

การสร้างน้ำมันระเหยในเซลล์เพาะเลี้ยงของโกจูพาล์พาจีน ยี่ห้อ กิมกิด และพิมเสน

นางศุภารรณ บุญระเทพ



วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรดุษฎีบัณฑิต

สาขาวิชาเกษตรเคมีและผลิตภัณฑ์ธรรมชาติ

คณะเกษตรศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย

ปีการศึกษา 2548

ISBN 974-17-55473

ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

ESSENTIAL OIL PRODUCTION IN CELL CULTURES OF *ARTEMISIA VULGARIS*
VAR. *INDICA*, *CUMINUM CYMINUM*, *FORTUNELLA JAPONICA*, AND *POGOSTEMON CABLIN*

Mrs. Supawan Bunrathep

A Dissertation Submitted in Partial Fulfillment of the Requirements for
the Degree of Doctor of Philosophy Program in Pharmaceutical Chemistry and Natural Products

Faculty of Pharmaceutical Sciences

Chulalongkorn University

Academic Year 2005

ISBN 974-17-5547-3

Thesis Title Essential oil production in cell cultures of *Artemisia vulgaris*
var. *indica*, *Cuminum cyminum*, *Fortunella japonica*, and
Pogostemon cablin

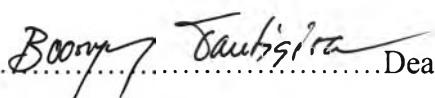
By Mrs. Supawan Bunrathep

Field of Study Pharmaceutical Chemistry and Natural Products

Thesis Advisor Associate Professor Nijssiri Ruangrungsi, Ph.D.

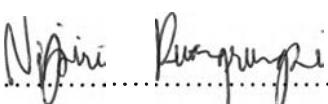
Thesis Co-advisor Thanapat Songsak, Ph.D.

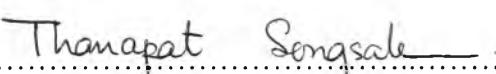
Accepted by the Faculty of Pharmaceutical Sciences, Chulalongkorn University in Partial Fulfillment of the Requirements for the Doctor's Degree

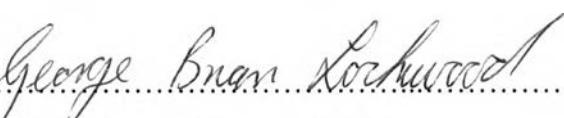
 Dean of the Faculty of Pharmaceutical Sciences
(Associate Professor Boonyong Tantisira, Ph.D.)

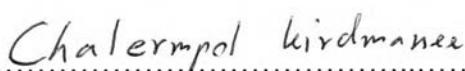
THESIS COMMITTEE

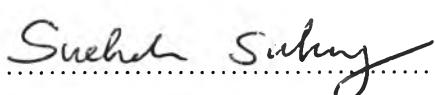
 Chairman
(Associate Professor Surattana Amnuoypol, Ph.D.)

 Thesis Advisor
(Associate Professor Nijssiri Ruangrungsi, Ph.D.)

 Thesis Co-advisor
(Thanapat Songsak, Ph.D.)

 Member
(George Brian Lockwood, Ph.D.)

 Member
(Chalermpol Kirdmanee, Ph.D.)

 Member
(Police Captain Suchada Sukrong, Ph. D.)

ศุภวรรณ บุญราษฎร์ : การสร้างน้ำมันระเหยในเซลล์เพาะเลี้ยงของโกฐุพาล้ำพาจีน ยี่หร่า กิมกิด และพิมเสน (ESSENTIAL OIL PRODUCTION IN CELL CULTURES OF *ARTEMISIA VULGARIS* VAR. *INDICA*, *CUMINUM CYMINUM*, *FORTUNELLA JAPONICA*, AND *POGOSTEMON CABLIN*) อ.ที่ปรึกษา : รศ.ดร.นิจศิริ เรืองรังษี, อ.ที่ปรึกษาร่วม : ดร.ธนกัตร ทรงสักดิ์, 199 หน้า. ISBN 974-17-5547-3

