CHAPTER IV

CONCLUSION

In this research, the woods of *Croton oblongifolius* Roxb. were extracted with hexane, dichloromethane, ethyl acetate and methanol, respectively. Five compounds were isolated from four crude extracts by column chromatography. They are a mixture of long chain aliphatic hydrocarbons (C_{27-33}), a mixture of long chain aliphatic carboxylic acids ($C_{18, 22-34}$), a mixture of steroids (stigmasterol, β -sitosterol and campesterol), a mixture of steroid glycosides (stigmasteryl-3-O- β -D-glucopyranoside, β -sitosteryl-3-O- β -D-glucopyranoside and campesteryl-3-O- β -D-glucopyranoside) and 7-hydroxy-6-methoxycoumarin ($C_{10}H_8O_4$).

All isolated substances and amounts are summerized in Table 19.

Table 19	Isolated Substances	from the	Woods o	of Croton oblongifolius Roxb.
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Compound	Name of Compound	Weight (mg)	% wt. by wt. of air-dried woods
1	a mixture of long chain aliphatic hydrocarbons (C ₂₇₋₃₃) white solid, mp. 57-58 [°] C	24.1	3.01x10 ⁻⁴
	a mixture of steroids (stigmasterol, β-sitosterol and campesterol) white needles, mp. 152-154 °C	998.2	1.25x10 ⁻²
111	a mixture of long chain aliphatic carboxylic acids(C _{18, 22-34}) white amorphous solid,mp.73-75 °C	29.9	3.74x10 ⁻⁴

Table 19Isolated Substances from the Woods of Croton oblongifolius Roxb.(continued).

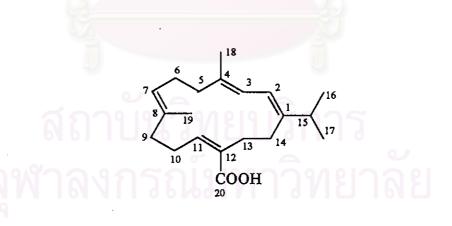
IV	7-hydroxy-6-methoxycoumarin	18.3	2.29x10 ⁻⁴
	(C ₁₆ H ₈ O ₄)		
	colourless needles, mp. 206-208 °C		
V	a mixture of steroid glycosides	203.8	2.55x10 ⁻³
	(stigmasteryl-3-O-β-D-		
	glucopyranoside, β-sitosteryl-3-O-β-D-		
	glucopyranoside and campesteryl-		
	3-O-β-D-glucopyranoside)		
	white amorphous solid,		
	mp. 271-273 °C		

No compound with a structure related to plaunotol which was diterpene containing double bonds and crotocembraneic acid which was 14-membered ring could be isolated or identified.

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย PART B

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

From previous research (19), a new compound from the bark of Croton oblongifolius Roxb. was found. This compound was crotocembraneic acid (1E,3E,7E,11Z-1-isopropyl-4,8-dimethylcyclotetradeca-1,3,7,11-tetraene-12-carboxylic acid) which has antitumor activity. The structure of crotocembraneic acid was 14-menbered ring containing four double bonds and substitution groups. Its structure was shown in Fig. 5. Naturally cembrene skeleton was found in marine creatures (27-29), tobacco (30-31), pine tree (30) and Croton pailanei (33). In 1993, the solution comformations of some cembrenoids have been studied by using NMR methods (30). This research studied the most stable conformation of this acid in solution. This conformation was the information to predict the products of reactions of crotocembraneic acid. The conformation of this compound was then studied by the dynamic NMR spectroscopy at various temperature. The temperatures of this measurement were ± 25 , ± 15 , ± 7 , 0 and -35 °C. The NMR spectra of crotocembraneic acid at various temperature were shown in Fig. 30-37. The results obtained were shown in Table 20.





The Structure of Crotocembraneic Acid.

Position	δ _H (ppm)								
	25 °C	15 °C	7°C	ວິດ	-7 °C	-15°C	-25°C	-35°C	
1	-	-	-	-	-	-	-	-	
2	6.03(d)	6.03(d)	6.03(d)	6.03(d)	6.03(d)	6.03(d)	6.03(d)	6.03(d)	
3	5.89(d)	5.89(d)	5.89(d)	5.89(d)	5.89(d)	5. 8 9(d)	5.89(d)	5.89(d)	
4	-	<u> </u>			-	-	-	-	
5	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.17(m)	2.17(m)	2.17(m)	
6	2.20(m)	2.20(m)	2.20(m)	2.20(m)	2.21(m)	2.21(m)	2.21(m)	2.21(m)	
7	5.10(t)	5.10(t)	5.11(t)	5.11(t)	5.11(t)	5.12 (t)	5.12(t)	5.12(t)	
8	-	· /	1.	5-	-	-	-	-	
9	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.17(m)	2.17(m)	2.17(m)	
10	2.69(q)	2.69(q)	2.70(q)	2.70(q)	2.70(q)	2.71(q)	2.71(q)	2.72(q)	
11	6.00(t)	6.00(t)	6.00(t)	6.00(t)	6.00(t)	6.00(t)	6.00(t)	6.00(t)	
12	-	-			-	-	-	-	
13	2.40(m)	2.40(m)	2.41(m)	2.41(m)	2.41(m)	2.41(s)	2.41(s)	2.41(s)	
14	2.40(m)	2.40(m)	2.41(m)	2.41(m)	2.41(m)	2.41(m)	2.41(m)	2.41(m)	
15	2.34(m)	2.34(m)	2.34(m)	2.34(m)	2.34(m)	2.34(m)	2.34(m)	2.34(m)	
16	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	
17	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	1.03(d)	
18	1.74(s)	1.75(s)	1.75(s)	1.75(s)	1.76 (s)	1.76 (s)	1.76 (s)	1.77(s)	
19	1.55(s)	1.55(s)	1.55(s)	1.55(s)	1.55(s)	1.55(s)	1.55(s)	1.55(s)	
20		-	-	-	•	_	-	-	
	-	-	4.94(s)	5.01(s)	5.30(s)	5.18(s)	5.32(s)	5.47(s)	
-	-	-	-	5.21(s)	-	5.42(s)	-		

