น้ำมันหอมระเหยที่มีฤทธิ์เป็นสารรมต่อด้วงงวงข้าว *Sitophilus oryzae* (Linnaeus)

นางสาว วชิราภรณ์ ฟูนัน

วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรมหาบัณฑิต

สาขาวิชาเทคโนโลยีชีวภาพ หลักสูตรเทคโนโลยีชีวภาพ

คณะวิทยาศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย

ปีการศึกษา 2549

ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

ESSENTIAL OILS AS FUMIGANT AGAINST RICE WEEVILS,

Sitophilus oryzae (Linnaeus)

Miss Wachiraporn Phoonan

A Thesis Submitted in Partial Fulfillment of the Requirements for the Degree of Master of Science Program in Biotechnology Faculty of Science Chulalongkorn University Academic Year 2006 Copyright of Chulalongkorn University

Thesis Title	ESSENTIAL OILS AS FUMIGANT AGAINST RICE
	WEEVILS, Sitophilus oryzae (Linnaeus)
Ву	Miss Wachiraporn Phoonan
Field of Study	Biotechnology
Thesis Advisor	Assistant Professor Warinthorn Chavasiri, Ph.D.
Thesis Co-advisor	Chatchawan Chaisuekul, Ph.D.

Accepted by the Faculty of Science, Chulalongkorn University in Partial Fulfillment of the Requirement for the Master's Degree

flens

(Professor Piamsak Menasveta, Ph.D.)

THESIS COMMITTEE

(Associate Professor Sirirat Kokpol, Ph.D.)

(Assistant Professor Warinthorn Chavasiri, Ph.D.)

Chutcher ChainderlThesis Co-advisor

(Chatchawan Chaisuekul, Ph.D.)

N. Ngamrojanavanieh Member

(Associate Professor Nattaya Ngamrojanavanich, Ph.D.)

Drouglehre Sittlicharoorchai Member

(Duangkae Sittijaroenchai, Ph.D.)

วชิราภรณ์ ฟูนัน : น้ำมันหอมระเหยที่มีฤทธิ์เป็นสารรมต่อด้วงงวงข้าว Sitophilus oryzae (Linnaeus) (ESSENTIAL OILS AS FUMIGANT AGAINST RICE WEEVILS, Sitophilus oryzae (Linnaeus)) อาจารย์ที่ปรึกษา : ผศ.ดร.วรินทร ชวศีริ, อาจารย์ที่ ปรึกษาร่วม : อ.ดร.ชัชวาล ใจซื่อกุล, 51 หน้า.

การประเมินฤทธิ์ในการเป็นสารรมของน้ำมันหอมระเหย 25 ชนิดต่อด้วงงวงข้าว Sitophilus oryzae Linn ซึ่งเป็นแมลงศัตรูในโรงเก็บที่สำคัญชนิดหนึ่ง พบว่าน้ำมันหอมระเหยจาก เมล็ดเทพธาโร (Cinnamomum porrectum Roxb Kosterm) แสดงฤทธิ์ในการเป็นสารรมต่อตัว เต็มวัยของด้วงงวงข้าวสูงที่สุดหลังการทดสอบเป็นเวลา 24 ชั่วโมง ตามด้วยน้ำมันหอมระเหยจาก สะระแหน่ญี่ปุ่น (Mentha arvensis Linn) และไพล (Zingiber cassumunar Roxb) ที่ค่า LC₅₀ เท่ากับ 79, 138 และ 260 µL/L air ตามลำดับ ได้วิเคราะห์องค์ประกอบหลักในน้ำมันหอมระเหย แต่ละชนิดด้วยเทคนิคทางสเปกโทรสโคปิกและหลังจากนั้นทดสอบฤทธิ์ในการเป็นสารรม พบว่า safrole องค์ประกอบหลักในน้ำมันหอมระเหยจากเมล็ดเทพธาโรแสดงประสิทธิภาพในการเป็น สารรมต่อด้วงงวงข้าวมากที่สุด และมี LC₅₀ = 57.9 µL/L air ที่ความเข้มข้น 100 µL/L air หลังการ ทดสอบเป็นเวลา 24 ชั่วโมง นอกจากนี้ ยังไม่พบสารตกค้างของ safrole บนข้าวที่ทำการรมแล้ว จากการศึกษาความสัมพันธ์ของประสิทธิภาพในการเป็นสารรมกับองค์ประกอบหลัก พบว่า menthol (LC₅₀ = 95.4 µL/L air) เป็นสารออกฤทธิ์ในน้ำมันหอมระเหยจากสะระแหน่ญี่ปุ่น ในขณะที่ terpinen-4-ol (LC₅₀ = 165.5 µL/L air) ซึ่งเป็นหนึ่งในองค์ประกอบหลักในน้ำมันหอม ระเหยจากไพลเป็นสารออกฤทธิ์ ดังนั้นน้ำมันหอมระเหยทั้งสามชนิดและองค์ประกอบหลัก เหล่านั้นอาจสามารถพัฒนาเป็นสารรมที่มีประสิทธิภาพสำหรับเมล็ดรัญพีขในโรงเก็บได้

สถาบนวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

หลักสูตรเทคโนโลยีชีวภาพ	ลายมือชื่อนิสิต	าซีราภาณ์	ป่งใน
สาขาวิชาเทคโนโลยีชีวภาพ	ลายมือชื่ออาจารย์ที่ป	รึกษา(Jon vote
ปีการศึกษา2549	ลายมือชื่ออาจารย์ที่ป	รึกษาร่วม	Bugons Dodogs

4772448123: MAJOR BIOTECHNOLOGY

KEY WORD: FUMIGANT ACTIVITY / ESSENTIAL OIL / Sitophilus oryzae Linn WACHIRAPORN PHOONAN : THESIS TITLE. ESSENTIAL OILS AS FUMIGANT AGAINST RICE WEEVILS, Sitophilus oryzae (Linnaeus) THESIS ADVISOR : ASSISTANT PROFESSOR WARINTHORN CHAVASIRI, Ph.D. THESIS CO-ADVISOR : CHATCHAWAN CHAISUEKUL, Ph.D., 51 pp.

The fumigant activity of twenty-five essential oils was evaluated on an important stored-product insect pest Sitophilus oryzae Linn. The essential oils from Cinnamomum porrectum (Roxb) Kosterm (Thep ta-ro) exhibited the highest fumigant toxicity against adults of S. oryzae after 24 h exposure, followed by those from Mentha arvensis Linn (Japanese mint) and Zingiber cassumunar Roxb (plai) with LC50 79, 138 and 260 µL/L air, respectively. The main constituents of each essential oil were analyzed by spectroscopic techniques and then tested in fumigation bioassay. Safrole, the major component of the essential oil from C. porrectum, exhibited the most potent fumigant (LC₅₀ = 57.9 μ L/L air) at the concentration of 100 μ L/L air after 24 h exposure. Additionally, the residue of safrole could not significantly be detected on the fumigated rice. The major component-fumigant activity relationship study revealed that menthol ($LC_{50} = 95.4 \,\mu L/L$ air) was an active component of the essential oil from *M. arvensis*, while terpinen-4-ol (LC₅₀ = 165.5 μ L/L air), one of the major constituents of the essential oil from Z. cassumunar, was an active component. Thus, the essential oils from all three plant sources and their major constituents may be developed as a potent fumigant for storage grains.

์ สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

Program ofBiotechnologyStudent's signature	W. Phoonan
Field of study BiotechnologyAdvisor's signature Academic year2006Co-advisor's signatu	re Chatre Chindel

ACKNOWLEDGEMENTS

The author would like to express her appreciation to her advisor and coadvisor, Assistant Professor Dr. Warinthorn Chavasiri and Dr. Chatchawan Chaisuekul for advice, assistance and comment during this research. Sincere thanks are extended to Natural Products Research Unit, Department of Chemistry, Faculty of Science, Chulalongkorn University for the support of chemicals and laboratory facilities. Many thanks to the thesis committee, Associate Professor Dr. Sirirat Kokpol, Associate Professor Dr. Nattaya Ngamrojanavanich and Dr. Duangkae Sittijaroenchai for their valuable discussion and suggestion.

The author also acknowledged Royal Forest Department and Thai-China Flavours and Fragrances industry Co., Ltd. for providing some essential oils used in this study. Moreover, the author would like to thanks Mrs. Nantaka Konchim and Miss Parichat Chaisaeng, staffs of Postharvest technology Research and Development, Department of Agriculture, Thailand for their kind gratitude of giving information in insect rearing and the colleagues at Natural Product Research Unit for assistances throughout this research.

In addition, appreciation is also extended the Graduate School of Chulalongkorn University and TRF-Master Research Grants for granting partial financial support to conduct this research. Finally, the author would like to express her gratitude to her parents and family members for their inspiration, understanding, great support and encouragement throughout the entire study.

สถาบนวทยบรการ จุฬาลงกรณ์มหาวิทยาลัย

CONTENT

Page

ABSTRACT IN THAI	iv
ABSTRACT IN ENGLISH	v
ACKNOWLEDGEMENTS	vi
CONTENT	vii
LIST OF TABLES	ix
LIST OF FIGURES	
LIST OF ABBREVIATIONS	xi

CHAPTER I INTRODUCTION

1.1 General characteristics of Sitophilus oryzae Linnaeus	2
1.2 Essential oils	8
1.3 Fumigants from plants against S. oryzae (L.)	11
1.4 Rationale	11
1.5 The goals of this research	11

CHAPTER II MATERIALS AND METHODS

2.1 Insects	12
2.2 Plant materials	12
2.3 Chemicals	14
2.4 General procedures of hydrodistillation	14
2.5 Fumigation bioassay	14
2.6 Screening of essential oils for fumigation bioassay	15
2.7 Isolation of major constituents from the potent essential oils	16
2.8 Spectroscopic analysis	16
2.9 Fumigation test of pure compounds	16
2.10 Residue determination	16
2.10 Statistical analysis	17

CHAPTER III RESULTS AND DISCUSSION

3.1 Hydrodistillation results	18
-------------------------------	----

Page

3.2 Screening of essential oils for fumigation bioassay	18
3.3 LC ₅₀ determination	21
3.4 Essential oil analysis	24
3.4.1 The essential oil from <i>M. arvensis</i>	24
3.4.2 The essential oil from Z. cassumunar	25
3.4.3 The essential oil from <i>C. porrectum</i>	28
3.5 Fumigation test of constituents	29
3.6 Residue analysis	35

