAPAKPHIRVSASNALRVDMEELKPSDQGEYVCSASNALGSASAATYF
 GTRALHRLHLFQHLLWFLGLLASLLGLGVWYAWRRGNFYKLRMGE
 YSVEMVSRKETTQMSTDQEEVTGIGDDAGSVNQAAFDPAHLCENTQSV
 KSTV

3. Thai-US-1-	MDFLLLLLASSALAGLASWTVSSPETVQGIKGSCIIPCTFRFPANVEVP
Sn (1,730 aa)	HGITAIWYYDYSRKRLVSHSRNPKVVENHFQGRALLGQVEQRTCSLLL KDLQPQDSGSYNFRFEISEGNRWSDVKGTVVTTEVPSVPTIALPAKLHE GMEVDFNCSTPYVCPTEPVNLQWQGQDPTRSVTSHLQKLEPSGTSHME TLHMAISWQDHGRILSCQVSAERRMQKEIHLQVQYAPKGVEILFSHSG RNVLPGDLVTLSCQVNSSNPQVSSVQWVKDGTKLKDKRVLQLRRAAW ADAGVYTCQAGNAVGSVSPPVSLHVFMAEVQVSPVGSILENQTVTLAC NTPKEAPSELRYSWYKNHALLEGSHSRTLRLHSVTRADSGFYFCEVQNA RGRERSPPVSVVSHPPLPDLTAFLETQAGLVGILQCSVVSEPPATLVLS

HGGLILASTSGEGDHSPRFSVASAPNSLRLEIQDLGPTDSGEYMCSSSS
LGNASSTLDFHANAARLLISPAAEVWEGQAVTLSRSSSLMPDTRFSWY
LNGALILEGPSSLLLPAASSTDAGSYHCRAQNSHSTSGPSSPAVLTVLY
APRQPVFTAQLDPDTAGAGAGRQGLLLCRVDSDPPAQLQLLHRGRVVA
SSLWGGGCCTCGGCFHRMKVTKAPNLLRVEIRDVLEDEGVYLCEAS
SALGNASASATLDAQATVLVITPSHTLQEGIEANLTCNVSREASGPANFS
WFRDGALWAQGPLDTVLLPVARTDAALYACRIVTEAGAGLSTPVALNV
LYPPDPPKLSALLDVDQGHTAVFVCTVDSRPLAQALFRGEHLLAASSAL
RLPPRGRQLQAKASANSLQLEVRDLSLGDSGSYHCEATNILGSANTSITF
QVRGAWRVSPSPELQEGQAWLSCQVPIGVLEGTSYRWYRDGQPLQE
STSATLRFAAITLSQAGAYHCQAQAPGSATTDLAAPVSLHVTYAPRQATL
TTLMDSGLGLRGLLLCRVNSDPPAQLRLLHGSRLVASTLQGVEELAGSS
PRLQVATAPNTLRLEIHNAVLEDEGVYTCEATNTLGQTLASAADFQAQAMR
VQWPNATVQEGQLVNLTCLVWTTHLAQLTYTWYRDQQQLPGAAHSIL
LPNVTVTDAAZYRCGILIPGQALRLSRPVALDVLYAPRRLRLTHLLESRGG
QLAVVLCTVDSRPAAQTLSHAGRLLASSTAASVPNTLRLELWEPRPSDE
GLYSCSARSPLGQANTSLELRLEGVQUALAPSATVPEGAPVTCEDPAA
RPPTLYVWYHNSRWLQEGSAASLSFPAATRAHAGAYTCQVQDAQGTRI
SQPAALHILYAPRDAVLSSFWDSRASPMAVVQCTVDSEPPAEMTLSHDG
KVLATSHGVHGLAVGTGHVQVARNALQLRVQNVPNSRDKDTYVCMDRNS
SGSVSTMGQLQPEGVHVVAEPGLDVPEGTALNLSCRPLSGPGHIGNSTF
AWFRNGRQLHTESVPTLTFTHVARAQAGLYHCQAELPAGAATSAPVLLR
VLYPPKTPTMTVFEPEGGIQGILDCRVDSEPLASLTLLHGSRLVASSQPQ
AAPAKPHIRVSASNALRVDMEELKPSDQGEYVCSASNALGSASAATYF
GTRALHRLHLFQHLLWFLGLLASLLFLLGLGVWYAWRRGNFYKLRMGE
YSVEMVSRKETTQMSTDQEEVTGIGDDAGSVNQAAFDPAHLCENTQSV
KSTV

	10 20 30 40 50
refSn	MDFLLLLLLL ASSALAGLAS WTVSSPETVQ GIKGSCLIIP CTEGF PANVE
Thai-HP-1-Sn	MDFLLLLLLL ASSALAGLAS WTVSSPETVQ GIKGSCLIIP CTERFPANVE
Thai-EU-1-Sn	MDFLLLLLLL ASSALAGLAS WTVSSPETVQ GIKGSCLIIP CTEGF PANVE
Thai-US-1-Sn	MDFLLLLLLL ASSALAGLAS WTVSSPETVQ GIKGSCLIIP CTERFPANVE

	60 70 80 90 100
refSn	VPHGITAIWY YDYSRKRLVV SHSRNPKVVE NHFQGRALLL GQVEQRTCSL
Thai-HP-1-Sn	VPHGITAIWY YDYSRKRLVV SHSRNPKVVE NHFQGRALLL GQVEQRTCSL
Thai-EU-1-Sn	VPHGITAIWY YDYSRKRLVV SHSRNPKVVE NHFQGRALLL GQVEQRTCSL
Thai-US-1-Sn	VPHGITAIWY YDYSRKRLVV SHSRNPKVVE NHFQGRALLL GQVEQRTCSL

	110 120 130 140 150
refSn	LLKDLQPQDS GSYNFRFEIS EGNRWSDVKG TVVTVTEVPS VPTIALPAKL
Thai-HP-1-Sn	LLKDLQPQDS GSYNFRFEIS EGNRWSDVKG TVVTVTEVPS VPTIALPAKL
Thai-EU-1-Sn	LLKDLQPQDS GSYNFRFEIS EGNRWSDVKG TVVTVTEVPS VPTIALPAKL
Thai-US-1-Sn	LLKDLQPQDS GSYNFRFEIS EGNRWSDVKG TVVTVTEVPS VPTIALPAKL

	160 170 180 190 200
refSn	HEGMEVDFNC STPYVCPTEP VNLQWQGQDP TRSVTSHLQK LEPSGTSHME
Thai-HP-1-Sn	HEGMEVDFNC STPYVCPTEP VNLQWQGQDP TRSVTSHLQK LEPSGTSHME
Thai-EU-1-Sn	HEGMEVDFNC STPYVCPTEP VNLQWQGQDP TRSVTSHLQK LEPSGTSHME
Thai-US-1-Sn	HEGMEVDFNC STPYVCPTEP VNLQWQGQDP TRSVTSHLQK LEPSGTSHME

	210 220 230 240 250
refSn	TLHMALSWQD HGRILSCQVS AAERRMQKEI HLQVQYAPKG VEILFSHSGR
Thai-HP-1-Sn	TLHMALSWQD HGRILSCQVS AAERRMQKEI HLQVQYAPKG VEILFSHSGR
Thai-EU-1-Sn	TLHMALSWQD HGRILSCQVS AAERRMQKEI HLQVQYAPKG VEILFSHSGR
Thai-US-1-Sn	TLHMALSWQD HGRILSCQVS AAERRMQKEI HLQVQYAPKG VEILFSHSGR

	260 270 280 290 300
refSn	NVLPGDLVTL SCQVNSSNPQ VSSVQWVKDG TKLKDKQKRVL QLRRAAWADA
Thai-HP-1-Sn	NVLPGDLVTL SCQVNSSNPQ VSSVQWVKDG TKLKDKQKRVL QLRRAAWADA
Thai-EU-1-Sn	NVLPGDLVTL SCQVNSSNPQ VSSVQWVKDG TKLKDKQKRVL QLRRAAWADA
Thai-US-1-Sn	NVLPGDLVTL SCQVNSSNPQ VSSVQWVKDG TKLKDKQKRVL QLRRAAWADA

	310 320 330 340 350
refSn	GVYTCQAGNA VGSSVSPPVS LHVFMAEVQV SPVGSILENQ TVTLACNTPK
Thai-HP-1-Sn	GVYTCQAGNA VGSSVSPPVS LHVFMAEVQV SPVGSILENQ TVTLACNTPK
Thai-EU-1-Sn	GVYTCQAGNA VGSSVSPPVS LHVFMAEVQV SPVGSILENQ TVTLACNTPK
Thai-US-1-Sn	GVYTCQAGNA VGSSVSPPVS LHVFMAEVQV SPVGSILENQ TVTLACNTPK

	360 370 380 390 400
refSn	EAPSELRYSW YKNHALLEGS HSRTLRLHSV TRADSGFYFC EVQNARGRER
Thai-HP-1-Sn	EAPSELRYSW YKNHALLEGS HSRTLRLHSV TRADSGFYFC EVQNARGRER
Thai-EU-1-Sn	EAPSELRYSW YKNHALLEGS HSRTLRLHSV TRADSGFYFC EVQNARGRER
Thai-US-1-Sn	EAPSELRYSW YKNHALLEGS HSRTLRLHSV TRADSGFYFC EVQNARGRER

	410 420 430 440 450
refSn	SPPVSVVVSH PPLTPDLTAF LETQAGLVGI LQCSVVSEPP ATLVLSHGGL
Thai-HP-1-Sn	SPPVSVVVSH PPLTPDLTAF LETQAGLVGI LQCSVVSEPP ATLVLSHGGL
Thai-EU-1-Sn	SPPVSVVVSH PPLTPDLTAF LETQAGLVGI LQCSVVSEPP ATLVLSHGGL
Thai-US-1-Sn	SPPVSVVVSH PPLTPDLTAF LETQAGLVGI LQCSVVSEPP ATLVLSHGGL

	460 470 480 490 500
refSn	ILASTSGEGD HSPRFSVASA PNSLRLEIQR LGPTDSGEYM CSASSSLGNA
Thai-HP-1-Sn	ILASTSGEGD HSPRFSVASA PNSLRLEIQR LGPTDSGEYM CSASSSLGNA
Thai-EU-1-Sn	ILASTSGEGD HSPRFSVASA PNSLRLEIQR LGPTDSGEYM CSASSSLGNA
Thai-US-1-Sn	ILASTSGEGD HSPRFSVASA PNSLRLEIQR LGPTDSGEYM CSASSSLGNA

	510 520 530 540 550
refSn	SSTLDFHANA ARLLISPAAE VVEGQAVTLS CRSSLSLMPD TRFSWYLNNGA
Thai-HP-1-Sn	SSTLDFHANA ARLLISPAAE VVEGQAVTLS CRSSLSLMPD TRFSWYLNNGA
Thai-EU-1-Sn	SSTLDFHANA ARLLISPAAE VVEGQAVTLS CRSSLSLMPD TRFSWYLNNGA
Thai-US-1-Sn	SSTLDFHANA ARLLISPAAE VVEGQAVTLS CRSSLSLMPD TRFSWYLNNGA

	560 570 580 590 600
refSn	LILEGPSSSL LLPAASSTD A GSYHCRAQNS HSTSGPSSPA VLTVLYAPRQ
Thai-HP-1-Sn	LILEGPSSSL LLPAASSTD A GSYHCRAQNS HSTSGPSSPA VLTVLYAPRQ
Thai-EU-1-Sn	LILEGPSSSL LLPAASSTD A GSYHCRAQNS HSTSGPSSPA VLTVLYAPRQ
Thai-US-1-Sn	LILEGPSSSL LLPAASSTD A GSYHCRAQNS HSTSGPSSPA VLTVLYAPRQ

	610 620 630 640 650
refSn	PVFTAQLDPD TAGAGAGRQG LLLCRVDSDP PAQLQLLHRG RVVASSLSWG
Thai-HP-1-Sn	PVFTAQLDPD TAGAGAGRQG LLLCRVDSDP PAQLQLLHRG RVVASSLSWG
Thai-EU-1-Sn	PVFTAQLDPD TAGAGAGRQG LLLCRVDSDP PAQLQLLHRG RVVASSLSWG
Thai-US-1-Sn	PVFTAQLDPD TAGAGAGRQG LLLCRVDSDP PAQLQLLHRG RVVASSLSWG

	660 670 680 690 700
refSn	GGCCTCGGCF HRMKVTKAPN LLRVEIRDPV LEDEGVYLCE ASSALGNASA
Thai-HP-1-Sn	GGCCTCGGCF HRMKVTKAPN LLRVEIRDPV LEDEGVYLCE ASSALGNASA
Thai-EU-1-Sn	GGCCTCGGCF HRMKVTKAPN LLRVEIRDPV LEDEGVYLCE ASSALGNASA
Thai-US-1-Sn	GGCCTCGGCF HRMKVTKAPN LLRVEIRDPV LEDEGVYLCE ASSALGNASA

	710 720 730 740 750
refSn	SATLDAQATV LVITPSHTLQ EGIEANLTCN VSREASGPAN FSWFRDGALW
Thai-HP-1-Sn	SATLDAQATV LVITPSHTLQ EGIEANLTCN VSREASGPAN FSWFRDGALW
Thai-EU-1-Sn	SATLDAQATV LVITPSHTLQ EGIEANLTCN VSREASGPAN FSWFRDGALW
Thai-US-1-Sn	SATLDAQATV LVITPSHTLQ EGIEANLTCN VSREASGPAN FSWFRDGALW

	760 770 780 790 800
refSn	AQGPLDTVTL LPVARTDAAL YACRIVTEAG AGLSTPVALN VLYPPDPK
Thai-HP-1-Sn	AQGPLDTVTL LPVARTDAAL YACRIVTEAG AGLSTPVALN VLYPPDPK
Thai-EU-1-Sn	AQGPLDTVTL LPVARTDAAL YACRIVTEAG AGLSTPVALN VLYPPDPK
Thai-US-1-Sn	AQGPLDTVTL LPVARTDAAL YACRIVTEAG AGLSTPVALN VLYPPDPK

	810 820 830 840 850
refSn	SALLDVDQGH TAVFVCTVDS RPLAQLALFR GEHLLAASSA LRLPPRGRLQ
Thai-HP-1-Sn	SALLDVDQGH TAVFVCTVDS RPLAQLALFR GEHLLAASSA LRLPPRGRLQ
Thai-EU-1-Sn	SALLDVDQGH TAVFVCTVDS RPLAQLALFR GEHLLAASSA LRLPPRGRLQ
Thai-US-1-Sn	SALLDVDQGH TAVFVCTVDS RPLAQLALFR GEHLLAASSA LRLPPRGRLQ

	860 870 880 890 900
refSn	AKASANSLQL EVRDLSLGDS GSYHCEATNI LGSANTSITF QVRGAWSRVS
Thai-HP-1-Sn	AKASANSLQL EVRDLSLGDS GSYHCEATNI LGSANTSITF QVRGAWSRVS
Thai-EU-1-Sn	AKASANSLQL EVRDLSLGDS GSYHCEATNI LGSANTSITF QVRGAWSRVS
Thai-US-1-Sn	AKASANSLQL EVRDLSLGDS GSYHCEATNI LGSANTSITF QVRGAWSRVS

	910 920 930 940 950
refSn	PSPELQEGQA VVLSCQVPIG VLEGTSYRWY RDGQPLQEST SATLRFAAIT
Thai-HP-1-Sn	PSPELQEGQA VVLSCQVPIG VLEGTSYRWY RDGQPLQEST SATLRFAAIT
Thai-EU-1-Sn	PSPELQEGQA VVLSCQVPIG VLEGTSYRWY RDGQPLQEST SATLRFAAIT
Thai-US-1-Sn	PSPELQEGQA VVLSCQVPIG VLEGTSYRWY RDGQPLQEST SATLRFAAIT

	960 970 980 990 1000
refSn	LSQAGAYHCQ AQAPGSATTD LAAPVSLHVT YAPRQATLTT LMDSGLGRGLG
Thai-HP-1-Sn	LSQAGAYHCQ AQAPGSATTD LAAPVSLHVT YAPRQATLTT LMDSGLGRGLG
Thai-EU-1-Sn	LSQAGAYHCQ AQAPGSATTD LAAPVSLHVT YAPRQATLTT LMDSGLGRGLG
Thai-US-1-Sn	LSQAGAYHCQ AQAPGSATTD LAAPVSLHVT YAPRQATLTT LMDSGLGRGLG

	1010 1020 1030 1040 1050
refSn	LLLCRVNSDP PAQLRLLHGS RLVASTLQGV EELAGSSPRL QVATAPNTR
Thai-HP-1-Sn	LLLCRVNSDP PAQLRLLHGS RLVASTLQGV EELAGSSPRL QVATAPNTR
Thai-EU-1-Sn	LLLCRVNSDP PAQLRLLHGS RLVASTLQGV EELAGSSPRL QVATAPNTR
Thai-US-1-Sn	LLLCRVNSDP PAQLRLLHGS RLVASTLQGV EELAGSSPRL QVATAPNTR

	1060 1070 1080 1090 1100
refSn	LEIHNAVLED EGVYTCEATN TLGQTLASAA FDAQAMRVQV WPNATVQEGQ
Thai-HP-1-Sn	LEIHNAVLED EGVYTCEATN TLGQTLASAA FDAQAMRVQV WPNATVQEGQ
Thai-EU-1-Sn	LEIHNAVLED EGVYTCEATN TLGQTLASAA FDAQAMRVQV WPNATVQEGQ
Thai-US-1-Sn	LEIHNAVLED EGVYTCEATN TLGQTLASAA FDAQAMRVQV WPNATVQEGQ

