

REFERENCES

- Andrade, J.D.; Vanwagenen, R.A.; Gregonis, D.E. (1985) Remote fiber-optic biosensors based on evanescent-excited fluoro-immunoassay: Concept and progress. *IEEE Trans. Electron. Devices* 32, 1175–1179.
- Barreiros dos Santos, M., Agusil, J.P., Prieto-Simon, B., Sporer, C., Teixeira, V. (2013) Highly sensitive detection of pathogen Escherichia coli O157:H7 by electrochemical impedance spectroscopy. *Biosensor and Bioelectronics* 45, 174-180.
- Billah, M.M., Hodges, C.S., Hays, H.C. and Millner, P.A. (2010) Directed immobilization of reduced antibody fragments onto a novel SAM on gold for myoglobin impedance immunosensing. *Bioelectrochemistry* 80(1), 49-54.
- Chaki, N.K., Aslam, M., Sharma, J., Vijayamohanan, K. (2001) Applications of self-assembled monolayers in materials chemistry. *Chemical Science* 113, 659-670.
- Chen, H., Heng, C.K., Puiu, P.D., Zhou, X.D., Lee, A.C., Lim, T.M., Tan, S.N. (2005) Detection of *Saccharomyces cerevisiae* immobilized on self-assembled monolayer (SAM) of alkanethiolate using electrochemical impedance spectroscopy. *Analytica Chimica Acta* 554, 52-59.
- Cooper, M.A. (2002) Optical biosensors in drug discovery. *Nat. Rev* 1, 515–528.
- Danielsson, B., Mosbach, K. (1988). Enzyme thermistors. *Methods in Enzymology* 137, 181–197.
- Danielsson, B., Mattiasson, B., Mosbach, K. (1981) Enzyme thermistor devices and their analytical applications. *Applied Biochemistry Bioengineering* 3, 97–143.
- Ding, S.-J., Chang, B.-W., Wu, C.-C., Lai, M.-F. and Chang, H.-C. (2005) Impedance spectral studies of self-assembly of alkanethiols with different chain lengths using different immobilization strategies on Au electrodes. *Analytica Chimica Acta* 554(1-2), 43-51.
- Eggins, B.R. (2002) *Chemical Sensors and Biosensors*. West Sussex: John Wiley.
- Fan, X., White, I.M., Shopova, S.I., Zhu H. (2008) Sensitive optical biosensors for unlabeled targets: A review. *Analytica Chimica Acta* 620, 8–26.

- Ganesh, V., Pal, S.K., Kumar, S. and Lakshminarayanan, V. (2006) Self-assembled monolayers (SAMs) of alkoxycyanobiphenyl thiols on gold--a study of electron transfer reaction using cyclic voltammetry and electrochemical impedance spectroscopy. *Journal of Colloid and Interface Science* 296(1), 195-203.
- Geng, P., Zhang, X., Meng, W., Wang, Q., Zhang, W., Jin, L., Feng, Z. and Wu, Z. (2008) Self-assembled monolayers-based immunosensor for detection of *Escherichia coli* using electrochemical impedance spectroscopy. *Electrochimica Acta* 53(14), 4663-4668.
- Grime, J.K. (1985) *Analytical Solution Calorimetry*. New York: Wiley.
- Hnaien, M., Diouani, M.F., Helali, S., Hafaid, I., Hassen, W.M., Renault, N.J., Ghram, A. and Abdelghani, A. (2008) Immobilization of specific antibody on SAM functionalized gold electrode for rabies virus detection by electrochemical impedance spectroscopy. *Biochemical Engineering Journal* 39(3), 443-449.
- Koyun, A., Ahlatcioglu, E., Ipek, Y.K. (2012) Biosensor and Their Principles. In Kara, S. (ed.) *A Roadmap of Biomedical Engineers and Milestones* (pp.115-142) Rijeka, Croatia: Intech.
- Long, F., Zhu, A., Shi, H. (2013) Recent Advances in Optical Biosensors for Environmental Monitoring and Early Warning. *Sensors* 13, 13928-13948.
- Long, F., He, M., Zhu, A.N., Shi, H.C. (2009) Portable optical immunosensor for highly sensitive detection of microcystin-LR in water samples. *Biosensors and Bioelectron* 24, 2346–2351.
- Luo, P., Liu, Y., Xia, Y., Xu, H. and Xie, G. (2014) Aptamer biosensor for sensitive detection of toxin A of *Clostridium difficile* using gold nanoparticles synthesized by *Bacillus stearothermophilus*. *Biosensor and Bioelectron* 54, 217-221.
- Mendes, R.K., Carvalhal, R.F. and Kubota, L.T. (2008) Effects of different self-assembled monolayers on enzyme immobilization procedures in peroxidase-based biosensor development. *Journal of Electroanalytical Chemistry* 612(2), 164-172.