CHAPTER IV CONCLUSION37Proposal for the future work.38REFERENCES.39APPENDIX.45VITAE51



LIST OF TABLES

		Page
Table 2.1	Sources of essential oils	13
Table 3.1	The hydrodistillation of some selected plants	18
Table 3.2	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 1000 μ L/L air after 24 h treatment	19
Table 3.3	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 400 µL/L air after 24 h treatment	20
Table 3.4	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 200 µL/L air after 24 h treatment	21
Table 3.5	The compositions of essential oil from <i>M. arvensis</i>	24
Table 3.6	The composition of essential oil from Z. cassumunar	25
Table 3.7	The ¹ H-NMR chemical shift assignments of sabinene	26
Table 3.8	The ¹ H-NMR chemical shift assignments of terpinene-4-ol	28
Table 3.9	The ¹ H-NMR chemical shift assignments of safrole	29
Table 3.10	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at the concentration of constituent at 100 μ L/L air after 24	
	h treatment	30
Table 3.11	LC ₅₀ values of main component in three essential oils against	
	S. oryzae	34
Table 3.12	Residues of menthol and safrole (mg/kg) on fumigated rice	
	comparison between groups of pre-aeration and 24 h aeration	35
Table A1	The LC ₅₀ of three effective essential oils against <i>S. oryzae</i>	49
Table A2	Residues of menthol and safrole (mg/kg) on fumigated rice	
	comparison between groups of pre-aeration and 24 h aeration	50

ix

LIST OF FIGURES

		Page
Figure 1.1	The adult of <i>Sitophilus oryzae</i> a) side view feature b) top view	
	Feature	3
Figure 1.2	Different stages of <i>S. oryzae</i> a) egg b) larva c) pupa d) adult	4
Figure 1.3	The infestations of S. oryzae (L.) in grains	5
Figure 1.4	Some common monoterpenes found in essential oils	9
Figure 1.5	Some common sesquiterpenes found in essential oils	10
Figure 3.1	Linear regression of concentrations of three highest potent	
	essential oils against S. oryzae at 24 h exposure	22
Figure 3.2	The gas chromatogram of the essential oil from <i>M. arvensis</i>	24
Figure 3.4	¹ H NMR spectrum of compound 1 (sabinene)	26
Figure 3.5	¹ H NMR spectrum of compound 2 (terpinen-4-ol)	27
Figure 3.6	The gas chromatogram of the essential oil from C. porrectum	28
Figure 3.7	¹ H NMR spectrum of safrole	29
Figure 3.8	The mortality percentages of adult S. oryzae at the concentration	
	of the effective constituents at 100 μ L/L air after 24 h treatment.	31
Figure 3.9	Structures of compounds used in this fumigation test	32
Figure 3.10	Residues of menthol and safrole (mg/kg) on fumigated rice	
	comparison between groups of pre-aeration and 24 h	
	aeration	36
Figure A1	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 1000 μ L/L air after 24 h treatment	46
Figure A2	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 400 μ L/L air after 24 h treatment	47
Figure A3	The fumigant activity of essential oils against Sitophilus oryzae	
	adults at concentration of 200 μ L/L air after treatment for 24 h	48

LIST OF ABBREVIATIONS

b.p.	boiling point
°C	degree Celsius
CDCl ₃	deuterated chloroform
cm	centimeter
¹³ C-NMR	proton nuclear magnetic resonance
D	dry
F	fresh
g	gram (s)
GC	gas chromatography
GC-MS	gas chromatography – mass spectroscopy
h	hour (s)
¹ H-NMR	carbon nuclear magnetic resonance
J	coupling constant
L	liter (s)
LC ₅₀	the concentration which causes 50% mortality
L:D	light:dark
mg	milligram (s)
MHz	megahertz
min	minute (s)
mL	milliliter (s)
mm 💽	millimeter (s)
n	number
ppm	part per million
\mathbf{R}_{f}	retardation factor
R.H.	relative humidity
SE	standard error
w/w	weight by weight
µg/mL	microgram per milliliter
μL	microlitre (s)
δ	chemical shift

CHAPTER I

INTRODUCTION

Grain products form a large part of the diet of the world's population, 64% of all food sources in the world, which divide into 50% of cereals and 14% of pulses. Therefore, the grain culturing area constitutes or occupied 75% of total culturing area worldwide. The agricultural grain products are rice, maize, wheat, sorghum, mung bean, soybean and groundnut etc. (Sukprakarn *et al.*, 1996). In the Southeast Asia, post harvest losses of grains have been estimated at 10-30%, caused mainly by improper drying and pest infestation during crop storage and distribution, and under hot and humid conditions (Hayashi *et al.*, 2004).

Rice is the most importance crop and staple food in Thailand. The rice is cultivated in all part of Thailand at about 10 million hectares and the annual production is about 19.5 million tons. Nearly all of the grain and other agricultural products are kept in the mills, godowns or silos (Sukprakarn, 1989). Storage facility operators must manage and protect the stored grain from various damages. The damages of rice grains are resulted from two important factors. The first factor is the physical factor, such as temperature and humidity, and the second factor is biological factor, such as insects, mites, fungi, birds, and rodents. Among these biological factors, insects are the most important pest, which cause the most damage to rice grain products (Sukprakarn *et al.*, 1996).

Infestation of stored product insects results in a variety of damage and economic loss, including the physical loss of commodity by insect direct consumption, the spoilage and loss of commodity, the encouragement of mold growth, the contamination of commodities with insect bodies and waste products which may be toxic, repulsive or allergenic. Moreover, the infested commodities may be rejected by consumers (both human and animal), and the resultant social and legal costs, restriction of trade and damage to economies will follow (Rees, 2004).

These are very serious problems for all agricultural countries, including Thailand. Nevertheless, chemical control agents, known as pesticides, have been the common practice of pest management and have contributed a major part in protecting crops from the damages inflicted by pests. In fact, most pesticides are obtained from synthetic agents. Most farmers usually employ synthetic agrochemicals since they are convenient, fast and easy to handle. However, synthetic agrochemicals have many drawbacks. The use of synthetic pesticides has caused some concerns regarding their adverse effects on the environment. These compounds are often not biodegradable and the residues are concentrated in food chains and accumulated in soil, water and plants, so they cause invariably environment pollution (Munakata, 1970). Furthermore, synthetic pesticides are generally toxic to human and non-target animals, and most of them are imported from foreign countries. According to the all of the previous reasons, novel alternatives from natural products seem to be promising to replace some synthetic chemicals. Naturally occurring compounds are considering friendly to environment because they are easy to decompose, low toxic to user, and inexpensive than imported pesticides (Lee *et al.*, 2001b).

1.1 General characteristics of Sitophilus oryzae Linnaeus

Many of the stored product insects are in the order Coleoptera at about 600 species (Sukprakarn *et al.*, 1996). The Curculionidae or true weevils is one of the largest families of beetles that can be found in a wide range of habitats, and many species are important pests of agriculture, horticulture and forestry. They can attack the stems, roots and seeds of plants, and some are wood borers. The head of adult weevil has a characteristic snout to bore into substrate. Members of the genus Sitophilus are among the most important pests of stored grain (Rees, 2004).

Sitophilus oryzae (Linnaeus) or rice weevil is a major important pest of paddy and polished rice in Thailand (Sukprakarn *et al.*, 1996) because this weevil is capable of successfully attacking and breeding in previously undamaged whole cereal and pulse grains. However, rice weevils are rarely successful on milled ground commodities. Rice weevils' infestation also allows the secondary pests to attack the damaged grains (Mutambuki, 2004). Therefore, the control of the weevil is necessary to manage the stored rice.

Morphological characteristics: Rice weevils vary in body color from reddish brown to nearly black and are usually marked on the back with four light- reddish or yellowish spots. The body length is 2.4-4.5 mm in adult. The size of adult varies somewhat with the seed size from which it emerged (Hayashi et al., 2004; Rees, 2004; Naunwat et al., 2005).

3

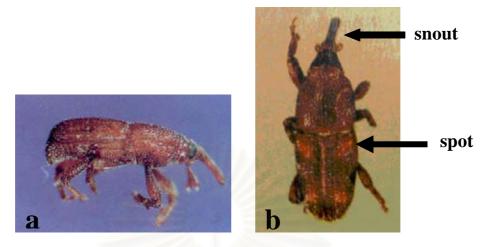


Figure 1.1 The adults of Sitophilus oryzae a) side view feature b) top view feature

Life cycle: Female rice weevil selects a spot on the grain surface then chews a small hole, into which she lays an egg. Eggs are laid singly into grains. The hole is then plugged with a waxy secretion. About 150-300 eggs are laid per female. The white legless larvae develop hiddenly within a cavity hollowed out within the grain. This stage takes approximately 25 days at 25 °C. Pupation takes place within the cavity made by the larva. Upon emergence from the pupa, the adult may spend several days within the cavity. Eventually, a newly molted adult will chew its way out, leaving a ragged hole. Adult weevils continue to feed on grain and may live about 4-12 months. The life cycle under favorable conditions can be completed within 30-35 days (Hayashi *et al.*, 2004; Rees, 2004; Naunwat *et al.*, 2005).

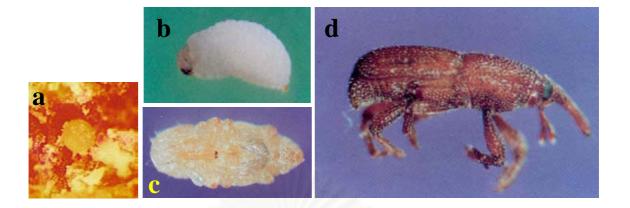


Figure 1.2 Different stages of *S. oryzae* a) egg b) larva c) pupa d) adult (Ministry of Agriculture and Cooperatives, Rice Department, 2006)

Behavior: The adults can fly but not often. Their behavior is always hidden in the grain pile. They will move out from their habitat and climb to the top when they are disturbed (Naunwat *et al.*, 2005). When they are threatened by enemies, they will feign death by enclosing their appendages with their body.

Infestation damage: Rice weevils are capable of infesting all cereal grains, but they prefer wheat, rice and other small grains. Feeding by larvae leaves large cavities inside grains and newly emerging adult leave behind large ragged emergence holes. Adults cause further damage by feeding, mainly by attacking previously damaged grain. Their infestations produce a lot of heat and moisture, and this encourages extensive quality loss, mold growth and growth of populations of other insect species (Rees, 2004).





Figure 1.3 The infestations of S. oryzae (L.) in grains.

(Ministry of Agriculture and Cooperatives, Rice Department, 2006)

The insect control of storage pests: The insect control can be divided into two strategies.