การศึกษาองค์ประกอบเคมีของน้ำมันระเหยของโกฐุพาล้ำพาจีน ยี่หร่า กิมกิด และพิมเสน ทำได้โดยกลั่นชิ้นส่วนต่างๆ ของพืชด้วยไอน้ำและนำน้ำมันระเหยที่ได้มาวิเคราะห์ด้วยวิธี Gas Chromatography-Mass Spectrometry พบว่าน้ำมันระเหยของพืชดังกล่าวประกอบด้วยสารประกอบเทอร์ปินอยค์หลายชนิด และมี (+)-davanone (71.59 %), cuminaldehyde (36.30 %), β -pinene (47.44 %), d-limonene (87.07 %) และ patchouli alcohol (60.30 %) เป็นองค์ประกอบเคมีหลักในใบโกฐุพาล้ำพาจีน ผลยี่หร่า ใบกิมกิด เปลือกผลกิมกิด และใบพิมเสน ตามลำดับ เพื่อที่จะศึกษาองค์ประกอบเคมีของน้ำมันระเหยของเซลล์เพาะเลี้ยงของพืชเหล่านี้ ชิ้นส่วนของพืชชนิดต่างๆ จึงถูกนำมาซึ่งที่พิเศษและซักนำให้เกิดเป็นแคลส์บันอาหารเพาะเลี้ยงกึ่งแข็งชนิด MS ที่ประกอบด้วยสารควบคุมการเจริญเติบโตชนิดต่างๆ แล้วจึงเพาะเลี้ยงในสภาวะที่เหมาะสมสำหรับพืชแต่ละชนิด หลังจากนั้นเปลี่ยนถ่ายแคลส์ที่สมบูรณ์ลงในอาหารเหลวชนิดเดียวกัน เพื่อซักนำให้เกิดเป็นเซลล์เพาะเลี้ยงแขวนลอย การศึกษาองค์ประกอบเคมีของน้ำมันระเหยของเซลล์เพาะเลี้ยงโกฐุพาล้ำพาจีน ยี่หร่า กิมกิด และพิมเสน ทำได้โดยนำเซลล์เพาะเลี้ยงแต่ละชนิดมาสักดิ้วย dichloromethane และนำสารสักดิ์ที่ได้ไปวิเคราะห์องค์ประกอบเคมีด้วยวิธี Gas Chromatography และ Gas Chromatography-Mass Spectrometry ผลการทดลองแสดงว่ามีน้ำมันระเหยจากเซลล์เพาะเลี้ยงเหล่านี้มีองค์ประกอบเคมีหลักที่เหมือนกับต้นจริง แต่มีปริมาณน้อยมาก และประกอบด้วยองค์ประกอบเคมีย่อยชนิดต่างๆ ในปริมาณเล็กน้อย ดังนั้นการทดลองนี้จึงศึกษาหารือเพิ่มปริมาณองค์ประกอบเคมีหลักของน้ำมันระเหย และศึกษาการเปลี่ยนแปลงทางชีวภาพของสารเทอร์ปินอยค์ต่างๆ ในเซลล์เพาะเลี้ยงพืช รวมทั้งหารือซักนำให้เกิดเป็นอวัยวะเพาะเลี้ยง เพื่อให้เป็นแหล่งสะสมน้ำมันระเหยในเซลล์เพาะเลี้ยงเหล่านี้ ผลการทดลองแสดงว่ามีที่สามารถเพิ่มปริมาณองค์ประกอบเคมีหลักได้คือการเติมสารตั้งด้านของการชีวสังเคราะห์ และการใช้โคโตชานเป็นสารกระตุ้นการสร้างสารทุติยภูมิ ส่วนวิธีอื่นๆ สามารถเพิ่มปริมาณองค์ประกอบเคมีย่อยได้บ้างเล็กน้อย

สาขาวิชาเภสัชเคมีและผลิตภัณฑ์ธรรมชาติ
ปีการศึกษา 2548

ลายมือชื่อนิสิต ศุภวรรณ บุญราษฎร์
ลายมือชื่ออาจารย์ที่ปรึกษา อ.ดร.นิจศิริ เรืองรังษี
ลายมือชื่ออาจารย์ที่ปรึกษาร่วม ดร.ธนกัตร ทรงสักดิ์

4576969033: MAJOR PHARMACEUTICAL CHEMISTRY AND NATURAL PRODUCTS
 KEYWORDS: *ARTEMISIA VULGARIS VAR. INDICA / CUMINUM CYMINUM / FORTUNELLA JAPONICA / AND POGOSTEMON CABLIN / ESSENTIAL OIL / PLANT CELL CULTURES*

