



REFERENCES

- Allcock, E. R., and D. R. Woods. 1981. Carboxymethyl cellulase and cellobiase production by *Clostridium acetobutylicum* in an industrial fermentation medium. *Appl. Environ. Microbiol.* 41(2): 539-41.
- Ando, S., H. Ishida, Y. Kosugi, and K. Ishikawa . 2002 . Hyperthermostable endoglucanase from *Pyrococcus horikoshii*. *Appl. Environ. Microbiol.* 68(1): 430-433 .
- Ash, C., J. A. E. Farrow, S. Wallbanks, and M. D. Collins. 1991. Phylogenetic heterogeneity of the genus *Bacillus* revealed by comparative analysis of small-subunitribosomal RNA. *Lett Appl Microbiol.* 13:202–206
- Ash, C., F. G. Priest, and M. D. Collins. 1993. Molecular identification of rRNA group 3 bacilli using a PCR probe test. Proposal for the creation of a new genus *Paenibacillus*. *Antonie van Leeuwenhoek.* 64:253–260
- Ash *et al.* 2005. JUDICIAL COMMISSION OF THE INTERNATIONAL COMMITTEE ON SYSTEMATICS OF PROKARYOTES: The type species of the genus *Paenibacillus*. *Int. J. Syst. Evol. Microbiol.* 55: 513.
- Au, K. S., and K. Y. Chan. 1987. Purification and properties of the endo-1,4- β -glucanase from *Bacillus subtilis*. *J. Gen. Microbiol.* 133:2155–2162.
- Aurilia, V., S. Y. Ding, M. T. Rincon, R. Lamed, J. C. Martin, S. I. McCrae, Y. Shoham, E. A. Bayer, and H. J. Flint. 2000. Cellulosomal Scaffoldin-Like Proteins from *Ruminococcus flavefaciens*. *J. Bacteriol.* 183(6): 1945-1953.
- Barrow, G. H., and R. K. A. Feltham. 1993. Cowan and Steel's Manual for Identification of Medical Bacteria. Third edition. Cambridge University Press, Cambridge. pp. 331.
- Beguin, P., and JP. Aubert. 1994. The biological degradation of cellulose. *FEMS*

- Microbiol Rev.* 13:25-58.
- Béguin, P., and H. Eisen. 1978. Growth properties of *Cellulomonas flavigena* mutants affected in cellulose utilization. *J. Bacteriol.* 133(2): 1044–1046.
- Belaich, A., G. Parsiegla, L. Gal, C. Villard, R. Haser, and J. Belaich. 2002. Cel9M, a new family 9 cellulase of the *Clostridium cellulolyticum* cellulosome. *J. Bacteriol.* 184(5): 1378-1384.
- Bergquist, P. L., M. D. Gibbs, D. D. Morris, V. S. J. Teo, D. J. Saul, and H. W. Morgan. 1999. Molecular diversity of thermophilic cellulolytic and hemicellulolytic bacteria. *Microbiol. Ecol.* 28: 99-110.
- Bhat, M. K. 2000. Cellulases and related enzymes in biotechnology. *Biotechnol. Advanc.* 18:355-383.
- Bronnenmeier, K., and L. Staudenbaur. 1990. Cellulose hydrolysis by a highly thermostable endo-1,4- β -glucanase (Avicelase I) from *Clostridium stecorarium*. *Enz. Microbiol. Tech.* 12:431–436.
- Chan, K. Y., and K. S. Au. 1987. Studies on cellulase production by *Bacillus subtilis*. *Antoine van Leeuwenhoek* . 53:125–136.
- Chen, S. F., L. Zhao, and D. H. Liu. 2004. Solid-state Fermentation for Cellulase Production by *Trichoderma viride*. *Food Ferment. Ind.* 30(1):9–12.
- Clarke, J. H., J. E. Rixon, A. Ciruela, H. J. Gilbert, and G. P. Hazlewood. 1997. Family-10 and family-11 differ in their capacity to enhance the bleachability of hardwood and softwood paper pulps. *Appl. Microbiol. Biotechnol.* 48:177–183.
- Claus, D., and R. C. W. Berkeley. 1987. Genus *Bacillus* Cohn In *Bergey's Manual of Determinative Bacteriology*, ed P.H.A.Sneath. 1105–1140. Baltimore: William & Wilkins.
- Chen, P., T. Wei, Y. Chang, and L. Lin. 2004. Purification and characterization of carboxymethyl cellulase from *Sinorhizobium fredii*. *Bot. Bull. Acad. Sin.*

- 45: 111-118.
- Coughlan, M. P., and L. G. Ljungdahl. 1988. Comparative biochemistry of fungal and bacterial cellulolytic enzyme systems. In *Biotechnology and Genetics of Cellulose Degradation* pp. 11–30. London: Academic Press.
- Csiszar, E., K. Urbanszki, and G. Szakacs. 2001. Biotreatment of desized cotton fabric by commercial cellulase and xylanase enzymes. *J. Mol. Catal. B: Enzym.* 11: 1065–1072.
- Deacon, J. W. (ed.) 2005-2006. The microbial world : *Armillaria mellea* and other wood-decay fungi. [Online]. <http://helios.bto.ed.ac.uk/bto/microbes/armill.htm>. Accessed 20 March 2006.
- Dehority, B. A. 1968. Mechanism of isolated hemicellulose and xylan degradation by cellulolytic rumen bacteria. *Appl. Environ. Microbiol.* 16(5): 781–786.
- Deman, J. M., De man, L., and Gupta, S. 1986. Texture and microstructure of soybean curd (tofu) as affected by different coagulants. *Food. Micro. Struct.* 5: 3-89.
- Ding, S. Y., E. A. Bayer, D. Steiner, Y. Shoham, and R. Lamed. 1999. A Novel Cellulosomal Scaffoldin from *Acetivibrio cellulolyticus* That Contains a Family 9 Glycosyl Hydrolase. *J. Bacteriol.* 181:6720-6729.
- Endo, K., Y. Hakamada, S. Takizawa, H. Kubota. 2001. A novel alkaline endoglucanase from an alkaliphilic *Bacillus* isolate: enzymatic properties, and nucleotide and deduced amino acid sequences. *Appl. Microbiol. Biotechnol.* 57:109-116.
- Felsenstein, J. 1985. Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, 39, 783-791. Forbes, L. 1981. Rapid flagella stain. *J. Clin. Microbiol.* 13:807-809.
- Fukumori, F., T. Kodo, and K. Horikoshi. 1985. Purification and properties of a cellulase from alkalophilic *Bacillus* sp. No. 1139. *J. Gen. Microbiol.* 131:129–135.
- Gardner, R. M., K. C. Doerner and B. A. White. 1987. Purification and characterization of

- an exo- β -1,4-glucanase from *Ruminococcus flavefaciens* FD-1. *J. Bacteriol.* 169:4581–4588.
- Garrity, G. M. 2001. Bergey's manual of systematic bacteriology. 2nd edn. Springer, Berlin Heidelberg New York.
- George, S. P., A. Ahmad, and M. B. Rao. 2001. Studies on carboxymethyl cellulase produced by an alkalothermophilic actinomycete. *Bioresource Technol.* 77: 171–175.
- Ghose, T. K. 1987. Measurement of Cellulase Activities. *Pure & Appl. Chem.* 59: 257-268.
- Gordon, R. E., W. C. Haynes, and C. Pang. 1973. *The Genus Bacillus. Agricultural Handbook No. 427*. Washington DC: The United States Department of Agriculture.
- Guedon, E., M. Desvaux, and H. Petitdemange. 2002. Improvement of cellulolytic properties of *Clostridium cellulolyticum* by metabolic engineering. *Appl. Environ. Microbiol.* 68(1): 53-58.
- Hakamada, Y., K. Endo, S. Takizawa, T. Kobayashi, T. Shirai, T. Yamane, and S. Ito. 2002. Enzymatic properties, crystallization, and deduced amino acid sequence of an alkaline endoglucanase from *Bacillus circulans*. *Biochim. Biophys. Acta.* 1570:174-180.
- Haki, G. D., and S. K. Rakshit . 2003 . Developments in industrially important thermostable enzymes. *Bioresource Technol.* 89: 17–34.
- Hao, C., B. Yu, and L. Yan. 2006 . Optimization of the medium for the production of cellulase by the mutant *Trichoderma reesei* WX-112 using response surface methodology. *Food Technol. Biotechnol.* 44(1): 89-94.
- Harjunpaa, V. 1998. Enzymes hydrolyzing wood polysaccharides. *Technical Research Centre of Finland, VTT Publications.* 372: 1-76.

- Hongpattarakere, T. 2002. Hyperthermostable cellulolytic and hemicellulolytic enzymes and their biotechnological applications. *Songklanakarin J. Sci. Technol.* 24(3): 481-491.
- Horikoshi, K., M. Nakao, Y. Kurono, and N. Sashihara. 1984. Cellulases of an alkalophilic *Bacillus* strain isolated from the soil. *Canadian Journal of Microbiology*. 30: 774–779.
- Hoshino, E., and S. Ito. 1997. Application of alkaline cellulases that contribute to soil removal in detergents. In: van Ee J. H., Misset O., Bass E. J.(eds) Enzymes in detergency. Marcel Dekker, New York, pp 149-174.
- Howard, T., and A. White. 1988. Molecular cloning and expression of cellulase genes from *Ruminococcus albus* 8 in *Escherichia coli* Bacteriophage λ . *Appl. Environ. Microbiol.* 54(7): 1752–1755.
- Huang, L., and W. Forsberg . 1987. Isolation of a cellobextrinase from *Bacteroides succinogenes*. *Appl. Environ. Microbiol.* 53(5): 1034-1041.
- Immanuel, G., R. Dhanusa, P. Prema, A. Palavesam. 2006. Effect of different growth parameters on endoglucanase enzyme activity by bacteria isolated from coir retting effluents of estuarine environment. *Int. J. Environ. Sci. Tech.* 3(1):25-34.
- Irwin, D., D. H. Shin, S. Zhang, B. K. Barr, J. Sakon, P. A. Karplus, and D. B. Wilson. 1998. *J. Bacteriol.* 180:1709–1714.
- Ito, S., S. Shikata, K. Ozaki, S. Kawai, K. Okamoto, S. Inque, A. Takel, Y-I. Ohta, and T. Satoh. 1989. Alkaline cellulase for laundry detergents: Production by *Bacillus* sp. KSM-635 and enzymatic properties. *Agricultural and Biological Chemistry*. 53:1275–1281.
- Lynd, L. R., J. H. Cushman, R. J. Nichols, and C. E. Wyman. 1991. Fuel ethanol from cellulosic biomass. *Science* 251:1318–1323.
- Johnson, E. A., M. Sakajoh, G. Halliwell, A. Madia, and A. L.Demain. 1982.