- 1) Non-chemical control
 - *Store hygiene*: The primary control tactic targets to minimizing insect pest infestation in stored grain through storage hygiene. Pit stores need to be thoroughly cleaned of insect pests prior to filling. Old grain should be checked, and if necessary, re-dried and cleaned to control existing infestations. Different products should be stacked separately. The storage structures should be closed off to prevent entry by pests, airtight silos with good thermal insulation offer the best protection (Mutambuki, 2004).
 - *Repellent herbs*: Admixture the grain or seed with inert substances such as dust or plant parts could prevent the grains from insect damage for some period (Sukprakarn, 1989).

- *Temperature Control*: Since most stored product insects cannot tolerate extreme temperature, heating and cooling are logical approaches to insect control. To some extent, it has been a common practice to superheat some commodities for insect control. The temperatures of 55-60°C maintained for 10 to 12 hours are effective. Low temperature is probably the most important single factor in making long term storage possible and economical. The insects become inactive and eventually die at a temperature below 12°C (Sukprakarn, 1989).
- *Moisture Control:* Most of the stored grain insects are unable to survive and reproduce in grain whose moisture content is below 9%. Most favorable grain moistures for insect development ranges from 12 to 15%. Various means, such as drying, and refrigerator curing, can be employed to reduce and maintain the moisture below than favorable for reproduction and development to control the insects (Sukprakarn, 1989).
- 2) Chemical Control

This strategy is commonly used in insect control. For the protection of stored produce against the insects, the following groups of pesticides are used:

- a) Contact insecticides: They can be either liquid or dust formulations. They can kill by penetrating the insect body being ingested or vapor inhalation. The lower toxicity insecticides in the group of organophosphate are fernitrothion (Sumithion[®]), chlorpyrifos methyl (Reldan[®]), methacrifos (Damfin[®]) and Dichlorvos. The commonly used synthetic pyrethroids are permithrin, cypermethrin (K-orthene[®]), deltamehrin (Ripcord[®]) and betacyfluthrin. These can be sprayed on coated or admixed with grain for long-term protection (Ministry of Agriculture and Cooperatives, Rice Department, 2006).
- b) *Fumigants*: These are chemicals which are toxic to insects in the vapour and smoke forms. They can be supplied in various formulations (solid, liquid or gas), when they are in contact with moist air, release highly toxic respiratory poison gases. In order for the fumigant to be effective, the commodity or space being treated must be properly sealed. The most common used fumigants in storage are 1) gasses fumigants and 2) liquid

fumigants. Gasses fumigants, such as methylbromide and phosphine are highly effective, but they have some constraints. Use of methylbromide will soon be restricted due to its potential ozone depleting properties (World Meteorological Organization [WMO], 1995). Moreover, it is highly toxic to warm-blooded animals including human (Dansi et al., 1984) and reducing the rate of seed germination (Ministry of Agriculture and Cooperatives, Department of Agriculture, 2005). Phosphine fumigation, which is widely used, may become increasingly limited in use because the resistance of stored-grain insects to phosphine has now been discovered in more than 45 countries. In addition, phosphine has been argued to be genotoxic to occupationally exposed fumigators (Bell, 2000). Liquid fumigants, such as carbon tetrachloride or mixture of carbon disulphide, ethylene dibromide or ethylene dichloride, and carbon tetrachloride, are easier to handle as they are less toxic to man. The liquid is poured on the produce or left in trays to evaporate. Such a fumigation takes several days depending on the temperature and quantity of fumigant used (Sukprakarn, 1989).

In many storage systems, fumigants are the most economical and convenient tool for managing stored-grain insect pests not only because of their ability to kill a broad spectrum of pests but because of their easy penetration into the commodity while leaving minimal residues (Bell, 2000). Moreover, they have potential to kill all insects in a single application (Sukprakarn *et al.*, 1996). According to the constraints of the synthetic fumigants, we try to find the new safer fumigants from the natural products.

The synthetic standard compound we used in this study, Dichlorvos (dimethyl-2,2-dichlorovinyl phosphate, often referred to as DDVP), has high vapor pressure and high insecticidal activity in vapor phase comparing to other insecticides. It is widely used in household, food storage, for the treatment of companion animals and for the control stored-product pests. Dichlorvos has short residue stability, especially at higher temperatures and higher moisture levels. It is readily metabolized and rapidly excreted. Many researches usually use dichlorvos as the fumigant against the storedinsect pests (Lee *et al.*, 2003a; Rowlands, 1970).

1.2 Essential oils

Plant essential oils are secondary metabolites synthesized by plants. Plants containing essential oil belong to both Gymnospermae (Pines and Cycads) and Angiospermae (flowering plants). The essential oils are produced in glandular hair, schizogenous canals, ducts, cavities of heartwood, oil glands present in leaves, buds and in some cases, oleoresins. Essential oils occur commonly in the families Lamiaceae, Verbenaceae, Valerianaceae, Araliaceae, umbelliferae, Myrtaceae, Cistaceae, Violaceae, Thymelaceae, Anacardiaceae, Rutaceae, Burseraceae, Cneoraceae, Meliaceae, Magnoliaceae, Santalaceae, Betulaceae, Juglandaceae, Myricaceae, Salicaceae, Asteraceae, Ericaceae, Poaceae, Araceae, Pandanaceae, Cyperaceae, Zingiberaceae and Orchidaceae (Bhat, 2005). Many essential oils can be utilized as antiseptic agents, perfume industry, food additives and aromatherapy. Essential oils are abundant sources of terpenoids. They consist of a complex mixture of mono- and sesquiterpenes, alcohols, ketones, acids and esters. There are four general methods for the extraction of essential oils: expression, steam distillation, extraction with volatile solvents and resorption in purified fats (Ikan, 1969).

Classifications of essential oil

Monoterpenes: Monoterpenes are widespread and tend to occur as major components of essential oils (Ikan, 1969). They consist of two isoprene units. Their molecular formular is $C_{10}H_{16}$. The boiling point of monoterpenes is about 140-180°C (Ikan, 1969). The monoterpenes can be subdivided into three groups: acyclic, monocyclic and bicyclic (Ikan, 1969) as shown in Figure 1.4

ุ่ลฬาลงกรณ่มหาวิทยาลัย

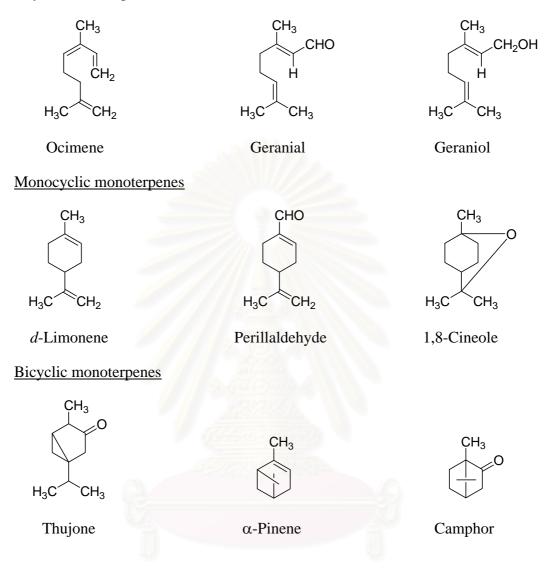


Figure 1.4 Some common monoterpenes found in essential oils (Ikan, 1969)



Sesquiterpenes: Sesquiterpenes are forming the higher-boiling fraction of essential oils. They are formed by the union of three isoprene units. Sesquiterpenes are unsaturated compounds and may be acyclic, monocyclic, bicyclic and tricyclic (Ikan, 1969) (Figure 1.5).

Acyclic sesquiterpenes

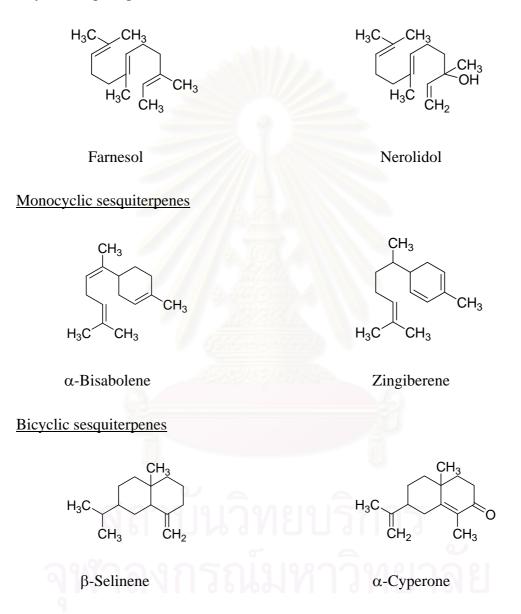


Figure 1.5 Some common sesquiterpenes found in essential oils (Ikan, 1969).

1.3 Fumigants from plants against S. oryzae (L.)

Several essential oils have been previously reported to effectively kill adult rice weevils, such as *Labiatae* sp. oil ZP51 (Shaaya *et al.*, 1997), *Mentha arvensis* L. var *piperascens* (Lee *et al.*, 2001a), eucalyptus, rosemary (Lee *et al.*, 2001b), *Foeniculum vulgare* fruit (Kim and Ahn, 2001), cinnamon, horseradish, mustard (Kim *et al.*, 2003), *Acorus gramineus* rhizome (Park *et al.*, 2003b), and plants in family Myrtaceae; *Eucalyptus nicholii, E. conodocarpa, E. blakelyi, Callistemon sieberi, Melaleuca fulgens*, and *M. armillaris* (Lee *et al.*, 2004a; Lee *et al.*, 2004b).

Moreover, the fumigant activity of the essential oil constituents was also reported, such as menthone (Lee *et al.*, 2001a), 1,8-cineole, benzaldehyde (Lee *et al.*, 2001b), (*E*)-anethole, estragole and (+)-fenchone (Kim and Ahn, 2001), *l*-fenchone, limonene and pulegone (Lee *et al.*, 2003a), and (*Z*)-asarone (Park *et al.*, 2003b). This study investigated the fumigant toxicity of various essential oils and their constituents from Thai plants towards *S. oryzae*. From these reports, oxygenated monoterpenoids tend to exhibit higher activity than hydrocarbons (Papachristos *et al.*, 2004). Ketone compounds were found more toxic than other monoterpenoids (Lee *et al.*, 2003a).