	1110 1120 1130 1140 1150
refSn	LVNLTCLVWT THLAQLTYTW YRDQQQLPGA AHSILLPNVT VTDAASYRCG
Thai-HP-1-Sn	LVNLTCLVWT THLAQLTYTW YRDQQQLPGA AHSILLPNVT VTDAASYRCG
Thai-EU-1-Sn	LVNLTCLVWT THLAQLTYTW YRDQQQLPGA AHSILLPNVT VTDAASYRCG
Thai-US-1-Sn	LVNLTCLVWT THLAQLTYTW YRDQQQLPGA AHSILLPNVT VTDAASYRCG

	1160 1170 1180 1190 1200
refSn	ILIPGQALRL SRPVALDVLY APRRLRLTHL LESRGQQLAV VLCTVDSRPA
Thai-HP-1-Sn	ILIPGQALRL SRPVALDVLY APRRLRLTHL LESRGQQLAV VLCTVDSRPA
Thai-EU-1-Sn	ILIPGQALRL SRPVALDVLY APRRLRLTHL LESRGQQLAV VLCTVDSRPA
Thai-US-1-Sn	ILIPGQALRL SRPVALDVLY APRRLRLTHL LESRGQQLAV VLCTVDSRPA

	1210 1220 1230 1240 1250
refSn	AQLTLSHAGR LLASSTAASV PNTLRLELWE PRPSDEGLYS CSARSPLGQA
Thai-HP-1-Sn	AQLTLSHAGR LLASSTAASV PNTLRLELWE PRPSDEGLYS CSARSPLGQA
Thai-EU-1-Sn	AQLTLSHAGR LLASSTAASV PNTLRLELWE PRPSDEGLYS CSARSPLGQA
Thai-US-1-Sn	AQLTLSHAGR LLASSTAASV PNTLRLELWE PRPSDEGLYS CSARSPLGQA

	1260 1270 1280 1290 1300
refSn	NTSLELRLEG VQVALAPSAT VPEGAPVTVT CEDPAARPPT LYVWYHNSRW
Thai-HP-1-Sn	NTSLELRLEG VQVALAPSAT VPEGAPVTVT CEDPAARPPT LYVWYHNSRW
Thai-EU-1-Sn	NTSLELRLEG VQVALAPSAT VPEGAPVTVT CEDPAARPPT LYVWYHNSRW
Thai-US-1-Sn	NTSLELRLEG VQVALAPSAT VPEGAPVTVT CEDPAARPPT LYVWYHNSRW

	1310 1320 1330 1340 1350
refSn	LQEGSAASLS FPAATRAHAG AYTCQVQDAQ GTRISQPAAL HILYAPRDAV
Thai-HP-1-Sn	LQEGSAASLS FPAATRAHAG AYTCQVQDAQ GTRISQPAAL HILYAPRDAV
Thai-EU-1-Sn	LQEGSAASLS FPAATRAHAG AYTCQVQDAQ GTRISQPAAL HILYAPRDAV
Thai-US-1-Sn	LQEGSAASLS FPAATRAHAG AYTCQVQDAQ GTRISQPAAL HILYAPRDAV

	1360 1370 1380 1390 1400
refSn	LSSFWDSRAS PMAVVQCTVD SEPPAEMTLS HDGKVLATSH GVHGLAVGTG
Thai-HP-1-Sn	LSSFWDSRAS PMAVVQCTVD SEPPAEMTLS HDGKVLATSH GVHGLAVGTG
Thai-EU-1-Sn	LSSFWDSRAS PMAVVQCTVD SEPPAEMTLS HDGKVLATSH GVHGLAVGTG
Thai-US-1-Sn	LSSFWDSRAS PMAVVQCTVD SEPPAEMTLS HDGKVLATSH GVHGLAVGTG

	1410 1420 1430 1440 1450
refSn	HVQVARNALQ LRVQNVPSPRD KDTYVCMDRN SLGSVSTMHQ LQPEGHVVA
Thai-HP-1-Sn	HVQVARNALQ LRVQNVPSPRD KDTYVCMDRN SSGSVSTMHQ LQPEGHVVA
Thai-EU-1-Sn	HVQVARNALQ LRVQNVPSPRD KDTYVCMDRN SLGSVSTMHQ LQPEGHVVA
Thai-US-1-Sn	HVQVARNALQ LRVQNVPSPRD KDTYVCMDRN SSGSVSTMHQ LQPEGHVVA

	1460 1470 1480 1490 1500
refSn	EPGLDVPEGT ALNLSCRILPS GPGHIGNSTF AWFRNQRQLH TESVPTLTFT
Thai-HP-1-Sn	EPGLDVPEGT ALNLSCRILPS GPGHIGNSTF AWFRNQRQLH TESVPTLTFT
Thai-EU-1-Sn	EPGLDVPEGT ALNLSCRILPS GPGHIGNSTF AWFRNQRQLH TESVPTLTFT
Thai-US-1-Sn	EPGLDVPEGT ALNLSCRILPS GPGHIGNSTF AWFRNQRQLH TESVPTLTFT

	1510 1520 1530 1540 1550
refSn	HVARAQAGLY HCQAELPAGA ATSAPVLLRV LYPPKPTPTMT VFVEPEGGIQ
Thai-HP-1-Sn	HVARAQAGLY HCQAELPAGA ATSAPVLLRV LYPPKPTPTMT VFVEPEGGIQ
Thai-EU-1-Sn	HVARAQAGLY HCQAELPAGA ATSAPVLLRV LYPPKPTPTMT VFVEPEGGIQ
Thai-US-1-Sn	HVARAQAGLY HCQAELPAGA ATSAPVLLRV LYPPKPTPTMT VFVEPEGGIQ

	1560 1570 1580 1590 1600
refSn	GILDCRVDSE PLASLTLHLG SRLVASSQPQ AAPAKPHIRV SASPNALRVD
Thai-HP-1-Sn	GILDCRVDSE PLASLTLHLG SRLVASSQPQ AAPAKPHIRV SASPNALRVD
Thai-EU-1-Sn	GILDCRVDSE PLASLTLHLG SRLVASSQPQ AAPAKPHIRV SASPNALRVD
Thai-US-1-Sn	GILDCRVDSE PLASLTLHLG SRLVASSQPQ AAPAKPHIRV SASPNALRVD

	1610 1620 1630 1640 1650
refSn	MEELKPSDQG EYVCSASNAL GSASAATYFG TRALHRLHLF QHLLWFLGLL
Thai-HP-1-Sn	MEELKPSDQG EYVCSASNAL GSASAATYFG TRALHRLHLF QHLLWFLGLL
Thai-EU-1-Sn	MEELKPSDQG EYVCSASNAL GSASAATYFG TRALHRLHLF QHLLWFLGLL
Thai-US-1-Sn	MEELKPSDQG EYVCSASNAL GSASAATYFG TRALHRLHLF QHLLWFLGLL

	1660 1670 1680 1690 1700
refSn	ASLLFLLLGL GVWYAWRRGN FYKLRMGEYS VEMVSRKETT QMSTDQEEVT
Thai-HP-1-Sn	ASLLFLLLGL GVWYAWRRGN FYKLRMGEYS VEMVSRKETT QMSTDQEEVT
Thai-EU-1-Sn	ASLLFLLLGL GVWYAWRRGN FYKLRMGEYS VEMVSRKETT QMSTDQEEVT
Thai-US-1-Sn	ASLLFLLLGL GVWYAWRRGN FYKLRMGEYS VEMVSRKETT QMSTDQEEVT

	1710 1720 1730
refSn	GIGDDAGSVN QAAFDPAHLC ENTQSVKSTV
Thai-HP-1-Sn	GIGDDAGSVN QAAFDPAHLC ENTQSVKSTV
Thai-EU-1-Sn	GIGDDAGSVN QAAFDPAHLC ENTQSVKSTV
Thai-US-1-Sn	GIGDDAGSVN QAAFDPAHLC ENTQSVKSTV

Figure 4.10 Deduced amino acid alignments of the reference Sn (refSn) (accession number is NP_999511) with recombinant plasmids of Thai-HP-1-pCR®-XL-TOPO®-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR®-XL-TOPO®-Sn (Thai-EU-1-Sn) and Thai-US-1-pCR®-XL-TOPO®-Sn (Thai-US-1-Sn), respectively. The amino acid changes are boxed.

2.2 Nucleotide and deduced amino acid sequences of recombinant porcine Sn (N-terminal domain) plasmids

The author selected 3 recombinant plasmids, Thai-HP-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-US-1-N-terminal-Sn), respectively, to sequence. The complete sequence of 3 recombinant porcine Sn (N-terminal domain) plasmids were successfully sequenced at the same length of 1,024 bp (Table 4.5) and were analyzed the alignment with the available data bases of refSn cDNA (accession number is NM_214346) form the PAM of normal pigs as shown in Figure 4.11. The results showed that the nucleotide sequences of 3 recombinant porcine Sn (N-terminal domain) plasmids were changed in 2 positions of 130 and 762.

The deduced amino acid sequences and alignment of recombinant porcine Sn (N-terminal domain) plasmids from PRRVS infected PAM (341 amino acids) and refSn cDNA (accession number is NP_999511) from normal PAM (341 amino acids) were shown in Table 4.6 and Figure 4.12, respectively. The results indicated that the amino acid sequences of 3 recombinant porcine Sn (N-terminal domain) plasmids were changed in a position of 44 (Glycine to Arginine).

Table 4.5 Nucleotide sequences of 3 recombinant porcine Sn (N-terminal domain) plasmids

Product's name	Sequence
1. Thai-HP-1- N-terminal-Sn (1,024 bp)	atggactcctgctccgtcctccctggctcatctgccttagcaggcctggcctcgacggtt ccagccccgagaccgtgcagggcatcaaggcctgcctcatatcccctgcacccctccgctcc cggccaacgtggaggtgccccatggcatcacagccatctggtaactatgactactcaggcaagcgc ctggtagtgagccactccaggaacccaaagggtggagaaccactccaaggccggccctg ctgtggggcaggttgaacagaggacgtgcagcctgctgaaggacctgcagccccaggact cgggctcctataactccgcttgagatcagcgagggcaaccgctggtcagatgtcaaaggcacag ttgtcaccgtgacagaggtgcccagcgtgcccaccattgcctgccagccaagctgcattgaggc atggaggtggactcaactgctccactccctatgtgtgcccacggagccgtcaacctacagtgg caaggccaggatcccacccgctccgtcacccacccatccagaagctgagccctcgccacca gccacatggagaccctgcacatggccctgtcctggcaggaccatggccggatctgagctgcca ggtctcagcagccgaacgcaggatgcagaaggagattcacctcaagtgcagtatgcccccaag ggtgtggagatccttcagccactccggacggacgtccttctggatctggtcaccctcagctg ccaggtgaatagcagcaaccctcaggtcagttccgtgcagtgggtcaaggatggacgaagctc aaagaccagaaaacgtgtactgcagttgcgcggcggcagcctgggtatgtggcgtctacacctg ccaagccggaaatgccgtggcttcagtcaccctccgtcagcctccacgtctcatggctga ggtccaggtaaaggccctgtgggtccatcctggagaaccagacgg
2. Thai-EU-1- N-terminal-Sn (1,024 bp)	atggactcctgctccgtcctccctggctcatctgccttagcaggcctggcctcgacggtt ccagccccgagaccgtgcagggcatcaaggcctgcctcatatcccctgcacccctccgctcc cggccaacgtggaggtgccccatggcatcacagccatctggtaactatgactactcaggcaagcgc ctggtagtgagccactccaggaacccaaagggtggagaaccactccaaggccggccct cgtgtggggcaggttgaacagaggacgtgcagcctgctgaaggacctgcagccccaggac tcgggctcctataactccgcttgagatcagcgagggcaaccgctggtcagatgtcaaaggcac ttgtcaccgtgacagaggtgcccagcgtgcccaccattgcctgccagccaagctgcattgagg catggaggtggactcaactgctccactccctatgtgtgcccacggagccgtcaacctacagtgg

	gcaaggccaggatcccacccgtccgtcacccacccacccagaagctgagccctggcacc agccacatggagaccctgcacatggccctgtccgtggcaggaccatggccggatccgtgactgcc aggctcagcagccgaacgcaggatgcagaaggagattcacccaagtgcagtatgccccaa gggtgtggagatccttcagccactccggacggacgtcctccaggatctggtaccctcagct gccaggtaatagcagcaaccctcaggtcagttccgtgcagtgggtcaaggatggacgaagct caaagaccagaaacgtgtactgcagttgcgcggcagccctggctgtgctggcgtctacacct gccaaggccggaaatgccgtggcttcagtcaccctccggcagccctccacgtctcatggct aggccaggtaagccctgtggctccatcctggagaaccagacgg
3. Thai-US-1- N-terminal-Sn (1,024 bp)	atggactccctgctctgtcctccctggctcatctgccttagcaggccctggcctcgacggtt ccagccccgagaccgtgcagggcatcaagggtcctgcctcatcatccctgcacccctccgtcc cggccaaacgtggaggtgccccatggcatcacagccatctggtactatgactactcaggcaagcgc ctggtagtgagccactccaggaacccaaagggtggagaaccactccaaggccggccctg ctgtggggcaggttaacagaggacgtgcagcctgctgaaggacctgcagccccaggact cgggctctataactccgccttgagatcagcgagggcaaccgctggtcagatgtcaaaggcacag ttgtcaccgtacagagggtgcccagcgtgcccaccattgcctgccagccaagctgcattggc atggaggtggacttcaactgccttactccatgtgtgcggacggagccggtaacccatgtgg caaggccaggatcccacccgtccgtcacccacccacccagaagctgagccctgggacca gccacatggagaccctgcacatggccctgtccgtggcaggaccatggccggatccgtgactgca ggtctcagcagccgaacgcaggatgcagaaggagattcacccaagtgcagtatgccccaa gggtgtggagatccttcagccactccggacggacgtcctccgtgatctggtaccctcagct ccaggtaatagcagcaaccctcaggtcagttccgtgcagtgggtcaaggatggacgaagct aaagaccagaaacgtgtactgcagttgcgcggcagccctggctgtgctggcgtctacacct gccaaggccggaaatgccgtggcttcagtcaccctccggcagccctccacgtctcatggct ggccaggtaagccctgtggctccatcctggagaaccagacgg

Figure 4.11 Nucleotide sequence alignments of the reference Sn (refSn) (accession number is NM_214346) with recombinant plasmids of Thai-HP-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-US-1-N-terminal-Sn), respectively. The nucleotide changes are boxed.

Table 4.6 Deduced amino acid sequences of 3 recombinant porcine Sn (N-terminal domain) plasmids

Product's name	Sequence
1. Thai-HP-1- N-terminal-Sn (341 aa)	MDFLLLLLASSALAGLASWTVSSPETVQGIKGSCLIIPCTFGFPANVEVPH GITAIWYYDYSRKRLVSHSRNPKVVENHFQGRALLLGQVEQRTCSLLLKDL QPQDSGSYNFRFEISEGNRWSDVKGTVVTTEVPSVPTIALPAKLHEGMEVD FNCSTPYVCPTEPVNLQWQGQDPTRSVTSHLQKLEPSGTRHMETLHMALS WQDHGRILSCQVSAEERRMQKEIHLQVQHAPKGVEILFSHSGRNVLPGDLV TLSCQVNSSNPQVSSVQWVKDGTLKDQKCVLQLRRAAWADAGVYTCQA GNAVGSVSPPVSLHVFMAEVQVSPVGSILENQT
2. Thai-EU-1- N-terminal-Sn (341 aa)	MDFLLLLLASSALAGLASWTVSSPETVQGIKGSCLIIPCTFGFPANVEVPH GITAIWYYDYSRKRVVSHSRNPKVVENHFQGRALLLGQVEQRTCSLLLKDL QPQDSGSYNFRFEISEGNRWSDVKGTVVTTEVPSVPTIALPAKLHEGMEVD FNCSTPYVCPTEPVNLQWQGQYPTRSVTSHLQKLEPSGTRHMETLHMALS WQDHGRILSCQVSAEERRMQKEIHLQVQHAPKGVEILFSHSGRNVLPGDLV TLSCQVNSSNPQVSSVQWVKDGTLKDQKCVLQLRRAAWADAGVYTCQA GNAVGSVSPPVSLHVFMAEVQVSPVGSILENQT
3. Thai-US-1- N-terminal-Sn (341 aa)	MDFLLLLLASSALAGLASWTVSSPETVQGIKGSCLIIPCTFGFPANVEVPH GITAIWYYDYSRKRLVSHSRNPKVVENHFQGRALLLGQVEQRTCSLLLKDL QPQDSGSYNFRFEISEGNRWSDVKGTVVTTEVPSVPTIALPAKLHEGMEVD FNCSTPYVCPTEPVNLQWQGQDPTRSVTSHLQKLEPSGTRHMETLHMALS WQDHGRILSCQVSAEERRMQKEIHLQVQHAPKGVEILFSHSGRNVLPGDLV TLSCQVNSSNPQVSSVQWVKDGTLKDQKCVLQLRRAAWADAGVYTCQA GNAVGSVSPPVSLHVFMAEVQVSPVGSILENQT