- Saccharification of complex cellulosic substrate by the cellulase system of *Clostridium thermocellum*. *Appl. Environ. Microbiol.* 43:1125–1132.
- Johnson, P. E., E. Brun, L. F. MacKenzie, S.G. Withers, and L.P. McIntosh .1999 . The cellulose-binding domains from *Cellulomonas fimi* beta-1, 4-glucanase CenC bind nitroxide spin-labeled cellooligosaccharides in multiple orientations. *Mol Biol.* 287(3): 609-25.
- Jørgensen, H., and L. Olsson. 2005. Production of cellulases by *Penicillium brasiliense* IBT 20888-effect of substrate on hydrolytic performance. *Enz. Microbiol. Technol.* 38(3-4): 381-390.
- Julliard, H., A. D. Vaux, L. Millet, and G. Fonty. 1999. Identification of *Ruminococcus flavefaciens* as the predominant cellulolytic bacterial species of the equine cecum. *Appl. Environ. Microbiol.* 65(8): 3738-3741.
- Kawai, S., H. Okoshi., K. Ozaki, S. Shikata, K.Ara, and S. Ito. 1988. Neutrophilic *Bacillus* strain, KSM-522, that produces an alkaline carboxymethyl cellulase. *Agric. Biol. Chem.* 52:1425–1431.
- Khan, A. W., M. Asther, and C. Giuliano. 1984. Utilization of steam and explosion-decompressed aspen wood by some anaerobes. *Journal of Fermentation Technology*. 62:335–339.
- Kim, C. H., and D. S. Kim. 1993. Extracellular cellulolytic enzymes of *Bacillus circulans* are present as two multiple-protein complexes. *Appl. Biochem. Biotechnol.* 42:83–94.
- Kim, H., and M. Y. Pack. 1988. Endo-1,4- β -glucanase encoded by *Bacillus subtilis* gene cloned in *Bacillus megaterium*. *Enz. Microbial. Technol.* 10:347–351.
- Komagata, K., and K. Suzuki. 1987. Lipid and cell-wall analysis in bacterial systematics. *Methods microbiol.*19 : 161-207.
- Kumar, S., D. Bouzida, R. H. Swendsen, P. A. Kollman, and J. M. Rosenberg. 1992. The

- weighted histogram analysis method for free-energy calculations on biomolecules.
- I. The method. *J. Comp. Chem.* 13:1011-1021.
- Leatherwood, J. M. 1965. Cellulase from *Ruminococcus albus* and mixed rumen microorganisms. *Appl. Environ. Microbiol.* 13(5): 771-775.
- Lednická, D., J. Mergaert, M. C. Cnockaert, and J. Swings. 2000. Isolation and identification of cellulolytic bacteria involved in the degradation of natural cellulosic fibres. *Syst Appl Microbiol.* 23:292-299.
- Lee, B. H., and T. H. Blackburn, 1975. Cellulase production by a thermophilic *Clostridium* species. *Appl. Environ. Microbiol.* 30(3): 346-353.
- Logan, N. A., E. D. Clerck, L. Lebbe, A. Verhelst, J. Goris, G. Forsyth, M. Rodriguez-Diaz, M. Heyndrickx, and P. D. Vo. 2004. *Paenibacillus cineris* sp. nov. and *Paenibacillus cookii* sp. nov., from Antarctic volcanic soils and a gelatin-processing plant. *Int. J. of Syst. Evol. Microbiol.* 54:1071-1076.
- Lu, W. J., H. T. Wang, S. J. Yang, Z. C. Wang, and Y. F. Nie. 2005. Isolation and characterization of mesophilic cellulose-degrading bacteria from flower stalks-vegetable waste co-composting system. *Gen. Appl. Microbiol.* 51: 353-360.
- Lucana, D. O. O., T. Schaa , and H. Schrempf. 2004. The novel extracellular *Streptomyces reticuli* haem-binding protein HbpS influences the production of the catalase-peroxidase CpeB . *Microbiol.* 150: 2575-2585.
- Lynd, L. R., P. J. Weimer, W. H. V. Zyl, and S. P. Isak. 2002. Microbial Cellulose Utilization: Fundamentals and Biotechnology. *Microbiol. Molecular Biol Rev.* 66: 506-577
- Rajoka, M. I., and K. A. Malik. 1984. Cellulase and hemicellulase production by *Cellulomonas flavigena* NIAB 441. *Biotechnol Lett.* 6:597-600.
- Mandels, M. 1982. Cellulases. In *Annual reports on fermentation processes*, Volume 5, ed TsaoG.T. pp. 35–78 New York: Academic Press.

- Marmur, J., and P. Doty. 1962. Determination of the base composition of deoxyribonucleic acid from its thermal denaturation temperature. *J. Mol. Biol.* 5:109-118.
- Miller, G. L., R. Blum, W. E. Glennon, and A. L. Burton. 1960. Measurement of carboxymethyl cellulase activity. *Analyt. Biochem.* 2:127–132.
- Mosier, N., C. Wyman, B. Dale, R. Elander, Y. Y. Lee, M. Holtzapple, and M. Ladisch. 2005. Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technol.* 96 : 673–686.
- Murashima, K., A. Kosugi, and R. H. Doi. 2002. Synergistic effects on crystalline cellulose degradation between cellulosomal cellulases from *Clostridium cellulovorans*. *Bacteriol.* 184(18): 5088-5095.
- Muzariri, C.C., J. Mapingire, J. Mazorodze, and L. Mandikutse. 2001. Isolation and screening of microorganisms for potential application in remediation of effluent water from the pulp and paper industry. The 2nd WARFSA/WaterNet Symposium: Integrated Water Resources Management: Theory, Practice, Cases; Cape Town. pp. 242-250.
- Nakamura, K., and K. Kitamura. 1983. Purification and some properties of a cellulase active on crystalline cellulose from *Cellulomonas uda*. *J. Ferment. Technol.* 61:379–382.
- Nelson, N. 1944. A photometric adaption of the Somogyi methode for the determination of glucose. *Biol. Chem.* 153: 375-380.
- Ng, T. K., A. Ben-Bassat , and J. G. Zeikus. 1981 . Ethanol production by thermophilic bacteria: fermentation of cellulosic substrates by cocultures of *Clostridium thermocellum* and *Clostridium thermohydrosulfuricum* . *Appl. Environ. Microbiol.* 41(6): 1337-1343.
- Ohara, H., S. Karita, T. Kimura, K. Sakka, K. Ohmiya. 2000. Characterization of the

- cellulolytic complex (cellulosome) from *Ruminococcus albus*. *Biosci Biotechnol Biochem* 64:254–260
- Ogawa, A., A. Suzumatsu, S. Takizawa, H. Kuboto, K. Sawada, Y. Hakamada, S. Kawai, T. Kobayashi, and S. Ito. 2007. Endoglucanases from *Paenibacillus* spp from a new clan in glycoside hydrolase family 5. *J. Biotechnol.* Article in press.
- Ojuma, T. V., B. O. Solomon, E. Betiku, S. K. Layokun, and B. Amigun. 2003. Cellulase production by *Aspergillus flavus* linn isolate NSPR 101 fermented in sawdust, bagasse and corncob. *African Biotechnol.* 2(6):150–152.
- Okoshi, H., K. Ozaki, S. Shikata, K. Oshini, S. Kawai, and S. Ito. 1990. Purification and characterization of multiple carboxymethyl cellulase from *Bacillus* sp. KSM-522. *Agric. Biol. Chem.* 54:83–89.
- Paradis, F. W., R. A. Warren, D. G. Kilburn, and R. C. Miller. 1987. The expression of *Cellulomonas fimi* cellulase genes in *Brevibacterium lactofermentum*. *Gene*. 61(2): 199-206.
- Patel, J. B., J. Richard, J. R. Wallace, B. A. Brown-Elliott, T. Taylor, C. Imperatrice, D. G. B. Leonard, R. W. Wilson, L. Mann, K. C. Jost, and I. Nachamkin. 2004. Sequence-Based Identification of Aerobic Actinomycetes. *J. Clin. Microbiol.* 42(6): 2530–2540.
- Priest, F. G. 1977. Extracellular enzyme synthesis in the genus *Bacillus*. *Bacteriol. Rev.* 41:711-753.
- Rao, M., V. V. Deshpande, and C. Mishra. 1986. Purification, characterization and synergistic action of endoglucanases from *Fusarium lini*. *Biotech. Bioeng.* 28:1100–1105.
- Robson, L. M. and G. H. Chambliss. 1984. Characterization of the cellulolytic activity of a *Bacillus* isolate. *Appl. Environ. Microbiol.* 47:1039–1046.

- Ruttersmith, L. D., and R. M. Daniel. 1993. Thermostable α -glucosidase and α -xylosidase from *Thermotoga* sp. Strain FjSS3B.1. *Biochim. Biophys. Acta.* 1156:167-172.
- Saito, H., and K. Miura. 1963. Preparation of transforming DNA by phenol treatment. *Biochem. Biophys. Acta.* 72: 619-629.
- Saitou, N., and M. Nei. 1987. The neighboring-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4: 406-425.
- Sa-pereira, P., A. Mesquita, J. C. Duarte, M. R. A. Barros, and M. Costa-Ferreira. 2002. Rapid production of thermostable cellulase-free xylanase by a strain of *Bacillus subtilis* and its properties. *Enz. Micro. Technol.* 30: 924-933.
- Salmon, L., U. Sahlberg, A. Oscarsson. 1997. The fibrillar orientation in the S2-layer of wood fibres as determined by x-ray diffraction analysis. *Wood Sci. Technol.* 31: 77-86.
- Sharma, P., J. K. Gupta, D. V. Vadhera, and D. K. Dube. 1990. Purification and properties of an endoglucanase from a *Bacillus* isolate. *Enz. Microbial. Technol.* 12:132-137.
- Shikata, S., K. Saeki, H. Okoshi, T. Yoshimatsu, K. Ozaki, S. Kawai, and S. Ito. 1990. Alkaline cellulase for laundry detergents: production by alkalophilic strains of *Bacillus* and some properties of crude enzymes. *Agri. Biol. Chem.* 54:91-96
- Sissons, C. H., K. R. Sharrock, R. M. Daniel, and H. W. Morgan. 1987. Isolation of cellulolytic anaerobic extreme thermophiles from New Zealand thermal sites. *Appl. Environ. Microbiol.* 53: 832-838.
- Shida, O., H. Takagi, K. Kadowaki, and K. Komagata. 1996. Proposal for two new genera, *Brevibacillus* gen. nov. and *Aneurinibacillus* gen. nov. *International Journal of Systematic Bacteriology.* 46(4):936-946.
- Shida, O., H. Takagi, K. Kadowaki, L. K. Nakamura, and K. Komagata. 1997. Transfer of *Bacillus alginolyticus*, *Bacillus chondroitinus*, *Bacillus curdlanolyticus*, *Bacillus glucanolyticus*, *Bacillus kobensis*, and *Bacillus thiaminolyticus* to the Genus

- Paenibacillus* and Emended Description of the Genus *Paenibacillus*. *Int. J. Syst. Bacteriol.* 47(2):289–298.
- Sternberg, D. 1976. Beta-glucosidase of *Trichoderma*: its biosynthesis and role in saccharification of cellulose. *Appl. Environ. Microbiol.* 31(5): 648–654.
- Stutzenberger, F. J. 1971. Cellulase production by *Thermomonospora curvata* isolated from municipal solid waste compost. *Appl. Environ. Microbiol.* 22(2): 147–152.
- Somogyi, M. 1952. Notes on sugar determination. *Biol. Chem.* 195: 19-23.
- Tamaoka, J., and K. Komagata. 1984. Determination of DNA base comparision by reverse-phase high-performance liquid chromatography. *FEMS. Microbiol. Lett.* 25: 125-128.
- Teather, R. M., and P. J. Wood. 1982. Use of congo red-polysaccharide interactions in enumeration and characterization of cellulolytic bacteria from bovine rumen. *Appl. Environ. Microbiol.* 43:777– 780.
- Ten, L. N., W. T. Im, M. K. Kim, M. S. Kang, and S. T. Lee. 2004. Development of a plate technique for screening of polysaccharide-degrading microorganisms by using a mixture of insoluble chromogenic substrates. *Microbiol. Methods.* 56(3): 375-382.
- Thompson, J. D., T. J. Gibson, F. Plewniak, F. Jeanmougin, and D. G. Higgins. 1997. The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.* 25: 4876-4882.
- Tindall, B. J. 2000. What is the type species of the genus *Paenibacillus*? Request for an Opinion. *Int. J. Syst. Evol. Microbiol.* 50: 939-940.
- Vielle, C., and G. J. Zeikus. 2001. Hyperthermophilic Enzymes: Sources, Uses, and Molecular Mechanisms for Thermostability. *Microbiol. Molecular Biol. Reviews.* 65(1):1-34.