1.4 Rationale

The use of synthetic fumigants for managing the stored-product insect pest at the present time normally causes harmful effects to the environment and other animals. Essential oils are considered to be the natural fumigant controlling storedproduct insect pests because of their high volatility, toxicity to stored-grain insect pest, biodegradability and eco-friend. In this research, the fumigant activity of selected essential oils was examined against rice weevil, *Sitophilus oryzae* L and their major constituents were investigated comparing with the synthetic fumigant as standard.

1.5 The goals of this research

- 1. To study the fumigation activity of various essential oils against S. oryzae.
- 2. To study the fumigation activity of their major constituents against S. oryzae.
- 3. To analyze the residues of fumigants on the fumigated rice.

CHAPTER II

MATERIALS AND METHODS

2.1 Insects

Rice weevils were obtained from the colony maintained at the Department of Biology, Faculty of Science, Chulalongkorn University. Weevils were reared on the polished rice in the 8 ounce glass jar; 100 adult insects per 500 g of rice grain under conditions of 30 ± 2 °C, 70 ± 10 % relative humidity (R.H.) and a photoperiod of 12:12h (L:D). The rearing jars were sealed with the blotting paper to give air flow. Cultures were changed every two weeks and newly emerging adult rice weevils were transferred into a new jar containing fresh rice grain. The 7-14 days adults of emerging rice weevils were used as tested specimen.

2.2 Plant materials

Sources of plant samples and commercial-grade essential oils used in this study are presented in Table 2.1.

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

No	Family	Scientific name	Common name	Plant part
1	Annonaceae	Cananga odorata (Lamk.)	Ylang ylang	Flowers
		Hook. P. et Th.		
2	Graminae	Vertivevia zizanioides	Vertiver	Fibrous root
3	Illiciaceae	Illicium verum Hook.f.	Star anise	Fruits
4	Lamiaceae	Mentha arvensis Linn.	Japanese mint	Flowering
5		Rosemarinus officinalis Linn.	Rosemary	Flowering
				tops
6	Lauraceae	Cinnamomum cassia Presl.	Cinnamon	Leaves
7		Cinnamomum porrectum	Thep Ta-ro	Seeds
		(Roxb.) Kosterm		
8	Myrtaceae	Eugenia caryophyllata Thumb	Clove	Flowers
9				Leaves
10		Melaleuca cajuputi Powell	Cajeput	Leaves
11		Eucalyptus globules Labille	Eucalyptus	Leaves
12	Piperaceae	Piper betle Linn.	Betal vine	Leaves
13		Piper nigrum Linn.	Black pepper	Peppercorns
14	Rutaceae	Citrus hystrix DC.	Kaffir lime	Leaves
15				Fruits
16		Citrus aurantifolia Swing.	Lime	Fruits
17		Zanthoxylum limonella Alston	Ma khaen	Fruits
18		Citrus reticulata Blanco	Tangerine	Fruits
19	Scrophulariaceae	Limnophila aromatica Merr.	Phak khayaeng	Aerial part
			(Thai name)	
20	Zingiberaceae	Zingiber cassumunar Roxb	Plai	Rhizomes
			(Thai name)	
21		Curcuma spp.	Khamin dam	Rhizomes
			(Thai name)	
22		Zingiber spp.	Plai khaew	Rhizomes
			(Thai name)	

Table 2.1 Sources of essential oils

Table 2.1 (continued)

No	Family	Scientific name	Common name	Plant part
23	Zingiberaceae	Alpinia nigra (Gaertn.)	Ginger	Rhizomes
		B.L.Burtt		
24		Boesenbergia pandurata	Lesser galangal	Rhizomes
25		Curcuma domestica Valeton.	Turmeric	Rhizomes

Note: The essential oils No 1-6, 8-9, 11-16, 18 and 24-25 were obtained from Thai-China Flavours and Fragrances industry Co., Ltd., Nontaburi and those of No 7 and 10 were supported by Royal Forest Department, Bangkok. The rest essential oils were gained by hydrodistillation.

2.3 Chemicals

Merck's TLC (aluminium sheet, silica gel 60 F254 pre-coated 20×20 cm, layer thickness 0.2 mm) was used for the compound separation. All solvents were purified by distillation, except diethyl ether and acetone which were analytical grade. Certain monoterpenes were purchased from Fluka Chemies A.G. (Switzerland).

2.4 General procedures for hydrodistillation

Some essential oils were obtained by hydrodistillation (Dean-stark distillation) (Vogel, 1980). Each sample was finely chopped and put into a 1000 mL round bottom flask. The deionized water was added into the flask to about 500 mL. The flask was connected to the Dean-stark apparatus for hydrodistillation. The hydrodistillation was carried out for approximately 4 h or until no oil come out with the distillate. After cooling, the distillate was extracted by diethyl ether twice. After extraction, the solvent phase was dried over anhydrous sodium sulfate and was concentrated by rotary evaporator. The obtained essential oil was then collected and stored in the dark at 4°C to avoid the oxidation until being tested for the fumigant activity.

2.5 Fumigation bioassay

The fumigation bioassay was adapted from the method used in Park *et al.* (2003a). Ten 7-14 days adults *S. oryzae* were put into the $2.5 \times 2.5 \times 2.0$ cm plastic box together with 0.7 g of the polish rice. The mesh cloth was covered over the body part of the box by adhesive tape to prevent any direct contact of the weevils with the tested

compounds. Then, 50 μ L of the essential oil or pure compound solution diluted by acetone for each concentration was applied into the 2.3×2.3 cm filter paper (Whatman No.1, Whatman International Ltd., Maidstone, England). The applied paper was left out in the fume hood for a minute to evaporate the solvent. After that, the paper was placed on the mesh cloth and then the lid was closed immediately. The parafilm was bound down the connection between the box body and the lid to prevent the leaking of fumigant and bound cover again with the adhesive tape. Then, tested boxes were turned over to allow the fumigant paper lay in the bottom side. The control groups were the boxes which did not have any treatment (control A) and the boxes which contained only solvent (control B). All treatments were replicated three times. Those tested boxes were observed at the same conditions with the insect rearing conditions. The mortality was collected after 24 h exposure.

% mortality = $(O/T) \times 100$

O: Observed number of the dead S. oryzae

T: Total number of S. oryzae

The dead adult *S. oryzae* was taken into the account by shaking the box or prodding with paintbrush to observe the moving of appendages of the insect. If its appendages did not move, that insect was considered to be dead. It revealed that the insect died (Kim and Ahn, 2001)

2.6 Screening of essential oils for fumigation bioassay

Twenty-five essential oils from Thai plants were tested as fumigants at the concentration of 1000 μ L/L air as described above. The mortality percentage was observed after 24 h exposure.

The essential oils which caused 100% mortality were selected to test at lower concentrations (400 and 200 μ L/L air). After that, the LC₅₀ of the most potential essential oils were computed by Probit analysis (Finney, 1971).

2.7 Isolation of major constituents from the potent essential oils

A selected essential oil was fractionally distilled. Each fraction was collected upon boiling range and analyzed by gas chromatography (GC) comparing with the crude essential oil. The fractions containing major constituent were re-separated by column chromatography. The pure compounds were characterized by spectroscopic techniques including ¹H- and ¹³C- NMR and reconfirmed the purity by GC.

2.8 Spectroscopic analysis

The GC analysis was performed on Varian, CP-3800 gas chromatograph. The essential oil mixture was subjected to the GC analysis using CP-sil 5 column. The temperature program was increased from 50 to 65°C with a rate of 1.5°C per min as step 1 and from 65 to 160°C with a rate of 15°C per min as step 2. The injection temperature and the detector temperature was 250°C. The components of essential oil were determined by comparing with external standards.

The ¹H and ¹³C nuclear magnetic resonance including 2D-NMR experiments were carried out with a Jeol 400 MHz JNM-A500 FT-NMR spectrometer.

2.9 Fumigation test of pure compounds

The isolated constituents of the effective essential oils were tested for fumigation bioassay comparing with fumigant standard—dichlorvos at concentration of 100 μ L/L air. The exposure time was 24 h. The LC₅₀ of main components of each essential oil were determined, subsequently.

2.10 Residue determination

Residues in the polished rice were determined in menthol and safrole at 400 μ L/L air. Three groups of rice were compared. The first group was the rice sample mixed with fumigant directly as positive control. The second group was negative control fumigated with only acetone and the last group was the fumigated rice sample. Each group was sampled twice at pre-aeration and 24 h aeration in fume hood. The rice samples (2.1 g) were extracted with 5 mL acetone for 1 h as adapted from Lee *et al.* (2003b). Analysis of fumigant residue was performed by GC under the same

program as previously described in 2.8. The residue quantity was determined from calibration curve of each compound.

2.11 Statistical analysis

The percentage mortality was determined and transformed to arcsine square-root values for analysis of variance (ANOVA). Treatment means were compared and separated by Duncan's test at P = 0.05. The LC₅₀ was calculated by Probit analysis (Finney, 1971). All computations were done using the SPSS program. Means (± SE) of untransformed data are reported. Corrected mortality was calibrated by Abbott's formula as follows:

Corrected % mortality = $(T-C) \times 100$ 100 - C

T: % mortality of treatment group

C: % mortality of control group

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

CHAPTER III

RESULTS AND DISCUSSION

3.1 Hydrodistillation results

Some selected plants were obtained from hydrodistillation according to the procedure described in Chapter II. The results of hydrodistillation are presented in Table 3.1.

Table 3.1 The hydrodistillation of some selected plants

No	Scientific name	Plant part	plant weight [*]	Oil weight (g),
INU	Scientific name		(g)	(% w/w)
1	Alpinia nigra	Rhizomes	2,000 F	1.64 (0.08%)
2	Curcuma spp.	Rhizomes	1,300 F	1.67 (0.13%)
3	Limnophila aromatica	Aerial part	2,400 F	1.92 (0.08%)
4	Zanthoxylum limonella	Fruits	700 D	3.49 (0.50%)
5	Zingiber cassumunar	Rhizomes	3,000 F	13.25 (0.44%)
6	Zingiber spp.	Rhizomes	1,100 F	0.85 (0.08%)

* F = fresh weight and D = dry weight.

The hydrodistillation of some selected Thai plants yielded variable amounts of essential oils ranging from 0.08 to 0.50% (w/w) of fresh (or dry) weight. *A. nigra*, *L. aromatica* and *Zingiber* spp. provided the lowest amount whereas the highest amount of essential oil was achieved from the fruits of *Z. limonella*.

3.2 Screening of essential oils for fumigation bioassay

Six essential oils extracted as aforementioned, together with nineteen commercial essential oils were preliminarily screened for fumigant activity against *Sitophilus oryzae* at 1000 μ L/L air. The exposure time was at 24 h. The results are shown in Table 3.2.