SUPAWAN BUNRATHEP : ESSENTIAL OIL PRODUCTION IN CELL CULTURES OF *ARTEMISIA VULGARIS VAR. INDICA, CUMINUM CYMINUM, FORTUNELLA JAPONICA, AND POGOSTEMON CABLIN.* THESIS ADVISOR: ASSOC. PROF. NIJSIRI RUANGRUNGS, Ph.D., THESIS CO-ADVISOR: THANAPAT SONGSAK, Ph.D., 199 pp. ISBN 974-17-5547-3

Study on chemical constituents of essential oils of *Artemisia vulgaris* var. *indica*, *Cuminum cyminum*, *Fortunella japonica*, and *Pogostemon cablin* was done by hydrodistillation on each explant and then analysed by Gas Chromatography-Mass Spectrometry. It was found that individual essential oil contained terpenoid compounds of which (+)-davanone (71.59 %), cuminaldehyde (36.30 %), β-pinene (47.44 %), d-limonene (87.07 %), and patchouli alcohol (60.30 %) are major constituents of leaves of *Artemisia vulgaris* var. *indica*, fruits of *Cuminum cyminum*, leaves of *Fortunella japonica*, peels of *Fortunella japonica* and leaves of *Pogostemon cablin*, respectively. In order to study chemical constituents of essential oil of these plant cell cultures, each explant was surface sterilised and callus cultures initiated on MS media containing various plant growth regulators, followed by incubation in suitable culture conditions. Cell suspension cultures were initiated by subculturing each cell cultures into new liquid media and maintained in the same conditions. Study on chemical constituents of essential oils produced by these cell cultures was done by extraction with dichloromethane and extracts analysed by Gas Chromatography and Gas Chromatography-Mass Spectrometry. The results showed that essential oil obtained from these cultures had contained same major constituents as in the intact plant but the level was low, and also contained a small amount of minor constituents. Methods for improving the major constituents of these essential oils, and biotransformation of terpenoids in individual plant cell cultures, including methods for organ culture initiation for use as accumulation sites in cultures had been studied in this experiment. It was found that feeding precursors of biosynthesis and elicitation with chitosan can improve the yield of major constituents successfully, whilst other methods can improve a small amount of minor constituents.

Field of study	Pharmaceutical Chemistry and Natural Products	Student's signature	Supawan Bunrathip
Academic year	2005	Advisor's signature	Nijsiri Ruangrungs
		Co-advisor's signature	Thanapat Songsak

ACKNOWLEDGEMENTS



The success of this dissertation would not be realized without the support and assistance of persons and various institutions to whom I would like to express my sincere and profound gratitude:

Associate Professor Dr. Nijsiri Ruangrungsi of the Department of Pharmacognosy, Faculty of Pharmaceutical Sciences, Chulalongkorn University, my thesis advisor, for his helpful suggestions, guidance, continual interest and encouragement throughout the course of this work,

Dr. Thanapat Songsak of the Department of Pharmacognosy, Faculty of Pharmacy, Rangsit University, my thesis co-advisor, for his helpful guidance and keen interesting during the course of this work,

Dr. George Brian Lockwood of the School of Pharmacy and Pharmaceutical Sciences, The University of Manchester, United Kingdom, for providing research opportunities and invaluable suggestions during my stay in Manchester, United Kingdom,

The Thailand Research Fund for a 2002 Royal Golden Jubilee Scholarship for granting a whole financial support throughout the course of this work

The Tissue Culture Unit of the Department of Pharmacognosy, Faculty of Pharmacy, Rangsit University for providing laboratory facilities during the course of this work,

The thesis committee for their constructive suggestions and critical review of this thesis,

All of my friends in Chulalongkorn University, Rangsit University and The University of Manchester for their kindness, friendship and encouragement,

Finally, the most special thanks are due to my family for their love, understanding and encouragement until this work had been finished.