- Wang, W., and J. A. Thomson. 1990. Nucleotide sequence of the *celA* gene encoding a cellulase of *Ruminococcus flavefaciens* FD-1. *Molec. Gen. Gene.* 22:265–269.
- Yazdi, M. T., J. R. Woodward, and A. Radford. 1990. The cellulase of *Neurospora crassa*: activity, stability and release. *J. Gen. Microbiol.* 136:1313–1319.
- Zhu, Y. S., Y. Q. Wu, W. Chen, C. Tan, J. H. Gao, J. X. Fei, and C.N. Shih. 1982. Induction and regulation of cellulase synthesis in *Trichoderma pseudokoningii* mutants EA₃-867 and N₂-78. *Enz. Microbial. Tech.* 4:3–12.
- Zverlov, V., S. Mahr, K. Riedel, and K. Bronnenmeier. 1998. Properties and gene structure of a bifunctional cellulolytic enzyme (CelA) from the extreme thermophile ' *Anaerocellum thermophilum*' with separate glycosyl hydrolase family 9 and 48 catalytic domains . *Microbiol.* 144: 457-465.

APPENDICES

APPENDIX A

Instruments and chemical reagents

1. Instruments

- Analytical balance: Mettler Toledo model AG204, Switzerland.
- Autoclave: Tomy model SS-325, Japan.
- Centrifuges: Beckman model Avanti J25, U.S.A; Eppendorf model 5430, Germany; Sorvall model RC-5C Plus and Sorvall tabletop centrifuge model RC-5C Plus, USA.
- Circulating Water Bath: Techre model TE8 A, UK.
- Freeze Dryer: Savant model Super Modulya 233, USA.
- Freezer : Sharp model FC27 (-20°C), Japan and Deep Freezer : REVCO model ULT1790-7-V12 (-80°C), USA.
- Hot plate and stirrer: Thermolyne model Crimarec2, USA.
- Incubator: Memmert model BE500(30°C, 37°C, 45°C, 50°C, and 55°C), Germany.
- Incubator shaker: New Brunswick Scientific model innova4300, U.S.A
- Magnetic stirrer: Ika model RO-10, Malaysia.
- Microwave: Sanyo model EM-815FW, Japan.
- Oven: Memmert UE 600, Germany.
- pH Meter: Mettler Toledo model CH-8603, Switzerland.
- Pipetteman: Gilson, Villiers-Le-Bel, France.
- Precision balance: Mettler Toledo model PB3002, Switzerland.
- Shaking Water Bath: Memmert, model WB22, Germany.
- Spectrophotometer: Sherwood Scientific model259, Cambridge, UK.
- Vortex mixer: Barnstead/Thermolyne model M37610-26, Iowa, USA.

2. Chemicals

Chemicals	Company	Grade
Acetone	Merck	Analytical
L-arginine monohydrochloride	Fluka	Analytical
Bovine serum albumin	Sigma	Analytical
Chloroform	Mallinckrodt	Analytical
Copper (II) sulfate pentahydrate	Sigma	Analytical
Ethanol	Carlo Erba	Analytical
Ethylene diamine tetraacetic acid (EDTA)	Merck	Analytical
Ferric sulfate sevenhydrate	Carlo Erba	Analytical
Folin-Ciocalteu's phenol	Merck	Analytical
Hydrochloric acid	Merck	Analytical
Magnesium sulfate heptahydrate	Sigma	Analytical
Methanol	Merck	Analytical
Phenol	Carlo Erba	Analytical
Potassium hydrogen sulfate	Merck	Analytical
Di-potassium tartate	Carlo Erba	Analytical
Sodium chloride	Carlo Erba	Analytical
Tri-sodium citrate dihydrate	Merck	Analytical
Sodium dodecyl sulfate	Fluka	Analytical
Sodium hydroxide	Merck	Analytical
Sodium potassium tartate	Merck	Analytical
Trichloroacetic acid	Merck	Analytical
Trisma base	Merck	Analytical
Tyrosine	Sigma	Analytical
Carboxymethyl cellulose (CMC)	Merck	Analytical
Cellulose powder	Merck	Analytical
Magnesium sulfate	Sigma	Analytical

Potassium chloride	Merck	Analytical
Di-Ammonium sulfate	Merck	Analytical
Ferric citrate	Merck	Analytical

APPENDIX B

Culture Media

All media were dispensed and sterilized in autoclave at 120° C, 15 pounds/inch pressure for 15 min except the medium for acid from carbon sources testing which were sterilized at 110° C, 10 pounds/inch pressure for 10 min.

1. C medium

Polypeptone	5	g
Yeast extract	1	g
K ₂ HPO ₄	4	g
MgSO ₄ .7H ₂ O	1	g
KCl	0.2	g
FeSO ₄ .7H ₂ O	0.02	g
Agar	15	g
Distilled water	1000	ml
Dissolve and adjust to pH 7.0		

2. Cellulose powder (CP) medium

Cellulose powder	1	g
Peptone	5	g
Yeast extract	1	g
K ₂ HPO ₄	4	g
MgSO ₄ .7H ₂ O	1	g
KCl	0.2	g
FeSO ₄ .7H ₂ O	0.02	g
Agar	15	g
Distilled water	1000	ml
Dissolve and adjust pH to 7.0		

3. Carboxymethyl cellulose (CMC) medium

CMC (Carboxymethyl cellulose)	1	g
Peptone	5	g
Yeast extract	1	g
K ₂ HPO ₄	4	g
MgSO ₄ .7H ₂ O	1	g
KCl	0.2	g
FeSO ₄ .7H ₂ O	0.02	g
Agar	15	g
Distilled water	1000	ml

Dissolve and adjust pH to 7.0

4. Carboxymethyl cellulose (CMC-basal) medium

(NH ₄) ₂ SO ₄	1	g
CMC	5	g
Yeast extract	1	g
Agar	10	g
Distilled water	1000	ml

5. L-arginine agar medium

Phenol red, 1.0% aq.solution	1.0	ml
L(+)arginine monohydrochloride	10.0	g
Agar	3.0	g
C medium	1000	ml

Dissolve the solids in the C medium, adjust to pH to 7.2

6. Aesculin broth

Aesculin	1	g
Ferric citrate	0.5	g
C medium	1000	ml

Dissolve the aesculin and iron salt in the C medium, adjust pH to 7.4 and sterilized at 110 °C for 10 min.

7. Casein agar

Skim milk	10	g
C medium	1000	ml
Agar	15	g

Dissolve and adjust pH to 7.2.

8. Gelatin agar

Gelatin	10	g
C medium	1000	ml
Agar	15	g

Dissolve and adjust pH to 7.2.

9. Motility test medium

Motility medium (Difco)	20	g
Distilled water	1000	ml

Dissolve and adjust pH to 7.2

10. Simmon Citrate agar

Simon citrate agar (Difco)	24.2	g
Distilled water	1000	ml

Dissolve and adjust pH to 6.8

11. Starch agar

Starch	10	g
C medium	1000	ml
Agar	15	g

Dissolve and adjust pH to 7.2.

12. Triple sugar iron agar

Triple sugar iron agar (Difco)	60	g
Distilled water	1000	ml

Dissolve and adjust pH to 7.4.

13. Tyrosine agar

Tyrosine	50	g
C medium	1000	ml
Agar	15	g

Dissolve and adjust pH to 7.2.

14. Deoxyribonuclease (DNase) media

DNase test agar (Difco)	42	g
Distilled water	1000	ml

Adjust pH to 7.3 and heat to boiling to dissolve completely

15. Indole test

Tryptone	10	g
Meat extract	3	g
Distilled water	1000	ml

Dissolve and adjusted pH to 7.4

16. Nitrate broth

Meat extract	3	g
Peptone	10	g
KNO ₃	1	g
Distilled water	1000	ml
Dissolve and adjusted pH to 7.2		

17. Tween 80 agar medium

Tween 80	2	ml
C medium	1000	ml
Agar	15	g
Dissolve and adjusted pH to 7.2		

18. Urea agar medium

Urea	20	g
C medium	1000	ml
Agar	15	g
Dissolve and adjusted pH to 7.2		

19. MR-VP broth

MR-VP medium (Merck)	17	g
Distilled water	1000	ml
Dissolve and adjusted to pH 6.9		

APPENDIXC

Reagents and Buffers

1. Reducing sugar

Standards of 0, 0.1, 0.2, 0.3, 0.4, 0.5, 0.7, 0.8, 0.9 and 1.0 mg/ml were prepared from glucose. The reactions were carried out with the same procedure as described by Somogyi and Nelson method (1952)

2. Flagella staining

Basic fuchisin	0.5	g
Tannic acid	0.2	g
Aluminium sulfate	0.5	g

Solvent was composed of a mixture of 2.0 of 95% ethanol, 0.5 ml of glucerol, and 7.5 ml of tris(hydroxymethyl)aminomethane(tris)buffer.

3. Kovacs'reagent

O-dimethylaminobenzaldehyde	5	g
Amyl alcohol	75	g
Conc. HCl	25	ml

Dissolve the aldehyde in the alcohol by gently warming in a water bath (about 50-55 °C). Cool and the acid with care. Protect from light and store at 4 °C.

4. Nitrate test reagent

Solution A

0.33% sulphanilic acid in 5 N- acetic acid

Dissolve by gentle heating

Solution B

0.6% dimethyl- α -naphthylaminein 5 N-acetic acid

Dissolve by gentle heating in a fume hood.

Add two drops of sulphanilic acid solution and three drops of *N,N*-dimethyl-l-naphthylamine into peptone nitrate broth inoculating with the test microorganisms.

5. 6 N HCl

Conc. HCl	60	ml
Distiller water	. 60	ml

Add conc. HCl into the distilled water

6. 2 N H₂SO₄

Conc. H ₂ SO ₄	2	ml
Distiller water	34	ml

Add conc. HCl into the distilled water

7. Ninhydrin solution

Ninhydrin	0.3	g
l-Butanol	100	ml
Glacial acetic acid	3	ml

8. Phenol : Chloroform (1:1 v/v)

Crystalline phenol was liquidified in water bath at 65° C and mixed with chloroform in the ratio of 1:1 (v/v). The solution was stored in a light tight bottle.

9. 0.5M EDTA (pH 8.0)

800 ml of distilled water, 186.1 g of disodium ethylenediaminetetraacetate.2H₂O was added and stirred vigorously on a magnetic stirrer. The pH was adjusted to 8.0 with NaOH (20 g of NaOH pellets). The volume was adjusted to 1 litre. The solution was dispensed into aliquots and sterilized by autoclaving for 15 minutes at 15 lb/in².