No	Scientific name	Mean \pm SE**, $n = 3$	Toxicity Class***	
1	Cananga odorata	0.0 ± 0.0 ^a		
2	Vertivevia zizanioides	0.0 ± 0.0 $^{\mathrm{a}}$		
3	Piper betle	0.0 ± 0.0 a		
4	Zingiber spp.	0.0 ± 0.0 $^{\mathrm{a}}$	Low	
5	Curcuma domestica	0.0 ± 0.0 ^a		
6	Curcuma spp.	3.3 ± 3.3^{a}		
7	Limnophila aromatica	16.7 ± 12.0^{ab}		
8	Citrus reticulata	43.3 ± 21.9 bc	Medium	
9	Citrus hystrix (fruits)	53.3 ± 17.6 ^{cd}		
10	Cinnamomum cassia	63.3 ± 6.7 ^{cd}		
11	Piper nigrum	63.3 ± 12.0 ^{cd}		
12	Eugenia caryophellata (flowers)	$70.0 \pm 10.0^{\text{de}}$	Medium-high	
13	Zanthoxylum limonella	70.0 ± 15.3 de		
14	Citrus aurantifolia	73.3 ± 8.8^{de}		
15	Illicium verum	90.0 ± 5.8 ^{ef}		
16	Eugenia caryophellata (leaves)	90.0 ± 5.8 ^{ef}		
17	Melaleuca cajuputi	$93.3\pm6.7^{\rm \ f}$		
18	Boesenbergia pandurata	$93.3 \pm 3.3^{\rm f}$		
19	Mentha arvensis	100.0 ± 0.0 f		
20	Rosemarinus officinalis	$100.0\pm0.0~{\rm f}$	High	
21	Cinnamomum porrectum	$100.0\pm0.0^{\rm \ f}$		
22	Eucalyptus globules	$100.0\pm0.0^{\rm \ f}$		
23	Citrus hystrix (leaves)	$100.0\pm0.0~^{\rm f}$		
24	Zingiber cassumunar	$100.0\pm0.0~^{\rm f}$		
25	Alpinia nigra	$100.0\pm0.0^{\rm \ f}$		

Table 3.2 The fumigant activity of essential oils against *Sitophilus oryzae* adults atconcentration of 1000 μ L/L air after 24 h treatment.*

* Each datum represents the means of three replicates, each set up with 10 adults.

** Means within a column followed by the same letter are not significantly different at P = 0.05 (Duncan's test). Mortalities were transformed to arcsine square-root before ANOVA. Means (±SE) of untransformed data are reported.

*** The low toxicity class is < 25 % mortality; the medium toxicity class is 25-50 % mortality; the medium-high toxicity class is 51-74 % mortality and the high toxicity class is \geq 75 % mortality.

From Table 3.2, twenty-five essential oils from Thai species were screened against *S. oryzae* by a fumigation method at 1000 μ L/L air and %mortality was observed after 24 h treatment. Eleven essential oils: *I. verum*, *E. caryophellata* (leaves), *M. cajuputi*, *B. pandurata*, *M. arvensis*, *R. officinalis*, *C. porrectum*, *E. globules*, *C. hystrix*, *Z. cassumunar* and *A. nigra*, caused mortality in *S. oryzae* higher than 75%, ranging from 90-100%. This group exhibited high significant potent toward *S. oryzae*, thus was selected for further testing at the lower concentration of 400 μ L/L air. The results are presented in Table 3.3.

Table 3.3 The fumigant activity of essential oils against *Sitophilus oryzae* adults at concentration of 400 μL/L air after 24 h treatment.*

No	Scientific name	Mean \pm SE**, $n = 3$	Toxicity Class***
1	Melaleuca cajuputi	17.8 ± 9.1^{ab}	Low
2	Alpinia nigra	30.0 ± 15.8 ^{ab}	
3	Boesenbergia pandurata	31.1 ± 23.3 ^{ab}	Medium
4	Rosemarinus officinalis	45.3 ± 23.2 ^{ab}	
5	Citrus hystrix (leaves)	56.9 ± 3.2 ^{ab}	
6	Eugenia caryophellata (leaves)	$57.2 \pm 3.9^{\ ab}$	
7	Eucalyptus globules	68.9 ± 19.4 bc	Medium-high
8	Illicium verum	69.5 ± 6.3 bc	
9	Mentha arvensis	100.0 ± 0.0 ^c	
10	Cinnamomum porrectum	100.0 ± 0.0 ^c	High
11	Zingiber cassumunar	100.0 ± 0.0 $^{\rm c}$	

* Each datum represents the means of three replicates, each set up with 10 adults.
** Means within a column followed by the same letter are not significantly different at P = 0.05 (Duncan's test). Mortalities were transformed to arcsine square-root before ANOVA. Means (±SE) of untransformed data are reported.

*** The low toxicity class is < 25 % mortality; the medium toxicity class is 25-50 % mortality; the medium-high toxicity class is 51-74 % mortality and the high toxicity class is \geq 75 % mortality.

From Table 3.3, the essential oils from *M. arvensis, C. porrectum*, and *Z. cassumunar* resulted in 100% mortality after 24 h exposure. Five essential oils: *M. arvensis, C. porrectum, Z. cassumunar, I. verum* and *E. globules* were further selected for testing at lower concentration of 200 μ L/L air. These were classified in the medium-high and the high class of toxicity. The results are presented in Table 3.4.

Table 3.4 The fumigant activity of essential oils against *Sitophilus oryzae* adults atconcentration of 200 μ L/L air after 24 h treatment.*

No	Scientific name	$Mean \pm SE^{**}, n = 3$	Toxicity Class***	
1	Eucalyptus globules	$0.0 \pm 0.0^{\ a}$	Ŧ	
2	Illicium verum	20.0 ± 0.0^{a}	Low	
3	Zingiber cassumunar	76.7 ± 23.3 ^b		
4	Cinnamomum porrectum	83.3 ± 3.3^{b}	High	
5	Mentha arvensis	100.0 ± 0.0 ^b		

* Each datum represents the means of three replicates, each set up with 10 adults.

** Means within a column followed by the same letter are not significantly different at P = 0.05 (Duncan's test). Mortalities were transformed to arcsine square-root before ANOVA. Means (±SE) of untransformed data are reported.

*** The low toxicity class is < 25 % mortality; the medium toxicity class is 25-50 % mortality; the medium-high toxicity class is 51-74 % mortality and the high toxicity class is \geq 75 % mortality.

From fumigation testing at concentration of 200 μ L/L air, the results showed that three essential oils from *Z. cassumunar*, *C. porrectum* and *M. arvensis* exhibited the highest activity toward *S. oryzae* at exposure time 24 h. The essential oil from *M. arvensis* revealed the complete mortality while those of *Z. cassumunar* and *C. porrectum* showed 76.7 % and 83.3 % mortality, respectively. These three essential oils were categorized in the high toxicity class.

3.3 LC₅₀ determination

The concentration of effective essential oils which caused 50 % mortality of *S. oryzae* was investigated by fumigation bioassay. LC_{50} was estimated by Probit analysis (Finney, 1971). Probit linear regression curves of three essential oils are shown in Figure 3.1.

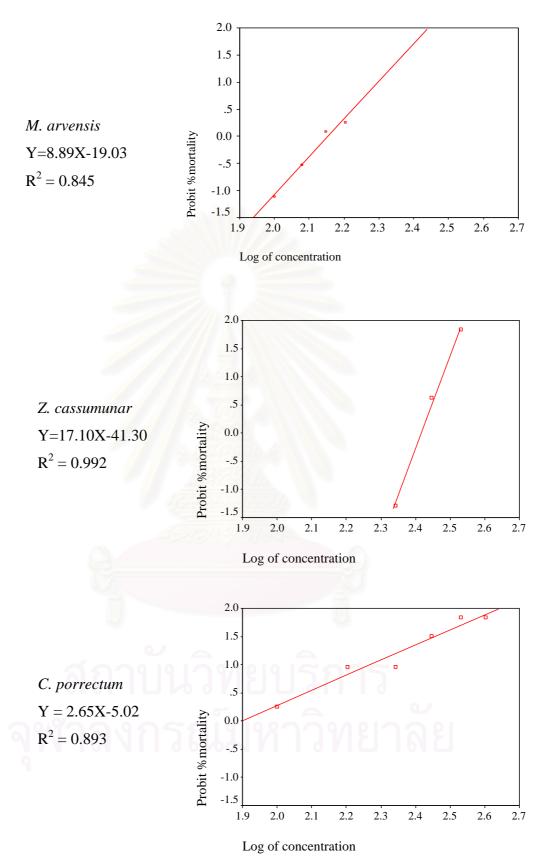


Figure 3.1 Linear regressions of concentrations of three highest potent essential oils against *S. oryzae* at 24 h exposure.

According to the LC₅₀ analysis curves (Figure 3.1), the concentration causing 50% mortality of *S. oryzae* after 24 h exposure and at 95% of confidential limit (*P*=0.05) of the essential oils from *M. arvensis*, *Z. cassumunar* and *C. porrectum* were 138, 260 and 79 μ L/L air, respectively. Their regression equations of mortality were Y=8.89X-19.03, Y=17.10X-41.30 and Y=2.65X-5.02, respectively. The higher value of LC₅₀ showed the lower efficiency of that compound. This result indicated that the essential oil from *C. porrectum* exhibited the highest activity against *S. oryzae* while that from *Z. cassumunar* displayed the lowest one.

The fumigant activity against S. oryzae of essential oil from M. arvensis has been reported by Lee et al. (2001a) and Singh et al. (1995), while essential oils from C. porrectum and Z. cassumunar had not been previously reported. Their studies reported that essential oil from *M. arvensis* gave high potent in fumigation against S. oryzae at 24 h exposure time. Its LC₅₀ values were varied from 45.5 μ L/L air (Lee et al., 2001a) to 229.8 µL/L air (Singh et al., 1995). The LC₅₀ of essential oil from M. arvensis in our study were higher than that in the report of Lee et al. (2001a) but comparable to LC_{50} from the study of Singh *et al.* (1995) The reasons may be from different strain of S. oryzae or different in major constituents in M. arvensis essential oil. This discrepancy was comparable to that reported by Lee et al. (2000) that Oryzaephilus surinamensis L., a chlorpyrifos-methyl-resistant strain, was crossresistant to essential oil from Eucalyptus globules Labill and its primary monoterpenes, 1,8-cineole. Compared to our study, the essential oils from E. globules and Rosemary officinalis which were rich in 1,8-cineole gave high value of LC₅₀ (non effective). Another reason may derive from the difference in major constituents of essential oil. Thus, it is essential to examine fumigant activity of major constituents against S. oryzae.