CONTENTS

	Page
ABSTRACT (Thai).....	iv
ABSTRACT (English).....	v
ACKNOWLEDGEMENTS.....	vi
CONTENTS.....	vii
LIST OF TABLES.....	xiii
LIST OF FIGURES.....	xvii
ABBREVIATIONS.....	xxi
CHAPTER I INTRODUCTION.....	23
1.1 The importance of the plant kingdom.....	23
1.2 Plant cell and tissue cultures.....	23
1.3 History and perspective.....	24
1.4 Definition.....	25
1.5 Callus culture.....	26
1.6 Cell suspension culture.....	26
1.7 The requirements of cultures.....	27
1.8 Culture media.....	28
1.8.1 Media composition.....	29
1.8.2 Plant growth regulators.....	29
1.8.2.1 Auxins.....	30
1.8.2.2 Cytokinins.....	31
1.9 External environment conditions.....	32
1.10 Applications.....	33
1.11 Secondary metabolites.....	34
1.11.1 Definition.....	34
1.11.2 Biosynthetic pathway of secondary metabolites.....	35
1.11.3 Natural products.....	35
1.12 Production of secondary metabolites by plant cell and tissue cultures.....	37
1.13 Effect of culture conditions on secondary product synthesis.....	38

	Page
1.14 Strategies for improving the production of secondary metabolites in cell cultures.....	39
1.14.1 Selection of high-yielding cell lines.....	39
1.14.2 Cell immobilization.....	42
1.14.3 Morphological modifications.....	45
1.14.4 Precursor feeding and biotransformation.....	49
1.14.5 Elicitation.....	52
1.14.6 Permeabilisation.....	54
1.14.7 Two-phase system.....	54
1.14.8 Use of adsorbent.....	57
1.15 Essential oils.....	57
1.16 Biosynthesis of essential oil constituents.....	58
1.17 Medicinal and commercial uses.....	62
1.18 Production of essential oil from plant cell cultures.....	62
CHAPTER II HISTORICAL.....	66
2.1 <i>Artemisia vulgaris</i> L. var. <i>indica</i> Maxim.....	66
2.2 <i>Cuminum cyminum</i> L.....	70
2.3 <i>Fortunella japonica</i> (Thunb.) Swingle.....	73
2.4 <i>Pogostemon cablin</i> (Blanco) Benth.....	77
CHAPTER III MATERIALS AND METHODS.....	82
3.1 Chemicals.....	82
3.2 Plant materials.....	82
3.3 Essential oil hydrodistillation.....	83
3.4 Identification of essential oil obtained from hydrodistillation by Gas Chromatography-Mass Spectrometry (GC-MS).....	84
3.5 Media preparation for plant tissue culture experiments.....	85
3.5.1 Nutrition media.....	85
3.5.2 General preparation of semi-solid and liquid media.....	85
3.6 Aseptic work.....	85
3.7 Germination of seedling.....	86
3.8 Preparation of leaf explants and surface sterilization.....	86
3.9 Initiation and maintenance of callus.....	86

	Page
3.10 Initiation and maintenance of suspension cultures.....	87
3.11 Measurement of growth parameter.....	87
3.11.1 Fresh and dry weigh measurement.....	87
3.11.2 Pack cell volume (PCV) or cell volume after sedimentation (CVS).....	88
3.12 Study on the effect of plant growth regulators on callus formation and appearance.....	89
3.13 Study on the effect of light on callus formation and appearance.....	89
3.14 Study on chemical constituents of essential oil produced by plant cell cultures.....	89
3.14.1 Extraction of chemical constituents of essential oil produced by plant cell cultures.....	89
3.14.2 Analysis of essential oil constituents by Gas Chromatography (GC).....	90
3.14.3 Identification of essential oil constituents produced by plant cell culture by Gas Chromatography-Mass Spectrometry (GC-MS).....	90
3.15 Shoot regeneration (Organogenesis).....	90
3.16 Methods for improving chemical constituents of essential oil produced by plant cell cultures.....	91
3.16.1 Feeding precursor and biotransformation.....	91
3.16.1.1 Substrate feeding and extraction of biotransformation products from suspension cultures.....	91
3.16.1.2 Time-course studies.....	91
3.16.1.3 Feeding precursors of each major chemical constituents in individual cell suspension cultures.....	91
3.16.1.4 Use of HEMA co-polymers for biotransformation.....	92
3.16.1.4.1 Preparation of p-HEMA discs.....	92
3.16.1.4.2 Substrate feeding using p-HEMA discs.....	93