10. 2xPBS

8 mM Na₂HPO₄

1.5 mM KH₂PO₄

137 mM NaCl

2.7 mM KCl

The 2xPBS was adjusted the pH to 7.0 with 1N NaOH or 1N HCL. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in².

11. 10 mg/ml Salmon sperm DNA

A 10 mg of Salmon sperm DNA was dissolved in 1 ml of 10 mM TE buffer pH 7.6. Boiling for 10 minutes, immediately cooling in ice and sonication for 3 minutes.

12. 3 M Sodium acetate pH 5.2

To 800 ml of distilled water, 408.1 g of sodium acetate was added and adjusted the pH to 5.2 with glacial acetic acid. The volume was adjusted to 1 litre. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in².

13. 10% Sodium dodecyl sulphate (SDS)

The stock solution of 10% SDS was prepared by dissolved 10 g of sodium dodecyl sulphate in 100 ml sterilized distilled water. Sterilization is not required for the preparation of this stock solution.

14. 20xSSC

3 M NaCl

0.1 M Tri-sodiumcitrate

The 20xSSC was adjusted the pH to 7.0 with 1N NaOH. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in².

15. 1 M Tris-HCl pH 8.0

The 1M Tris was prepared by dissolving 121.1 g of Tris base in 800 ml of distilled water. The pH was adjusted to the desired value by adding conc. HCL (pH 8.0, 42 ml of HCl). The solution was cooled to room temperature before making final adjustment to the desired pH. The volume of the solution was adjusted to 1 litter with distilled water and sterilized by autoclaving.

16. RNase A solution

RNase A	20 mg
---------	-------

0.15 M NaCl	10 ml
-------------	-------

Dissolve 20 mg of RNase A in 10 ml 0.15 M NaCl and heat at 95° C for 5-10 minutes. Keep RNase A solution in -20°C.

17. RNase T₁ solution

RNase T ₁	80	μl
----------------------	----	----

0.1 M Tris-HCl (pH 7.5)	10	ml
-------------------------	----	----

Mix 80 μl of RNase T₁ in 10 ml of 0.1 M Tris-HCl (pH 7.5) and heat at 95°C for 5 minutes. Keep RNase T₁ solution in -20°C.

18. Proteinase K

Proteinase K (Sigma)	4	mg
----------------------	---	----

50 mM Tris-HCl (pH 7.5)	1	ml
-------------------------	---	----

Use freshly prepared solution.

19. Nuclease P₁ solution

Nuclease P1	0.1	mg
-------------	-----	----

40 mM CH ₃ COONa+12 mM ZnSO ₄ (pH5.3)	1	ml
---	---	----

Store at 4°C.

20. Alkaline phosphatase solution

Alkaline phosphatase	2.4	units
----------------------	-----	-------

0.1 M Tris-HCl (pH 8.1)	1	ml
-------------------------	---	----

21. 0.1 M Tris-HCl buffer, pH 9

Tris	1.21 mg
Distilled water	100 ml
Adjust the pH to 9 with HCl.	

22. TE buffer

10 mM Tris HCl (pH 8.0)

1 mM Na₂-EDTA (pH 8.0)**23. TE buffer + RNase A**

TE buffer	960 ml
RNase A (2 mg/ml)	100 µl

24. Saline-Na₂ EDTA

0.1 M NaCl

50 mM EDTA.2Na (pH 8.0)

25. Reagents and buffers for DNA-DNA hybridization**25.1 Prehybridization solution**

100xDenhardt solution	5 ml
10 mg/ml Salmon sperm DNA	1 ml
20xSSC	10 ml

Formamide	50
ml	
Distilled water	34 ml

25.2 Hybridization solution

Prehybridization solution	100 ml
Dextran sulfate	5 g

25.3 Solution I

Bovine serum albumin (Fraction V)	0.25 g
Triton X-100	50 μ l
PBS	50 ml

25.4 Solution II

Streptavidin-POD	1 μ l
Solution I	4 ml

25.5 Solution III

3,3',5,5'-Tetramethylbenzidine (TMB)	100 μ l
(10 mg/ml in DMFO)	
0.3% H ₂ O ₂	100 μ l
0.4 M Citric acid + 0.2 M Na ₂ HPO ₄ buffer	100 μ l

pH 6.2 in 10% DMFO

25.6 2 M H₂SO₄

H₂SO₄ 22 ml

Distilled water 178 ml

The solution was sterilized by autoclaving.

26. Fehling's solution

Coppersulfate 34.64 g

Sodiumpotassiumtartate 173 g

Sodiumhydroxide 50 g

Solvent was composed of a mixture 500 ml of coppersulfate and 500 ml of mixture sodiumtatare and sodiumhydroxide.

APPENDIX D

Physiological and biochemical characteristics of isolates

Primers, 16S rDNA nucleotide sequences and DNA G+C contents

Physiological and biochemical characteristics of isolates

Isolate no.	Growth In (%NaCl)		Growth at pH					Growth at °C									Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis
	3	5	5	6	8	9	10	15	20	45	50	55	60																		
P5-5	-	-	+	+	+	-	-	-	+	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-				
P6-5	-	-	+	+	+	-	-	-	+	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-				
P1-4	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	-	+	-	+	+	+	+	+	-	-	-				
P1-9	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	+	-	+	+	+	+	+	-	+	+	-				
P2-1	+	+	+	+	+	-	-	-	+	+	+	+	-	+	+	+	-	+	+	-	+	+	-	+	+	-	-				
P3-2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	-	-	+	+	+	+	+	-				
P5-7	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	-	-	-	+	-	+	-	-	-				
P5-8	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	-	-	+	+	-	+	+	-	-				
P6-6	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	+	-	+	+	-	+	+	-	-				
S8-1	+	+	+	+	+	+	-	-	+	+	-	-	-	+	+	-	-	+	-	-	-	+	+	-	-	-	-				
S10-2	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	-	-	-	-	+	-	-	-	-	-				

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth In (%NaCl)		Growth at pH					Growth at °C										Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis	
	3	5	5	6	8	9	10	15	20	45	50	55	60																				
P1-2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P1-3	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P2-2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P6-2	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P6-3	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P7-4	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P7-5	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P7-6	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
P7-7	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-			
S8-4	+	+	+	+	+	+	-	-	+	+	-	-	-	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-			
S10-4	+	+	+	+	+	+	-	-	+	+	-	-	-	+	+	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-			

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth In (%NaCl)		Growth at pH					Growth at °C										Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Aesculin	L-arginine	Hydrolysis			
	3	5	5	6	8	9	10	15	20	45	50	55	60																					
P1-7	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	+	+	+	+	-	+	+	-	-	-	-	-				
P1-11	+	+	+	+	+	+	+	-	-	+	+	+	-	+	+	-	-	+	+	+	-	+	+	-	+	+	-	-	-	-				
P2-3	+	+	+	+	+	+	+	-	-	+	+	+	-	+	+	-	-	-	+	+	-	-	+	+	-	+	-	-	-	-				
P4-6	+	+	+	+	+	+	+	-	-	+	+	+	-	+	+	-	-	+	+	+	-	+	+	-	+	-	-	-	-	-				
P4-7	+	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	-	-	+	+	-	-	-	-	-	-				
P4-11	+	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	-	-	+	+	-	-	-	-	-	-				
P5-3	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	+	+	-	+	+	-	+	-	-	-	-	-				
P5-6	+	+	+	+	+	+	+	-	-	+	+	+	-	+	+	-	-	+	+	-	+	+	-	+	-	-	-	-	-	-				
P6-4	+	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	+	+	-	-	+	+	-	-	-	-	-	-				
P6-9	+	+	+	+	+	+	-	-	+	+	+	-	-	+	+	-	-	-	+	+	-	-	+	+	-	-	-	-	-	-				
P6-10	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	+	+	-	-	+	+	-	-	-	-	-	-				

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth In (%NaCl)		Growth at pH					Growth at °C									Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis		
	3	5	5	6	8	9	10	15	20	45	50	55	60																				
P7-2	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	+																	
S10-3	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+																	
P1-1	+	-	+	+	+	+	-	+	+	+	+	-	-	+	+	+	+																
P4-1	+	-	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P4-3	+	-	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P4-4	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P4-5	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P4-9	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P4-10	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
P5-1	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																
S8-3	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	+																

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth in (%NaCl)		Growth at pH					Growth at °C									Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis
	3	5	5	6	8	9	10	15	20	45	50	55	60																		
S9-2	-	-	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-				
S10-1	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-				
S11-1	+	+	+	+	+	+	-	+	+	+	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-				
P1-6	-	-	+	+	+	+	-	+	+	+	+	+	-	+	+	W	-	-	-	-	+	+	-	-	-	-	-	-			
P1-8	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	-	-	-	-	+	+	-	+	-	-	-				
P1-10	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	-	-	-	-	+	+	-	+	-	-	-				
P1-12	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	-	+	+	-	+	-	-	-				
P2-4	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	W	-	+	-	+	+	-	+	-	-				
P2-5	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	W	-	+	-	+	+	-	+	-	-				
P3-1	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	-	-	-	-	+	+	+	-	+	-	-				
P3-4	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	-	+	+	+	-	+	-	-				

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth In (%NaCl)		Growth at pH		Growth at °C									Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis		
	3	5	5	6	8	9	10	15	20	45	50	55	60																	
P3-S	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	+	+	-	+	+	-	+	+	-	+		
P4-2	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	+	-	-	+	W	W	-	+	+	+	
P4-12	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	+	+	-	+	+	+	-	-	
P4-13	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	-	+	-	+	+	-	+	+	+	-	-	
PS-4	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	+	-	+	+	+	-	+	+	+	-	-	
PS-9	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	-	+	+	-	+	+	-	+	+	-	+	-	-	
PS-10	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	+	-	+	+	+	+	-	-	
PS-11	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	+	-	+	+	-	+	-	-	
PS-12	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	+	-	+	+	-	+	+	+	-	-	
PS-13	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	+	+	-	+	+	+	-	-	
PS-14	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	-	+	-	+	-	+	+	+	-	-	-	

Physiological and biochemical characteristics of isolates (Cont)

Isolate no.	Growth In (%NaCl)		Growth at pH				Growth at °C										Catalase test	Oxidase test	Anaerobic growth	Methyl red	Voges-Proskauer	DNAase	Urease	Indole production	Citrate	TSI	Nitrate reduction	Asculin	L-arginine	Dihydroxyacetone	Hydrolysis
	3	5	5	6	8	9	10	15	20	45	50	55	60																		
P6-1	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-			
P6-8	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-			
S8-2	+	+	+	+	+	+	-	-	+	+	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-			
P3-3	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	-	-	-	-	+	+	+	+	+	+	+	+			
P4-8	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	-	+	+	+	-	+	+	+	+	+	-	-			
P5-2	+	+	+	+	+	+	-	+	+	+	+	-	-	+	+	-	-	+	+	+	-	+	+	+	+	+	-	-			
P7-1	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	-	-	+	+	-	-	+	+	+	-	+	+	-			
P7-3	+	+	+	+	+	+	-	-	+	+	+	+	-	+	+	-	-	+	+	-	+	+	+	-	+	+	-	+			
P1-S	+	+	+	+	+	-	-	+	+	+	+	-	-	+	+	-	-	+	+	-	-	+	+	-	+	+	-	-			
P6-7	+	+	+	+	+	-	+	+	+	+	+	-	-	+	+	-	-	+	-	-	-	+	+	-	+	+	-	-			