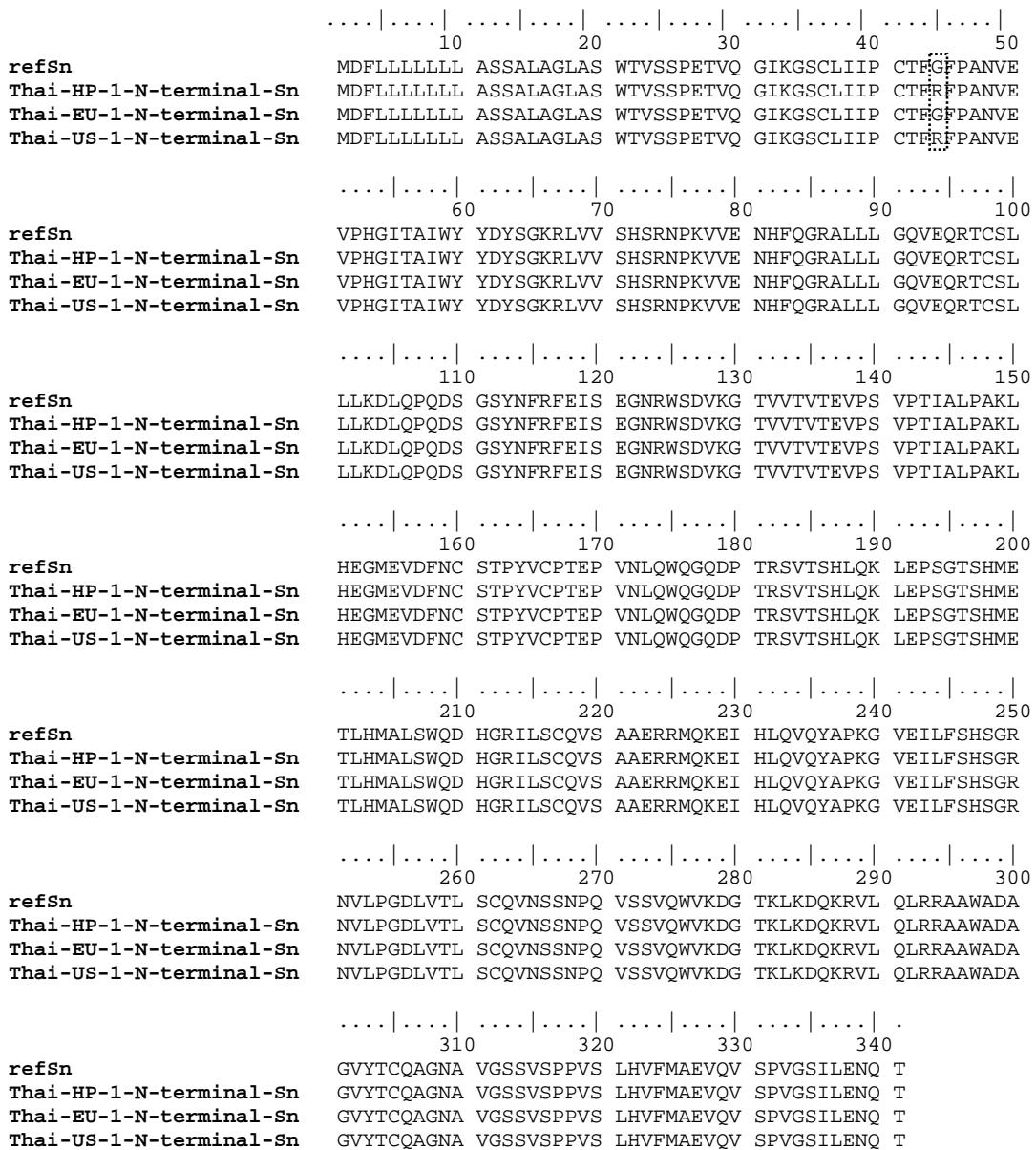


Figure 4.12 Deduced amino acid alignments of the reference Sn (refSn) (GenBank accession number is NP_999511) with recombinant plasmids of Thai-HP-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR®8-GW-TOPO®-N-terminal Sn (Thai-US-1-N-terminal-Sn), respectively. The amino acid change is boxed.

2.3 Nucleotide and deduced amino acid sequences of recombinant porcine CD163 (full) plasmids

The author selected 3 recombinant plasmids of Thai-HP-1-pCR®-XL-TOPO®-CD163, (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163) and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163) to sequence. The 3 recombinant porcine CD163 (full) plasmids were successfully sequenced (3,400 bp) and were analyzed alignment with the available data bases of the reference CD163 (refCD163) cDNA (accession number is NM_213976) form the PAM of normal pigs as shown in Table 4.7 and Figure 4.13, respectively. The results revealed that the nucleotide sequences of 3 recombinant porcine CD163 (full) plasmids were showed similar pattern and changed in 11 positions at 796; 960; 1,370; 1,965; 2,437; 2,455; 2,700; 2,798; 2,897; 2,898 and 3,240, respectively.

The deduced amino acid sequences and alignment of 3 recombinant porcine CD163 (full) plasmids from PRRVS infected PAM (1,133 amino acids) and the refCD163 (accession number is NP_999141) from normal PAM (1,133 amino acids) were shown in Table 4.8 and Figure 4.14, respectively. The results showed that the amino acid sequences of 3 recombinant porcine CD163 (full) plasmids were changed in 6 positions at 266 (Leucine to Valine); 457 (Aspartate to Alanine); 655 (Aspartate to Glutamate); 813 (Serine to Proline); 819 (Arginine to Glycine) and 933 (Serine to Phenylalanine), respectively.

Table 4.7 Nucleotide sequences of 3 recombinant porcine CD163 (full) plasmids

Transformant's name	Sequence
1. Thai-HP-1- CD163 (3,400 bp)	atgggtctacttgaagactctggatctgcagacttagaagatgttctgcccatttaagttccctcactt ttgcttagtcgtcgcttcagtgcctgctggacttagtctcttggaggaaaagacaaggagct gaggctaacgggtggtaaaacaagtgtcgttggaaagagtggagggtgaaagtgcaggagga gtggggactgtgtataatggctggacatggatgtggctctgttgtttaggcagctgg atgtccaactgctatcaaagccactggatggcttaatttagtgcagggtctggacgcattggatg gatcatgttctgtcgagggaatgagtcagctctggactgcaaacatgtatggatggggaaa gcataactgtactcaccaacaggatgtggagtaacctgctcagatggatctgatttagagatga ggctggtaatggagggaaaccggcttaggaagaatagaagtc当地caagagcggtg ggaacagtgtgtatgataactcaacataatcatgcttctgtggttgtaaacaactgtatgg aagtgtcgtcagttctgtggctagctaattggagaaggttctggaccaatctggttgtatctt gtatgcaatggaaatgagtcagctctggactgcaaacatgtatggatggggaaaagcacaat tgcgatcatgtcgaggatgtggagtgattgttggactggcagacactgagatgg atggagtcactgaatgttcaggaagatggaaatccaaggagaatggggaaaact ctgtgtatggctggatagtgtatgtatgtatgtatgtatgtatgtatgtatgtatgt ctgtcaactgccattggtcgagtaacgccagtggactggcacattggcttgcacactgtttc ttgccatggacacgagtctgtctggcagtgttagacaccatgtatggggaaaagcattattgca atcataatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgt gaggcagccactgtgtggacagtggagggtggaaattcagaaactggtagggaaaagtgt gatagaagctggggactgaaagaagactgtatgtggttgcaggcagctggatgtggatctgca ctcaaaaatcatcaatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgt ctgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgtatgt cactatgacgaagccaaaattacctgctcagcccacaggaaaccaggctgggtggaggg acattccctgtctggcgtgttgaagtacaacatggagacacgtggggcaccgtctgtattctg acttctctgtggaggcggccagcgtgctgcaggaaactacagtgcggcactgtggttccctc

ctggggggagctcacttggagaaggaagtggacagatctggctgaagaattccagtgttag
ggcacgagtcccacccactctgcccagtagcacccgcctgacggacatgtacca
cagcaggacgtcgccgtagtcgtcaagatacacacacaaatccgctggtaatggcaaga
ccccatgtgaaggaagagtgaggactcaacattctggctggggccctctgcaactctca
ctgggacatggaagatgcccattttatgccagcagctaaatgtggagttgccttctatcccg
ggaggagcaccttggaaaggaagtggagcaggtctggaggcacatgttcaactgcactgg
actgagaagcacaatgggagactgtccgtactgctggcgcacactctgttctcaggca
agtggcctctgtaatctgctcaggaaaccagagtcagacactatccccgtgcaattcatcatcct
cgaccatcaagctctattttcagaagaaagtgggtgcctgcataggagtgcaacttc
gcctggtcgatggagggtggcgttgtgctggagagtagaggctatcctgggcattcctggg
caccatctgtgatgacagctgggacctgaatgatgcccattgtggtgcaaacagctgagctg
ggatggccattaatgccactggctgctcatttggaaaggaacagggccattggctgga
tgagataaactgtaatggaaaagaatctcatattggcaatgccactcacatggtgccccggc
acaattgcaggcataaggaggatgcaggagtcatctgctcagagttcatgtctctggactgatc
agtaaaaacagcagagagacctgtgcagggcgcctggaagtttacaacggagctgggg
cagcgtggcaggaatagcatgtctccagccacagtgggggtgtatgcaggcagctggct
gtgcagacagaggggacatcagccctgcatctcagacaagacagtgtccaggcacatgtgg
gtggacaatgttcagtgtcctaaaggacactgacacactatggcagtgccatcatccatggaa
gaagagactggccagccctcagaggagacatggatcacatgtgccaacaaaataagacttc
aagaaggaaacactaattttggacgtgtggagatctgtacggagttccctgggcactgtg
tgtgacgactcctggacacttgaagatgctcagggtgtgcccacagctggctaaggctca
gctttggaggcagggaaagagccgcattggccagggactggccatatggctcaatga
agtgaagtgcaggggaatgaaccctcgtggattgtcctgcccacatctggggccacagt
gactgtggacacaaggaggatgctgtgacgtgctcagaaattgcaaaagagccgagaatc
cctacatgccacaggcgtcatcttgcacttgcaatcttgggtcattctgttggcctgtctc
atcgcatcatttgcactcagaagcgaagacagaggcagcggctcagttctcaggag
gagagaattctgtacatcaaattcaaatccgggagatgaattctgcctgaaagcagatgaaacg

	gatatgtctaaatccctcaggagaccactctgaagtacaatgaaaaggaaaatggattataac ctggtagttcagccctgatgaagacacctggacta
2. Thai-EU-1-	atgggtctacttgaagactctggatctgcagactttagaagatgtctgcccatttaagttccttcactt
CD163	ttgctgttagtcgcttgttcagtgccctgctggactagttcttggaggaaaagacaaggagct
(3,400 bp)	gaggctaacgggtggtaaaacaagtgcctgaaagagtggaggtgaaagtgcaggagga gtggggactgtgttaataatggctggacatggatgtggctctgttgttaggcagctgg atgtccaactgctatcaaagccactggatggctaaatttagtgcaggctggacgcattggatg gatcatgttcttgcagggatagactcagctctggactgc当地atggatggatggggaaa gcataactgtactcaccaacaggatgc当地ggactgc当地atggatctgatggatggggaaa ggctggtaatggaggaaaccggcttaggaagaatagaagtcaaatcaagagcgggtgg ggAACAGTGTGATGATAACTCAACATAATCATGCCTCTGGTTGTAACACAATTGAATGTGG AAGTGCTGTCAGTTCTGGTCAGCTAAATTGGAGAAGGTTCTGGACCAATCTGGTTGATGATCT GTATGCAATGAAATGAGTCAGCTCTGGAACTGCAAACATGAAGGATGGGGAAAGCACAAT TGCGATCATGCTGAGGATGCTGGAGTGATTGCTAAATGGAGCAGACCTGAAACTGAGAGTGG AGATGGAGTCACTGAATGTTAGGAAGATTGGAAGTGAAATTCCAAGGAGAATGGGGAAACAAT CTGTGATGGCTGGGATAGTGATGATGCCGCTGGCATGTAAGCAACTGGGATGCTCAACTG CTGTCAGGCCATTGGTCAGTTAACGCCAGTGAGGGAACTGGTCACATTGGCTGACAGTGTTC TTGCCATGGACACGAGTCAGCTCTGGCAGTGAGACACCATGAATGGGGAAAGCATTATTGCA ATCATAATGAAGATGCTGGTGTACATGTTCTGATGGATCAGATCTGGAACTGAGACTAAAGGTG GAGGCAGCCACTGTGCTGGGACAGTGGAGGTGGAATTCAAGAACACTGGTAGGAAAAGTGTG GATAGAAGCTGGGGACTGAAAGAAGCTGATGTTGAGGCACTGGGATGTTGATCTGCA CTCAAAACATCATCAAGTTATTCCAACCAAGGCAACAAACACATGGCTGTTGTAAGCAG CTGTAATGAAATGAAACTCTTGGACTGCAAGAATTGGCAGTGAGGGTGGACTTAGTGTGAT CACTATGACGAAGCCAAAATTACCTGCTCAGCCCACAGGAAACCCAGGCTGGTTGGAGGG ACATTCCCTGCTGGTCAGCAGTGCTGAGGAAACTACAGTGCAGGCACTGTGGTTCCCT ACTTCTCTGGAGGCGGCCAGCAGTGCTGAGGAAACTACAGTGCAGGCACTGTGGTTCCCT CTGGGGGAGCTCACTTGGAGGAAGTGGACAGATCTGGCTGAAAGAATTCCAGTGTGAG

ggcacgagtcccacttcactctgccagtagcacccgcctgacggacatgtacca
cagcaggacgtcgcgtagtctgctcaagatacacacacaatccgctggtaatggcaaga
ccccatgtgaaggaagagtgagtcacatttgcggcctggggccctctgcaactctca
ctgggacatggaagatgcccatttgcggcactgctggggccatcttcatcccg
ggaggagcaccttggaaaggaagtgagcaggtctggaggcacatgttcaactgcactgg
actgagaagcacaatggagagtgtccgtcactgtctggcgcatcactctgttctcaggca
agtggcctctgtaatctgctcaggaaaccagagtcagacactatccccgtgcaattcatcatcct
cgaccatcaagctctattttcagaagaaagtgggtgcctgcataggagtgcaacttc
gcctggatggagggtggcgtgtgctggagagtagaggctatcctgggcattctgggg
caccatctgtatggcactgggtctgctcatttggaaaggaacaggccattggctgga
tgagataaactgtaatggaaaagaatctcatattggcaatgccactcacatggggggcggc
acaattgcaggcataaggaggatgcaggagtcatgtccagagttcatgtctggactgat
cagtggaaacagcagagagacgtgcaggcgcctggaaagtttacaacggagctgg
gcagcgtggcaggaatagcatgtccagccacagtgggggtgtatgcaggcagctggc
tgtcagacagaggggacatcagccctgcatttcagacaagacagtgtccaggcacatgt
ggtggacaatgttcagtgcttgcatttggacactgacacactatggcagtgccatcatccatgga
agaagagactggccagccctcagaggagacatggcactgtgccaacaaaataagactt
caagaaggaaacactaattttctggacgtgtggagatctggatcggaggctggggactg
tgtgtacgactcctgggacatttgcatttggacgtgtggagatctggatcggaggctgg
agctttggaggcagggaaagagccgcattggccaggggactggccatggctcaatg
aagtgaagtgcacggggaaatgaacccttgcatttggatcggaggctggggccaca
gtgactgtggacacaaggaggatgtgtgcacgtgtcagaaattgcaaaagagccgagaat
ccctacatgccacaggctgcattttgtgcatttgcacatttgcatttggggcatttgc
catcgcatcctcatttggactcagaagcgaagacagaggcagcggctcagtttcagga
ggagagaattctgtacatcaaattcaataccggggagatgaattctgcctgaaagcagatgaaac
ggatgtctaaatccctcaggagaccactgtcagttacaatgaaaaaggaaaatggaaattataa

	cctggtagttcagccgtatgaagacacctggacta
3. Thai-US-1-	atggtgctacttgaagactctggatctgcagactttagaagatgtctgcccatttaagtccctcactt
CD163	ttgctgttagtcgttgttcagtcgtcgtggacttagttcttggaggaaaagacaaggagct
(3,400 bp)	gaggctaacgggtggtaaaacaaggactctggaaagagtggagggtggaaatgtcaggagga gtggggactgtgtataatggctggacatggatgtggctctgttgttaggcagctgg atgtccaactgttatcaaagccactggatggctaattttatgtcaggtctggacgcattggat gatcatgttcttgcaggaaatgagtcagctctggactgcaaacatgtggatggggaaa gcataactgtactcaccaacaggatgctggagtaacctgctcagatggatctgatttagatga ggctggtaatggaggaaaccgggtgttaggaagaatagaaggtaatcaagagcggtg ggAACAGTGTGATGATAACTCAACATAATCATGCTCTGTGGTTGAAACAACATGAATGTGG AAGTGCTGTCAGTTCTGGTCAGCTAATTGGAGAAGGTTCTGGACCAATCTGGTTGATGATCTT GTATGCAATGAAATGAGTCAGCTCTGGAACTGCAAACATGAAGGATGGGGAAAGCACAAT TGCATGCTGAGGATGCTGGAGTGATTGCTTAATGGAGCAGACCTGAAACTGAGAGTGG AGATGGAGTCACTGAATGTTAGGAAGATTGGAGTGAATCCAAGGAGAATGGGGAAACAT CTGTGATGGCTGGGAGTAGTGATGATGCCGCTGGCATGTAAGCAACTGGGATGTCCAAC CTGTCACTGCCATTGGTCAGTTAACGCCAGTGAGGGAACTGGTCACATTGGCTGACAGTGT TTGCCATGGACACGAGTCGCTCTGGCAGTGTAGACACCATGAATGGGGAAAGCATTATTGCA ATCATAATGAAGATGCTGGTGTACATGTTCTGATGGATCAGATCTGGAACTGAGACTTAAGGT GAGGCAGCCACTGTGCTGGGACAGTGGAGGGTGGAAATTCAAGAACACTGGTAGGAAAAGTGT GATAGAAGCTGGGGACTGAAAGAAGCTGATGTTGAGGAGCTGGGATGTGGATCTGCA CTCAAAACATCATCAAGTTATTCCAACCAAGGCAACAAACACATGGCTGTTGAAGCAG CTGTAATGAAATGAAACTCTTGGACTGCAAGAATTGGCAGTGGGGTGGACTTAGTTGCT CACTATGACGAAGCCAAAATTACCTGCTCAGCCCACAGGAAACCCAGGCTGGTTGGAGGG ACATTCCCTGCTGGTCGTGTTGAAGTACAACATGGAGACACGTGGGGCACCGTGTGATTCTG ACTTCTCTGGAGGGCGGCCAGCGTGTGAGGAAACTACAGTGCAGGACTGTGGCAGTGGTT CTGGGGGGAGCTCAGTTGGAGAAGGAAGTGGACAGATCTGGCTGAAGAATTCCAGTGT GGCACGAGTCCCACCTTCACTGCCAGTAGCACCCCCGCCCTGACGGGACATGTAGCCA