	Page
3.16.2 Plant growth regulators.....	93
3.16.3 Light.....	93
3.16.4 Elicitation.....	93
3.16.4.1 Elicitation with chitosan.....	94
3.16.4.2 Elicitation with methyl jasmonate (MEJA).....	94
3.16.5 Permeabilisation.....	94
3.16.6 <i>In situ</i> product removal (two-phase system).....	95
CHAPTER IV RESULTS.....	96
4.1 Identification of essential oil obtained from hydrodistillation.....	96
4.1.1 <i>Artemisia vulgaris</i> var. <i>indica</i>	96
4.1.2 <i>Cuminum cyminum</i>	96
4.1.3 <i>Fortunella japonica</i>	96
4.1.3.1 Leaves.....	96
4.1.3.2 Peels.....	97
4.1.4 <i>Pogostemon cablin</i>	97
4.2 Determination of germination and growth of seedling.....	103
4.3 Surface sterilization of leaf explants.....	104
4.4 Growth and appearance of callus cultures.....	104
4.5 Effect of plant growth regulators on callus formation and appearance...108	108
4.6 Effect of light on callus formation and appearance.....	108
4.7 Growth and appearance of cell suspension cultures.....	109
4.8 Identification the essential oil constituents produced by plant cell culture by Gas Chromatography (GC) and Gas Chromatography- Mass Spectrometry (GC-MS).....	112
4.9 Time-course studies of volatiles constituent content during the growth cycle of plant cell cultures.....	114
4.10 Shoot regeneration (organogenesis).....	123
4.11 Methods for improving chemical constituents of essential oil produced by plant cell cultures.....	124
4.11.1 Study on feeding precursor and biotransformation.....	124

	Page
4.11.1.1 Effect of substrate concentration on biotransformation.....	124
4.11.1.2 Time-course study of monoterpene feeding to suspension cultures of <i>Fortunella japonica</i>	125
4.11.1.3 Study of relationship between citral, geraniol, nerol, geranyl acetate, and neryl acetate in cell suspension cultures of <i>Fortunella japonica</i>	125
4.11.1.4 Substrate feeding in cell suspension cultures.....	130
4.11.1.4.1 Biotransformation of acyclic terpenes.....	130
4.11.1.4.2 Biotransformation of cyclic monoterpenes.....	149
4.11.1.5 Improve yield of biotransformation products by p-HEMA discs.....	160
4.11.1.5.1 Time-course study of feeding of p-HEMA discs containing monoterpenes to cell suspension cultures.....	160
4.11.1.6 Feeding precursors of each major chemical constituents in individual cell suspension cultures.....	160
4.11.2 Elicitation.....	162
4.11.2.1 Elicitation with chitosan.....	162
4.11.2.1.1 Determination of optimum chitosan concentration.....	162
4.11.2.1.2 Determination of optimum period of elicitation.....	163
4.11.2.2 Elicitation with methyl jasmonate (MEJA).....	164
4.11.2.2.1 Determination of optimum concentration of methyl jasmonate.....	164
4.11.2.2.2 Effect of methyl jasmonate on essential oil constituents product in <i>Fortunella japonica</i> cell suspension cultures...164	164
4.11.3 Permeabilisation.....	166
4.11.3.1 Determination of optimum Tween-20 concentration.....	166
4.11.3.2 Effect of Twee-20 on essential oil constituents in <i>Fortunella japonica</i> cell suspension cultures.....	166

	Page
4.11.4 <i>In situ</i> product removal (two-phase system).....	168
CHAPTER V DISCUSSION.....	169
REFERENCES.....	174
APPENDICES.....	191
APPENDIX A Plant Tissue Culture Terms.....	192
APPENDIX B Murashige and Skoog's Basal Media.....	197
APPENDIX C Surface Sterilising Agent.....	198
VITA.....	199

LIST OF TABLES

Table	Page
1. Common auxins used in plant cell cultures.....	30
2. Common cytokinin used in plant cell cultures.....	31
3. Secondary metabolites produced in high levels by plant cell and tissue cultures.....	38
4. Selected examples of plant species, secondary metabolites, and selection methods studied in plant cell cultures.....	41
5. Selected examples of plant species, secondary metabolites, and immobilisation methods studied in plant cell cultures.....	44
6. Selected examples of plant species, secondary metabolites, and organogenesis studied in plant cell cultures.....	48
7. Selected examples of plant species, secondary metabolites, and biotransformation products studied in plant cell cultures.....	51
8. Selected examples of plant species, secondary metabolites, and elicitors studied in plant cell cultures.....	53
9. Selected examples of plant species, secondary metabolites, and permeabilisation studied in plant cell cultures.....	56
10. Selected examples of plant species, secondary metabolites, and two-phase system studied in plant cell cultures.....	56
11. Selected examples of essential oil production in plant cell cultures.....	64
12. Selected examples of <i>Artemisia</i> spp. had been studied in essential oil production from cell and tissue cultures.....	69
13. Plant species and sources.....	82
14. Explants of individual plant species selected for essential oil hydrodistillation.....	83
15. Precursor fed into individual cell suspension culture attempt to increase level of major constituent.....	92
16. Essential oil constituents of <i>Artemisia vulgaris</i> var. <i>indica</i> obtained by fresh leaves hydrodistillation.....	98

