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APPENDICES

1.4

APPENDIX A

Picture	Name	Scientific name	Part of
			Use
	Pak Chee Lao (Dill) ผักซีลาว	Anethum graveolens Linn	Aerial part
	Pak Ka Yang (ผักแขยง)	Limnophila aromatica Merr	Aerial part
	Pak Pai (ผักไผ่)	Polygonum odoratum Lour.	Aerial part
	Pak Gud (ผักกูด)	<i>Diplazium esculentum</i> (Retz.)	Aerial part
	Pak Krad Hua Wan (ผักคราดหัวแหวน)	Spilanthes acmella Murr .	Aerial part

APPENDIX B

1.	Preparation of stock solution and Media (Maron and Ames, 1983)	
	1.1. Voget Bonner media E stock salt solution (VB salt)	
	Use : Minimal agar	
	Ingredient	Per liter
	Warm distilled water (45 °C)	670 ml
	Magnesium sulfate (MgSO ₄ 7H ₂ O)	10 g
	Cirtric acid monohydrate	100 g
	Potassium phosphate, dibasic (anhydrous) (K ₂ HPO ₄)	500 g
	Sodium ammonium phosphate (NaNH ₄ HPO ₄ .4H ₂ O)	175 g
	Add salts in the order indicated to warm water and allow each salt to dis	solve
	completely before adding the next. Adjust the volume to 1 liter. Filter the	solutions
	and then autoclave at 121 °C for 15 min.	

1.2. Minimal glucose agar plate

Use : Mutagenicity assay	
Ingredient	Per liter
Bacto agar	15 g
Distilled water	930 ml
VB salts	20 ml
40% glucose	50 ml

Add agar to distilled water in a glass bottle. Autoclave at 121 °C for 15 min. when the solution has cooled slightly, add sterile VB salts and sterile 40% glucose. Mix and pour 30 ml into each sterile petri plant. Minimal glucose agar plates were kept in incubator at 37 °C before using. (the VB salts and 40% glucose should be autoclaved separately)

1.3. Oxoid nutrient broth No.2

Use : Growing culture

Dissolve 2.5 g of nutrient broth No.2 in 100 ml distilled water. Transfer 12 ml nutrient broth for each 50 ml Erlenmeyer flask (covered with sterile gauze). Autoclave at 121°C for 15 min.

1.4. Top agar

Use : Mutagenic assay		
Ingredient	100 ml	
Bacto agar	0.6 g	
Sodium chloride (NaCl)	0.5 g	
Distilled water	100 ml	
Dissolve ingredients in distilled water. Store in a glass bottle. Autoclave for 15 min.		
At 121 °C and then add 10 ml of 0.5 mM L-histidine/biotin solution and mixed		

thoroughly by swirling.

1.5. 0.1 M L-histidine HCI stock

Use : Fortification of minimal agar plate		
Ingredient	10 ml	
L-histidine HCI	0.2096 g	
Distilled water	10 ml	
Dissolve 0.2096 g of L-histidine HCI (MW 209.63) in 10 ml distilled water. Autoclave		
at 121 °C for 15 min. Store in a glass bottle 4 °C.		

1.6. 1 mM L-histidine HCI stock

Use : Fortification of minimal agar plate.		
Ingredient	100 ml	
0.1 M L-histidine HCI	1 ml	
Distilled water	99 ml	
Dilute 1 ml of 0.1 M L-histidine HCl in 99 ml of distilled water. Autoclave at 121 °C for		
15 min.		

1.7. 1 mM biotin stock

Use : Fortification of minimal agar plate.		
Ingredient		100 ml
Biotin		24.43 mg
Distilled water		100 ml
Dissolve biotin (WM 244.3) in distilled H ₂ O. Warm it until dissolve completely.		

Autoclave at 121 °C for 15 min.

1.8. 0.5 mM L-histidine / biotin solution.

Use : Mutagenicty assay (add 10 ml to 100 ml of Top agar)	
Ingredient	200 ml
1 mM L-histidine HCI	100 ml
1 mM biotin	100 ml
Mix and autoclave at 121 °C for 15 min.	

1.9. 1 M potassium chloride

Use : Na ₃ PO ₄ -KCI buffer for mutagenicity assay	
Ingredient	100 mł
Potassium chloride	7.456 g
Distilled water	100 ml
Mix and autoclave at 121 °C for 15 min.	