cagcagggacgtcgccgtagtctgctcaagatacacacaatccgctggtaatggcaaga
ccccatgtgaaggaagagtggagactcaacattctggcctggggccctgcactctca
ctgggacatgaaagatgccatgtttatgccaggcagctaaatgtggagttgccttcatccg
ggaggagcaccttggaaaggaagtggagactgtccgtactgctggcgcactcactgttcaggca
actgagaagcacatgggagactgtccgtactgctggcgcactcactgttcaggca
agtggcctctgtaatctgctcaggaaaccagactcagacactatccccgtcaattcatcatcct
cggaccatcaagctctattttcagaagaaagtgggttgccataggagttcaacttc
gcctggatggagggtggcgttgtgctggagagtagaggctatcctgggcatcctggg
caccatctgtgatgacagactgggacctgaatgatgcccattgtgggtgcaaacagctgagctg
ggatggccattaatgccactgggtctgctcatggaaaggaacaggccattggctgga
tgagataaaactgtaatggaaaagaatctcatattggcaatgccactcacatggttggggcggc
acaattgcaggcataaggaggatgcaggactcatctgctcagagttcatgtctgagactgatc
agtggaaaacagcagagagacctgtgcagggcgcctggaaagttttacaacggagctggg
cagcgtggcaggaatagcatgtccagccacagtgggggtggatgcaggcagctggc
gtgcagacagagggacatcagccctgcatctcagacaagacactgtccaggcacatgtgg
gtggacaatgttcagtgtcctaaaggacctgacacactatggcagtgccatcatccatggaa
gaagagactggccagccctcagaggagacatggatcacatgtgccaacaaaataagactc
aagaaggaaacactaatgttctggacgtgtggagatctgtacggagttcctggggactgt
gtgtgacgactcctgggacctgaaagatgctcagggtgtgcccacagctggctgtggctca
gtttggaggcagggaaagagccgcattggccaggggactggccatatggctcaatga
agtgaagtgcagggaaatgaaccctcctgtggattgtcctgcccacatctggggccacagt
gactgtggacacaaggaggatgctgtgacgtgctcagaaattgcaagagccgagaatc
cctacatgccacaggctcgtcatcttgcacttgcaatcttggggcattctgttggcctgtctc
atcgcatcctcattggactcagaagcgaagacagaggcagcggctctcagtttcagggag
gagagaattctgtacatcaaattcaataccgggagatgaattctgctgaaagcagatgaaacg
gatatgctaaatccctcaggagaccactgaaatgaaaaggaaatggaaattataac
ctggtagttcagccctgatgaagacactggact

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 810 820 830 840 850

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

TGGAAATGAG TCAGCTCTCT GGAACGTCAA ACATGAAGGA TGGGGAAAGC
 TGGAAATGAG TCAGCTCTCT GGAACGTCAA ACATGAAGGA TGGGGAAAGC
 TGGAAATGAG TCAGCTCTCT GGAACGTCAA ACATGAAGGA TGGGGAAAGC
 TGGAAATGAG TCAGCTCTCT GGAACGTCAA ACATGAAGGA TGGGGAAAGC

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 860 870 880 890 900

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

ACAATTGCGA TCATGCTGAG GATGCTGGAG TGATTTGCTT AAATGGAGCA
 ACAATTGCGA TCATGCTGAG GATGCTGGAG TGATTTGCTT AAATGGAGCA
 ACAATTGCGA TCATGCTGAG GATGCTGGAG TGATTTGCTT AAATGGAGCA
 ACAATTGCGA TCATGCTGAG GATGCTGGAG TGATTTGCTT AAATGGAGCA

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 910 920 930 940 950

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GACCTGAAAC TGAGAGTGTT AGATGGACTC ACTGAATGTT CAGGAAGATT
 GACCTGAAAC TGAGAGTGTT AGATGGAGTC ACTGAATGTT CAGGAAGATT
 GACCTGAAAC TGAGAGTGTT AGATGGAGTC ACTGAATGTT CAGGAAGATT
 GACCTGAAAC TGAGAGTGTT AGATGGAGTC ACTGAATGTT CAGGAAGATT

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 960 970 980 990 1000

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GGAAGTGAAA TTCCAAGGAG AATGGGGAAC AATCTGTGAT GATGGCTGGG
 GGAAGTGAAA TTCCAAGGAG AATGGGGAAC AATCTGTGAT GATGGCTGGG
 GGAAGTGAAA TTCCAAGGAG AATGGGGAAC AATCTGTGAT GATGGCTGGG
 GGAAGTGAAA TTCCAAGGAG AATGGGGAAC AATCTGTGAT GATGGCTGGG

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 1010 1020 1030 1040 1050

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

ATAGTGATGA TGCCGCTGTG GCATGTAAGC AACTGGGATG TCCAAGTGCT
 ATAGTGATGA TGCCGCTGTG GCATGTAAGC AACTGGGATG TCCAAGTGCT
 ATAGTGATGA TGCCGCTGTG GCATGTAAGC AACTGGGATG TCCAAGTGCT
 ATAGTGATGA TGCCGCTGTG GCATGTAAGC AACTGGGATG TCCAAGTGCT

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 1060 1070 1080 1090 1100

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GTCACTGCCA TTGGTCGAGT TAACGCCAGT GAGGGAACTG GACACATTTG
 GTCACTGCCA TTGGTCGAGT TAACGCCAGT GAGGGAACTG GTCACATTTG
 GTCACTGCCA TTGGTCGAGT TAACGCCAGT GAGGGAACTG GTCACATTTG
 GTCACTGCCA TTGGTCGAGT TAACGCCAGT GAGGGAACTG GTCACATTTG

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 1110 1120 1130 1140 1150

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GCTTGACAGT GTTCTTGCC ATGGACACGA GTCTGCTCTC TGGCAGTGTA
 GCTTGACAGT GTTCTTGCC ATGGACACGA GTCTGCTCTC TGGCAGTGTA
 GCTTGACAGT GTTCTTGCC ATGGACACGA GTCTGCTCTC TGGCAGTGTA
 GCTTGACAGT GTTCTTGCC ATGGACACGA GTCTGCTCTC TGGCAGTGTA

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 1160 1170 1180 1190 1200

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GACACCATGA ATGGGGAAAG CATTATTGCA ATCATAATGA AGATGCTGGT
 GACACCATGA ATGGGGAAAG CATTATTGCA ATCATAATGA AGATGCTGGT
 GACACCATGA ATGGGGAAAG CATTATTGCA ATCATAATGA AGATGCTGGT
 GACACCATGA ATGGGGAAAG CATTATTGCA ATCATAATGA AGATGCTGGT

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 1210 1220 1230 1240 1250

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1260 1270 1280 1290 1300

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1310 1320 1330 1340 1350

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1360 1370 1380 1390 1400

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1410 1420 1430 1440 1450

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1460 1470 1480 1490 1500

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1510 1520 1530 1540 1550

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

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 1560 1570 1580 1590 1600

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GTGACATGGTT CTGATGGATC AGATCTGGAA CTGAGACTTA AAGGTGGAGG

 CAGCCACTGT GCTGGGACAG TGGAGGTGGA ATTCAAGAAA CTGGTAGGAA

 AAGTGTGTGA TAGAAGCTGG GGACTGAAAG AAGCTGATGT GGTTTCAGG

 CAGCTGGGAT GTGGATCTGC ACTCAAAACA TCATATCAAG TTATTCCAA

 AACCAAGGCCA ACAAACACAT GGCTGTTGT AAGCAGCTGT AATGGAAATG

 AAACCTTCTCT TTGGGACTGC AAGAATTGGC AGTGGGTGG ACTTAGTTGT

 GATCACTATG ACGAAGCCAA AATTACCTGC TCAGCCCACA GGAAACCCAG

 GCTGGTTGGA GGGGACATTC CCTGCTCTGG TCGTGTGAA GTACAACATG
 GCTGGTTGGA GGGGACATTC CCTGCTCTGG TCGTGTGAA GTACAACATG
 GCTGGTTGGA GGGGACATTC CCTGCTCTGG TCGTGTGAA GTACAACATG
 GCTGGTTGGA GGGGACATTC CCTGCTCTGG TCGTGTGAA GTACAACATG

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 1610 1620 1630 1640 1650

refCD163 GAGACACGTG GGGCACCGTC TGTGATTCTG ACTTCTCTCT GGAGGCAGGCC
Thai-HP-1-CD163 GAGACACGTG GGGCACCGTC TGTGATTCTG ACTTCTCTCT GGAGGCAGGCC
Thai-EU-1-CD163 GAGACACGTG GGGCACCGTC TGTGATTCTG ACTTCTCTCT GGAGGCAGGCC
Thai-US-1-CD163 GAGACACGTG GGGCACCGTC TGTGATTCTG ACTTCTCTCT GGAGGCAGGCC

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 1660 1670 1680 1690 1700

refCD163 AGCGTGCTGT GCAGGGAAC ACAGTGCAGC ACTGTGGTT CCCTCCTGGG
Thai-HP-1-CD163 AGCGTGCTGT GCAGGGAAC ACAGTGCAGC ACTGTGGTT CCCTCCTGGG
Thai-EU-1-CD163 AGCGTGCTGT GCAGGGAAC ACAGTGCAGC ACTGTGGTT CCCTCCTGGG
Thai-US-1-CD163 AGCGTGCTGT GCAGGGAAC ACAGTGCAGC ACTGTGGTT CCCTCCTGGG

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 1710 1720 1730 1740 1750

refCD163 GGGAGCTCAC TTTGGAGAAG GAAGTGGACA GATCTGGGCT GAAGAATTCC
Thai-HP-1-CD163 GGGAGCTCAC TTTGGAGAAG GAAGTGGACA GATCTGGGCT GAAGAATTCC
Thai-EU-1-CD163 GGGAGCTCAC TTTGGAGAAG GAAGTGGACA GATCTGGGCT GAAGAATTCC
Thai-US-1-CD163 GGGAGCTCAC TTTGGAGAAG GAAGTGGACA GATCTGGGCT GAAGAATTCC

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 1760 1770 1780 1790 1800

refCD163 AGTGTGAGGG GCACGAGTCC CACCTTCAC TCTGCCAGT AGCACCCCCGC
Thai-HP-1-CD163 AGTGTGAGGG GCACGAGTCC CACCTTCAC TCTGCCAGT AGCACCCCCGC
Thai-EU-1-CD163 AGTGTGAGGG GCACGAGTCC CACCTTCAC TCTGCCAGT AGCACCCCCGC
Thai-US-1-CD163 AGTGTGAGGG GCACGAGTCC CACCTTCAC TCTGCCAGT AGCACCCCCGC

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 1810 1820 1830 1840 1850

refCD163 CCTGACGGGA CATGTAGCCA CAGCAGGGAC GTGGCGTAG TCTGCTCAAG
Thai-HP-1-CD163 CCTGACGGGA CATGTAGCCA CAGCAGGGAC GTGGCGTAG TCTGCTCAAG
Thai-EU-1-CD163 CCTGACGGGA CATGTAGCCA CAGCAGGGAC GTGGCGTAG TCTGCTCAAG
Thai-US-1-CD163 CCTGACGGGA CATGTAGCCA CAGCAGGGAC GTGGCGTAG TCTGCTCAAG

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 1860 1870 1880 1890 1900

refCD163 ATACACACAA ATCCGCTTGG TGAATGGCAA GACCCCATGT GAAGGAAGAG
Thai-HP-1-CD163 ATACACACAA ATCCGCTTGG TGAATGGCAA GACCCCATGT GAAGGAAGAG
Thai-EU-1-CD163 ATACACACAA ATCCGCTTGG TGAATGGCAA GACCCCATGT GAAGGAAGAG
Thai-US-1-CD163 ATACACACAA ATCCGCTTGG TGAATGGCAA GACCCCATGT GAAGGAAGAG

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 1910 1920 1930 1940 1950

refCD163 TGGAGCTCAA CATTCTTGGG TCCTGGGGGT CCCTCTGCAA CTCTCACTGG
Thai-HP-1-CD163 TGGAGCTCAA CATTCTTGGG TCCTGGGGGT CCCTCTGCAA CTCTCACTGG
Thai-EU-1-CD163 TGGAGCTCAA CATTCTTGGG TCCTGGGGGT CCCTCTGCAA CTCTCACTGG
Thai-US-1-CD163 TGGAGCTCAA CATTCTTGGG TCCTGGGGGT CCCTCTGCAA CTCTCACTGG

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 1960 1970 1980 1990 2000

refCD163 GACATGGAAG ATGCCCATGT TTTATGCCAG CAGCTTAAAT GTGGAGTTGC
Thai-HP-1-CD163 GACATGGAAG ATGCCCATGT TTTATGCCAG CAGCTTAAAT GTGGAGTTGC
Thai-EU-1-CD163 GACATGGAAG ATGCCCATGT TTTATGCCAG CAGCTTAAAT GTGGAGTTGC
Thai-US-1-CD163 GACATGGAAG ATGCCCATGT TTTATGCCAG CAGCTTAAAT GTGGAGTTGC

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 2010 2020 2030 2040 2050

refCD163 CCTTTCTATC CGGGGAGGAG CACCTTTGG GAAAGGAAGT GAGCAGGTCT
Thai-HP-1-CD163 CCTTTCTATC CGGGGAGGAG CACCTTTGG GAAAGGAAGT GAGCAGGTCT
Thai-EU-1-CD163 CCTTTCTATC CGGGGAGGAG CACCTTTGG GAAAGGAAGT GAGCAGGTCT
Thai-US-1-CD163 CCTTTCTATC CGGGGAGGAG CACCTTTGG GAAAGGAAGT GAGCAGGTCT

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 2060 2070 2080 2090 2100

refCD163 GGAGGCACAT GTTTCACTGC ACTGGGACTG AGAAGCACAT GGGAGACTGT
Thai-HP-1-CD163 GGAGGCACAT GTTTCACTGC ACTGGGACTG AGAAGCACAT GGGAGACTGT
Thai-EU-1-CD163 GGAGGCACAT GTTTCACTGC ACTGGGACTG AGAAGCACAT GGGAGACTGT
Thai-US-1-CD163 GGAGGCACAT GTTTCACTGC ACTGGGACTG AGAAGCACAT GGGAGACTGT

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 2110 2120 2130 2140 2150

refCD163 TCCGTCACTG CTCTGGGCGC ATCACTCTGT TCTTCAGGGC AAGTGGCCTC
Thai-HP-1-CD163 TCCGTCACTG CTCTGGGCGC ATCACTCTGT TCTTCAGGGC AAGTGGCCTC
Thai-EU-1-CD163 TCCGTCACTG CTCTGGGCGC ATCACTCTGT TCTTCAGGGC AAGTGGCCTC
Thai-US-1-CD163 TCCGTCACTG CTCTGGGCGC ATCACTCTGT TCTTCAGGGC AAGTGGCCTC

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 2160 2170 2180 2190 2200

refCD163 TGTAATCTGC TCAGGGAAACC AGAGTCAGAC ACTATCCCCG TGCAATTCTCAT
Thai-HP-1-CD163 TGTAATCTGC TCAGGGAAACC AGAGTCAGAC ACTATCCCCG TGCAATTCTCAT
Thai-EU-1-CD163 TGTAATCTGC TCAGGGAAACC AGAGTCAGAC ACTATCCCCG TGCAATTCTCAT
Thai-US-1-CD163 TGTAATCTGC TCAGGGAAACC AGAGTCAGAC ACTATCCCCG TGCAATTCTCAT

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 2210 2220 2230 2240 2250

refCD163 CATCCTCGGA CCCATCAAGC TCTATTATTT CAGAAGAAAG TGTTGTTGCC
Thai-HP-1-CD163 CATCCTCGGA CCCATCAAGC TCTATTATTT CAGAAGAAAG TGTTGTTGCC
Thai-EU-1-CD163 CATCCTCGGA CCCATCAAGC TCTATTATTT CAGAAGAAAG TGTTGTTGCC
Thai-US-1-CD163 CATCCTCGGA CCCATCAAGC TCTATTATTT CAGAAGAAAG TGTTGTTGCC

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 2260 2270 2280 2290 2300

refCD163 TGCATAGGGA GTGGTCAACT TCGCCTGGTC GATGGAGGTG GTCGTTGTGC
Thai-HP-1-CD163 TGCATAGGGA GTGGTCAACT TCGCCTGGTC GATGGAGGTG GTCGTTGTGC
Thai-EU-1-CD163 TGCATAGGGA GTGGTCAACT TCGCCTGGTC GATGGAGGTG GTCGTTGTGC
Thai-US-1-CD163 TGCATAGGGA GTGGTCAACT TCGCCTGGTC GATGGAGGTG GTCGTTGTGC

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 2310 2320 2330 2340 2350

refCD163 TGGGAGAGTA GAGGTCTATC CTGGGGCATIC CTGGGGCACC ATCTGTGATG
Thai-HP-1-CD163 TGGGAGAGTA GAGGTCTATC CTGGGGCATIC CTGGGGCACC ATCTGTGATG
Thai-EU-1-CD163 TGGGAGAGTA GAGGTCTATC CTGGGGCATIC CTGGGGCACC ATCTGTGATG
Thai-US-1-CD163 TGGGAGAGTA GAGGTCTATC CTGGGGCATIC CTGGGGCACC ATCTGTGATG