Table	Page
17. Essential oil constituents of <i>Cuminum cyminum</i> obtained by fruits hydrodistillation.....	99
18. Essential oil constituents of <i>Fortunella japonica</i> obtained by fresh leaves hydrodistillation.....	100
19. Essential oil constituents of <i>Fortunella japonica</i> obtained by peels hydrodistillation.....	101
20. Essential oil constituents of <i>Pogostemon cablin</i> obtained by fresh leaves hydrodistillation.....	102
21. Surface sterilisation fruits of <i>Cuminum cyminum</i> and seeds of <i>Fortunella japonica</i>	103
22. Germination of <i>Cuminum cyminum</i> and <i>Fortunella japonica</i>	103
23. Surface sterilisation leaves of <i>Artemisia vulgaris</i> var. <i>indica</i> and <i>Pogostemon cablin</i>	104
24. Appearance and callus growth of individual species.....	105
25. List of plant growth regulators used for maintenance callus cultures.....	108
26. Light condition used for maintenance callus cultures.....	108
27. Appearance and growth of suspension cultures of individual species.....	109
28. Essential oil constituents of <i>Artemisia vulgaris</i> var. <i>indica</i> callus cultures obtained by dichloromethane extraction.....	112
29. Essential oil constituents of <i>Artemisia vulgaris</i> var. <i>indica</i> cell suspension cultures obtained by dichloromethane extraction.....	112
30. Essential oil constituents of <i>Cuminum cyminum</i> callus cultures obtained by dichloromethane extraction.....	112
31. Essential oil constituents of <i>Cuminum cyminum</i> cell suspension cultures obtained by dichloromethane extraction.....	113
32. Essential oil constituents of <i>Fortunella japonica</i> callus cultures obtained by dichloromethane extraction.....	113
33. Essential oil constituents of <i>Fortunella japonica</i> cell suspension cultures obtained by dichloromethane extraction.....	113
34. Essential oil constituents of <i>Pogostemon cablin</i> callus cultures obtained by dichloromethane extraction.....	113

Table	Page
35. Essential oil constituents of <i>Pogostemon cablin</i> cell suspension cultures obtained by dichloromethane extraction.....	114
36. Fresh weight, dry weight and davanone content in <i>Artemisia vulgaris</i> var. <i>indica</i> callus cultures.....	115
37. Fresh weight, dry weight and davanone content in <i>Artemisia vulgaris</i> var. <i>indica</i> cell suspension cultures.....	116
38. Fresh weight, dry weight and cuminaldehyde content in <i>Cuminum cyminum</i> callus cultures.....	117
39. Fresh weight, dry weight and cuminaldehyde content in <i>Cuminum cyminum</i> cell suspension cultures.....	118
40. Fresh weight, dry weight and d-limonene content in <i>Fortunella japonica</i> callus cultures.....	119
41. Fresh weight, dry weight and d-limonene content in <i>Fortunella japonica</i> cell suspension cultures.....	120
42. Fresh weight, dry weight and patchouli alcohol content in <i>Pogostemon cablin</i> callus cultures.....	121
43. Fresh weight, dry weight and patchouli alcohol content in <i>Pogostemon cablin</i> cell suspension cultures.....	122
44. Biotransformation products from a range of acyclic terpenes fed to individual cell suspension cultures.....	131
45 Biotransformation products from a range of cyclic terpenes fed to individual cell suspension cultures.....	150
46. Yield of major chemical constituent of individual plant after precursor feeding experiment.....	161
47. Detected essential oil constituents after day 14 of elicitation of methyl jasmonate in <i>Fortunella japonica</i>	164
48. Detected essential oil constituents after day 21 of elicitation of methyl jasmonate in <i>Fortunellajaponica</i>	165
49. Detected essential oil constituents after day 14 of permeabilisation of various Tween-20 concentrations in <i>Fortunella japonica</i>	167