Acid from carbohydrates

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellubiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melcitoose	D-Methyl-D-glucoside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbital	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
P5-5	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-5	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-4	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-9	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P2-1	+	+	+	+	+	+	-	-	W	W	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P3-2	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P5-7	-	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P5-8	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-6	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S8-1	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S10-2	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellubiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	α -Methyl - D - glucoside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trichalose	D-Xylose
P1-2	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-3	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P2-2	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-2	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-3	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-4	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-5	-	-	+	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-6	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-7	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S8-4	-	+	+	-	+	+	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S10-4	W	+	+	+	+	+	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellulbiose	D-Fuctose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	α -Methyl - D - glucoside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
P1-7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P2-3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellobiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycitol	Inositol	Inulin	Lactose	D-Maltose	W	D-Mannitol	D-Mannose	D-Melibiose	D-Micetitose	Q - Methyl - D - glucide	Raffinose	L-Rhamnose	D-ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trichalose	D-Xylose
P7-2	-	-	+	+	-	+	-	-	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
S10-3	-	-	+	+	-	+	-	-	-	-	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
P1-1	+	-	+	+	-	+	-	+	-	-	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
P4-1	+	-	-	+	-	+	-	+	-	-	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
P4-3	+	-	+	+	-	+	+	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-4	+	-	+	+	-	+	+	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-5	+	-	+	+	-	+	-	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-9	+	-	+	+	-	+	+	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-10	+	-	+	+	-	+	+	+	-	W	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-1	+	-	+	+	-	+	-	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
S8-3	+	-	+	+	-	+	-	+	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellobiose	D-Fructose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melezitose	α -Methyl-D-glucide	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
S9-2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S10-1	-	-	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S11-1	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-6	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-8	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-10	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-12	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P2-4	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P2-5	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P3-1	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P3-4	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates(Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellubiose	D-Fuctose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Mezitose	D-Methyl-D-gluside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
P3-5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-2	-	-	-	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-12	-	-	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-13	-	-	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-4	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-9	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-10	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-11	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-12	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-13	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-14	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Acid from carbohydrates (Cont)

Isolate no.	D-Amygdalin	L-Arabinose	D-Cellubiose	D-Fuctose	D-Galactose	D-Glucose	Gluconate	Glycerol	Inositol	Inulin	Lactose	D-Maltose	D-Mannitol	D-Mannose	D-Melibiose	D-Melcitoose	D-Methyl-D-gluside	Raffinose	L-Rhamnose	D-Ribose	Salicin	D-Sorbitol	L-Sorbose	Sucrose	D-Trehalose	D-Xylose
P6-1	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-8	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S8-2	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P3-3	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P4-8	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
PS-2	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-1	-	-	-	+	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P7-3	-	-	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P1-5	-	-	-	+	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
P6-7	-	-	-	+	-	+	-	W	W	-	-	+	-	-	-	-	-	-	-	-	-	-	+	W	-	-
KCTC 3135 ^T	-	-	-	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	+	-	+	-
KCTC 3998 ^T	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+

1. Primers for 16S rDNA amplification and sequencing

9F	5'-GAGTTGATCCTGGCTCAGGACGAACGCTGGCGCGTGCCTA
1541R	5'-AAGGAGGTGATCCAGCC-3'
357R	5'-CTGCTGCCTCCCGTAG-3'
802R	5'-TACCAGGGTATCTAATCCC-3'
530F	5'-GTGCCAGCAGCCGG-3'

2. 16S rDNA nucleotide sequences

2.1 The 16S rDNA nucleotide sequence of strain P1-5

TTTGAGTTTGATCCTGGCTCAGGACGAACGCTGGCGCGTGCCTA@TACATGCAAGTCGAGCGGACAGATGGGAGCTTGTCTCCCTGATGTAAGCGG
 CGGACGGGTGAGTAACACGGTAACTCGCTGTAAGACTGGGATAACTCCGGAAACCGGGGCTAATACCGGATGGTTGAACCGCATGGT
 CAGACATAAAAGGTGGCTCGGCTACCACTTACAGATGGACCCGCGCGCATTAGCTAGTTGGTAGGTAACGGCTACCAAGGCAACGATGCGTAG
 CCGACCTGAGAGGGTGTACGGCACACTGGACTGAAGACACGGCCAGACTCCTACGGGAGGCAGCAGTAGGAAATCTCCGCAATGGACGAA
 AGTCTGACGGAGCAACCCCGTGAGTGTAGAAGGTTTCCGATCGTAAAGCTCTGGTTAGGAAAGAACAGTGCCTCAAATAGGGCGGACC
 TTGACGGTACCTAACAGAAAGCCACGGCTAACTACGTGCCAGCAGCGCGTAATACGTAGGTGGCAAGCGTTCCCGAATTATTGGCGTAAAG
 GGCTCGCAGCGGTTCTTAAGTCTGTAGTGAAGGCCCCGGCTAACCGGGGAGGGTCAATTGAAACTGGGAACTTGAGTGCAGAAGAGGAG
 TGGAAATCCACGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCGTAGGCGAAGGCAGCTCTGGTCTGTAACTGACGCTGAGGAGCGA
 AGCGTGGGGAGCGAACAGGATTAGATACCCCTGGTAGTCCACGCCGTAACCGATGAGTGCTAAGTGTAGGGGTTCCGCCCTTAGTGCAGCT
 AACGCATTAAGCACTCCGCTGGGAGTACGGTCGCAAGACTCAAAGGAATTGACGGGGCCCGACAAGCGGTGGAGCATGTGGTTAA
 TTCGAAGCAACCGGAAGAACCTTACCGGGTCTGACATCCTCTGACAATCCTAGAGATAGGACGTCCCTCGGGGAGGTGACAGTGGTGCAT
 GGGTGTGTCAGCTCGTGTGAGATGTGGGTTAAGTCCCGAACCGAGCGAACCCCTGATCTTAGTGCAGCTACACCGTCTACAATGGACAGAAC
 GACTGCCGGTACAAACCGGAGGAAGGTGGGATGACGTCAAATCATGCCCTTATGACCTGGCTACACCGTCTACAATGGACAGAAC
 AGGGCAGCGAACCGCGAGGTTAAGCCAATCCCACAAATCTGTTCTCAGTCGGATCGCAGTCTGCAACTCGACTGCGTGAAGCTGGAATCGCTAGT
 AATCGCGGATCAGCATGCCCGGTGAATCGTCCCGGCTTGTACACACCGCCCGTACACCACGAGAGTTGTAACACCCGAAGTCGGTGGAGGT
 AACCTTATGGAGCCAGCCCGAAGGTGGACAGATGATTGGGAAGTCGACCGA

2.2 The 16S rDNA nucleotide sequence of strain P2-1

GAAAAAAAACGTTGAAGGGATTTATTAGCTTACCGACCCTGGCGGGCCTAACCTACAATTCTCGAATTGAGGAGAACCTTGCTCTCT
TAATGTTAGCGGGAACCGTTGATTTAACATGTAGAAAACCTCTCAAGACGGATAACCCAGAAATTGAGCTAACCGGGATATCTCATTTCCCT
CTCCCCCGGGAAATAAAAGACGGAGCATTTGTCACTTGCCTGGATGGGCTCGGCCATTAGCTAGTTGGTAGGGTAACGCTACCAAGGCACAAATGG
GTAGCCGACCTGAAGAGGGTAACCGGCCACACTGGACTGAGACACGCCAGAC TCC1 .CGGGAGGCACCAAGTGGAACTTCCGCAATGGG
CGAAAGCCTGACGGAGCACGCCCGTGAAGTTGAGTGTAAAGCTGTGCCCAGGGAAAGAACGCTCCGGTAGAGTAACGTG
ACCGGAGTGACGGTACCTGAGAAGAAAGCCCCGGCTAACACGTGCCAGCAGCCCGGTATACGTAGGGGCAAGCGTTGCTCGGAATTATTG
GGCGTAAAGCGCGCAGGGGTCTTAAGTCTGGTTAAGGCCAAGGCTAACCTGGTCTGCACTGGAAACTGGGTGACTTGAGTGCAGAAG
AGGAGAGTGAATTCCACGTGAGCGGTAAAGCTGAGATATGTGGAGGAACCCAGTGGCGAAGGCAGTCTCGGCTGTAACGTGAGCTGAG
GCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCCTGGTAGTCCACGCCGTAACGATGAATGCTAGGTGTTAGGGGTTGACCGTGGC
CGAAGTAAACACATTAAGCATTCCGCTGGGAGTACGGTCGAAGACTGAAACTCAAAGGAATTGACGGGACCCGACAAGCAGTGGAGTATGT
GGTTAATTGCAAGCAACCGGAAGAACCTTACCAAGGCTTGACATCCCTGACCGGTATAGAGATAACCTTCCGGAACAGGGAGACAGGT
GGTGCATGGTTGCAAGCTGTGAGATGTTGGTTAAGTCCGCAACGAGCGAACCTTGATTTAGTGCAGCAGTCCGGTGGC
TCTAGAATGACTGCCGTGACAACCGGAGGAAGGCAGGGATGACGTCAAATCATCATGCCCTTATGACCTGGCTACACAGTACTACAATGGC
AGTACAACCGGAAGCGAAGCCGAGGTGGAGCCAATCTATCAAAGCTGGTCAGTCAGGCTGCACTCGCCTGCATGAAGTCGAA
TTGCTAGTAATCGGGATCAGCATGCCGGTGAATACGTTCCGGGTTGTACACACCAGCGTACACCACGAGAGTTAACACACCGAAGTCG
GTGAGGTAACCGCAAGGAGCCAGCCGCCAGGTGGTAGATGTTGGAAAAGTCG

2.3 The 16S rDNA nucleotide sequence of strain P2-3

CTGGCGCGTGCCTACATGCAAGTCGAGCGGACAGATGGGAGCTTGCCTGATGTTAGCGGGACGGGTGAGTAACACGTGGTAACCTGCC
TGTAAGACTGGATAACTCCGGAAACCGGGCTAACCGGATGGTTGTTAACCGCATGGTCAGACATAAAAGGTGGCT
TCGGCTACCACTACAGATGGACCCCGCGCATTAGCTAGTTGGTAGGGTAACGGCTACCAAGGCACGATGCGTAGCCGACCTGAGAGGGTGATC
GGCCACACTGGGACTGAGACACGCCAGACTCCTACGGGAGGCAGCAGTAGGGAACTCCGCAATGGACGAAAGTCTGAC
GGAGCAACGCCGTGAGTGAAGGTTTGGATCGTAAAGCTCTGGTAGGGAAAGAACAGTGGCTCAATAGGGGGCACCTGACGGT
ACCTAACCAAGAAAGCACGGCTAACACGGTCAAGCTGCCAGCACCGCGGTATACGTAGGTGGCAAGCGTTGCCAAATTGGC
GTAAAGGGCTCGCAGCGGTTCTTAAGTCTGATGTGAAAGCCCCGGCTAACCGGGAGGGTCAAGGGAAACTGGGAACTTGAGTGCAGAAGA
GGAGAGTGGAAATTCCACGTGAGCGGTAAATGCGTAAGATGTGGAGGAACACCAGTGGCGAAGGCAGTCTGGTCTGAA
CTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCCTGGTAGTCCACGCCGTAACGATGAGTGTAAAGTGTAGGGGTTCC
GCCCTTATGTCAGCTAACGCTAACGACTCCGCTGGGAGTACGGTCGCAAGACTGAAACCTAACAGGAATTGACGGGG
GCCCGACAACGGGGACATGTGGTTAATTCAACACCCAAAACCTTACAGGTCTGACTCCTGACATCTAAAGGAATTGACGGGG
CAAATGACGGGGTGCATGTTCCCTCTGTCGAATGTTGGTAATCCCA