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 2360 2370 2380 2390 2400

refCD163 ACAGCTGGGA CCTGAATGAT GCCCATGTGG TGTGCAAACA GCTGAGCTGT
Thai-HP-1-CD163 ACAGCTGGGA CCTGAATGAT GCCCATGTGG TGTGCAAACA GCTGAGCTGT
Thai-EU-1-CD163 ACAGCTGGGA CCTGAATGAT GCCCATGTGG TGTGCAAACA GCTGAGCTGT
Thai-US-1-CD163 ACAGCTGGGA CCTGAATGAT GCCCATGTGG TGTGCAAACA GCTGAGCTGT

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 2410 2420 2430 2440 2450

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GGATGGGCCA TTAATGCCAC TGGTTCTGCT CATTGGGG AAGGAACAGG
 GGATGGGCCA TTAATGCCAC TGGTTCTGCT CATTGGGG AAGGAACAGG
 GGATGGGCCA TTAATGCCAC TGGTTCTGCT CATTGGGG AAGGAACAGG
 GGATGGGCCA TTAATGCCAC TGGTTCTGCT CATTGGGG AAGGAACAGG

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refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GCCCATTGG CTGGATGAGA TAAACTGTAA TGGAAAAGAA TCTCATATTT
 GCCCATTGG CTGGATGAGA TAAACTGTAA TGGAAAAGAA TCTCATATTT
 GCCCATTGG CTGGATGAGA TAAACTGTAA TGGAAAAGAA TCTCATATTT
 GCCCATTGG CTGGATGAGA TAAACTGTAA TGGAAAAGAA TCTCATATTT

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 2510 2520 2530 2540 2550

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GGCAATGCCA CTCACATGGT TGGGGCGGC ACAATTGCAG GCATAAGGAG
 GGCAATGCCA CTCACATGGT TGGGGCGGC ACAATTGCAG GCATAAGGAG
 GGCAATGCCA CTCACATGGT TGGGGCGGC ACAATTGCAG GCATAAGGAG
 GGCAATGCCA CTCACATGGT TGGGGCGGC ACAATTGCAG GCATAAGGAG

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refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GATGCAGGAG TCATCTGCTC AGAGTTCATG TCTCTGAGAC TGATCAGTGA
 GATGCAGGAG TCATCTGCTC AGAGTTCATG TCTCTGGGAC TGATCAGTGA
 GATGCAGGAG TCATCTGCCC AGAGTTCATG TCTCTGGGAC TGATCAGTGA
 GATGCAGGAG TCATCTGCTC AGAGTTCATG TCTCTGAGAC TGATCAGTGA

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 2610 2620 2630 2640 2650

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

AAACAGCAGA GAGACCTGTG CAGGGCGCCT GGAAGTTTT TACAACGGAG
 AAACAGCAGA GAGACCTGTG CAGGGCGCCT GGAAGTTTT TACAACGGAG
 AAACAGCAGA GAGACCTGTG CAGGGCGCCT GGAAGTTTT TACAACGGAG
 AAACAGCAGA GAGACCTGTG CAGGGCGCCT GGAAGTTTT TACAACGGAG

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 2660 2670 2680 2690 2700

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

CTTGGGGCAG CGTTGGCAGG AATAGCATGT CTCCAGCCAC AGTGGGGGTG
 CTTGGGGCAG CGTTGGCAGG AATAGCATGT CTCCAGCCAC AGTGGGGGTG
 CTTGGGGCAG CGTTGGCAGG AATAGCATGT CTCCAGCCAC AGTGGGGGTG
 CTTGGGGCAG CGTTGGCAGG AATAGCATGT CTCCAGCCAC AGTGGGGGTG

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 2710 2720 2730 2740 2750

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

GTATGCAGGC AGCTGGGCTG TGCAGACAGA GGGGACATCA GCCCTGCATC
 GTATGCAGGC AGCTGGGCTG TGCAGACAGA GGGGACATCA GCCCTGCATC
 GTATGCAGGC AGCTGGGCTG TGCAGACAGA GGGGACATCA GCCCTGCATC
 GTATGCAGGC AGCTGGGCTG TGCAGACAGA GGGGACATCA GCCCTGCATC

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 2760 2770 2780 2790 2800

refCD163
Thai-HP-1-CD163
Thai-EU-1-CD163
Thai-US-1-CD163

TTCAGACAAG ACAGTGTCCA GGCACATGT GGTGGACAAT GTTCAGTGTG
 TTCAGACAAG ACAGTGTCCA GGCACATGT GGTGGACAAT GTTCAGTGTG
 TTCAGACAAG ACAGTGTCCA GGCACATGT GGTGGACAAT GTTCAGTGTG
 TTCAGACAAG ACAGTGTCCA GGCACATGT GGTGGACAAT GTTCAGTGTG

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 2810 2820 2830 2840 2850

refCD163 CTAAGGACC TGACACACTA TGGCAGTGCC CCTCATCTCC ATGGAAGAAC
Thai-HP-1-CD163 CTAAGGACC TGACACACTA TGGCAGTGCC CATCATCTCC ATGGAAGAAC
Thai-EU-1-CD163 CTAAGGACC TGACACACTA TGGCAGTGCC CATCATCTCC ATGGAAGAAC
Thai-US-1-CD163 CTAAGGACC TGACACACTA TGGCAGTGCC CATCATCTCC ATGGAAGAAC

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 2860 2870 2880 2890 2900

refCD163 AGACTGGCCA GCCCCTCAGA GGAGACATGG ATCACATGTG CCAACAAAAT
Thai-HP-1-CD163 AGACTGGCCA GCCCCTCAGA GGAGACATGG ATCACATGTG CCAACAAAAT
Thai-EU-1-CD163 AGACTGGCCA GCCCCTCAGA GGAGACATGG ATCACATGTG CCAACAAAAT
Thai-US-1-CD163 AGACTGGCCA GCCCCTCAGA GGAGACATGG ATCACATGTG CCAACAAAAT

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 2910 2920 2930 2940 2950

refCD163 AAGACTTCAA GAAGGAAACA CTAATTGTTG TGGACGTGTG GAGATCTGGT
Thai-HP-1-CD163 AAGACTTCAA GAAGGAAACA CTAATTGTTG TGGACGTGTG GAGATCTGGT
Thai-EU-1-CD163 AAGACTTCAA GAAGGAAACA CTAATTGTTG TGGACGTGTG GAGATCTGGT
Thai-US-1-CD163 AAGACTTCAA GAAGGAAACA CTAATTGTTG TGGACGTGTG GAGATCTGGT

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 2960 2970 2980 2990 3000

refCD163 ACGGAGGTTG CTGGGGCACT GTGTGTGACG ACTCCTGGGA CCTTGAAGAT
Thai-HP-1-CD163 ACGGAGGTTG CTGGGGCACT GTGTGTGACG ACTCCTGGGA CCTTGAAGAT
Thai-EU-1-CD163 ACGGAGGTTG CTGGGGCACT GTGTGTGACG ACTCCTGGGA CCTTGAAGAT
Thai-US-1-CD163 ACGGAGGTTG CTGGGGCACT GTGTGTGACG ACTCCTGGGA CCTTGAAGAT

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 3010 3020 3030 3040 3050

refCD163 GCTCAGGTGG TGTGCCGACA GCTGGGCTGGT GGCTCAGCTT TGGAGGCAGG
Thai-HP-1-CD163 GCTCAGGTGG TGTGCCGACA GCTGGGCTGGT GGCTCAGCTT TGGAGGCAGG
Thai-EU-1-CD163 GCTCAGGTGG TGTGCCGACA GCTGGGCTGGT GGCTCAGCTT TGGAGGCAGG
Thai-US-1-CD163 GCTCAGGTGG TGTGCCGACA GCTGGGCTGGT GGCTCAGCTT TGGAGGCAGG

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 3060 3070 3080 3090 3100

refCD163 AAAAGAGCCC GCATTGGCC AGGGGACTGG GCCCATATGG CTCAAATGAAG
Thai-HP-1-CD163 AAAAGAGCCC GCATTGGCC AGGGGACTGG GCCCATATGG CTCAAATGAAG
Thai-EU-1-CD163 AAAAGAGCCC GCATTGGCC AGGGGACTGG GCCCATATGG CTCAAATGAAG
Thai-US-1-CD163 AAAAGAGCCC GCATTGGCC AGGGGACTGG GCCCATATGG CTCAAATGAAG

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 3110 3120 3130 3140 3150

refCD163 TGAAGTGCCTTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-HP-1-CD163 TGAAGTGCCTTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-EU-1-CD163 TGAAGTGCCTTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-US-1-CD163 TGAAGTGCCTTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC

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 3160 3170 3180 3190 3200

refCD163 TGGGGCCACCA GTGACTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-HP-1-CD163 TGGGGCCACCA GTGACTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-EU-1-CD163 TGGGGCCACCA GTGACTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC
Thai-US-1-CD163 TGGGGCCACCA GTGACTGTGG ACACAAGGAG GATGCTGCTG TGACGTGCTC

	3210 3220 3230 3240 3250
refCD163	AGAAAATTGCA AAGAGCCGAG AATCCCTACA TGCCACAGGT CGCTCATCTT
Thai-HP-1-CD163	AGAAAATTGCA AAGAGCCGAG AATCCCTACA TGCCACAGGT CGCTCATCTT
Thai-EU-1-CD163	AGAAAATTGCA AAGAGCCGAG AATCCCTACA TGCCACAGGT CGCTCATCTT
Thai-US-1-CD163	AGAAAATTGCA AAGAGCCGAG AATCCCTACA TGCCACAGGT CGCTCATCTT

	3260 3270 3280 3290 3300
refCD163	TTGTTGCACT TGCAATCTTT GGGGTCACTTG TGTTGCCCTG TCTCATCGCA
Thai-HP-1-CD163	TTGTTGCACT TGCAATCTTT GGGGTCACTTC TGTTGCCCTG TCTCATCGCA
Thai-EU-1-CD163	TTGTTGCACT TGCAATCTTT GGGGTCACTTC TGTTGCCCTG TCTCATCGCA
Thai-US-1-CD163	TTGTTGCACT TGCAATCTTT GGGGTCACTTC TGTTGCCCTG TCTCATCGCA

	3310 3320 3330 3340 3350
refCD163	TTCCTCATTT GGACTCAGAA GCGAAGACAG AGGCAGCGGC TCTCAGTTTT
Thai-HP-1-CD163	TTCCTCATTT GGACTCAGAA GCGAAGACAG AGGCAGCGGC TCTCAGTTTT
Thai-EU-1-CD163	TTCCTCATTT GGACTCAGAA GCGAAGACAG AGGCAGCGGC TCTCAGTTTT
Thai-US-1-CD163	TTCCTCATTT GGACTCAGAA GCGAAGACAG AGGCAGCGGC TCTCAGTTTT

	3360 3370 3380 3390 3400
refCD163	CTCAGGAGGA GAGAATTCTG T ACATCAAAT TCAATACCGG GAGATGAATT
Thai-HP-1-CD163	CTCAGGAGGA GAGAATTCTG T ACATCAAAT TCAATACCGG GAGATGAATT
Thai-EU-1-CD163	CTCAGGAGGA GAGAATTCTG T ACATCAAAT TCAATACCGG GAGATGAATT
Thai-US-1-CD163	CTCAGGAGGA GAGAATTCTG T ACATCAAAT TCAATACCGG GAGATGAATT

	3410 3420 3430 3440 3450
refCD163	CTTGCCTGAA AGCAGATGAA ACGGATATGC TAAATCCCTC AGGAGACCAC
Thai-HP-1-CD163	CTTGCCTGAA AGCAGATGAA ACGGATATGC TAAATCCCTC AGGAGACCAC
Thai-EU-1-CD163	CTTGCCTGAA AGCAGATGAA ACGGATATGC TAAATCCCTC AGGAGACCAC
Thai-US-1-CD163	CTTGCCTGAA AGCAGATGAA ACGGATATGC TAAATCCCTC AGGAGACCAC

	3460 3470 3480 3490 3500
refCD163	TCTGAAGTAC AATGAAAAGG AAAATGGGAA TTATAACCTG GTGAGTTCA
Thai-HP-1-CD163	TCTGAAGTAC AATGAAAAGG AAAATGGGAA TTATAACCTG GTGAGTTCA
Thai-EU-1-CD163	TCTGAAGTAC AATGAAAAGG AAAATGGGAA TTATAACCTG GTGAGTTCA
Thai-US-1-CD163	TCTGAAGTAC AATGAAAAGG AAAATGGGAA TTATAACCTG GTGAGTTCA

	3510 3520 3530
Sus scrofa	CCTTTAACAGAT ACCTTGATGA AGACCTGGAC TA
Thai-HP-1-CD163	CCTTTAACAGAT ACCTTGATGA AGACCTGGAC TA
Thai-EU-1-CD163	CCTTTAACAGAT ACCTTGATGA AGACCTGGAC TA
Thai-US-1-CD163	CCTTTAACAGAT ACCTTGATGA AGACCTGGAC TA

Figure 4.13 Nucleotide sequence alignments of the reference CD163 (refCD163) (accession number is NM_213976) with recombinant plasmids of Thai-HP-1-pCR®-XL-TOPO®-CD163 (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163) and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163), respectively. The nucleotide changes are boxed.

Table 4.8 Deduced amino acid sequences of 3 recombinant porcine CD163 (full) plasmids

Product's name	Sequence
1. Thai-HP-1- CD163 (1,133 aa)	MVLLEDSGSADFRRCSAHLSSFTFAVVAVLASACLVTSSLGGKDKEELRLTGGE NKCSRVEVKQEEWGTVCNNGWDMDVSVCRQLGCPTAIKATGWANFS AGSGRIWMDHVSCRGNESALWDCKHDGWGKHNCTHQQDAGVTCSDGSD LEMRLVNGGNRCLGRIEVKFQERWGTVCDDNFNINHASVCKQLECGSAVS FSGSANFGEGSGPIWFDDLVCNGNESALWNCKHEGWGKHNCDAEDAGV ICLNGADLKLRLVVDGVTECSGRLEVFKFQGEWGTICDDGWDSDDAAVACKQ LGCPTAVTAIGRVNASEGTGHIWLDVSCHGHESALWQCRHHEWGKHYCN HNEDAGVTCSDGSDLELRLKGGGSHCAGTVEVEIQKLVGKVCDRSWGLKE ADVVCRQLGCGSALKTSYQVYSKTAKNTWLFVSSCNGNETSLWDCKNWQ WGGLSCAHYDEAKITCSAHRKPRLVGGDIPCSGRVEVQHGDTWGTVCDSDF SLEAASVLCRELQCGTVWSLLGGAHFGEKGQIWAEEFQCEGHESHLSLC PVAPRPDGTCHSRDGVVCSRYTQIRLVNGKTPCEGRVELNILGSWGLCN SHWDMEDAHLVLCQQLKCGVALSIPGGAPFGKGSEQVWRHMFHCTGTEKH MGDCSVTALGASLCSSGQVASVICSGNQSQTSPCNSSSDPSSIIEESGV ACIGSGQLRLVDGGRCAGRVEVPGASWGTICDDSWDLNDAHVCKQLS CGWAINATGSAHFGEGTGPIWLDEINCNGKESHIWQCHSHGWGRHNCRHK EDAGVICSEFMSLGLISENSRETCAGRLEVFYNGAWGSVGRNSMSPATVGVV CRQLGCADRGDISPASSDKTVSRHMWVDNVQCPKGPDTLWQCPSSPWKK RLASPSEETWITCANKIRLQEGNTNCFGRVEIWYGGSWGTVCDDSWDLEDA QVVCRQLGGSALEAGKEPAFGQGTGPIWLNEVKCKGNEPSLWDCPARSWG HSDCGHKEDAATCSEIAKSRESLHATGRSSFVALAIFGVILLACLIAFLIWTQ KRRQRQRQLSVFSGGENSVHQIQYREMNSCLKADETDMLNPSGDHSEVQ*K GKWEL*PGEFSL*DTLMKTWT
2. Thai-EU-1-	MVLLEDSGSADFRRCSAHLSSFTFAVVAVLASACLVTSSLGGKDKEELRLTGGE