- Mosbach, K., Danielsson, B. (1974) An enzyme thermistor. Biochimica et Biophysica Acta 364, 140–145.
- Narayanaswamy, R., Wolfbeis, O.S. (2004) Optical Sensors, Springer, New York.
- Neria-Gonzalez, I., Wang, E.T., Ramirez, F., Romero, J.M. and Hernandez-Rodriguez, C. (2006) Characterization of bacterial community associated to biofilms of corroded oil pipelines from the southeast of Mexico. Anaerobe 12(3), 122-133.
- Park, B.W., Yoon, D.Y., Kim, D.S. (2011) Formation and modification of a binary self-assembled monolayer on a nano-structured gold electrode and its structural characterization by electrochemical impedance spectroscopy. Journal of Electroanalytical Chemistry 661(2), 329-335.
- Pissinis, D.E., Linarez Pérez, O.E., Cometto, F.P. and López Teijelo, M. (2014) Preparation and characterization of self-assembled monolayers of 2-mercaptopicotinic acid on Au(111). Journal of Electroanalytical Chemistry 712, 167-177.
- Pramanik, S., Pingguan-Murphy, B., Osman, A.A. (2013) Developments of Immobilized Surface Modified Piezoelectric Crystal Biosensors for Advanced Applications. Electrochemical Science 8, 8863-8892.
- Ribuat, C., Reybier, K., Torbiero, B., Launay, J., Valentin, A., Reynes, O., Fabre, P.-L., Nepveu, F. (2008) Strategy of red blood cells immobilization onto a gold electrode: Characterization by electrochemical impedance spectroscopy and quartz crystal microbalance. IRBM 29, 141-148.
- Sassolas, A., Blum, L.J., Leca-Bouvier, B.D. (2011) Immobilization Strategies to Develop Enzymatic. Biosensors Biotechnology Advances 30(3), 489-571.
- Shokrollahzadeh, S., Azizmohseni, F., Golmohammad, F., Shokouhi, H. and Khademhaghigat, F. (2008) Biodegradation potential and bacterial diversity of a petrochemical wastewater treatment plant in Iran. Bioresource Technology 99(14), 6127-6133.
- Spink, C., Wadso, I. (1976) Calorimetry as an analytical tool in biochemistry and biology. Methods of Biochemical Analysis. 23, 1–159.
- Sowards, J.W., Williamson, C.H.D., Weeks, T.S., McColsky, J.D. and Spear, J.R. (2014) The effect of Acetobacter sp. and a sulfate-reducing bacterial

- consortium from ethanol fuel environments on fatigue crack propagation in pipeline and storage tank steels. Corrosion Science 79, 128-138.
- Thevenot, D.R., Toth, K., Durst, R.A., Wilson, G.S. (1999) Electrochemical Biosensors: Recommended Definitions and Classification. Pure Apply Chemistry 7, 2333-2348.
- Vo-Dinh, T. (2008) Nanosensing at the single cell level. Spectrochimica Acta Part B: Atomic Spectroscopy 63, 95–103.
- Xiao, F., Zhang, N., Gu, H., Qian, M., Bai, J., Zhang, W., Jin, L. (2011) A monoclonal antibody-based immunosensor for detection of Sudan I using electrochemical impedance spectroscopy. Talanta 84, 204-211.
- Zheng, M., Chen, Y., Zhou, Y., Tang, Y. and Lu, T. (2010) The effect of the surface coverage of the phosphonic acid terminated self-assembled monolayers on the electrochemical behavior of dopamine. Talanta 81(3), 1076-1080.