3.4 Essential oil analysis

Essential oil was generally recognized to consist of a mixture of various compounds, particularly, terpenes (Ikan, 1969). Thus, it is important to analyze the active principle responsible for the interest biological activity. In this research, three effective essential oils are selected to examine for active ingredients effecting on *S. oryzae*. The GC and ¹H-NMR techniques were exploited.

3.4.1 The essential oil from M. arvensis

The gas chromatogram (Figure 3.2) of the essential oil from *M. arvensis* revealed the presence of two major components at retention times of 20.54 and 19.57 min, respectively. The comparison of the retention times of each component with those of standards revealed that these two major compounds were menthol and menthone, respectively. % Composition is displayed in Table 3.5.

Table 3.5 The compositions of essential oil from A	М.	arvensis
--	----	----------

Component	Retention time (min)	% composition
Menthol	20.54	76.64
Menthone	19.57	8.34

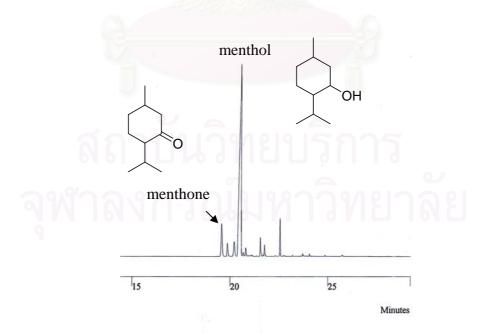


Figure 3.2 The gas chromatogram of the essential oil from *M. arvensis*

3.4.2 The essential oil from Z. cassumunar

The gas chromatogram (Figure 3.3) of the essential oil from *Z. cassumunar* revealed two major components at the retention times of 8.85 and 20.52 min. After comparing with authentic samples, the former was identified as sabinene, while the latter was terpinen-4-ol. The % composition is displayed in Table 3.6.

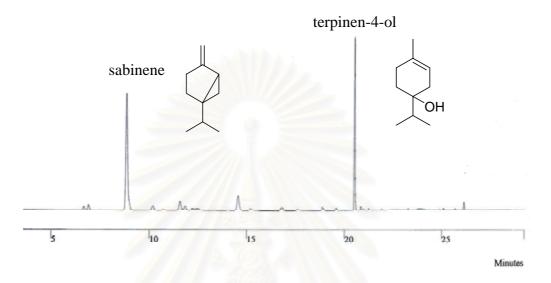


Figure 3.3 The gas chromatogram of the essential oil from Z. cassumunar

Component	Retention time (min)	% composition
Sabinene	8.85	47.36
Terpinen-4-ol	20.52	31.69

Table 3.6 The composition of the essential oil from Z. cassumunar

The essential oil from *Z. cassumunar* (450 g) was further fractionally distilled. Three fractions: 178 g, 86 g and 145 g with the boiling range of 160-170°, 170-200° and > 200°C were collected. The first fraction contained mainly sabinene according to the comparison with the standard compound. This fraction was carefully redistilled to collect the distillate with boiling range of 163-165°C. After analyzed by GC, more than 95% sabinene was obtained. The identity of this compound was confirmed by ¹H-NMR as shown in Figure 3.4 and Table 3.7.

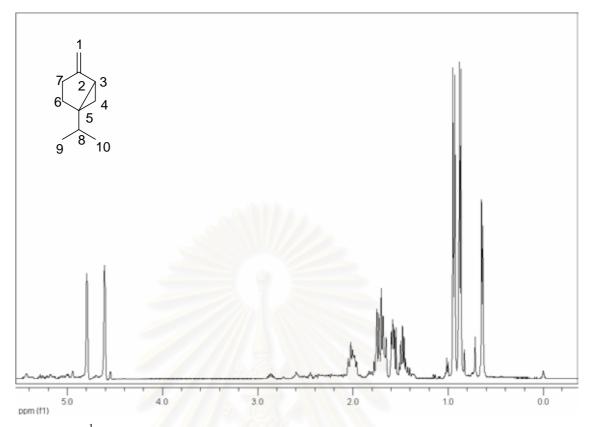


Figure 3.4 ¹H-NMR spectrum of compound 1 (sabinene)

Position	Chemical shift (ppm)					
1 0510011	Compound 1	Sabinene ^a				
1	4.79 (1H, s)	4.73, 4.54 (2H, C=CH ₂)				
	4.61 (1H, s)					
3	1.59-1.54 (1H, m)	A -				
4	0.64 (2H, d, <i>J</i> =5.5 Hz)	0.66, 0.60 (2H, C-6 H's)				
6	1.78-1.65 (2H, m)	<u> </u>				
7	2.05-1.96 (1H, m)	าวทยาลย				
8	1.51-1.43 (1H, m)					
9	0.94 (3H, d, <i>J</i> =6.9 Hz)	0.90 (3H, d, <i>J</i> =6.0 Hz, CH-CH ₃)				
10	0.87 (3H, d, <i>J</i> =6.9 Hz)	0.82 (3H, d, <i>J</i> =7.0 Hz, CH-CH ₃)				

Table 3.7 The ¹H-NMR chemical shift assignments of sabinene.

^a Reference from Fanta and Erman (1968).

The second fraction containing a mixture of compounds was analyzed by GC. This fraction was therefore not examined further for the constituents in details. The last fraction (50 g) was subjected to silica gel column chromatography using 1 : 1.5 of a mixture of CH₂Cl₂-hexane as an eluent to obtain the pure compound exhibiting on TLC at R_f 0.40 (50% hexane: CH₂Cl₂). The confirmation of this compound was performed by GC as terpinen-4-ol. The ¹H-NMR spectrum is displayed in Figure 3.5 and Table 3.8.

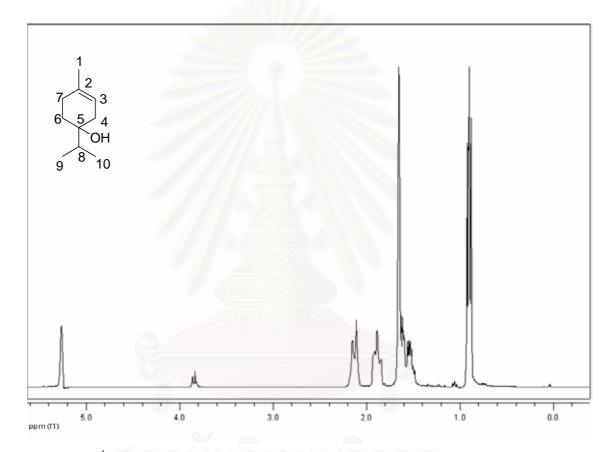


Figure 3.5 ¹H-NMR spectrum of compound 2 (terpinen-4-ol)

จุฬาลงกรณ์มหาวิทยาลย

Position	Chemical shift (ppm)					
rosition	Compound 2	Terpinen-4-ol ^a				
1	1.65 (3H, s)	1.70 (3H, s)				
3	5.27 (1H, s)	5.31 (1H, dd, <i>J</i> =2.3, 0.7 Hz				
4	2.13 (2H, d, <i>J</i> =15.8 Hz)	-				
6	1.56-1.49 (2H, m)	-				
7	1.92-1.85 (2H, m)	-				
8	1.63-1.60 (1H, m)					
9, 10	0.90 (6H, dd, <i>J</i> =7.0, 10.0 Hz)	0.96 (3H, d, <i>J</i> =6.9 Hz)				
		0.93 (3H, d, <i>J</i> =6.9 Hz)				

Table 3.8 The ¹H-NMR chemical shift assignments of terpinen-4-ol.

^aReference from Ngo and Brown (1998)

3.4.3 The essential oil from C. porrectum

The gas chromatogram of the essential oil from *C. porrectum* revealed that a major component presented at the retention time of 22.40 min with 65.70% composition (Figure 3.6). The major peak was compared with authentic sample and found that it gave the same retention time as safrole. This essential oil was further purified by silica gel column chromatograph eluting with 1 : 1 of CH_2Cl_2 : hexane. The main compound, safrole was confirmed its identity by ¹H-NMR (Figure 3.7 and Table 3.9).

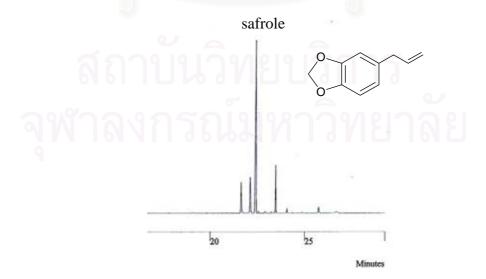


Figure 3.6 The gas chromatogram of the essential oil from C. porrectum

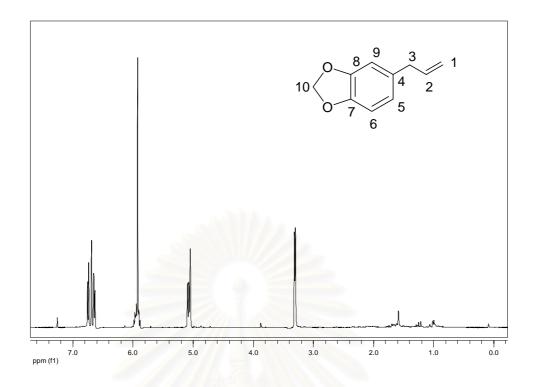


Figure 3.7 ¹H-NMR spectrum of safrole.

Position _	Chemical shift (ppm)					
	safrole	safrole ^a				
1	5.10-5.05 (2H, m)	5.10 (2H, m)				
2	5.99-5.89 (1H, m)	5.96 (1H, m)				
3	3.31 (2H, d, <i>J</i> =6.6 Hz)	3.34 (2H, d, <i>J</i> =6.7 Hz)				
5	6.50 (1H, d, <i>J</i> =7.8 Hz)	6.67 (1H, dd, <i>J</i> =7.9, 1.7 Hz)				
6	6.75 (1H, d, <i>J</i> =7.9 Hz)	6.78 (1H, d, <i>J</i> =7.9 Hz)				
9 6	6.69 (1H, s)	6.72 (1H, d, <i>J</i> =1.4 Hz)				
10	5.92 (2H, s)	5.95 (2H, s, O-CH ₂ -O)				

Table 3.9 The ¹H-NMR chemical shift assignments of safrole.