1.10. 0.5 M sodium phosphate pH 7.4

 $\text{Use}:\text{Na}_3\text{PO}_4-\text{KCI}$ buffer for mutagenicity assay

Ingredient

0.5 M Sodium dihydrogen phosphate (NaH₂ PO₄) (MW 120)

(30 g / 500 ml)

0.5 M Disodium hydrogen phosphate dehydrate (NaH₂ PO₄ .2 H₂O) (MW 177.99) (44.5 g / 500 ml)

Dissolve 44.5 g disodium hydrogen phosphate dehydrate in 300 ml of distilled water. Add 0.5 M sodium dihydrogen phosphate until to pH 7.4, and then adjust volume to 500 ml. Steriliize by autoclaving for 15 min at 121 °C

1.11. Na ₃ PO ₄ – i∖Cl Juffer	
Use : mutaenicity assay	
Ingredient	330 ml
0.5 M Sodium Phosphate (Na₃PO₄) pH 7.4	100 ml
1 M Potassium Chloride (KCI)	16.5 ml
Distilled water	213.5 ml
Autoclave for 15 min at 121 °C	

2. Recipes for Some Reagents and Test Chemicals

2.1. 2 M sodium nitrite	
Use : Nitrosation	
Ingredient	10 ml
Sodium nitrite	1.38 g
Distilled water to	10 ml
Mix and autoclave for 15 min at 121 °C	

2.2. 2 M ammonium sulfamate

Use : reaction mixture		
Ingredient	10 ml	
Ammonium sulfamate	2.28 g	
Distilled water to	10 ml	
Dissolve ammonium sulfamate in distilled water and adjust volume. Autoclave for 15		

min at 121 °C

2.3. 0.2 N Hydrochloric acid

Use : reaction mixture

Ingredient	100 ml	
Conc. Hydrochloric acid	1.66 ml	
Sterile distilled water	98.34 ml	
Dissolve conc.hydrochloric acid in sterile water. Store in sterile glass tubes or bottle		

with screw caps.

2.4. 0.3 mg/ml amonopyrene

Use : standard solution for mutagenicity assay

Ingredient	1	ml
Aminopyrene	3	mg
Acetonitrile	1	ml
Dissolve aminopyrene in acetonitrile. Store in sterile vial with screw caps	; in	the
freezer. Preparation of this solution must use sterile technique.		
Ingredient	1	ml
3 mg/ml aminopyrene	0.	1 ml
Acetonitrile	0.9	9 ml
Dissolve 3 ma/ml aminonyrene in acetonitrile. Store in sterile vial with sci	r OIM	i can i

Dissolve 3 mg/ml aminopyrene in acetonitrile. Store in sterile vial with screw cap in freezer. There preparation must be used sterilr technique.

2.5. 8 mg/ml ampicillin solution

Ingredient	10	ml
Ampicillin sodium	80	mg
Distilled water	10	ml

2.6. 0.1% crystal violet

Ingredient	10	ml
Crystal violet	10	mg
Distilled water to	10	ml
2.5 And 2.6: Store at 4 C in glass bottle with screw cap.		

3. Procedure for Reisolation and Growing Culture

Tester strains, TA 98 and TA 100 are grown in Oxoid nutrient broth No.2 and incubated overnight in a 37 $^{\circ}$ C shaking water bath . The growth period should not exceeded 16 h. these culture are re-isolated by streaking on minimal glucose agar plates which the surface were spread with 0.1 ml of 8 mg/ml ampicillin, 0.3 ml of 0.1 M histidine HCl and 0.1of 1 mM biotin. There plates are incubated at 37 $^{\circ}$ C for 48 h. After incubation, the 4 single colonies per strain TA 98 and TA 100 are pick and grown in Oxiod nutrient broth No.2 overnight at 37 $^{\circ}$ C in shaking water bath. Each culture is confirmed genotype of the strains and kept the cultures as the source of bacteria for mutagenicity testing. For each 1.0 ml of culture, add 0.09 ml of spectrophotometric grade DMSO. Combine the culture and DMSO in a sterile tub and distribute 40 µl of the culture aseptically into sterile cryotubes (Nunc). The tubes then transfer to a -80 $^{\circ}$ C freezer.

4. Confirming Genotype of Tester strains

The broth cultures of TA 98 and TA 100 are used to confirm genetype in the following ways.