APPENDIX

These Figures show the electrochemical impedance plot (Nyquist plot) of the SAMs with different alkyl chain lengths after immobilizing antibodies and binding Claudin 4 Recombinant Proteins at different concentrations

1. Mercaptopropionic acid (3MPA)

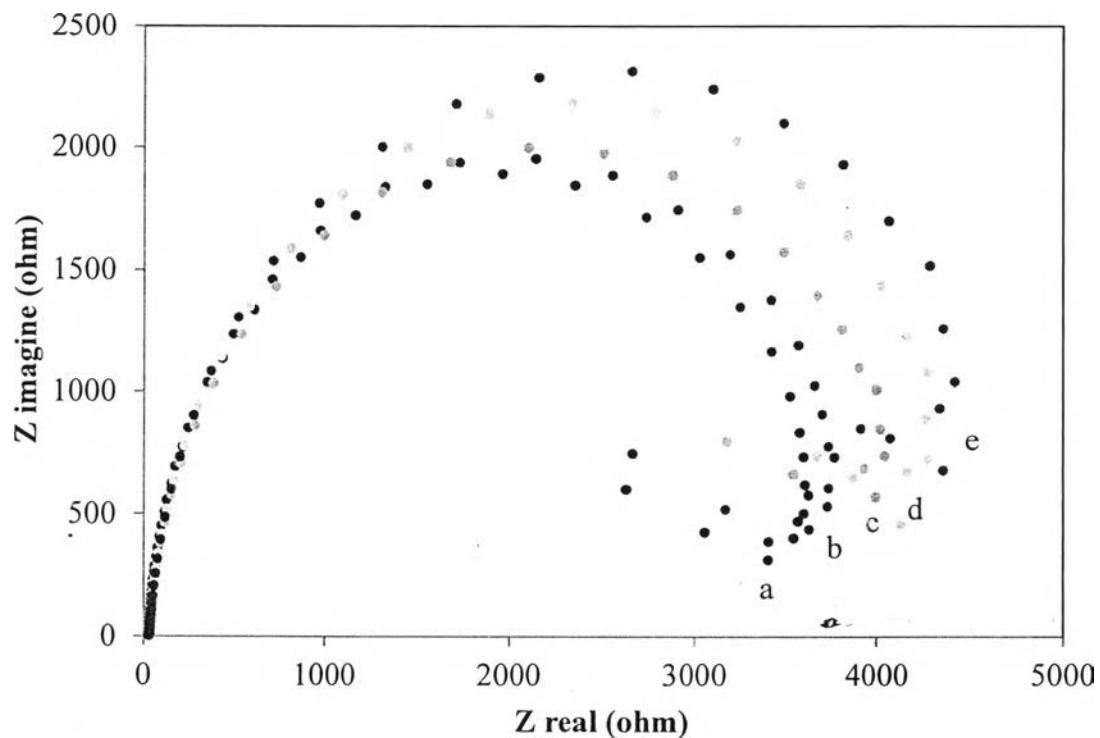


Figure A1 Impedance plot (Nyquist plot) of the sensor with 3MPA thiolate SAM in 0.1 M KCl, 5 mM $K_3Fe(CN)_6$ and 5 mM $K_4Fe(CN)_6 \cdot 3H_2O$ in 0.05 M phosphate buffer pH 7 as the supporting electrolyte for (a) SAM (b) after antibodies immobilization (c) 0.5 μ g/ml antigen, (d) 1 μ g/ml antigen , (e) 2 μ g/ml antigen.

