REFERENCES

- Alvira, P., Tomas-Pejo, E., Ballesteros, M., and Negro, M.J. (2009) Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. <u>Bioresource Technology</u>, 101, 4851-4861.
- Chen, W., Pen, B., Yu, C., and Hwang, W. (2011) Pretreatment efficiency and structural characterization of rice straw by an integrated process of diliteacid and steam explosion for bioethanol production. <u>Bioresource Technology</u>, 102, 2916–2924.
- Cuiping, L., Chuangzhi, W., Yanyongjie., and Haitao, H. (2004) Chemical elemental characteristics of biomass fuels in China. <u>Biomass and Bioenergy</u>, 27, 119 –130.
- Gabhane, J., William, S.P.M., Vaidya, A., Mahapatra, K., and Chakrabarti, T. (2010)

 Influence of heating source on the efficacy of lignocellulosic pretreatment—A cellulosic ethanol perspective. <u>Biomass and Bioenergy</u>, 1—7.
- Keshwani, D.P., Cheng, J., Burns, J.S., Li, L.G., and Chiang, V.C. (2007)

 Microwave Pretreatment of Switchgrass to Enhance Enzymatic

 Hydrolysis. <u>Biological Systems Engineering</u>, 0771277.
- Kim, T. H. and Lee Y. Y. (2005) Pretreatment and fractionation of corn stover by ammonia recycle percolation process. <u>Bioresource Technology</u>, 96, 2007–2013
- Lenihan, P., Orozco, A., O'Neill, E., Ahmad, M.N.M., Rooney, D.W., and Walker, G.M. (2010) Dilute acid hydrolysis of lignocellulosic biomass. <u>Chemical Engineering Journal</u>, 156, 395–403.
- Lin, L., Yan, R., Liu, Y., and Jiang W. (2010) In–depth investigation of enzymatic hydrolysis of biomass wastes based on three major components: Cellulose, hemicellulose and lignin. <u>Bioresource Technology</u>, 101, 8217–8223.
- Lloyd, T. and Wyman, C. (2005) Combined sugar yields for dilute sulfuric acid pretreatment of corn stover followed by enzymatic hydrolysis of the remaining solids. <u>Bioresource Technology</u>, 96, 1967–1977.

- McIntosh, S. and Vancov, T. (2010) Enhanced enzyme saccharification of Sorghum bicolor straw using dilute alkali pretreatment. <u>Bioresource Technology</u>, 101, 6718–6727.
- Miller, G. (1959) Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. <u>Analytical Chemistry</u>, 31 (3), 426–428.
- Palmqvist, E. and Hagerdal, B. (2000) Fermentation of lignocellulosic hydrolyzate I: inhibition and detoxification. <u>Bioresource Technology</u>, 74, 17–24.
- Redding, A., Wang, Z., Keshwani, D., and Cheng, J. (2011) High temperature dilute acid pretreatment of coastal Bermuda grass for enzymatic hydrolysis. Bioresource Technology, 102, 2916–2924.
- Saha, B., Iten. L., Cotta, M., and Wu, Y. (2005) Dilute acid pretreatment, enzymatic saccharification and fermentation of wheat straw to ethanol. <u>Process</u> Biochemistry, 40, 3693–3700.
- Saha, P., Manna, S., Chowdhury, S., Sen, R., Roy, D., and Adhikari, B. (2010) Enhancement of tensile strength of lignocellulosic jute fibers by alkalisteam treatment. <u>Bioresource Technology</u>, 101, 3182–3187.
- Sierra, R.C., Smith, A.R., Granda, C.S., and Holtzapple, M. (2009) Producing Fuels and Chemicals from Linocellulosic Biomass. <u>SBE Special Section</u>—Biofuel, S10–S18.
- Sumphanwanich, J., Leepipatpiboon, N., Srinorakutara, T., and Akaravharanya, A. (2008) Evaluation of dilute-acid pretreated bagasse, corn cobs and rice straw for ethanol fermentation by Saccharomyces Cerevisiae. <u>Annals of Microbiology</u>, 58, 219–225.
- Sun, Y. and Cheng, J.Y. (2002). Hydrolysis of lignocellulosic materials for ethanol production: A review. <u>Bioresource Technology</u>, 83, 1–11.
- Yang, H., Yan, R., Chen, H., Lee, D., and Zheng, C. (2007) Characteristics of hemicellulose, cellulose and lignin pyrolysis. <u>Fuel</u>, 86, 1781–1788.
- Available from: http://www.allaboutfeed.net/news/update-thailand-corn-production-and-trade-id1077.html
- Available from: http://as.doa.go.th/fieldcrops/cfc/sit/006.html

APPENDICES

Appendix A Autoclave

Batch reactor system used to study the effects of added acid on pretreatment. Autoclave (Parr reactor), which can operate in high temperature and high pressure, was created in order to use with heater stirrer because of the homogeneous system. The reactor was created from Stainless steel type 316 in order to withstand with acid solution. Autoclave was shown in Figure A1.