^a Reference from Mohottalage *et al.* (2007)

3.5 Fumigation test of constituents

Five main constituents found in the three effective essential oils, including ten commercial monoterpenoids consisted of each essential oil (Pappas, 2006; Bruneton, 1995), were tested against *S. oryzae* in fumigation bioassay described above in Chapter II. The concentration of 100 μ L/L air was selected for this test. The synthetic

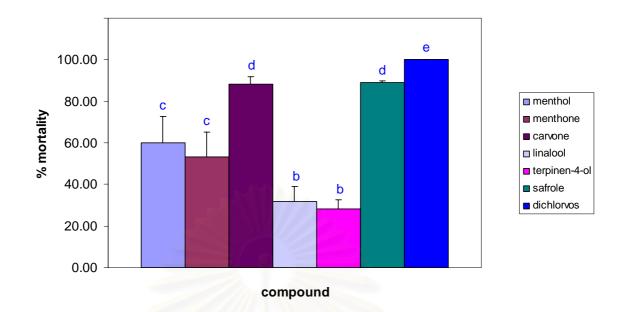
Compound	Mean \pm SE**, $n = 3$	Toxicity Class***		
1. menthol	60.0 ± 12.6 ^c	Medium-high		
2. menthone	53.3 ± 11.7 ^c			
3. limonene	0.0 ± 0.0 ^a	Low		
4. 1,8-cineole	0.0 ± 0.0 a	Low		
5. carvone	88.3 ± 3.3 ^d	High		
6. β-pinene	0.0 ± 0.0 ^a	Low		
7. α-pinene	0.0 ± 0.0 a	Low		
8. linalool	31.7 ± 7.3 ^b	Medium		
9. sabinene	0.0 ± 0.0 ^a	Low		
10. terpinen-4-ol	28.3 ± 4.4 ^b	Medium		
11. α-terpinene	0.0 ± 0.0 ^a			
12. γ-terpinene	0.0 ± 0.0 ^a	Low		
13. <i>p</i> -cymene	0.0 ± 0.0 $^{\mathrm{a}}$	Low		
14. terpinolene	0.0 ± 0.0 $^{\mathrm{a}}$			
15. safrole	88.9 ± 1.1 ^d	Iliah		
16. dichlorvos	100.0 ± 0.0^{e}	High		

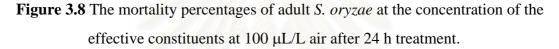
Table 3.10 The fumigant activity of essential oils against *Sitophilus oryzae* adults atthe concentration of constituents at 100 μ L/L air after 24 h treatment.*

* Each datum represents the means of three replicates, each set up with 10 adults.

** Means within a column followed by the same letter are not significantly different at P = 0.05 (Duncan's test). Mortalities were transformed to arcsine square-root before ANOVA. Means (\pm SE) of untransformed data are reported.

*** The low toxicity class is < 25 % mortality; the medium toxicity class is 25-50 % mortality; the medium-high toxicity class is 51-74 % mortality and the high toxicity class is \geq 75 % mortality.





From Table 3.10, the compounds number 1-8 were found in essential oil from *M. arvensis* (Bruneton, 1995) and compounds number 5, 7, 8 and 9-14 were found in essential oil from *Z. cassumunar* (Pappas, 2006). The structures of these compounds are shown in Figure 3.9.



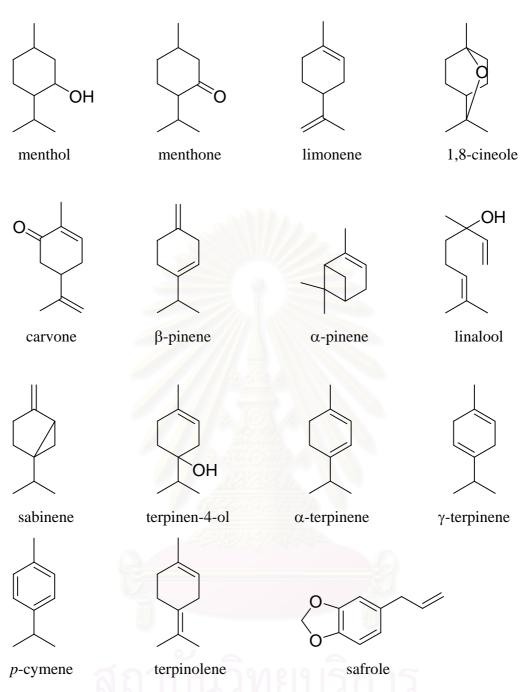
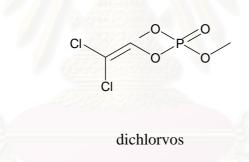


Figure 3.9 Structures of compounds used in this fumigation test

The results from Table 3.10 and Figure 3.8 showed the concentration of 100 μ L/L air of safrole gave the highest mortality of *S. oryzae* among natural chemicals. In the other hand, safrole was not significantly different to carvone and showed activity against *S. oryzae* less than the synthetic fumigant, dichlorvos. Three compounds, safrole, carvone and dichlorvos, showed high toxicity, while limonene, 1,8-cineole, β-pinene, α-pinene, sabinene, α-terpinene, γ-terpinene, *p*-cymene and

terpinolene appeared non toxic to *S. oryzae* in fumigation. These results were comparable to the report of Papachristos *et al.* (2004) that oxygenated monoterpenoids exhibited higher activity than hydrocarbons. Six active compounds in this study (menthol, menthone, carvone, linalool, terpinen-4-ol and safrole) showed oxygen atom in their structures, especially safrole and dichlorvos which have two and four atom of oxygen, respectively and caused the higher mortality. It was possible that much more number of oxygen atoms in the compound structure might cause higher mortality of insect, so it was interested to further study about this point.

Moreover, there are four constituents (menthol, menthone, carvone and linalool) of essential oil from *M. arvensis* effecting toward *S. oryzae*, while only one constituent (terpinen-4-ol) of essential oil from *Z. cassumunar* effected to this insect. Thus, those resulted in the essential oil from *M. arvensis* had higher effective than *Z. cassumunar* oil. The reason might be from the synergistic phenomena of oxygenated monoterpenoids containing in the essential oil (Papachristos *et al.*, 2004)



Subsequently, the main constituents of each essential oil were evaluated LC_{50} against *S. oryzae* and compared the effect of first major component and second major component toward *S. oryzae*. The result is shown in Table 3.11.





LC₅₀ Concentration (µL/L air) compound 50 100 160 220 700 (µL/L air) 280 M. arvensis 1. menthol 95.4 0.0 ± 0.0 60.0 ± 12.6 86.7 ± 6.7 93.3 ± 6.7 100.0 ± 0.0 100.0 ± 0.0 2. menthone 3.33 ± 3.3 53.3 ± 11.7 60.0 ± 5.8 93.3 ± 3.3 100.0 ± 0.0 100.0 ± 0.0 104.8 Z. cassumunar 1. sabinene 0.0 ± 0.0 0.0 ± 0.0 0.0 ± 0.0 0.0 ± 0.0 0.0 ± 0.0 0.0 ± 0.0 >700 2. terpinen-4-ol 30.0 ± 11.5 0.0 ± 0.0 53.3 ± 8.8 53.3 ± 6.7 83.3 ± 3.3 100.0 ± 0.0 165.5 *C. porrectum* safrole 46.7 ± 3.3 83.3 ± 3.3 86.7 ± 6.7 96.7 ± 3.3 100.0 ± 0.0 100.0 ± 0.0 57.9

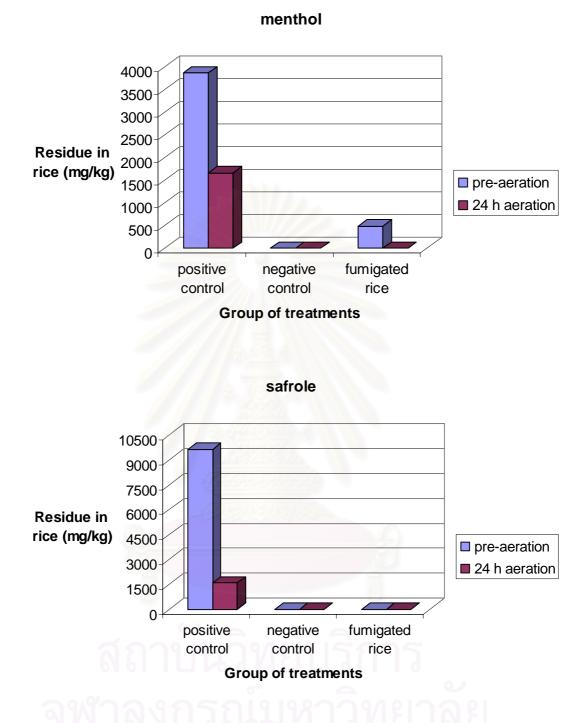
Table 3.11 LC₅₀ values of main components in three essential oils against *S. oryzae*.

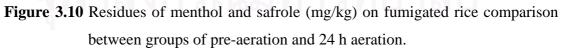


Safrole showed the highest effective against *S. oryzae* with LC₅₀ of 57.9 μ L/L air. Menthol, the first major component of *M. arvensis* essential oil, appeared non significant higher toxicity than menthone with LC₅₀ of 95.4 and 104.8 μ L/L air, respectively, while terpinen-4-ol, the second major constituent of *Z. cassumunar* oil caused significant higher mortality than sabinene (Duncan's test, *P* = 0.05). This result is in contrary with Lee *et al.* (2001a). Our result indicated that the major constituent was not necessarily to be the active ingredient of the essential oil.

3.6 Residue analysis

The levels of menthol and safrole residues in fumigated rice and control groups are shown in Figure 3.10. The residues of both fumigants were completely removed within 24 h aeration. In comparison with menthol, safrole left the lowest residue on the fumigated rice at pre-aeration. It was possible that safrole had lower absorption on surface rice. The results are consistent with those of wheat fumigated with 1,8-cineole (Lee et al., 2001a). European Commission 2002 (2002) reported the acute oral LD₅₀ of safrole for rats and mice to be 1950 and 2350 mg/kg of body weight, respectively. These indicated that safrole have low toxicity to mammals according to United States Environmental Protection Agency (2007). Additionally, maximum levels for safrole in foodstuffs and beverages can be added 1 mg/kg and the estimated average intake of safrole (for consumers) was assumed to be 1 mg/person/day (European Commission 2002, 2002). Thus, our study revealed that using safrole as fumigant for controlling S. oryzae should be safe to consumers because of no residues of safrole on rice after aeration for 24 h. In summary, safrole showed potential as the best alternative fumigant toward S. oryzae because it had more potential than other monoterpenes and in combination with low residue on the rice commodity.