2.4 The 16S rDNA nucleotide sequence of strain P4-7

GATTGAGTTGACCCGGCTAGGACGAACGCTGGCGCGTGCCTAAATACATGCAAGTCGAGCGGACAGATGGAAGCTGCTCCCTGATGTTAGC
 GGC GGACGGGTGAGTAACACGTGGTAACCTGCCTGTAAGACTGGATAACTCCGGAAACCGGGGCTAATACCGGATGGTTGTTGAACCGCATGG
 TTCAGACATAAAAGGTGGCTCGGCTACCACTAACAGATGGACCCGCGCATTAGCTAGTGGTAGGTAACGGCTACCAAGGAACAGATGCGT
 AGCCGACCTGAGAGGGTGTACGCCAACACTGGGACTGAGACACGCCAGACTCTCACGGGAGGCAGCAGTAGGGATTCTCCGAAATGGACG
 AAAGTCTGACGGAGCAACCCCGGTGAGTGTAGGTTGGATTCTGAAAAGCTCTGGTAGGAGAAGAACAGTGCCTCAAATAGGGCG
 CACCTTGACGGTACCTAACAGAAAGCCACGGCTAACAGTGGCCAGCGCAGGTAACCGTAGGTGGCAAGGGTGTCCGAAATTGGCGT
 AAAGGGCTCGCAGGGGTTCTTAAGTGTAGGAAAGCCCCGGCTCAACCGGGAGGGTCACTGGAAACTGGGAACTTGAGTGCAGAAGAGG
 AGAGTGGAAATCCACATTGTAGCGTAATAATGACGTAGAGATGTGGAGGAACACCAGTGGCAAGGCAGTCTGGTGTAACTGACGCTGAGG
 AAGCGAAAAGCGTGGGAGCGAACAGGATTAGATACCCCTGGTAGTCCACGCCAACAGATGAGTGCTAAGTGTAGGGGTTCCCCCTTAGT
 GCTCAGCTAACGCTAACGACTCCCGCTGGGAGTACGGTGCAGACTGAAACTCAAAGGAATTGACGGGGCCGACAAGCGGTGGAGCA
 TGTGGTTAATCGAAGCAACGCGAACAAACCTTACCGGTCTGACATCCTCTGACAATCTAGAGATAGGACGTCCTCGGGGAGGTGACA
 GGTGGTGCATGGTGGCAACAGCTCGTGTACGTGAGATGTGGTTAAGTCCCACGAGCGCAACCCCTGATCTAGTGGCCAGCATTGAGT
 GGCACACTAACGGTACTGCCGTGACAACCGGAGGAAGGTGGGAGTACGTCAAATCATGCCCCATTGACCTGGCTACACACGTGCTAAC
 TGGACAGAACAAAGGGCAGCGAACCGCGAGGTAAAGCAATCCCACAAATCTGTTCTAGTTCGATCGCAGTCTGCAACTCGACTGCGTGAAG
 GGAATCGCTAGTAATCGCGATCAGCATGCCCGGTGAATACGTTCCGGGCTTGTACACAACCGCCGTACACCACGAGAGTTGTAACACCG
 AAGTCGGTAGGTAACCTTATGGAGCCAGCCGAAAGTGGGAGATCCTGGAAAAGTCCTCATG

2.5 The 16S rDNA nucleotide sequence of strain P4-8

GCAAAGAAAATTATTCCTAGTAAATTAAAAATTGCCCCCCCCGGGGAAAGGTTTTTAACCTTCAGACAAAACCGGGGGCG
 TGCCTAAATACCTCAAGTCGAGCGGACAGAGGGAGCTTGCCTCCTGATTTAGCGGCGGACGGGTGAGTAACAGTGGTAACCTCC
 GTAAGACTGGGATAACTCCGGGAAACCCGGGGCTAACCGAATGTTGTTGAACCGCATGGTTAGCATAAAAGGGGTTGGTACCA
 TTACAGATGGAGCCCGCGGCCATTAACTTAGTTGGTAGGTACGGCTACCCAAGGAACGATGGGTAGCCGACCCCTGAGAGGGTGA
 TCGCCCAACTGGGACTGAAGACACCCGCCCCAGACTCCTACGGAGGCAGCAGTAGGGAACTTCCGCAATGGACGAAAGTCTGACGG
 AGCAACCCCCCGGTGGAGTGTAGGTTTCGGATCGTAAAGCTCTGTTAGGAAAGAACAGTGGCCTCAATAGGGCGGACC
 TTGACGGTACCTAACAGAAAGCCACGGTAACACTACGTGCCAGCAGCCCGGTAAACGTAGGTGGCAAGCGTTGTCCGAATTGGC
 GTAAAGGGCTCGAGCGTGTCTTAAGTGTAGTGAAGGCCCCGGCTAACCGGGAGGGTCACTGGAAACTGGGAACTTAAAGTGC
 GAAGAGGAGAGTGGAAATCCACGTGTAGCGGTGAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCAGTCTGGTGTAACT
 GACGCTGAGGAGCGAACCGTGGGAGCGAACCGAGGATTAGATACCCCTGGTAGTCCCACGCCAACGATGAGTGTAACTGTTAGGGG
 TTTCCGCCCCTTAGTGTGCTGAGCTAACGCATTAAAGCACTCCGCTGGGAGTACGGTCGAAGACTGAAACTCAAAGGAATTGACGGGG
 CCCACAAAGGGTGGAGCATGTGGTTAACCGAAGCAACCGGAAGAACCTTACCGGTCTGACATCCTCTGACAATCCCTAGAGATAGG
 ACGTCCCCCTCCGGGGCAAGTGTACAGGTGGCATGGTGTAACTAGCTCGATGTCGTAAAGATGTGGTTAACGTCCGCAACGAGC
 GCAACCCCTGATTAGTGTGCTGAGCTAACACGTCTAACATGGACAGAACAAAGGGCAGCGAACCGGAGGAAGGTGGGATGACGTCAAATC
 ATCATGCCCTTATGACCTGGCTACACACGTCTAACATGGACAGAACAAAGGGCAGCGAACCGGAGGGTAAGCCAATCCCACAAATCT
 GTTCTCAGTCGGATCGCAGTGTGCAACTCGACTGCCGTGAAGCTGGAATCGCTAGTAATCGGGATCAGCATGCCGGTGAATCGTCCC
 GGGCATGTACACCCGCCGTACACCAACGAGAGTTGTAACACCGAAGTCGGTAGGTAACCTTATGGAGCCAGCCCCGAAGGTGG
 CACAGAGAGGATGATTGAGGAAGGAAAGCCTCCATGG

2.6 The 16S rDNA nucleotide sequence of strain P5-2

TTAGTTTGATCCTGGCTCAGGACGAACGCTGGCGCGTGCCTAACGATACATGCAACTCAAGCGGACAGATGGGAGCTGCTCCCTGATGTTAGCGG
 CGGACGGGTGAGTAACACGTGGTAACCTGCCGTAAAGACTGGATACTCCGGAAACCGGGGCTAACACCGGATGGTTGTTGAACCGCATGGTT
 CAAACATAAAAGGTGGCTCGGUTACLAACCTACAGATGGACCCCGGGCGCATTAGCTAGTTGGTAGGGTAACGGCTCACCAAGGCAACGATGCGTAG
 CCAACCTGAGAGGGTGTAGGGCACACTGGGAATGAGACACGGCCAAGACTCCTACGGGGAGGCCAGCAGTTAGGGATCTTCCCCAATGGACG
 AAAAGTCTGACGGAGCAACCCCGGTGAGTGTAGGAAGGTTTCGGATCGTAAAGCTGTGTTAGGGAAAGAACAGTACCGTCAATAGGGC
 GTACCTGACGGTACCTAACAGAAAGCCACGGCTAACACGTGCCAGCAGCGCGTAATACGTAAGTGGCAACGGTGTCCGGAATTATTGGCG
 TAAAGGGCTCGCAGCGGTTCTAACGTTAGCTGTGAAAGCCCCGGCTAACCGGGGAGGGTCATTGGAAACTGGGGACTTGAGTGCAGAAAAA
 GAAAGTGGAAATTCAACGTGTAGCGGTAAATCGTAGAGATGTGGAGGAACACCGTGGCGAACGGCAACTCTGGTCTGTAACGTACGCTGAGGA
 GCGAAAGCGTGGGGAGCGAACAGGATTAGATAACCCCTGGTAGTCCCACCCGTAACCGGGGAGGGTCATTGGAAACTGGGGACTTGAGTGCAG
 TGCACTAACGCTAACCAACTCCCCCTGGGGAGTACGGTGCAGACTGAAACTCAAAGGAATTGACGGGGCCCGAACAGCGTGGAGCAT
 GTGGTTAATCGAAGCAACCGAACAGTCTTACAGGTCTGACATCTCTGACAATCCTAGAGATAGGACGTCCCCCTGGGGCAGAGTGACAG
 GTGGTGCATGGTGTGTAAGCTCGTGTGTAAGCTCGTGTGAAAATGTTGGTTAAGTCCCACCGCAACGAGCGAACCCCTGATCTTAGTGCAGCATTGAGTGG
 CACTCTAACGGTACTGCCGGTACAAACCGAGGAAGGGGGATGACGTCAAATCATGCCCCCTATGACCTGGCTACACAGTGCTAACATG
 GACAGAACAAAGGGCAGCGAACCGCGAGGTTAACGCAATCCCACAAATCTGTTCTAGTTGGATCGCAGTGTGCAACTCGACTGCGTGAAGCTGG
 AATCGTAGTAATCGGGATCAGCATGCCCGGTAAATCGTCCCCGGCTTGACACACCAGGGTACACACCGAGAGTTGTAACACCCGAAG
 TCGGTGAGGTAACCTTTAGGAGGCCAGCCGCCAACGGGGACAGATGATTGGGTGAAGTCGTCGAACTTC