2. Mercaptohexanoic acid (6MOA)

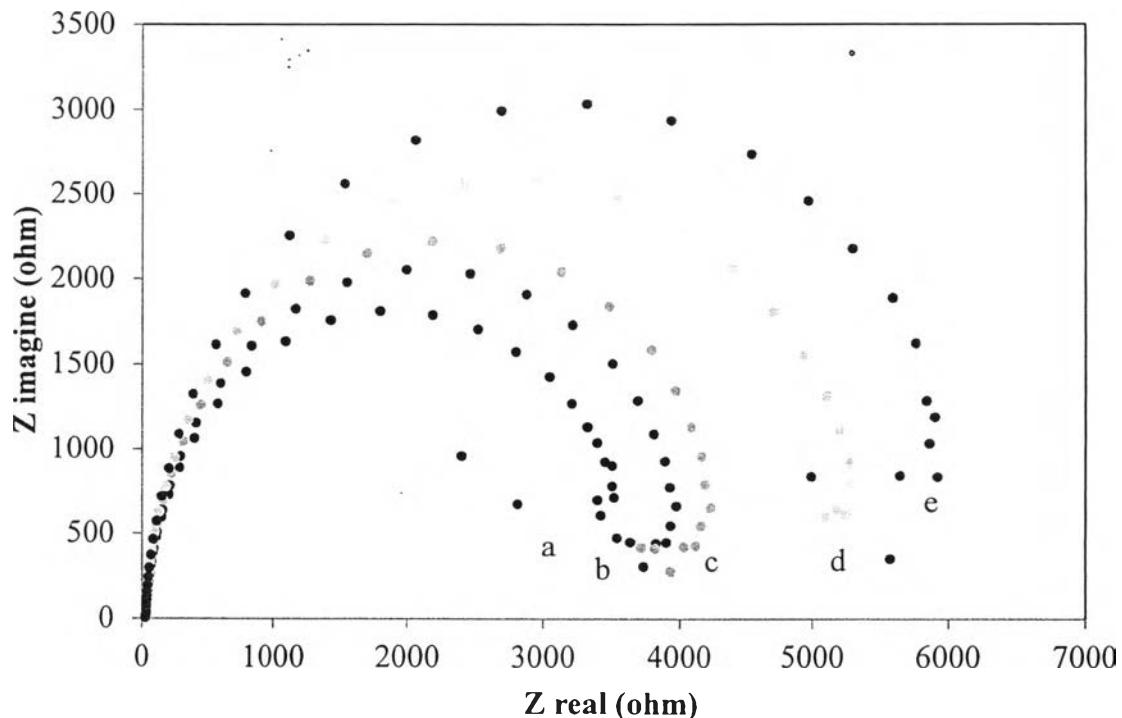


Figure A2 Impedance plot (Nyquist plot) of the sensor with 6 MHA thiolate SAM in 0.1 M KCl, 5 mM $K_3Fe(CN)_6$ and 5 mM $K_4Fe(CN)_6 \cdot 3H_2O$ in 0.05 M phosphate buffer pH 7 as the supporting electrolyte for (a) SAM (b) after antibodies immobilization (c) 0.5 μ g/ml antigen, (d) 1 μ g/ml antigen , (e) 2 μ g/ml antigen.

3. Mercaptooctanoic acid (8MOA)

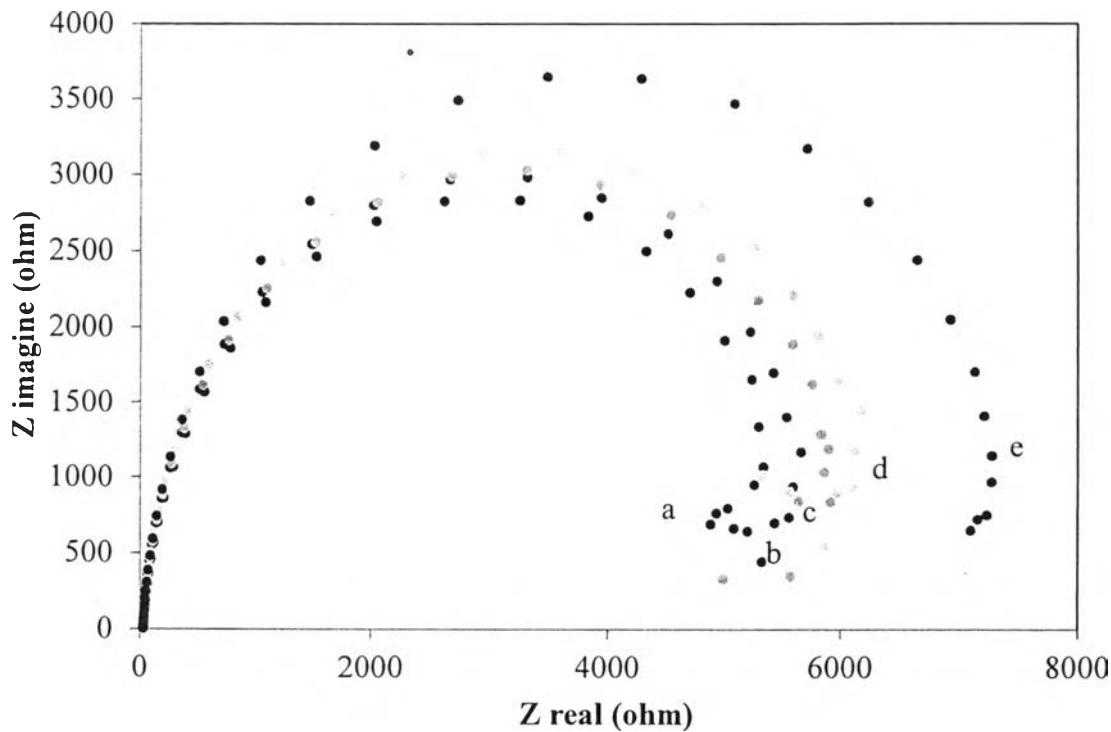


Figure A3 Impedance plot (Nyquist plot) of the sensor with 8 MOA thiolate SAM in 0.1 M KCl, 5 mM K₃Fe(CN)₆ and 5 mM K₄Fe(CN)₆·3H₂O in 0.05 M phosphate buffer pH 7 as the supporting electrolyte for (a) SAM (b) after antibodies immobilization (c) 0.5 µg/ml antigen, (d) 1 µg/ml antigen , (e) 2 µg/ml antigen.