Table	Page
50. Detected essential oil constituents after day 21 of permeabilisation using various Tween-20 concentrations in <i>Fortunella japonica</i>	168
51. The chemical constituents of Murashige and Skoog's basal media.....	197

LIST OF FIGURES

Figure	Page
1. Chemical structures of common auxins used in plant cell cultures.....	30
2. Chemical structures of common cytokinins used in plant cell cultures.....	32
3. Schematic diagram of the biosynthesis pathway for secondary metabolites.....	36
4. Chemical structure of thidiazuron (TDZ).....	46
5. Biosynthesis pathway of terpenes via acetate-mevalonic acid.....	60
6. Biosynthesis pathway phenylpropanoid compounds.....	61
7. <i>Artemisia vulgaris</i> L. var. <i>indica</i> Maxim. (syn. <i>Artemisia dubia</i> Wall. ex Bess.).....	67
8. Chemical structure of (+)-davanone.....	68
9. Possible biosynthesis pathway of davanone.....	69
10. <i>Cuminum cyminum</i> L.....	70
11. Chemical structure of cuminaldehyde.....	71
12. <i>Fortunella japonica</i> (Thunb.) Swingle (syn. <i>Citrus japonica</i> Thunb.).....	73
13. Chemical structure of d-limonene.....	75
14. Biosynthesis pathway of d-limonene via geranyl pyrophosphate formation.....	75
15. Biosynthesis pathway of linalyl pyrophosphate.....	76
16. <i>Pogostemon cablin</i> (Blanco) Benth. (syn. <i>Pogostemon patchouli</i> Hook.).....	77
17. Biosynthesis pathway of patchouli alcohol, seychellene, and cycloseychellen in <i>Pogostemon cablin</i>	79
18. Biosyntheisi pathway of α -pathoulene, β -pathoulene, γ -pathoulene, δ -pathoulene in <i>Pogostemon cablin</i>	80
19. Clevenger type apparatus.....	84
20. The apparatus for measuring cell growth of <i>Cuminum cyminum</i> cell suspension cultures.....	88
21. Chemical structures of 2-hydroxyethyl methacrylate (HEMA) and ethylene glycol dimethacrylate (EGDMA).....	92
22. Callus cultures of <i>Artemisia vulgaris</i> var. <i>indica</i>	106
23. Callus cultures of <i>Cuminum cyminum</i>	106

Figure	Page
24. Callus cultures of <i>Fortunella japonica</i>	107
25. Callus cultures of <i>Pogostemon cablin</i>	107
26. Cell suspension cultures of <i>Artemisia vulgaris</i> var. <i>indica</i>	110
27. Cell suspension cultures of <i>Cuminum cyminum</i>	110
28. Cell suspension cultures of <i>Fortunella japonica</i>	111
29. Cell suspension cultures of <i>Pogostemon cablin</i>	111
30. Time-course of growth and the formation of davanone in <i>Artemisia vulgaris</i> var. <i>indica</i> callus cultures.....	115
31. Time-course of growth and the formation of davanone in <i>Artemisia vulgaris</i> var. <i>indica</i> cell suspension cultures.....	116
32. Time-course of growth and the formation of cuminaldehyde in <i>Cuminum cyminum</i> callus cultures.....	117
33. Time-course of growth and the formation of cuminaldehyde in <i>Cuminum cyminum</i> cell suspension cultures.....	118
34. Time-course of growth and the formation of d-limonene in <i>Fortunella japonica</i> callus cultures.....	119
35. Time-course of growth and the formation of d-limonene in <i>Fortunella japonica</i> cell suspension cultures.....	120
36. Time-course of growth and the formation of patchouli alcohol in <i>Pogostemon cablin</i> callus cultures.....	121
37. Time-course of growth and the formation of patchouli alcohol in <i>Pogostemon cablin</i> cell suspension cultures.....	122
38. Fresh weight and dry weight of <i>Fortunella japonica</i> cell suspension cultures after feeding various concentration of geraniol.....	124
39. Time-course study of geraniol levels after feeding various concentrations in <i>Fortunella japonica</i> cell suspension culture.....	125
40. Terpenoid concentrations after feeding 100 ppm citral in <i>Fortunella japonica</i> cell suspension cultures.....	126
41. The possible biotransformation pathway of citral in <i>Fortunella japonica</i> cell suspension cultures.....	126
42. Terpenoid concentrations after feeding 100 ppm geraniol in <i>Fortunella japonica</i> cell suspension cultures.....	127