4.1 Histidne requirement

The his character of the strains confirmed by demonstrating the histidine requirement for growth on the minimal glucose agar plates enriched with histidine and biotin.

Procedure :

Plate A : no histidine and biotin

Plate B: 0.1 ml of 1 mM biotin

Plate C: 0.3 ml of 0.1 M His-HCl

Plate D: 0.3 ml of 0.1 M His-HCl + 0.1 ml of 1 mM biotin

Four minimal glucose agar plates are required for each tester strains. Each plate is applied on the surface with 0.1 ml of 1 mM biotin or no application (plate b,c,d,a, respectively) Made a single streak of each strains across these plates. Four strains

1

could be tested on the same plate Incubated at 37 °C for 24 h. The growing of bacteria on histidine plus biotin plate is the result of histidine requirement.

4.2 R-factor

The R-factor strains (TA 97, TA 98, TA 100 and TA 102) should the tosted routinely for the presence of the ampicillin resistance factor because the plasmid is somewhat unstable and can be lost from the bacteria.

Procedure: For each tester strain, add 0.3 ml of fresh overnight culture to a tube containing 0.1 ml of 0.1 M histidine – HCl followed by adding 20 ml of molten top agar containing 0.5 mM, mixed and poured on a minimal glucose agar plant. Rotated the plate to distribute the mixtures and allowed several minutes for agar to become firm. R-factor an rfa mutation (se the next section) are performed in the same plant by dividing the plate into 2 areas, one for R-factor and the other for rfa mutation. For R-factor, commercial ampicillin disc or filter paper disc containing 8 mg/ml ampicllin is applied on the surface of the agar by using sterile forceps. The dish is pressed lightly to embed in the overlay. The plates are incubated at 37 °C for 24 h. The absence of the clear zones of inhibition around the disc indicates resistance to ampicillin.

4.3 Rfa Mutation

Strains having the deep rough (rfa) character should be tested for crystal violet sensitivity.

Procedure : Pipetted 0.1% solution of crystal violet to the sterile filter paper disc (1/4 inch) and transferred the disc to plates, seeded with bacteria (the procedure is similar to R-factor). Incubated at 37 °C for 48 h. The clear zone appeared around the disc indicated the presence of the rfa mutation that permitted crystal violet to enter and kill bacteria.

5. Spontaneous Reverion

Spontaneous reversion of the tester strains to histidine independence is measured routinely in mutagenicity experiments and is expressed as the number of spontaneous revertants per plate. The revertant colonies are clearly visible in a uniform background lawn of auxotropic bacteria. Each tester strain reverts spontaneously at a frequency that is characteristic of the strain. Nevertheless, there is variability in the number of spontaneous revertants form one experiment to another and from one plate to another, and it is advisable to include at least 2-3 spontaneous mutation control plates for each strain ina mutagenicity assay.

Procedure: 0.2 ml of sterile distilled water (solvent in the experiment) is added to cap tube. Add 0.5 ml of NaPO4-KCl buffer pH 7.4 in the absence of metabolic activation, 0.1 ml of fresh overnight culture of TA 98 or TA 100, followed plates and left it to become harden. Incubated at 37 °C for 48 h. and the his^{*} reveratant colonies were counted.

6. The Response to Standard carcinogen.

Standard carcinogens or positive carcinogen are used routinely in mutagenicity experiment to confirm the reversion property and specificity of each strain. The standard mutagen which used in these experiments is aminopyrene in the absence of metabolic activation. Tester strain which highly response to positive mutagens must be collected.

Procedure: The procedure is as described in spontaneous reversion except aminopyrene (0.06, and 0.12 μ I per plate for TA 98 and TA 100, respectively) are used instead of sterile distilled water in absence S-9 mix, respectively. The characteristic of stock culture for TA 98 and TA 100 as the source of bacteria for mutagenicity is

- a) Contained R-factor (pKM 101) and rfa mutation
- b) His+ requirement
- c) low spontaneous reversion
- d) highly response to standard carcinogen

After the characteristic of the culture was tested, the mutagenicity test was started.

VITA

Miss Nisita Komsakorn was born on August 11, 1978 in Bangkok, Thailand. She received her Bachelor's degree of Sciences in Pharmacy in 2000 from the Faculty of pharmaceutical sciences, Chulalongkorn University, Thailand. She has five-year experiences in hospital pharmacy. Now she is working as marketing executive for the British Dispensary (L.P.) Co., Ltd.