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Table 20

The results from dynamic NMR, at $\delta 2.41$ ppm were proton of $-CH_2$ -, it changed from multiplet to singlet at temperature -15, -25 and -35 °C. At 7 °C found new peak at $\delta 4.94$ ppm as a singlet, 0 °C found new peak at $\delta 5.01$ and 5.21 ppm as singlet, -7 °C found new peak at $\delta 5.30$ ppm as a singlet, -15 °C found new peak at $\delta 5.18$ and 5.42 ppm as singlet, -25 °C found new peak at $\delta 5.32$ ppm as a singlet and -35 °C found new peak at $\delta 5.47$ ppm as a singlet. This results were suggest that they were the signal of proton of -OH group.

So methylester of crotocembraneic acid was prepared and measured with the dynamic NMR spectroscopy at ± 25 , ± 15 , ± 7 , 0 and -35 °C. The NMR spectra of methylester of crotocembraneic acid at various temperature were shown in Fig. 38-45. The results obtained were shown in Table 21.

Table 21The Chemical Shift of Methylester of Crotocembraneic Acid atVarious Temperature.

Position	δ _H (ppm)								
	25 °C	15 °C	7 °C	0°C	-7 °C	-15 [°] C	-25°C	-35 [°] C	
1	- 6		-	-		-	-	*	
2	5. 98 (d)	5. 98 (d)	5.98(d)	5.98(d)	5.98(d)	5.97(d)	5.97(d)	5.97(d)	
3	5.86(d)	5.86(d)	5.86(d)	5.85(d)	5.85(d)	5.85(d)	5.84(d)	5.84(d)	
4	30				004	-	-	-	
5	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.15(m)	2.15(m)	2.15(m)	
6	2.21(m)	2.21(m)	2.21(m)	2.21(m)	2.21(m)	2.21(m)	2.21(m)	2.20(m)	
7	5.08(t)	5.07(t)	5.07(t)	5.07(t)	5.07 (t)	5.07 (t)	5.06(t)	5.06(t)	
8	-	-	-	-	-	-	-	-	
9	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.16(m)	2.15(m)	2.15(m)	2.15(m)	
10	2.5 8 (q)	2.58(q)	2.58(q)	2.58(q)	2.58(q)	2.57(q)	2.57(q)	2.57(q)	
11	5.91(t)	5.90(t)	5.90(t)	5.90(t)	5.90(t)	5.89(t)	5.89(t)	5.89(t)	

Position	δ _H (ppm)								
	25 °C	15 °C	7°C	0°C	-7 °C	-15 °C	-25 °C	-35 °C	
12	-	-		-	-	+	-	-	
13	2.35(m)	2.35(m)	2.35(m)	2.35(m)	2.34(m)	2.34(m)	2. 3 4(m)	2.34(m)	
14	2.35(m)	2. <mark>35(m)</mark>	2.35(m)	2.35(m)	2.34(m)	2.34(m)	2.34(m)	2.34(m)	
15	2.29(m)	2.29(m)	2.28(m)	2.28(m)	2.28(m)	2.27(m)	2.27(m)	2. 2 6(m)	
16	1.00(d)	1.00(d)	0.99(d)	0.99(d)	0.98(d)	0.9 8 (d)	0.9 8 (d)	0.97(d)	
17	1.05(d)	1.04(d)	1.04(d)	1.04(d)	1.03(d)	1.03(d)	1.03(d)	1.02(d)	
18	1.71(s)	1.71(s)	1.71(s)	1.71(s)	1.71 (s)	1.71(s)	1.71(s)	1.71(s)	
19	1.52(s)	1.51(s)	1.51(s)	1.50(s)	1.50 (s)	1.50(s)	1.49(s)	1.48(s)	
20	-			-	-	-	-	-	
OC <u>H</u> 3	3.70(s)	3.6 <mark>9</mark> (s)	3.69(s)	3.69(s)	3.69 (s)	3.69(s)	3.69(s)	3.69(s)	
-	-	-		-	5.02 (s)	5.30(s)	5.26(s)	5.40(s)	
-	- 🔎	-	-	-	-0	-	5.47(s)	5.66(s)	

Table 21The Chemical Shift of Methylester of Crotocembraneic Acid atVarious Temperature (continued).

From the results of NMR spectra of methylester of crotocembraneic acid, at -7 °C found singlet peak at δ 5.02 ppm, at -15 °C found singlet peak at δ 5.30 ppm, at -25 °C found singlet peak at δ 5.26 and 5.47 ppm and -35 °C found singlet peak at δ 5.40 and 5.66 ppm. Therefore, the suggestion that new peaks in ¹H-NMR spectra of crotocembraneic acid were the signal of proton of -OH group, was not corrected.