Figure	Page
43. The possible biotransformation pathway of geraniol in <i>Fortunella japonica</i> cell suspension cultures.....	127
44. Terpenoid concentrations after feeding 100 ppm nerol in <i>Fortunella japonica</i> cell suspension cultures.....	128
45. The possible biotransformation pathway of nerol in <i>Fortunella japonica</i> cell suspension cultures.....	128
46. Terpenoid concentrations after feeding 100 ppm geranyl acetate in <i>Fortunella japonica</i> cell suspension cultures.....	129
47. The possible biotransformation pathway of geranyl acetate in <i>Fortunella japonica</i> cell suspension cultures.....	129
48. Biotransformation products of citral in individual cell suspension cultures....	131
49. Biotransformation products of citronellal in individual cell suspension cultures.....	133
50. Biotransformation products of citronellol in individual cell suspension cultures.....	135
51. Biotransformation products of Z,Z-farnesol in individual cell suspension cultures.....	136
52. Biotransformation products of Z,Z-farnesyl acetate in individual cell suspension cultures.....	137
53. Biotransformation products of geraniol in individual cell suspension cultures.....	139
54. Biotransformation products of geranyl acetate in individual cell suspension cultures.....	141
55. Biotransformation products of linalool in individual cell suspension cultures..	143
56. Biotransformation products of linalyl acetate in individual cell suspension cultures.....	146
57. Biotransformation products of nerol in individual cell suspension cultures....	148
58. Biotransformation products of l-bornyl acetate in individual cell suspension cultures.....	150
59. Biotransformation products of l-borneol in individual cell suspension cultures.....	151

Figure	Page
60. Biotransformation products of fenchone in individual cell suspension cultures.....	152
61. Biotransformation products of fenchyl acetate in individual cell suspension cultures.....	153
62. Biotransformation products of menthone in individual cell suspension cultures.....	154
63. Biotransformation products of menthol acetate in individual cell suspension cultures.....	155
64. Biotransformation products of d-limonene in individual cell suspension cultures.....	156
65. Biotransformation products of α -pinene in individual cell suspension cultures.....	157
66. Biotransformation products of β -pinene in individual cell suspension cultures.....	158
67. Biotransformation products of verbenol in individual cell suspension cultures.....	159
68. The effect of various chitosan concentrations on cell growth and d-limonene production in <i>Fortunella japonica</i> cell suspension cultures after 7 days.....	162
69. d-Limonene and α -terpineol concentrations in elicited cell cultures of <i>Fortunella japonica</i> compare to control experiment.....	163

ABBREVIATIONS

%	=	Percent (part per 100); percentage
μg	=	Microgram(s)
μl	=	Microlitre(s)
μm	=	Micrometre(s)
/	=	Per
2,4-D	=	2,4-Dichlorophenoxyacetic acid
AOAC	=	Association of Official Analytical Chemists
BA	=	6-Benzylaminopurine or N ⁶ -benzyladenine
°C	=	Degree Celsius
cm	=	Centimetre(s)
cm ²	=	Centimetre square(s)
CVS	=	Cell volume after sedimentation
DW	=	Dry weight
ed(s)	=	Editor(s)
e.g.	=	For example
EI-MS	=	Electron impact mass spectra
EO	=	Essential oil
<i>et al.</i>	=	Et alii
eV	=	Electron volt
FW	=	Fresh weight
FID	=	Flame ionization detector
Fig.	=	Figure
g	=	Gram(s)
GC	=	Gas Chromatography
GC-MS	=	Gas Chromatography-Mass Spectrometry
h	=	Hour(s)
IC ₅₀	=	50% Inhibitory concentration
Kn	=	Kinetin or 6-furfurylaminopurine
l	=	Litre(s)
M	=	Molar
MEJA	=	Methyl jasmonate

mg/l	=	Milligram per litre
min	=	Minute(s)
ml	=	Millilitre(s)
mm	=	Millimetre(s)
MS	=	Murashige and Skoog's media
NAA	=	Naphthaleneacetic acid
PGR	=	Plant growth regulator
pH	=	The negative logarithm of the concentration of hydrogen ions
ppi	=	Pore per inch
ppm	=	Part per million
rpm.	=	Round per minute or revolution per minute
SD	=	Standard deviation
sec	=	Secound(s)
TDZ	=	Thidiazuron
v/v	=	Volume over volume or volume by volume
w/v	=	Weight over volume or weight by volume