CD163 (1,133 aa)	NKC S GRVEVKVQEEWGTVCNNGWDMDVVS V CRQLGCPTAIKATGWANFS AGSGRIWMDHV S CRGNESALWDCKHDG W GKHNC T HQQDAGVT C SDGSD LEMRLVNGGNRCLGRIEVKFQERWGT C DDNFNINHASVVCKQLECGSAVS FSGSANFGE G SGPIWF D DLVCNGNESALWNCKHEG W GKHNC D HAEDAGV ICLNGADLKLRVVDGVTECSGRLEVKFQGEWGT I DDGWD S DDAAVACKQ LGCPTAVTAIGRVNASEGTGHIWLD S V S CH G HESALWQCRH H EWGKHYCN HNEDAGVT C SDGSDLELRLKG GG SHCAGTVEVEIQKLVGK V CD R SWGLKE ADV V CRQLGCGSALKTSYQVYS K T A NTWLFVSSCNGNETSLWDCKNWQ WGGLSCDHYDEAKITCSAHRK P RLVGGDIPCSGRVEVQHGDTWGT V CDSD FSLEAASVLCRELQCGTV S LLGGAHFGEGSGQIWAEEFQCEGHESHLSLC PVAPRPDGTC H SRDVGVVCSRYTQIRLVNGKTPCEGRVELNILGSWGSLCN SHWDMEDAHVLCQQLKCGVALSIPGGAPFGKGSEQVWRHMFHCTGTEKH MGECSV T ALGASLCSSGQVASVICSGNQS Q TLSPCN SS DP SS I E ESGV ACIGSGQLRV D GGRCAGRVEVPGASWGT I DDSWDLND A HVVCKQLS CGWAINATGSAHFGE G TGPIWLDEINCNGKESHIWQCHSHGWGRHNC R HK EDAGVICPEFMSLGLISENSRETCAGRLEV F YNGAWGSVGRNSMSPATGVV CRQLGCADRGDISPASSDKTV S RHMWVDNVQCPKG P DTLWQCPSSPWKK RLASPSEETWITCANKIRLQEGNTNC S GRVEIWYGGSWGT C DDSWDLEDA QVVCRQLGCGSALEAGKEPAFGQGTGPIWLNEVKCKGNEPSLWDCPARSW GHSDCGHKEDA A AVTCSEIAKSRESLHATGRSSFVALAIFGVILLACLIAFLIWT QKRRQRQR L S V SGGENSVHQI Q YREMNSCLKADETDMLNP S GDHSEVQ* KGK W E L *PGEFSL*DTLMKTWT
3. Thai-US-1- CD163 (1,133 aa)	MVLLED S GSADFRRCSAHLSSFTFAVVAVLSACLVTSSLGGKDKE L RTG G NKCSRVEVKVQEEWGTVCNNGWDMDVVS V CRQLGCPTAIKATGWANFS AGSGRIWMDHV S CRGNESALWDCKHDG W GKHNC T HQQDAGVT C SDGSD LEMRLVNGGNRCLGRIEVKFQERWGT C DDNFNINHASVVCKQLECGSAVS FSGSANFGE G SGPIWF D DLVCNGNESALWNCKHEG W GKHNC D HAEDAGV

ICLNGADLKLRVVDGVTECSGRLEVKFQGEWGTICDDGWDSDDAAVACKQ
LGCPTAVTAIGRVNASEGTGHIWLDVSCHGHESALWQCRHHEWGKHYCN
HNEDAGVTCSDGSDLELRKGGGSHCAGTVEVEIQKLVGKCDRSWGLKE
ADVVCRQLGCGSALKTSYQVYSKTAKNTWLFVSSCNGNETSLWDCKNWQ
WGGLSCAHYDEAKITCSAHRKPRLVGGDIPCSGRVEVQHGDTWGTVCDS
FSLEAASVLCRELQCGTVSLLGGAHFGEFGSQIWAEEFQCEGHESHLSC
PVAPRPDGTCSHSRDVGWCSRTQIRLVNGKTPCEGRVELNILGSWGSLCN
SHWDMEDAHLVLCQQLKCGVALSIPGGAPFGKGSEQVWRHMFHCTGTEKH
MGDCSVTALGASLCSSGQVASVICSGNQSQTLPNCNSSSDPSSIISEESGV
ACIGSGQLRLVDGGRCAGRVEVPGASWGTICDDSWDLNDAHWVCKQLS
CGWAINATGSAHFGEGTGPIWLDEINCNGKESHIWQCHSHGWGRHNCRHK
EDAGVICSEFMSLRLISENSRETCAGRLEVFYNGAWGSVGRNSMSPATGVW
CRQLGCADRGDISPASSDKTVSRHMWVDNVQCPKGPDTLWQCPSSPWKK
RLASPSEETWITCANKIRLQEGRNTNCSGRVEIWYGGSWGTVCDDSWDLEDA
QVVCRQLGCGSALEAGKEPAFGQGTGPIWLNEVKCKGNEPSLWDCPARSW
GHSDCGHKEDAATCSEIAKSRESLHATGRSSFVALAIFGVILLACLIAFLIWT
QKRRQRQRQLSVFSGGENSVHQIQYREMNSCLKADETDMLNP SGD HSEVQ*
KGKWEL*PGEFSL*DTLMKTWT

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 410 420 430 440 450

refCD163 ADVVCRQLGC GSALKTSYQV YSKTKATNTW LFVSSCNGNE TSLWDCKNWQ
Thai-HP-1-CD163 ADVVCRQLGC GSALKTSYQV YSKTKATNTW LFVSSCNGNE TSLWDCKNWQ
Thai-EU-1-CD163 ADVVCRQLGC GSALKTSYQV YSKTKATNTW LFVSSCNGNE TSLWDCKNWQ
Thai-US-1-CD163 ADVVCRQLGC GSALKTSYQV YSKTKATNTW LFVSSCNGNE TSLWDCKNWQ

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 460 470 480 490 500

refCD163 WGGLSCDHYD EAKITCSAHR KPRLVGGDIP CSGRVEVQHG DTWGTVCDS
Thai-HP-1-CD163 WGGLSCDHYD EAKITCSAHR KPRLVGGDIP CSGRVEVQHG DTWGTVCDS
Thai-EU-1-CD163 WGGLSCDHYD EAKITCSAHR KPRLVGGDIP CSGRVEVQHG DTWGTVCDS
Thai-US-1-CD163 WGGLSCDHYD EAKITCSAHR KPRLVGGDIP CSGRVEVQHG DTWGTVCDS

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 510 520 530 540 550

refCD163 FSLEAASVLC RELQCGTVVS LLGGAHFGE SGQIWAEEFQ CEGHESHLSL
Thai-HP-1-CD163 FSLEAASVLC RELQCGTVVS LLGGAHFGE SGQIWAEEFQ CEGHESHLSL
Thai-EU-1-CD163 FSLEAASVLC RELQCGTVVS LLGGAHFGE SGQIWAEEFQ CEGHESHLSL
Thai-US-1-CD163 FSLEAASVLC RELQCGTVVS LLGGAHFGE SGQIWAEEFQ CEGHESHLSL

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 560 570 580 590 600

refCD163 CPVAPRPDGT CSHSRDVGVV CSRYTQIRLV NGKTPCEGRV ELNILGSWGS
Thai-HP-1-CD163 CPVAPRPDGT CSHSRDVGVV CSRYTQIRLV NGKTPCEGRV ELNILGSWGS
Thai-EU-1-CD163 CPVAPRPDGT CSHSRDVGVV CSRYTQIRLV NGKTPCEGRV ELNILGSWGS
Thai-US-1-CD163 CPVAPRPDGT CSHSRDVGVV CSRYTQIRLV NGKTPCEGRV ELNILGSWGS

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 610 620 630 640 650

refCD163 LCNSHWDMED AHVLQQQLKC GVALSIPGGA PFGKGSEQVW RHMFHCTGTE
Thai-HP-1-CD163 LCNSHWDMED AHVLQQQLKC GVALSIPGGA PFGKGSEQVW RHMFHCTGTE
Thai-EU-1-CD163 LCNSHWDMED AHVLQQQLKC GVALSIPGGA PFGKGSEQVW RHMFHCTGTE
Thai-US-1-CD163 LCNSHWDMED AHVLQQQLKC GVALSIPGGA PFGKGSEQVW RHMFHCTGTE

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 660 670 680 690 700

refCD163 KHMGDCSVTA LGASLCSSGQ VASVICSGNQ SQTLPNCNS SSDPSSSIIS
Thai-HP-1-CD163 KHMGDCSVTA LGASLCSSGQ VASVICSGNQ SQTLPNCNS SSDPSSSIIS
Thai-EU-1-CD163 KHMGDCSVTA LGASLCSSGQ VASVICSGNQ SQTLPNCNS SSDPSSSIIS
Thai-US-1-CD163 KHMGDCSVTA LGASLCSSGQ VASVICSGNQ SQTLPNCNS SSDPSSSIIS

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 710 720 730 740 750

refCD163 EESGVACIGS GQLRIVDGGG RCAGRVEVYP GASWGTICDD SWDLNDAHVV
Thai-HP-1-CD163 EESGVACIGS GQLRIVDGGG RCAGRVEVYP GASWGTICDD SWDLNDAHVV
Thai-EU-1-CD163 EESGVACIGS GQLRIVDGGG RCAGRVEVYP GASWGTICDD SWDLNDAHVV
Thai-US-1-CD163 EESGVACIGS GQLRIVDGGG RCAGRVEVYP GASWGTICDD SWDLNDAHVV

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 760 770 780 790 800

refCD163 CKQLSCGWAI NATGSAHFGE GTGPIWLDEI NCNGKESHIW QCHSHGWGRH
Thai-HP-1-CD163 CKQLSCGWAI NATGSAHFGE GTGPIWLDEI NCNGKESHIW QCHSHGWGRH
Thai-EU-1-CD163 CKQLSCGWAI NATGSAHFGE GTGPIWLDEI NCNGKESHIW QCHSHGWGRH
Thai-US-1-CD163 CKQLSCGWAI NATGSAHFGE GTGPIWLDEI NCNGKESHIW QCHSHGWGRH

	810 820 830 840 850
refCD163	NCRHKEDAGV ICSEFMSI R L ISENSRETCA GRLEVFYNGA WGSVGRNSMS
Thai-HP-1-CD163	NCRHKEDAGV ICSEFMSI G L ISENSRETCA GRLEVFYNGA WGSVGRNSMS
Thai-EU-1-CD163	NCRHKEDAGV ICPEFMSI G L ISENSRETCA GRLEVFYNGA WGSVGRNSMS
Thai-US-1-CD163	NCRHKEDAGV ICSEFMSI R L ISENSRETCA GRLEVFYNGA WGSVGRNSMS

	860 870 880 890 900
refCD163	PATGVVVCRQ LGCADRGDIS PASSDKTVSR HMWVDNVQCP KGPDTLWQCP
Thai-HP-1-CD163	PATGVVVCRQ LGCADRGDIS PASSDKTVSR HMWVDNVQCP KGPDTLWQCP
Thai-EU-1-CD163	PATGVVVCRQ LGCADRGDIS PASSDKTVSR HMWVDNVQCP KGPDTLWQCP
Thai-US-1-CD163	PATGVVVCRQ LGCADRGDIS PASSDKTVSR HMWVDNVQCP KGPDTLWQCP

	910 920 930 940 950
refCD163	SSPWKKRLAS PSEETWITCA NKIRLQEGNT NC S GRVEIWY GGSWGTVCDD
Thai-HP-1-CD163	SSPWKKRLAS PSEETWITCA NKIRLQEGNT NC I GRVEIWY GGSWGTVCDD
Thai-EU-1-CD163	SSPWKKRLAS PSEETWITCA NKIRLQEGNT NC S GRVEIWY GGSWGTVCDD
Thai-US-1-CD163	SSPWKKRLAS PSEETWITCA NKIRLQEGNT NC S GRVEIWY GGSWGTVCDD

	960 970 980 990 1000
refCD163	SWDLEDAQVV CRQLGCGSAL EAGKEPAFGQ GTGPIWLNEV KCKGNEPSLW
Thai-HP-1-CD163	SWDLEDAQVV CRQLG*GSAL EAGKEPAFGQ GTGPIWLNEV KCKGNEPSLW
Thai-EU-1-CD163	SWDLEDAQVV CRQLGCGSAL EAGKEPAFGQ GTGPIWLNEV KCKGNEPSLW
Thai-US-1-CD163	SWDLEDAQVV CRQLGCGSAL EAGKEPAFGQ GTGPIWLNEV KCKGNEPSLW

	1010 1020 1030 1040 1050
refCD163	DCPARSWGHS DCGHKEDAAV TCSEIAKSRE SLHATGRSSF VALAIFGVIL
Thai-HP-1-CD163	DCPARSWGHS DCGHKEDAAV TCSEIAKSRE SLHATGRSSF VALAIFGVIL
Thai-EU-1-CD163	DCPARSWGHS DCGHKEDAAV TCSEIAKSRE SLHATGRSSF VALAIFGVIL
Thai-US-1-CD163	DCPARSWGHS DCGHKEDAAV TCSEIAKSRE SLHATGRSSF VALAIFGVIL

	1060 1070 1080 1090 1100
refCD163	LACLIAFLIW TQKRRQRQRL SVFSGGENSV HQIQYREMNS CLKADETDML
Thai-HP-1-CD163	LACLIAFLIW TQKRRQRQRL SVFSGGENSV HQIQYREMNS CLKADETDML
Thai-EU-1-CD163	LACLIAFLIW TQKRRQRQRL SVFSGGENSV HQIQYREMNS CLKADETDML
Thai-US-1-CD163	LACLIAFLIW TQKRRQRQRL SVFSGGENSV HQIQYREMNS CLKADETDML

	1110 1120 1130
refCD163	NPSGDHSEVQ *KGKWEL*PG EFSL*DTLMK TWT
Thai-HP-1-CD163	NPSGDHSEVQ *KGKWEL*PG EFSL*DTLMK TWT
Thai-EU-1-CD163	NPSGDHSEVQ *KGKWEL*PG EFSL*DTLMK TWT
Thai-US-1-CD163	NPSGDHSEVQ *KGKWEL*PG EFSL*DTLMK TWT

Figure 4.14 Deduced amino acid alignments of the reference CD163 (refCD163) (GenBank accession number is NP_999141) with recombinant plasmids of Thai-HP-1-pCR®-XL-TOPO®-CD163 (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163) and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163), respectively. The amino acid changes are boxed.

2.4 Nucleotide and deduced amino acid sequences of recombinant porcine CD163 (domain 5) plasmids

Three recombinant plasmids of Thai-HP-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5) were selected and sequenced. The complete sequences of 3 recombinant porcine CD163 (domain 5) plasmids were successfully sequenced at the same length of 395 bp and were analyzed alignment with the available databases of refCD163 cDNA (accession number is NM_213976) from the alveolar macrophages of normal pigs as shown in Table 4.9 and Figure 4.15, respectively. The results revealed that the nucleotide sequences of 3 recombinant porcine CD163 (domain 5) plasmids were the same with refCD163 cDNA (accession number is NM_213976).

The deduced amino acids sequences and alignment of recombinant porcine CD163 (domain 5) plasmids (131 amino acids) from PRRVS infected PAM with refCD163 (accession number is NP_999141) (131 amino acids) from normal PAM were shown in Table 4.10 and Figure 4.16, respectively. The results revealed that the amino acid sequences of 3 recombinant porcine CD163 (domain 5) plasmids were not different with the same refCD163 cDNA.

Table 4.9 Nucleotide sequences of 3 recombinant porcine CD163 (domain 5) plasmids

Product's name	Sequence
1. Thai-HP-1- CD163-DO5 (395 bp)	ggacattccctgctcggtcggtgaagtacaacatggagacacgtgggcaccgtctgtgattct gacttctctggaggcgccagcgtgctgcaggaaactacagtgcggcactgtggttccctc ctggggggagctacttggagaaggaagtggacagatctggctgaagaattccagtgtgagg ggcacgagtcccacccactctgcccagtagcacccgcctgacggacatgttagccacag cagggacgtcggcgtagtcgtcaagatacacacaaaatccgttggtaatggcaagaccca tgtgaaggaagagtggagctcaacatttggcctggggccctgtcaactctactggac atgg
2. Thai-EU-1- CD163-DO5 (395 bp)	ggacattccctgctcggtcggtgaagtacaacatggagacacgtgggcaccgtctgtgattct gacttctctggaggcgccagcgtgctgcaggaaactacagtgcggcactgtggttccctc ctggggggagctacttggagaaggaagtggacagatctggctgaagaattccagtgtgagg ggcacgagtcccacccactctgcccagtagcacccgcctgacggacatgttagccacag cagggacgtcggcgtagtcgtcaagatacacacaaaatccgttggtaatggcaagaccca tgtgaaggaagagtggagctcaacatttggcctggggccctgtcaactctactggac atgg
3. Thai-US-1- CD163-DO5 (395 bp)	ggacattccctgctcggtcggtgaagtacaacatggagacacgtgggcaccgtctgtgattct gacttctctggaggcgccagcgtgctgcaggaaactacagtgcggcactgtggttccctc ctggggggagctacttggagaaggaagtggacagatctggctgaagaattccagtgtgagg ggcacgagtcccacccactctgcccagtagcacccgcctgacggacatgttagccacag cagggacgtcggcgtagtcgtcaagatacacacaaaatccgttggtaatggcaagaccca tgtgaaggaagagtggagctcaacatttggcctggggccctgtcaactctactggac atgg

Figure 4.15 Nucleotide sequence alignments of the reference CD163 (refCD163) (accession number is NM_213976) with recombinant plasmids of Thai-HP-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5), respectively. The nucleotide change is boxed.

Table 4.10 Deduced amino acid sequences of 3 recombinant porcine CD163 (domain 5) plasmids

Product's name	Sequence
1. Thai-HP-1- CD163-DO5 (131 aa)	GHSLLWSC*STTWRHVGHRL*F*LLSGGGQRAVQGTTVRHCGFPPGGSSLWR RKWTDLG*RIPV*GARVPPFTLPSSTPP*RDM*PQQGRRRSLLKIHTNPLGEWQ DPM*RKSGAQHSWVLGVPLQLSLGH
2. Thai-EU-1- CD163-DO5 (131 aa)	GHSLLWSC*STTWRHVGHRL*F*LLSGGGQRAVQGTTVRHCGFPPGGSSLWR RKWTDLG*RIPV*GARVPPFTLPSSTPP*RDM*PQQGRRRSLLKIHTNPLGEWQ DPM*RKSGAQHSWVLGVPLQLSLGH
3. Thai-US-1- CD163-DO5 (131 aa)	GHSLLWSC*STTWRHVGHRL*F*LLSGGGQRAVQGTTVRHCGFPPGGSSLWR RKWTDLG*RIPV*GARVPPFTLPSSTPP*RDM*PQQGRRRSLLKIHTNPLGEWQ DPM*RKSGAQHSWVLGVPLQLSLGH

Figure 4.16 Deduced amino acid alignments of the reference CD163 (refCD163) (accession number is NP_999141) with recombinant plasmids of Thai-HP-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5), respectively. The amino acid change is boxed.