4. Mercaptoundecanoic acid (11MUA)

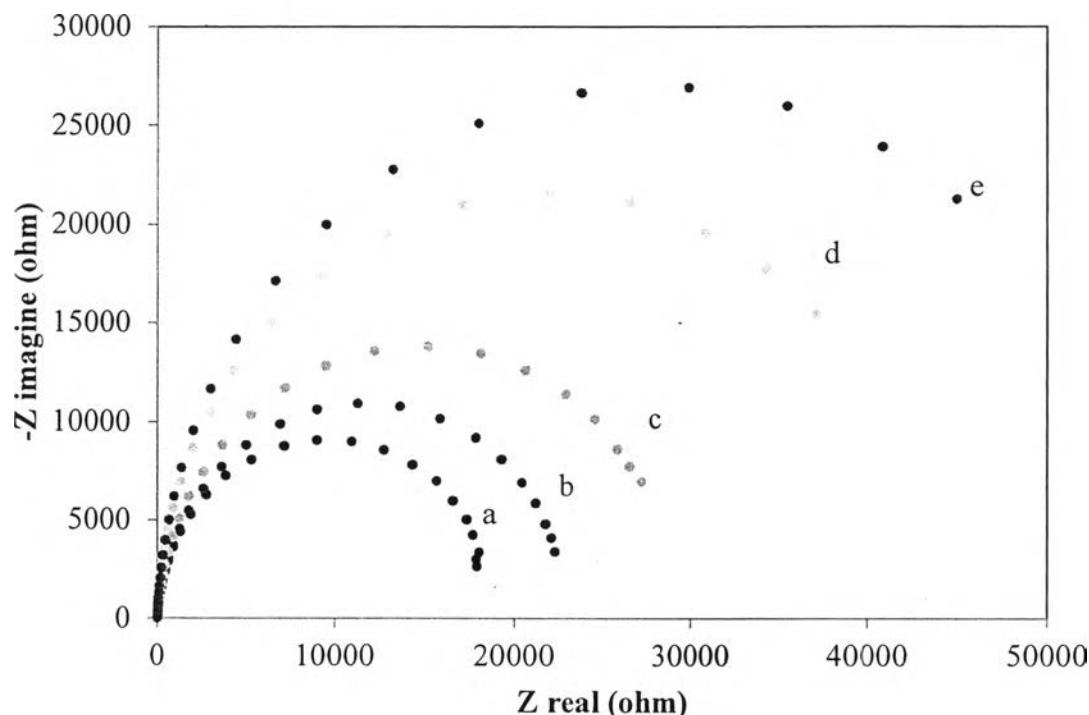


Figure A4 Impedance plot (Nyquist plot) of the sensor with 8 MOA thiolate SAM in 0.1 M KCl, 5 mM K₃Fe(CN)₆ and 5 mM K₄Fe(CN)₆·3H₂O in 0.05 M phosphate buffer pH 7 as the supporting electrolyte for (a) SAM (b) after antibodies immobilization (c) 0.5 µg/ml antigen, (d) 1 µg/ml antigen , (e) 2 µg/ml antigen.

CURRICULUM VITAE

Name: Mr. Chayut Yaempho

Date of Birth: June 9, 1991

Nationality: Thai

University Education:

2013-2015 Master Degree of Petrochemical Technology, Petroleum and Petrochemical Technology college, Chulalongkorn University, Bangkok, Thailand

2009-2013 Bachelor Degree of Science in Department of Chemical Technology, Faculty of Science, Chulalongkorn University, Bangkok, Thailand

Work Experience:

2013 Position: Student trainee

Company name: Thaioil Public Company Limited

Proceeding:

1. Yaempho, C., Kim, D. S., and Malakul, P. (2015, April 21) Electrochemical Biosensors to Detect Microbes in Petrochemical Processes. Proceedings of the 6th Proceedings of The 6th Research Symposium on Petroleum, Petrochemicals, and Advanced Materials and The 21th PPC Symposium on Petroleum, Petrochemicals, and Polymer, Bangkok, Thailand.