2.7 The 16S rDNA nucleotide sequence of strain P5-5

AACTGGCGCGTGCCTATACATGCAAGTCGAGCGAGGTCCCTCGGGGGCTAGCGTCGGACGGGTGGATAACACGGTAGGGCAACCTGGCCTCTC
 AGGACCGGGATAACTAGGGAAACTTATGCTAACACCGGATAGGTTTGGATCCATGATCCGAAAAGAAAAGATGGCTCGGCTATCACTGGGAGA
 TGGGCCTGCGCGCATTAGCTAGTTGGGGTAACGGCTACCAAGGGACATGCGTAGCCACCTGAAGGGTACCGGGCACACTGGGACTGAAAC
 CGGCCCAACTCCTACGGGAGGCAGCAGTAGGGATTTCCACAATGGACGAAAGTCTGATGGACAAACCCCGTAACGATGAAGGTCTCGGATTGTA
 AAGTCTGTTGCAAGGACAAAGTACCGTTCGAACAGGGGACCTTGACCGTACCTGACGAGAAAGGCCACGGCTAACTACTGCCAGCAGCGC
 GGTAAATAAAAGGGCTGGCCCTCAAGACAAACGTTGTCGGAATTATTGGCGTAAAGCGCGCGCAGGGGCTATGTAAGTCTGGTTAAAGCCG
 GGGCTCAACCCCGGTTCGCATCGAACACTGAGGCTTGAGTGCAAAAAGGAAAGCGGTATTCCACGTTGAGCGGTAAATCGTAAAGATGTG
 GAGGAACCCAGTGGCAAGGGCGCTTCTGGTCTGTAACGCGTACGGCGAACAGTGGGGAGCAACAGGATTAGATAACCTGGTAGTCC
 ACGCCGTAAACGAAGAGTGTAGGTGTTGGGTTCAATACCCCTAGTGGCGACGCTAACGCAATAAGCACTCCCCCTGGGGAGTACGTTCCAA
 GAGTGAACACTCAAAGGAATTGACGGGGCCCCACAGCGGTGGAGCATGTTAATCGAAGCAACCGCAAGAACCTTACAGGTCTGACATCC
 CGCTGACCGCCCTAAAGATAAGGCTTCCCTCGGGCTAGCGGTGACAGGTGGTCAAGTGGTTAAGTCACGCTCGTGTGAGATGTTGGGTTAAG
 TCCCGCAACGAGCGAACCCCTATTCTAGTGTGCAACACCGTGTACAATGGTTGCAACAGGGATGCTACCTCGCGAGAGGACCCAATCTGAA
 CGTCAAATCATGCCCCATTGACCTGGGTACACACGTGTACAATGGTTGCAACAGGGATGCTACCTCGCGAGAGGACCCAATCTGAA
 AACCAATCTAGTCGGATTGAGGCTGCAACTCGCCTACATGAAAGTCGAATCGCTAGTAATCGGGATCAGCATGCCCGGTAAATCGTCCCG
 GGCCTGTACACACCAGCCCGTACAACACCAGGGAGTTGCAACACCCGAAGTCGGTAGGTAACCCGAAGGGCCAGCCGCCGAAGGTGGG
 AAATGACTGGGTGAAGTCGTAACAAGGTAAACCGTAA

2.8 The 16S rDNA nucleotide sequence of strain P6-7

TGGAAAAGGGAGCCGGGGATTATTTGAGTATCGTCCTGGCTCAAGGACGAACGCCGGCGGCCTAATACATGCAAGTCAGCCGACAG
 ATGGGAGCTGCTCCCTGATGTTAGCGCGGACGGGTAGTAACACGTGGTAACCTGCCTGTAAGACTGGGATAACTCCGGAAACCGGGGCTAA
 TACCGGATGGTTGTTGAACCGCATGGTCAACATAAAAGGTGGCTCGGCTACCCACTACAGATGGACCCGCGCGCATTAGCTAGTGGTAG
 GTAACGGCTACCAAGGCAACGATGCGTAGCCGACCTGAGAGGTGATCGGCACACTGGGACTGAAGACACCGCCCCAGACTCCTACGGGAGGCAG
 CAGTAGGGAATCTCCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCGTAGTGATGAAGGTTTCGGATCGAAAGCTCTGTTAGGGAAAGA
 ACAAGTACCGTTCGAATAGGGCGGTACCTTGACGGTACCTAACAGAAAAGCCACGCCGTAACACTGCGCAGCAGCGCGTAATACGTAGTGGCAA
 GCGTTGTCGGAATTATGGGCTAAAGGGCTCGCAGCGGTTCTTAAGTCTGATGTGAAAGCCCCGGCTAACCGGGGAGGGTATTGGAAACT
 GGGGAACCTGAGTGAGAAGAGGAGAGTGGATTCCACGTGAGCGTAAATGCGTAGAGATGTGGAGGAACACAGTGGCGAAGGCAGCTCT
 GGTCTGTAACTGACGCTGAGGAGCGAAAGCGTGGGAGCGAACAGGATTAGATACCCGTGAGTCCACGCGTAAACGATGAGTGCTAAGTGGTAG
 GGGGTTCCCGCCCTAGTGTGAGCTAACGCACTAACGACTCCGCTGGGAGTGCGTAGGCTGAAAGACTCAAAGGAATTGACGGGGGCC
 GCACAAGCGGTGGAGCATGGTTAATTGAAGCAACCGCAAGAACCTTACCGTGTGACATCCTGACAATCTAGAGATAGGACGTCCC
 TCGGGGGCAGGTGACAGGGTGCATGGTTGCGTAGCTCGTGTGAGATGTTGGGTTAAGCCGCAACGAGCGAACCCCTGATCTTAGTT
 CCAGCATTCACTGGGACTCTAACGGTACTGCCGTGACAAACCGGAGGAAGGTGGGATGACGTCATCATGCCCCTATGACCTGGGCTA
 CACACGTCTACAATGGACAGAACAAAGGGCAGCAGAACCCCGAGGTTAAGCCAATCCCACAAATCTGTTCTCAGTTGGATCGCAGTCTGCAACT
 GACTGCGTGAAGCTGGAATCGTAGTAATCGGGATCAGCATGCCCGGTGAATACGTTCCGGGCTTGACACACCGCCGTCACACCACGAGAG
 TTGTAACACCGAAGTCGGTAGGTAACCTTTAGGAGCCAGCCGCGAAGGTGGGACAGATGATTGGAGAAGTCGTACGGA

2.9 The 16S rDNA nucleotide sequence of strain P6-8

GAGACTGTTGAAATTAAACGAAATGAAAAAGGTATAAAAAGATATTATTTATCCTGGCTCAGACGAACCCGGCGGGGTTCTTAATAC
 CTGCCAATCGAGCGGACAGATGGGAGCTCCTCCCTGATTAAGCGCGGACGGGTAGTAACACGTGGTACCTCTGTAACACTGGGATAA
 CTCCGGAAACCGGGCTAATACCGAATGGTTTAAACCGCATGGTCAAAACAAAAGGTGGCTACCCACTAACAGATGGACCG
 CGGGCCATTAACTTAGTGGTAGTAACGGCTCACCAAGGCAACGATCGGTTAGCCCGTGAGAGGTGATTGCCCACACTGGGATTGAA
 ACACGCCCCCAGACTCCTACGGGAGGGCACCACTGGGAGGGTAAACGTTCTGAGGAAAGTCTGACGGAGCAACCCCGCGTAGTGTGAA
 AGGTTTCGGATCGTAAAGCTTCTGTTAGGGAGAACACAAGTACCGTTGAAATGGCGGTACCTTGACGGTACCTAACAGAAACCC
 GGCTAAACTACGTGCCAGCGCCGGTAATACGTAGGTGCAAGCGTTGCGGAAATTATGGGCTAAAGGGCTCGCAGGGGTTCTTA
 AGTCTGATGTGAAAGCCCCGGCTAACCGGGAGGGTCACTGGAAACTGGGAACTTGAGTGAGAAGAGGAGAGTGAATTCCACGTG
 AGCGGTGAAATCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGGACTCTGGTCTGTAACTGACGCTGAGGAGCGAACAGCTGGG
 AGCGAACCCAGGATTAGATACCCGTAGTCCACGCCGAAACGATGAGTGCTAAGTGTAGGGGTTCCCGCCCTTAGTGTGAGCTAA
 CGCATTAAACCACTCCGCTGGGACTACGGTCGCAAGACTGAAACGTTGACGGGGCCGACAAGCGGTGGAGCATGTGGTT
 TAATTGCAAGCAACGCGAAGAACCTTACCAAGGTCTGACATCCTGACAATCCCTAGAGATAGGACGTTCCCGGGCAGAGTGACA
 GGTGGTGCATGGGGTAGTCAGCTCGATGTCGAAAGATGTTGGGTTAAGTCCCGCAACGAGCGAACCCCTGATCTTAGTGTGCA
 GTTGGGACTCTAACGGTACTGCCGTGACAAACCGGAGGAAGGTGGGATGACGTCATCATGCCCCTATGACCTGGGCTACACA
 CGTGCTACAATGGACAGAACAAAGGGCAGCGAACCCCGAGGTTAAGCCAATCCCACAAATCTGTTCTCAGTTGGATCGCAGTCTGCAACT
 CGACTGCGTGAAGCTGGAATCGCTAGTAATCGGGATCAGCATGCCCGGTGAATACGTTCCGGGCTTGACACACCGCCGTCACACC
 ACGAGAGTTGTAACACCGAAGTCGGTAGGTAACCTTTAGGAGCCAGCCGCCGAAGGTGGGACAGAGAGAAGTTGGAAGGAAGAGCC
 ATCCG

2.10 The 16S rDNA nucleotide sequence of strain P7-1

TGACTGGCGCCGTGCCATAACATGCAAGTCGAGCGGACAGATGGGAGCTTGCCTCTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAA
 TGCCTGTAAGACTGGATAACTCCGGAAACCGGGGCTAATACCGGATGGTTGAACCGCATGGTTCAAACATAAAAGGTGGCTCGCTACCA
 CTTACAGATGGACCCGGCGCATTAGCTAGTTGGTAGGGTAACCGCTACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTATCGGCCACACT
 GGGACTGAGACACGGCCCAGACTCCTACGGGAGGCACAGTAGGGAATCTCCGAATGGACGAAAGTCTGACGGAGCAACGCCGTGAGTGATG
 AAGGT;TTCGGATCGTAAAGCTCTGTTAGGGAAAGACAAGTACCGTTGAATAGGGCGTACCTTGACGGTACCTAACAGAAAGGCCACGGCTA
 ACTACGTGCCAGCAGCCGGTAACGTAGGTGGCAAGCGTTGCGGAATTATTGGGCTGAAAGGGCTCGCAGGGGTTCTTAAGTCTGATGTG
 AAAGCCCCCGCTCAACCGGGGAGGGTCACTGGAAACTGGGAACCTGAGTGAGTCAGAAAGAGGAGAGTGAATTCCACGTGAGGGTAAATGCGTA
 GAGATGTGGAGGAACACCAACTGGCGAAGGCAGCTCTGGTCTGTAACTGACGCTGAGGAGCGAAAGCGTGGGAGCGAACAGGATTAGATA
 GGTAGTCCACGCCGTAACGATGAGTCTAAGTGTAGGGGTTCCGCCCTTAGTGTGCACTAACGCTAACGCACTCCGCTGGGAGTACG
 GTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGGCCCACAAGCCGGTGAGCATGTTAATTCAAACACCCAAAACCTTACCAAGGTCT
 TGACATCCTCTGACAATCTAAATAGACTCCCTCGGGGAAATACGGTGCATGTTCTCTGCTGAATTGGTTAAATCCCC