3. Nucleotide and deduced amino acid compositions of recombinant porcine Sn and CD163 plasmids

3.1 Nucleotide compositions of recombinant porcine Sn plasmids

The nucleotide compositions of recombinant porcine Sn (full and N-terminal domain) plasmids were shown as in Table 4.11. The results indicated that the number of the Cytosine nucleotide of porcine Sn (full) from 3 recombinant plasmids of Thai-HP-1-pCR[®]-XL-TOPO[®]-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR[®]-XL-TOPO[®]-Sn (Thai-EU-1-Sn), Thai-US-1-pCR[®]-XL-TOPO[®]-Sn (Thai-US-1-Sn) and the refSn cDNA from normal PAM (accession number is NM_214346) was higher than that of the other nucleotides.

The number of Cytosine nucleotide compositions in porcine Sn (N-terminal domain) of 3 recombinant plasmids, Thai-HP-1-pCR[®]8-GW-TOPO[®]-N-terminal Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-US-1-N-terminal-Sn), from PRRSV infected PAM and the refSn cDNA from normal PAM was higher than that of the remaining nucleotides.

Table 4.11 Nucleotide compositions of recombinant porcine Sn plasmids

Nucleotide	Sn							
	Number of nucleotide							
	refSn	Full			refSn	N-terminal domain		
		Thai-HP-1	Thai-EU-1	Thai-US-1		Thai-HP-1	Thai-EU-1	Thai-US-1
A	924	923	926	923	203	202	203	202
C	1,803	1,806	1,801	1,804	336	337	336	337
G	1,486	1,485	1,485	1,486	293	292	293	292
T	980	979	981	980	192	193	192	193
Total	5,193	5,193	5,193	5,193	1,024	1,024	1,024	1,024

Note: (A: Adenine; C: Cytosine; G: Guanine; T: Thymine)

3.2 Deduced amino acid compositions of recombinant porcine Sn plasmids

The deduced amino acid compositions of recombinant porcine Sn (full and N-terminal domain) plasmids were shown in Table 4.12. The number of Leucine, Alanine and Serine of 3 recombinant porcine Sn (full) plasmids, Thai-HP-1-pCR[®]-XL-TOPO[®]-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR[®]-XL-TOPO[®]-Sn (Thai-EU-1-Sn) and Thai-US-1-pCR[®]-XL-TOPO[®]-Sn (Thai-US-1-Sn), and the refSn from normal PAM (accession number is NP_999511) was higher than that of the other amino acids.

The results of deduced amino acid compositions of recombinant porcine Sn (N-terminal domain) plasmids revealed that the number of the Valine, Leucine and Serine from 3 recombinant porcine Sn (N-terminal domain) plasmids, Thai-HP-1-pCR[®]8-GW-TOPO[®]-N-terminal Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-US-1-N-terminal-Sn), and the refSn from normal PAM (accession number is NP_999511) was higher than that of the remaining amino acids.

Table 4.12 Deduced amino acid compositions of recombinant porcine Sn plasmids

Amino acids	Abbreviation	Sn									
		Number of amino acids									
		Full			N-terminal domain						
		refSn	Thai-HP-1	Thai-EU-1	Thai-US-1	refSn	Thai-HP-1	Thai-EU-1	Thai-US-1		
Alanine	Ala	A	183	183	183	183	20	20	20	20	20
Cysteine	Cys	C	41	41	41	41	8	9	8	9	9
Aspartate	Asp	D	65	65	65	65	12	12	11	12	12
Glutamate	Glu	E	80	80	80	80	17	17	17	17	17
Phenylalanine	Phe	F	36	36	36	36	9	9	9	9	9
Glycine	Gly	G	124	123	124	123	23	22	23	22	22
Histidine	His	H	52	52	52	52	11	12	12	12	12
Isoleucine	Ile	I	34	34	34	34	11	11	11	11	11
Lysine	Lys	K	27	27	27	27	13	13	13	13	13
Leucine	Leu	L	219	218	219	218	36	36	35	36	36
Methionine	Met	M	20	20	20	20	6	6	6	6	6
Asparagine	Asn	N	50	50	50	50	12	12	12	12	12
Proline	Pro	P	111	111	111	111	20	20	20	20	20
Glutamine	Gln	Q	91	91	91	91	23	23	23	23	23
Arginine	Arg	R	97	98	97	98	14	15	14	15	15
Serine	Ser	S	173	174	173	174	36	35	35	35	35
Threonine	Thr	T	117	117	117	117	19	19	19	19	19
Valine	Val	V	145	145	145	145	37	37	38	37	37
Tryptophan	Trp	W	25	25	25	25	7	7	7	7	7
Tyrosine	Tyr	Y	40	40	40	40	7	6	7	6	6
Total			1,730	1,730	1,730	1,730	341	341	341	341	341

3.3 Nucleotide compositions of recombinant porcine CD163 plasmids

The nucleotide compositions of recombinant porcine CD163 (full and domain 5) plasmids were shown as in Table 4.13. The results indicated that the numbers of the Guanine nucleotide of 3 recombinant porcine CD163 (full) plasmids, Thai-HP-1-pCR®-XL-TOPO®-CD163 (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163), and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163), and the refCD163 cDNA from normal PAM (accession number is NM_213976) was the highest number. In contrast, the numbers of the Cytosine nucleotide was the lowest number.

The results of nucleotide compositions of recombinant porcine CD163 (domain 5) plasmids revealed that the numbers of the Guanine nucleotide from 3 recombinant porcine CD163 (domain 5) plasmids, Thai-HP-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5), and the refCD163 cDNA normal PAM (accession number is NM_213976) was the highest numbers and followed by the number of Cytosine nucleotide.

Table 4.13 Nucleotide compositions of recombinant porcine CD163 plasmids

Nucleotide	CD163							
	Number of nucleotide							
	refCD163	Full			N-terminal domain			
		Thai-HP-1	Thai-EU-1	Thai-US-1	refCD163	Thai-HP-1	Thai-EU-1	Thai-US-1
A	867	868	867	867	83	83	83	83
C	687	685	685	686	104	104	104	104
G	1,018	1,019	1,021	1,019	123	123	123	123
T	828	828	827	826	85	85	85	85
Total	3,400	3,400	3,400	3,400	395	395	395	395

Note: (A: Adenine; C: Cytosine; G: Guanine; T: Thymine)

3.4 Dduced amino acid compositions of recombinant porcine CD163 plasmids

The deduced amino acid compositions of recombinant porcine CD163 (full and domain 5) plasmids were shown as in Table 4.14. The number of the Glycine and Serine amino acids of 3 recombinant porcine CD163 (full) plasmids, Thai-HP-1-pCR®-XL-TOPO®-CD163 (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163), and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163), and the refCD163 from normal PAM (accession number is NP_999141) was higher than that of the other amino acids.

The results of deduced amino acid compositions of recombinant porcine CD163 (domain 5) plasmids revealed that the number of the Glycine and Leucine amino acid from 3 recombinant porcine CD163 (domain 5) cDNA plasmids, Thai-HP-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®8-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5), and the refCD163 from normal PAM (accession number is NP_999141) was higher than that of the remaining amino acids.

Table 4.14 Deduced amino acid compositions of recombinant porcine CD163 plasmids

Amino acids	Abbreviation	CD163								
		Number of amino acids								
		Full				Domain 5				
		ref	Thai-	Thai-	Thai-	ref	Thai-	Thai-	Thai-	Thai-
		CD163	HP-1	EU-1	US-1	CD163	HP-1	EU-1	US-1	
Alanine	Ala	A	72	73	72	73	3	3	3	3
Cysteine	Cys	C	76	75	76	76	2	2	2	2
Aspartate	Asp	D	66	65	65	65	3	3	3	3
Glutamate	Glu	E	70	70	71	70	1	1	1	1
Phenylalanine	Phe	F	25	26	25	25	3	3	3	3
Glycine	Gly	G	130	131	131	130	16	16	16	16
Histidine	His	H	42	42	42	42	7	7	7	7
Isoleucine	Ile	I	38	38	38	38	2	2	2	2
Lysine	Lys	K	47	47	47	47	3	3	3	3
Leucine	Leu	L	81	80	80	80	15	15	15	15
Methionine	Met	M	13	13	13	13	2	2	2	2
Asparagine	Asn	N	46	46	46	46	1	1	1	1
Proline	Pro	P	28	28	29	28	12	12	12	12
Glutamine	Gln	Q	40	40	40	40	7	7	7	7
Arginine	Arg	R	52	51	51	52	13	13	13	13
Serine	Ser	S	115	114	114	115	12	12	12	12
Threonine	Thr	T	51	51	51	51	8	8	8	8
Valine	Val	V	84	85	85	85	7	7	7	7
Tryptophan	Trp	W	45	45	45	45	6	6	6	6
Tyrosine	Tyr	Y	9	9	9	9	0	0	0	0
Total			1,133	1,133	1,133	1,133	131	131	131	131

4. The homology of nucleotide and deduced amino acid sequences of recombinant porcine Sn and CD163 plasmids

4.1 The homology of nucleotide and deduced amino acid sequences of 3 recombinant porcine Sn (full) plasmids

The recombinant porcine Sn (full) plasmids of Thai-HP-1-pCR®-XL-TOPO®-Sn (Thai-HP-1-Sn), Thai-EU-1-pCR®-XL-TOPO®-Sn (Thai-EU-1-Sn) and Thai-US-1-pCR®-XL-TOPO®-Sn (Thai-US-1-Sn) were further compared with the refSn cDNA from GenBank (accession number is NM_214346) by using Bioedit Sequence Alignment Editor software. The percentage homology among those recombinant plasmids and refSn database were demonstrated in Table 4.15. The results showed high nucleotide homology at 99.8 - 99.9 % identities. The deduced amino acid sequences revealed that among those recombinant plasmids, Thai-HP-1-pCR®-XL-TOPO®-Sn, Thai-EU-1-pCR®-XL-TOPO®-Sn and Thai-US-1-pCR®-XL-TOPO®-Sn, were similar at 99.8 - 100 % amino acid identities.

Table 4.15 Comparison homology (%) of nucleotide and deduced amino acid sequences of the refSn and 3 recombinant porcine Sn (full) plasmids (Nucleotide identity (%) in lower triangle of table; Deduced amino acid identity (%) in upper triangle of table)

Sequences	refSn	Thai-HP-1-Sn	Thai-EU-1-Sn	Thai-US-1-Sn
refSn		99.8	100	99.8
Thai-HP-1-Sn	99.8		99.8	100
Thai-EU-1-Sn	99.8	99.8		99.8
Thai-US-1-Sn	99.8	99.9	99.8	

4.2 The homology of nucleotide and deduced amino acid sequences of recombinant porcine Sn (N-terminal domain) plasmids

The recombinant porcine Sn (N-terminal domain) plasmids of Thai-HP-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-HP-1-N-terminal-Sn), Thai-EU-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-EU-1-N-terminal-Sn) and Thai-US-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn (Thai-US-1-N-terminal-Sn) were compared with refSn from GenBank (accession number is NM_214346) by using Bioedit Sequence Alignment Editor software. The number homology among these recombinant porcine Sn (N-terminal domain) plasmids, Thai-HP-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn, Thai-EU-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn and Thai-US-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn, and refSn were shown in Table 4.16. The results showed high nucleotide identities at 99.8 - 100 % homology. The deduced amino acid sequences revealed that the recombinant porcine Sn (N-terminal domain) plasmids, Thai-HP-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn, Thai-EU-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn and Thai-US-1-pCR[®]8-GW-TOPO[®]-N-terminal-Sn, were similar at 99.7 - 100 % of amino acid identities.

Table 4.16 Comparison homology (%) of nucleotide and deduced amino acid sequences of the refSn and 3 recombinant porcine Sn (N-terminal domain) plasmids (Nucleotide identity (%) in lower triangle of table; Deduced amino acid identity (%) in upper triangle of table)

Sequences	refSn	Thai-HP-1-N-terminal-Sn	Thai-EU-1-N-terminal-Sn	Thai-US-1-N-terminal-Sn
refSn		99.7	100	99.7
Thai-HP-1-N-terminal-Sn	99.8		99.7	100
Thai-EU-1-N-terminal-Sn	100	99.8		99.7
Thai-US-1-N-terminal-Sn	99.8	100	99.8	

4.3 The homology of nucleotide and deduced amino acid sequences of recombinant porcine CD163 (full) plasmids

The recombinant porcine CD163 (full) plasmids of Thai-HP-1-pCR®-XL-TOPO®-CD163 (Thai-HP-1-CD163), Thai-EU-1-pCR®-XL-TOPO®-CD163 (Thai-EU-1-CD163), and Thai-US-1-pCR®-XL-TOPO®-CD163 (Thai-US-1-CD163), were further analyzed with refCD163 (refCD163) from GenBank (accession number is NM_213976) by using Bioedit Sequence Alignment Editor software. The homology among these recombinant porcine CD163 (full) plasmids, Thai-HP-1-pCR®-XL-TOPO®-CD163, Thai-EU-1-pCR®-XL-TOPO®-CD163 and Thai-US-1-pCR®-XL-TOPO®-CD163, and refCD163 were shown in Table 4.17. The results of nucleotide sequences revealed high percent homology at 99.7 - 99.8 %

homology. The deduced amino acid sequences revealed that the recombinant porcine CD163 (full) plasmids of Thai-HP-1-pCR®-XL-TOPO®-CD163, Thai-EU-1-pCR®-XL-TOPO®-CD163 and Thai-US-1-pCR®-XL-TOPO®-CD163 were similar at 99.5, 99.6 and 99.7 % of amino acid identities, respectively.

Table 4.17 Comparison homology (%) of nucleotide and deduced amino acid sequences of the refCD163 and 3 recombinant porcine CD163 (full) plasmids (Nucleotide identity (%) in lower triangle of table; Deduced amino acid identity (%) in upper triangle of table)

Sequences	refCD163	Thai-HP-1-	Thai-EU-1-	Thai-US-1-
		CD163	CD163	CD163
refCD163		99.5	99.6	99.8
Thai-HP-1-CD163	99.7		99.5	99.7
Thai-EU-1-CD163	99.7	99.8		99.6
Thai-US-1-CD163	99.8	99.8	99.8	

4.4 The homology of nucleotide and deduced amino acid sequences of recombinant porcine CD163 (domain 5) cDNA plasmids

The recombinant porcine CD163 (domain 5) plasmids of Thai-HP-1-pCR®-GW-TOPO®-CD163-DO5 (Thai-HP-1-CD163-DO5), Thai-EU-1-pCR®-GW-TOPO®-CD163-DO5 (Thai-EU-1-CD163-DO5) and Thai-US-1-pCR®-GW-TOPO®-CD163-DO5 (Thai-US-1-CD163-DO5) were compared with refCD163 from GenBank (accession number is NM_213976) using Bioedit Sequence Alignment Editor software. The results of nucleotide sequences revealed at 100 % homology. The number homology of deduced amino acid among these

recombinant porcine CD163 (domain 5) plasmids of Thai-HP-1-pCR[®]-GW-TOPO[®]-CD163-DO5, Thai-EU-1-pCR[®]-GW-TOPO[®]-CD163-DO5 and Thai-US-1-pCR[®]-GW-TOPO[®]-CD163-DO5 and refCD163 were shown in Table 4.18. The results revealed high deduced amino acid at 100 % identities.

Table 4.18 Comparison homology (%) of nucleotide and deduced amino acid sequences of refCD163 and 3 recombinant porcine CD163 (domain 5) plasmids (Nucleotide identity (%) in lower triangle of table; Deduced amino acid identity (%) in upper triangle of table)

Sequences	refCD163	Thai-HP-1-CD163-DO5	Thai-EU-1-CD163-DO5	Thai-US-1-CD163-DO5
refCD163		100	100	100
Thai-HP-1-CD163-DO5	100		100	100
Thai-EU-1-CD163-DO5	100	100		100
Thai-US-1-CD163-DO5	100	100	100	

CHAPTER V

DISCUSSION

PRRS has caused severe economic loss in most swine-producing countries. For a basic research on pathogenesis of PRRSV infection in nursery pigs, porcine Sn and CD163 which have been identified as the major receptors for PRRSV (Calvert et al., 2007; Van Gorp et al., 2008) infection on the macrophages were studied. In this thesis, the author first reported in Thailand about the cloning using RT-PCR technique, the expression level using gel densitometry, the genetic and deduced amino acid sequences and both compositions of the porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) on alveolar macrophages of nursery pigs infected with 3 strains of PRRSV. The author also demonstrated the homology and the comparison of both receptor data among the 3 PRRSV strains.

In general, there are many methods to study the comparison of the gene expression level including gel densitometry (Dozois et al., 1997; Schmittgen et al., 2000; Etienne et al., 2004). The author selected the GAPDH gene as internal controls for levels of housekeeping genes because this gene was usually used for RT-PCR analysis in human tissues (Suzuki et al., 2000; Sila-Asna et al., 2007) and porcine tissues (Foss et al., 1998). The author found that US strain of PRRSV infected PAM samples were showed higher expression level of both porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) than that of EU and HP strains of PRRSV infected PAM. This finding in part suggested that higher expression of porcine Sn and CD163 were higher the susceptibility to PRRSV infection and the US strain may be easily infected than the others strains (Jiang et al., 2012). In addition, an increase in the percentage of infected cells was correlated with an enhanced expression CD163 of porcine monocytes/macrophages in the process of ASFV infection. The study of Patton et al. (2009) showed that the expression level of CD163 was also correlated well with the overall

level of PRRSV replication. Moreover, the high expression level of human Sn receptor was relatively enhanced HIV infection and infectivity (Rempe et al., 2008). In addition, the infection study of PRRSV in the Chinese Dapulian (DPL) pigs were showed the lower gene expression level of porcine Sn and CD163 on macrophages, the lower rectal temperature and the lower PRRSV viral copy number than that of the commercial Duroc x Landrace x Yorkshire (DLY) crossbred pig indicated that DPL pigs were more resistant than DLY pigs (Jiang et al., 2012). In this thesis, the PAM samples were obtained from the PRRSV outbreak farms and the field data may have many factors involved. According to the limitation of PAM samples from the farms in Ratchaburi province only, the samples from other pig farms from different parts of Thailand should be further studied. The viremic PRRSV in sera samples of Thai nursery pigs should not only confirm the positive with S/P ELISA and RT-PCR, but also quantitatively measure the viral mRNA concentration by using Real-time PCR technique. Moreover, the correlation between the level of receptor expression and the efficiency of PRRSV infection among 3 strains is necessary to obtain a complete study. Furthermore, the comparison study of two receptor gene expressions between normal PAM and PAM infected with PRRSV should be more elucidated.