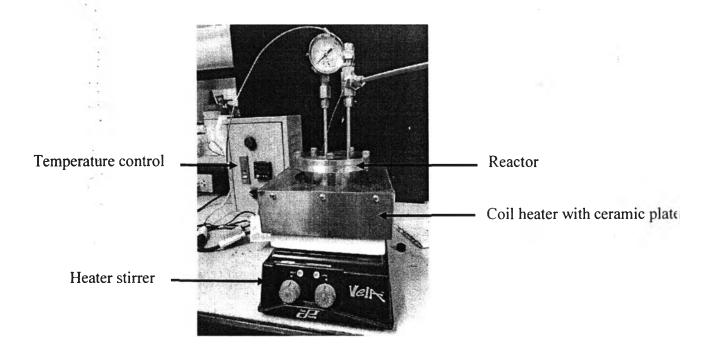


Figure A1 Autoclave.

Autoclave consisted of temperature control, coil heater with ceramic plate, heater stirrer, Reactor, pressure gauge, and pressure release value. About heating rate of autoclave, the researcher explored the heating rate of autoclave system by plotting the graph between temperature from room temperature to temperature setting point (100 °C, 120 °C, 140 °C, and 160 °C) and time (1 min interval). The result is shown showed in Figure A2.

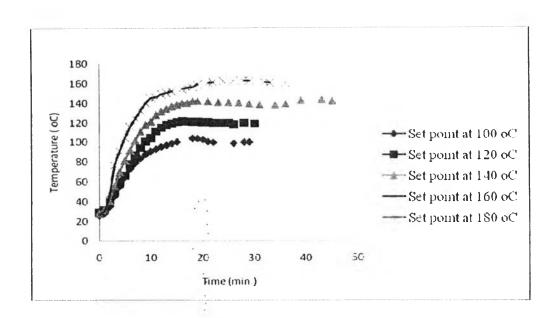


Figure A2 Heating rate of autoclave system.

Figure A2 showed that the system had approximately used 15 minutes to enter steady stage (Temperature setting point). Therefore, heating of system approximately equaled 5 °C/min.

Appendix B Activity of Enzyme

The enzymatic activity was determined according to method developed by T.K. Ghose. First, Glucose standard curve was plotted between amounts of glucose (micromole/ml) and value of absorbance of UV-VIS Spectrometer. The calibration curve is shown in B1.

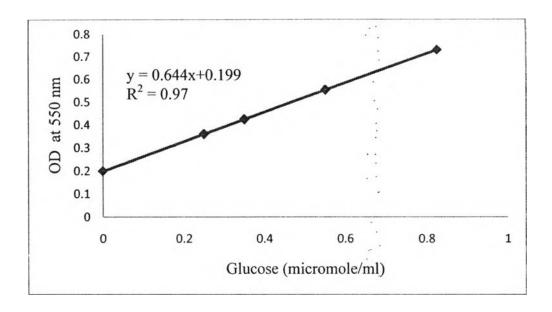


Figure B1 Glucose standard curve.

After plotting Glucose standard curve, cellulase (from Novozyme) and ACCELLERASE@1500 (from Genencor) was investigated activity of enzyme. The result is shown in Table B1.

Table B1 Activity of Enzyme

Type of Enzyme	Activity (unit/ml)
Cellulase	2889.27
ACCELLERASE@1500	2482.45

Table B1 showed that cellulase from Novozyme has activity higher than ACCELLERASE@1500 from Genencor because this T.K. Ghose's method can detect only cellulase. Therefore, it is possible that cellulase in ACCELLERASE@1500 might be less than cellulase from Novozyme.

Appendix C Standard Curve of HPLC

The fermentable sugars were identified and quantified by both Aminex column HPX-87H. The column was maintained at 65 °C and 20 µl of each sample and eluted by 0.005 M sulfuric acid at the flow rate 0.6ml/min and Lichrospher NH₂ 250*4mm column in HPLC-ELSD), and organic acid were identified and quantified by both Aminex column HPX-87H. The column was maintained at 25 °C and 10 µl of each sample and eluted by 84% of ACN: 16% of water at the flow rate 1.6 ml/min. Analytical grade glucose, xylose, mannose, galactose, and cellubiose were used as authentic standards. The result of standards was shown in Table C1 and Figure C1.

Table C1 Retention time of each fermentable sugar

Standard	Aminex column (min)	Lichrospher column (min)
Glucose	8.638	9.633
Xylose	9.216	5.466
Mannose	9.114	8.516
Galactose	9.137	10.466
Cellubiose	7.143	N/A
Arabinose	9.939	6.433
Rhamnose	N/A	4.350
Furfural	N/A	N/A

N/A; Not Available

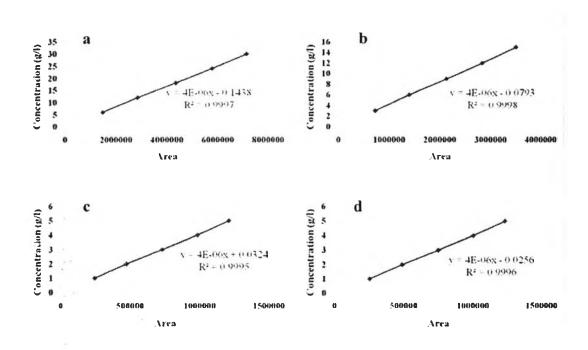


Figure C1 Standard curve of sugar for (a) standard glucose (b) standard xylose (c) standard cellubiose and (d) standard arabinose.

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 <u>Proceedings of The 2nd Research Symposium on Petroleum, Petrochemicals, and Advanced Materials and The 17th PPC Symposium on Petroleum, Petrochemicals, and Polymers, Bangkok, Thailand.

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