2.11 The 16S rDNA nucleotide sequence of strain P7-3

TGGCATGATCTCCCAGGGCCCCCTTGAGTTGAATCCTGGCTAGGACGAACGCTGGCGCGTGCCTTAATACATCCAAGTCGAGCGGACAA
 AATGGGAGCTGCTCCATAATGTAAGCGCGGACGGGTGAATTAAACACGTGGGTAACTCGCTTGAAGACTGGATAACTCCGGAAACCGGGC
 TAATACCGGATGGTTGTTGAACCAACATGGTCAAACATAAAAGGTGGCTCGCTACCAACTACAGATGGACCCCGCGCATTAGCTAGTTGGT
 AGGTAAACGGCTACCAAGGCAACGATCGCTAGCGACCTGAAAGGGTATCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAG
 CAGCAGTAGGGAATCTTCCGAATGGGACGAAAGTCTGACCGAACCGCCCGTGAGTGATGAAAGGTTTCGGATCGTAAAGCTCTGTTGGT
 AGGGAAAGAACCAAGTACCGTTGAATAGGGCGTACCTTGACGGTACCTAACAGAAAGCCACGGTAACACTGTGCCAGCAGCGCGTAATACG
 TAGGTGGCAAGCGTTGTCGGATTATTGGCGTAAGGGCTCGCAGGCAGGTTCTTAAGTCTGATGTGAAAGCCCCCGCTAACCGGGAGGGT
 ATTGGAAACTGGGAACTTGAGTGAGAAGAGGAGAGCGGAACTCCACGTGAGTCGATGAAATGGTAGAGATGTGGAGGAACACCAACTGGCGA
 GGCAGCTCTGGTCTGAAACTGACGCTGAGGAAGCGAAAGCGTGGGGAGCGAACCGAGGTTAAGATAACCTGGTAGTCCACGCCGAAACG
 ATGAGTCTAAGTGTAGGGGTTCCGCCCTTAGTGTGCACTAACGCTAACGCAACTCCCCCTGGGAGTACGGTGCAGACTGAAAGCT
 AAGGAATTGACGGGGGCCACAAGCGGTGGAGCATGTTTAATCGAAGCAACCGCAAGAACCTTACAGGTCTGACATCCTCTGACAATCC
 TAGAGATAGGACGTCCTCGGGGAGAATGACAGGGTGCATCGTGTGCTAACAGCTCGTATCGTGGAGATGTTGGTTAAAGTCCCGCA
 ACGAGCGCAACCCCTGATCTAGTGTGCACTAACGCTAACGCAAGTGGAGAACAAAGGGCAGCGAACCGCAGGTTAAGCCAATCCCACAA
 CATCATGCCCTTATGACCTGGCTACACACGTCTAACATGGACAGAACAAAGGGCAGCGAACCGCAGGTTAAGCCAATCCCACAA
 TCAGTTCGGATCGCAGTCTGCAACTCGACTCGCTGAAGCTGGAAATCGCTAGTAATCGCGGATCAGCATGCCGGTGAATACGTT
 CACACCGCCCGTACACCACGAGAGTTGTAACACCCGAAGTCGGTGAGGTAACCTTTAGGAGCCGGCGAAAGGTGGCATCCCTGGAA
 ACCTAAACA

2.12 The 16S rDNA nucleotide sequence of strain S9-2

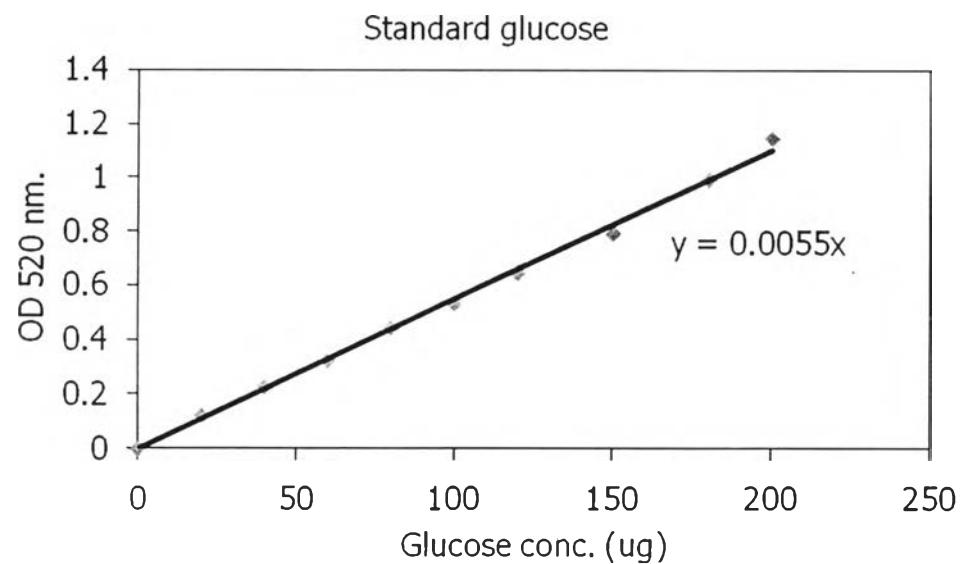
TGGTTCCCTTTGTTATACCCAGGGCCTCCAGGACGAAACGCCGGGGGTCTTAAACATCCAAGTCGAAGCGACAAAAGGGGAGTGCTTCCCC
 TGATTITAAGCGCGGACGGGTTAGAAACAGTGGTAACCTGCCCTGAAAGACTGGGACTCTCCGGGAAACCCGGGCTAATACCGGGATGTTGT
 TTAAACCCCATGGTCAACATAAAAGGGCTCGGCCTCCACTACAGATGGACCCCGCGCGATAAGCTAGTGGGTGAGGTAACGGCTACCAAG
 GCGACGATGTAGCCGACCTGAGAGGGGATCGCCCACACTGGGACTGAGACACGCCAGACTCCTACGGGAGGCAGCAGTAGGGATCTCCGCA
 ATGGGACGAAAGTCTGACGGAGCAACCCCGTGAGTGTGAAGGTTTCGGATCTAAAGCTCTGTTAGGAAGAACAGTACCGTCAATA
 GGGCGTACCTGACGGTACCTAACAGAAAGCCACGGCTAACTACGGCCAGCAGCGCGTAATACGTAGGGCAGCGTAGGGATCTCCGCA
 AGGGCGTAAAGGGCTCGCAGCGGTTCTTAAGTGTGAAAGCCCCGGCTAACCGGGGAGGGTCACTGGAAACTGGGAACTTGGTACTGAGTCAGA
 AGAGGAGAGTGGAACTCACGTGAGGGTGAATCGTAGAGATGTGGAGGAACACAGTGGAGAACGGCAGCTCTGGTCTGAACTGACGCTG
 AGGAGCGAAAAGCGTGGGGAGCGAACAGGATTAGATACCTGGTAGTCCACGCCGAAACGATGAGTGTAAAGTGTAGGGGTTCCGCCCTAG
 TGCTGCAGCTAACGCTAACGACTCCGCCGGAGTACGGTCGAAAGACTCAAAGGAAATTGACGGGGCCGACAAGCGTGGAGCA
 TGTGGTTAACGCAAGCAACCGAAGAACCTTACCAAGGCTTGACATCTCTGACAATCTAGAGATAGGACGCTCCCTCGGGGAGAGTGA
 GGTGGTGATGGTTGTCATCAGCTCTGCGTAGATGGTTAAGTCCCGAACAGCGAACCTTGATCTAGGGCTACACCGTCTGACAATGG
 TTCTAAGGTGACTGCCGGTACAACCCGGAGGAAGGTGGGATGACGTCAAATCATGCCCCATTGACCTGGCTACACCGTCTGACAATGG
 CAGAACAAAGGGCAGCGAACCCCGAGGTTAACCCAATCTGTTCTGAGCTCGGATCGCAGTCTGCAACTCGACTGCGTGAAGCTGGAA
 TCGCTAGTAATCGGGATCAGCATGCCCGGTGAATCGTCCCGGCTTGACACACGCCGTCACACCACGAGAGTTGAAACACCGAACGTC
 GGTGAGGTAACCTTTAGGAGGCCGGCAAGGTGGGAGCAGATGATTGGAGAACGTCGTTGAGAAGTGTGAGCTCGAGA

2.13 The 16S rDNA nucleotide sequence of strain S10-4

CAACTGCGCGTGCCTATCATGCAAGTCGAGCGGACTTGATGGAGAGCTTGCTCTCTGATGGTAGCGGCGGACGGGTGAGTAACACGTAGGCAAC
 CTGCCTGCAAGACCGGGATAACCCACGAAACGTGAGCTAAACCGGATATCTCATTTCTCTGAGGGGATGATGAAAGACGGAGAACCTG
 ACTTGCGGATGGGCCTGCGCGCATTAGCTAGTGGTAGGTAACGGCTACCAAGGCAGATGCGTAGCCGACCTGAGAGGGTAACGGCCACA
 CTGGGACTGAGACACGCCAGACTCCTACGGAGGCAGCAGTAGGAAATCTCCGAAATGGCGAAAGCCTGACGGAGCAACGCCGTGAGTGA
 TGAAGGTTTCGGATCGTAAAGCTCTGTTCCAGGGAAAGAACGTCGGTAGAGTAACGCTACCGGAGTGACGGTACCTGAGAAAGAACCCGGCT
 AACTACGTGCCAGCAGCGCGTNAATAATAGCNNNNNGCAGGAAGCGTTGCGGAAATTATGGCGTAAAGCGCGCAGCGGTCTTAA
 AGTCTGGTCTTAAGGCCAAGCTAACCTGGTCGACTGAAACTGGGTGACTTGAGTGCAGAAGAGGAGAGTGGAAATTCCACGTGAGCGT
 AAATGCGTAGATATGGAGGAACACCGATGCGAAGCGACTCTCTGGCTGTAACTGACGCTGAGGCGCGAACGCTGGGAGCAAACAGGATT
 AGATACCCCTGGTAGTCCACGCCGTAACCGATGAATGCTAAGGTGTTAGGGGTTGACACCTTGGTCCGAAAGTTAACACATTAAGCATCCCTG
 GGGAGTACNGGTGCAAGACTCAAAGGAATTGACGGGANCCCGACAAGCAGTGGAAATATGGNTTAATTGAAGCAACCCGAAAAA
 CCTTACCAAGGNTCTGACATCCCTGAAACCGGTCTAGAGATANANNNCCTTCGGACAAGAACGACANGGTGGCNATGGNTGCGT
 NCAGCTCGTNGTCGTGAGNATGGTTAAGTCCCCAACNNNNNNNNNGAGCGAACCCCTNNNNNNNNNCAGTTAGTGGCTACACAGTA
 GGGTGGCAGTCTAGAATGACTGCCGTGACAAACCGAGGTAAGGCGGGATGACGTCACATGCCCCATTGACCTGGCTACACAGTA
 CTACAATGCCAGTACAACGGGAAGCGAAGCCGAGGTGAGGCAATCTATCAAAGCTGGCTCAGTCGGATTCAGGCTCAACTGCCGT
 TGAAGTCGAAATTGCTAGTAATCGGGATCAGCATGCCCGGTGAATACGTTCCGGCTTGACACACGCCGTCACACCACGAGAGTTACAA
 CACCCGAAGTCGGTAGGTAACCGCCAAGGAAGCCAAGGCCGAAAGGTGGGAGTGTGAGTGGGTGAAGTCGAACAAGGTACCGTAAT

APPENDIX E

1. Standard curve of glucose



Poster presentation :

1. Thanawan Taprig , Ancharida Acharacharanya and Somboon Tanasupawat. 2005. Screening and identification of cellulase-producing bacteria from soil in NAN province. The Thai Society for Biotechnology. November 2-3, 2006 at the Montien Riversidde Hotel Bangkok, Thailand.

Biography

Miss Thanawan Taprig was born on June 20, 1983 in Bangkok, Thailand . she obtained a Bachelor of Science Degree in Microbiology from Srinakharinwirot university. Bankok, Thailand 2005.