For an application of these receptors in enhancement of the PRRS vaccine development, porcine Sn and CD163 cDNA were first amplified and cloned into the recombinant plasmids. In this study, the porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA were cloned into pCR®8-GW-TOPO® vectors. However, only two short fragments of the porcine Sn (N-terminal domain) and CD163 (domain 5) were transformed with positive colonies but the porcine Sn (full) and CD163 (full) were not successfully transformed (no positive colony). Finally, the author has successfully transformed the porcine Sn (full) and CD163 (full) cDNA into pCR®-XL-TOPO® vectors. These recombinant plasmids were useful for study and analysis of the genetic and deduced amino acid sequences and composition of those receptors. There are many factors concerning in selecting the cloning and expression system depending on the biological

properties of interested proteins, the requirement of an amount of recombinant proteins, and the nature of the experiments (Geisse et al., 1996). The porcine Sn and CD163 cDNA were studied in many expression systems (Vanderheijden et al., 2003; Calvert et al., 2007; Pérez et al., 2008). A wide variety of animal receptors were expressed in many expression vectors such as yeast and baculovirus (Tate et al., 2003), whereas and *E.coli* bacterial expressing system was selected because it is cheap, fast and easy to manipulate.

In this thesis, the full length of porcine Sn and CD163 cDNA from 3 strains of PRRSV infected PAM in Thai nursery pigs were completely sequenced and reported at the first time. The composition and homology of nucleotide and deduced amino acid sequences were analyzed. The composition and the homology were also revealed highly conserved in the sequences of porcine Sn and CD163. In addition, the variability of nucleotide and deduced amino acid sequences of porcine Sn and CD163 cDNA from PRRSV infected PAM were not different among 3 strains. The nucleotide and deduced amino acid sequences from recombinant plasmids of porcine Sn and CD163 from 3 strains of PRRSV infected PAM samples revealed no different from the refSn and refCD163 from normal PAM samples. This study showed the same results as the studies by Srikumaran (2006) about Sn and Pérez et al. (2008) about CD163. Both researchers showed that the complete sequences of porcine Sn (Srikumaran, 2006) and CD163 (Pérez et al., 2008) cDNA from PAM samples of healthy pigs by cloning into mammalian system revealed the homology with both receptor ancestors. These results indicated that porcine Sn and CD163 cDNA is the conserved genes. Moreover, the author investigated the domain of porcine Sn that is involved in the interaction with PRRSV. The sialic acid-binding activity of porcine Sn by site-directed mutagenesis and whether the absence of sialic acid-binding activity may be effective the interaction of the porcine arterivirus with Sn (Delputte et al., 2007). The N-terminal domain of Sn was considered to be involved in PRRSV attachment to macrophage cells. Furthermore, An et al. (2010) constructed a series of truncated fragments of porcine Sn and expressed them in the non-permissive PK 15 cell line to investigate whether the N-terminal domain of

Sn is sufficient for PRRSV attachment. Their result showed that the first 150 amino acids comprising the entire first domain of the porcine Sn (N-terminal region) was necessary for PRRSV binding to cells, and the N-terminal domain alone was sufficient for virus attachment. The study of CD163 protein domains involved in PRRSV infection, Van Gorp et al. (2009) created CD163 deletion mutants and chimeric mutants (replacement experiments) and found that scavenger receptor cysteine-rich (SRCR) domain 5 (SRCR 5) consisted of 100 amino acids is essential for PRRSV infection. In this thesis, the genetic and deduced amino acids sequences of porcine Sn (N-terminal domain) and CD163 (domain 5) were investigated. The porcine Sn (N-terminal domain) and CD163 (domain 5) were successfully amplified and the sequence homology was then analyzed. The author demonstrated that the recombinant plasmids of porcine Sn (N-terminal domain) and CD163 (domain 5) of 3 strains of PRRSV infected PAM from Thai nursery pigs were showed the same patterns of the sequences and composition of nucleotide and deduced amino acids sequences as the refSn or refCD163 of PAM isolated from normal pigs. No mutation of deduced amino acid from two receptors (Sn and CD163) among 3 strains of PRRSV was detected in this study. However, the PRRSV infected PAM samples collected from different pig farms of Thailand should be more studied. In this thesis, the data about the composition and deduce amino acid of receptors from 3 strains of PRRSV showed no mutation. This finding will be useful for the development of monoclonal anti-receptor antibody, antiviral drugs or SiRNA to block their cellular receptors in the future (Calvez et al., 2004). However, the PRRSV controlling strategy is still difficult as the same as HIV (Human Immunodeficiency Virus) in human. Many factors or co-factors involved in the PRRSV pathogenesis in pigs such as mutation or variation between host receptors and virus invasion are required additional studies.

Taken together, gained knowledge in this thesis may be useful for further expression and production of these recombinant porcine Sn and CD163 proteins. These results also provide a preliminary data about the sequences and composition of nucleotide and deduce amino acid of both porcine Sn and CD163 after PRRSV infected PAM in Thai nursery pigs

and allow to understand more on characterization of porcine Sn (N-terminal domain) and CD163 (domain 5). The obtained data of the expression level of porcine Sn and CD163 cDNA among three PRRSV strains infected PAM may be contributed the understanding on the inhibition of PRRS virus and the control of PRRS virus infection in the future.

Conclusion

This thesis, the author performed the study of molecular biology of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) cDNA from PRRSV infected PAM. The amplicons of those porcine Sn and CD163 cDNA were successfully amplified using RT-PCR technique. Furthermore, the author also successfully cloned and constructed the recombinant plasmids of porcine Sn (full and N-terminal domain) and CD163 (full and domain 5) vectors. The results from this thesis showed that the alignment and composition of nucleotide and deduced amino acid sequences from porcine Sn and CD163 cDNA of 3 strains of PRRSV infected PAM are the same. No mutation of deduced amino acid was observed. The obtained knowledge and tools in this study contributes to the basic research of porcine Sn and CD163 expression level, the basic data of nucleotides and deduced amino acid.

For future work, since the applied research for improving the susceptible cell lines to PRRSV infection and for enhancing vaccines production, so recombinant porcine Sn and CD163 expression vectors are further constructed and the expression, purification and reactivity of these recombinant proteins are further studied.

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APPENDICES

APPENDIX A
Instrument and chemical substances

1. A -20°C refrigerator, Model SF-C997 (Sanyo, Thailand)
2. A -80°C refrigerator, Model 905 (Thermofisher Scientific, USA)
3. Centrifuge and Microcentrifuge
4. Experimental glasswares
5. Gel document system, Model GVM 20 (Synoptics, UK)
6. Gel electrophoresis system, Model GE-100 (Bioer technology Co. Ltd., China)
7. Heat block (Labnet International Inc., USA)
8. Incubator, Model BE-400 (Memmert Inc., Germany)
9. Lamina air flow, Model Bio II A (Telstar, Spain)
10. Micropipette (Labnet, USA) and Micropipette tips
11. PCR assay
 - 11.1 Agarose gel (Molecular grade)
 - 11.2 1kb DNA marker
 - 11.3 1kb DNA marker (BioLab, USA)
 - 11.4 Ethidium bromide 10mg/ml (Sigma Aldrich Inc., USA)
 - 11.5 Gel electrophoresis buffer (TAE)
 - 11.6 Loading dye (Fermantas, Canada)
 - 11.7 Itaq DNA polymerase (iNtRon, South Korea)
12. PCR cabinet (Biometra, Germany)
13. PCR tube and Microcentrifuge tube
14. Vortex, Model K 550-GE (Scientific Inc., USA)
15. Shaking incubator, Model 311DS (Labnet International Inc., USA)

APPENDIX B
Reagents and preparations

Reagents for agarose gel electrophoresis

1. 10 mg/ml Ethidium bromide

- Ethidium bromide 1 g
- Distilled deionized water 100 ml

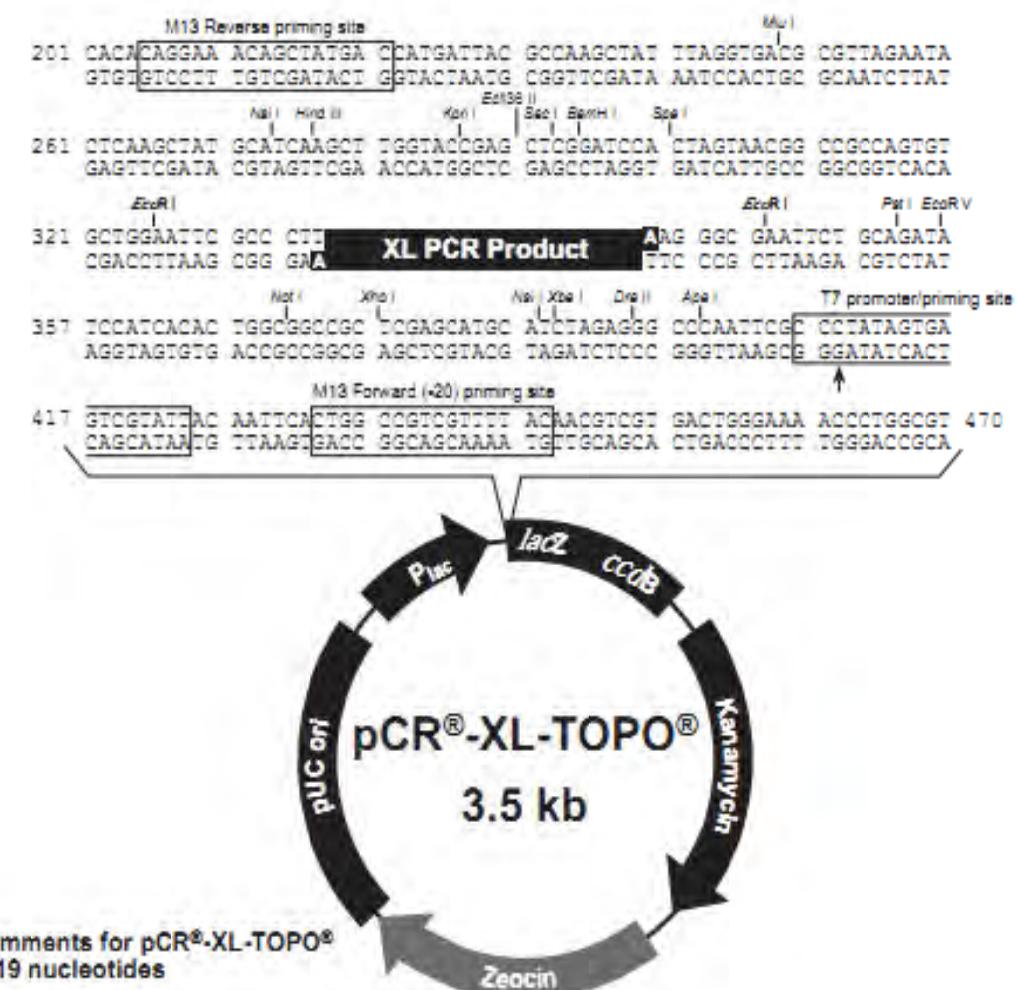
Add 1g of ethidium bromide to 100 ml of distill deionized water. Stir on a magnetic stirrer for several hours to ensure that the dye has dissolved. Wrap container in aluminum foil or transfer to a dark bottle and store at room temperature.

2. 25X TAE (Tris-Acetate buffer) 1,000 ml contains

- Tris base 242.0 g
- Glacial acetic acid 57.1 ml
- 0.5 M EDTA pH 8.0 100.0 ml
- Distilled deionized water (final volume) 1,000 ml

Add 242 g of Tris base, 57.1 ml of Glacial acetic acid and 100 ml of 0.5 M EDTA pH 8.0 to 500 ml of distilled deionized water and then adjusts the final volume to 1,000 ml. Sterilize the solution by autoclaving.

APPENDIX C
Physical map of plasmid pCR®-XL-TOPO®

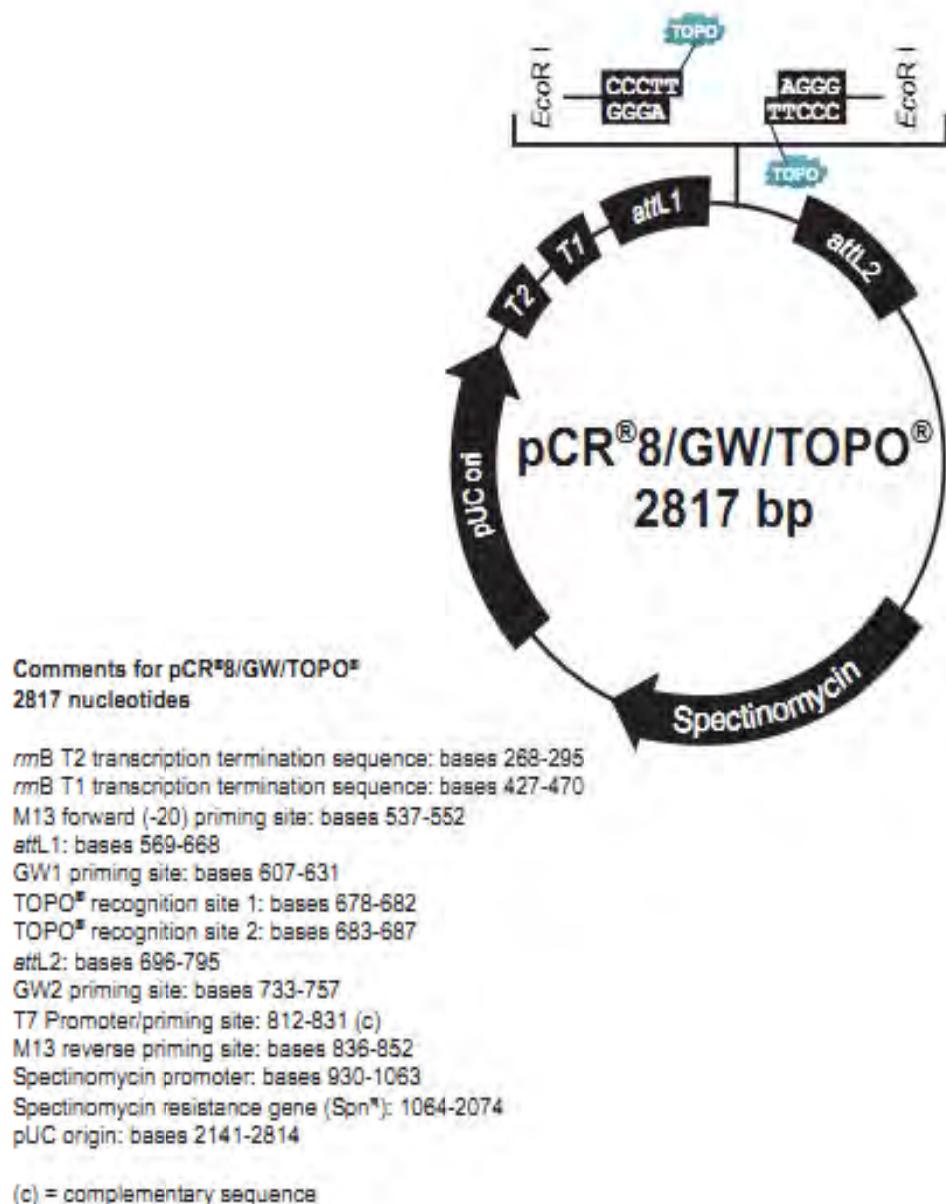


Comments for pCR®-XL-TOPO®
3519 nucleotides

Lac promoter/operator region: bases 95-216
 M13 Reverse priming site: bases 205-221
 Lac Zα ORF: bases 217-576
 Multiple Cloning Site: bases 248-399
 TOPO® Cloning site: bases 336-337
 T7 promoter/priming site: bases 406-425
 M13 Forward (-20) priming site: bases 433-448
 Fusion joint: bases 577-586
 ccdB lethal gene ORF: bases 586-888
 Kanamycin resistance ORF: bases 1237-2031
 Zeocin resistance ORF: bases 2238-2612
 pUC origin: bases 2680-3393

APPENDIX D

Physical map of plasmid pCR®8/GW/TOPO®



BIOGRAPHY

Mr. Vo Phong Vu Anh Tuan was born on 19 September 1978 in Chogao district, Tiengiang province, Vietnam. He is doctor of veterinary medicine, graduated from Faculty of Animal Science and Veterinary Medicine, Agriculture and Forestry University, Ho Chi Minh city, Vietnam in 2002. After graduation, he works as lecturer of major domesticated animal disease in department of Veterinary Medicine, Faculty of Animal Husbandry and Veterinary Medicine, Southern Agriculture College, The Ministry of Agriculture and Rural Development. In 2010, he applied and got the scholarship “The Graduate Scholarship Programs for Neighboring Countries” from Chulalongkorn University, Thailand. He studied in the field of Swine Medicine at Department of Veterinary Medicine, Faculty of Veterinary Science, Chulalongkorn University during 2010-2012.