CHAPTER IV

CONCLUSION

During the course of this research, the fumigant activity of twenty-five essential oils from Thai plants was screened against a stored-product insect pest, *Sitophilus oryzae* Linn (Rice weevil). As the results, three essential oils from *Cinnamomum porrectum* (Roxb) Kosterm (Thep ta-ro) exhibited highest fumigant toxicity against adults of *S. oryzae* after 24 h exposure, followed by those from *Mentha arvensis* Linn (Japanese mint) and *Zingiber cassumunar* Roxb (plai), respectively. Their LC₅₀ values ranged from 79 to 260 μ L/L air.

Moreover, chemical constituents and fumigant activity of each active essential oil were examined, as well as, the active components in each active essential oil were identified, fractionalized and purified. The essential oil from *M. arvensis* was found to be rich in menthol and followed by menthone, while essential oil from *Z. cassumunar* was rich in sabinene and terpinen-4-ol, and safrole appeared to be the largest component of essential oil from *C. porrectum*. The results of fumigation bioassay of those components revealed that safrole exhibited the most potent fumigant against *S. oryzae* at concentration of 100 μ L/L air after 24 h exposure.

The main component and its fumigant activity relationship were compared for mint and plai. The results revealed that menthol, the first major constituent, is active component of essential oil from *M. arvensis*, while terpinen-4-ol, the second major constituent of essential oil from *Z. cassumunar*, is its active component. Therefore, some major constituent was not necessarily to be the active ingredient.

In addition, the structures of compounds also affected to fumigant activity. Other essential oil's compound test indicated that the most of oxygenated monoterpenoids were more effective on *S. oryzae* than hydrocarbons. These effective oxygenated compounds were menthol, menthone, carvone, linalool, terpinen-4-ol and safrole. They tended to be much more effective depended upon the number of oxygen atom in their structures.

Residue analysis of two monoterpenes (menthol and safrole) revealed that the fumigated polished rice, which was not aerated, cannot be detected any residue of safrole. However, both of them did not leave residues on the fumigated polished rice after 24 h aeration.

In summary, safrole and its essential oil from *C. porrectum* showed promise as the best alternative safety fumigants toward *S. oryzae* because it had more potential than other monoterpenes and together with lower residue on the rice commodity.

Proposal for future work

This research provided the screening of the fumigant activity of crude essential oils. Further studies should involve the elucidation of all active constituents from active crude essential oils, modify structure and thoroughly study on structure activity relationship. Moreover, mode of action of essential oil on *S. oryzae* has not been illuminated. In addition the essential oils are not as broad spectrum as synthetic fumigants, but their efficacy can be improved by using them in conjunction with carefully designed packaging and developing new application methodology such as control release or encapsulation or synthesizing their active components as safely commercial fumigants.

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

REFERENCES

Bell, C. H. 2000. Fumigation in the 21st century. Crop Protection 19: 563-569.

- Bhat, S. V., Nagaampagi, B. A. and Sivakumar, M. 2005. Chemistry of Natural Products. Narosa publishing house, New Delhi, India: 840 pp.
- Bruneton, J. 1995. **Pharmacognosy, phytochemistry, medicinal Plants**. Paris: Lavoisier publishing.
- Chamratpan, S., Homchuen, S. 2005. Ethnobotany in upper northeastern Thailand.
 In Bernath, J., Nemeth, E., Craker, L. E. and Gardner, Z. E. (eds.), ISHS Acta
 Horticulturae 675: III WOCMAP Congress on Medicinal and Aromatic
 Plants-volume 1: Bioprospecting and Ethnopharmacology, Chiangmai,
 Thailand, 1 February 2005.
- Dansi, L., Van Velson, F. L., Vander Heuden, C. A. 1984. Methyl bromide: carcinogenic effects in the rat fore stomach. Toxicology Applied Pharmacology 72: 262-271.
- European Commission 2002. 2002. Opinion of the Scientific committee on food on the safety of the presence of safrole (1-allyl-3,4-methylene dioxy benzene) in flavourings and other food ingredients with flavouring properties. Scientific Committee on Food, Belgium, 9 January 2002.
- Fanta, W. I. and Erman, W. F. 1968. Total synthesis of *dl*-sabinene, *dl-trans*sabinene hydrate, and related monoterpenes. The Journal of Organic Chemistry 33: 1655-1658.
- Finney, D. J. 1971. **Probit analysis**. 3rd edition. London: Cambridge University Press.

- Hayashi, T., Nakamura, S., Visarathanonth, P., Uraichuen, J. and Kengkanpanich, R.
 2004. Stored rice insect pests and their natural enemies in Thailand.
 JIRCAS International Agricultural Series No. 13. Bangkok: Funny publishing.
- Ikan, R. 1969. Natural Product: a Laboratory Guide. London: Academic Press.
- Kim, D. H. and Ahn, Y. J. 2001. Contact and fumigant activities of constituents of *Foeniculum vulgare* fruit against three coleopteran stored-product insects.
 Pest Management Science 57: 301-306.
- Kim, S. I., Roh, J. Y., Kim, D. H., Lee, H. S. and Ahn, Y. J. 2003. Insecticidal activities of aromatic plant extracts and essential oils against *sitophilus oryzae* and *Callosobruchus chinensis*. Journal of Stored Products Research 39: 293-303.
- Lee, S. E., Choi, W. S., Lee, H. S. and Park, B. S. 2000. Cross-resistance of a chlorpyrifos-methyl of *Oryzaephilus surinamensis* (Coleoptera: Cucujidae) to fumigant toxicity of essential oil extracted from *Eucalyptus globules* and its major monoterpene, 1,8-cineole. Journal of Stored Product Research 36: 383-389.
- Lee, S. E., Lee, B. H., Choi, W. S., Park, B. S., Kim, J. G. and Campbell, B. C.
 2001a. Fumigant toxicity of volatile natural products from Korean spices and medicinal plants towards the rice weevil, *Sitophilus oryzae* (L). Pest
 Management Science 57: 548-553.
- Lee, B. H., Choi, W. S., Lee, S. E. and Park, B. S. 2001b. Fumigant toxicity of essential oils and their constituent compounds towards the rice weevil, *Sitophilus oryzae* (L.). Crop Protection 20: 317-320.

- Lee, S., Peterson, C. J. and Coats, J. R. 2003a. Fumigation toxicity of monoterpenoids to several stored product insects. Journal of Stored Products Research 39: 77-85.
- Lee, B. H., Annis, P. C. and Tumaalii, F. 2003b. The potential of 1,8-cineole as a fumigant for stored wheat. In Wright, E. J., Webb, M. C. and Highley, E. (eds.), Stored grain in Australia 2003. 230-234. Proceeding of the Australian postharvest technical Conference, Canberra, 25-27 June 2003.
- Lee, B. H., Annis, P. C., Tumaalii, F. and Choi, W. S. 2004a. Fumigant toxicity of essential oils from the Myrtaceae family and 1,8-cineole against 3 major stored-grain insects. Journal of Stored Products Research 40: 553-564.
- Lee, B. H., Annis, P. C., Tumaalii, F. and Lee, S. E. 2004b. Fumigant toxicity of *Eucalyptus blakelyi* and *Melaleuca fulgens* essential oils and 1,8-cineole against different development stages of the rice weevil *Sitophilus oryzae*.
 Phytoparasitica 32: 498-506.
- Ministry of Agriculture and Cooperatives, Department of Agriculture. 2002.
 Suggestions for management of animal and insect pests 2545. 13th edition.
 Bangkok: The printing house of the Agricultural Co-operative Federation of Thailand. (Printed in Thai).
- Ministry of Agriculture and Cooperatives, Rice Department. 2006. **Rice Knowledge Bank**[online]. Available from: <u>http://www.ricethailand.go.th/rkb/data_005/rice_xx2-05_bug19.html</u>[2006, August 21].
- Mohottalage, S., Tabacchi, R. and Guerin, P. M. 2007. Components from Sri Lankan *Piper betle* L. leaf oil and their analogues showing toxicity against the housefly, *Musca domestica*. Flavour and Fragrance Journal 22: 130-138.

- Munakata, K. 1970. Insect Antifeedants in Plants. In D. L. Wood, R. M. Silverstein and M. Nakajima (eds.), Control of insect behavior by natural products. New York: Academic Press.
- Mutambuki, K. 2004. Reference manual on: The major insect pests of stored cereal and pulse grains in Somalia and their control. In A. J. Harberd (ed.),
 Integrated pest management project in Somalia[online]. Available from Email: <u>IPMNairobi@una.org</u>.
- Naunwat, K., Visarathanonth, P., Chankaewmanee, B., Uraichuen, C., Kengkanpanich, R., Pengkum, K. and Tongpan, J. 2005. Paddy rice insect pests and management. Bangkok: The printing house of the Agricultural Cooperative Federation of Thailand. (Printed in Thai).
- Ngo, K. S. and Brown, G. D. 1998. Stilbenes, monoterpenes, diarylheptanoids, labdanes and chalcones from *Alpinia katsumadai*. Phytochemistry 47: 1117-1123.
- Palanuvej, C., Werawatganone, P. and Ruangrungsi, N. 2005. Chemical composition and antifungal activity of essential oil from the leaves of *Cinnamomum porrectum*. 31st Congress on science and technology of Thailand at Suranaree University of Technology, 18-20 October 2005.
- Papachristos, D. P., Karamanoli, K. I., Stamopoulos, D. C. and Spiroudi, U. M. 2004. The relationship between the chemical composition of three essential oils and their insecticidal activity against *Acanthoscelides obtectus* (Say). Pest management science 60: 514-520.
- Pappas, R. S. 2006. Plai. In: Applied Essential Oil Research[online]. Available from: <u>www.essentialoils.org/plai.htm.</u> [2006, October 23].
- Park, I. K., Lee, S. G., Choi, D. H., Park, J. D. and Ahn, Y. J. 2003a. Insecticidal activities of constituents identified in the essential oil from leaves of

Chamaecyparis obtusa against *Callosobruchus chinensis* (L.) and *Sitophilus oryzae* (L.). Journal of Stored Products Research 39: 375-384.

- Park, C., Kim, S. I. and Ahn, Y. J. 2003b. Insecticidal activity of asarones identified in *Acorus gramineus* rhizome against three coleopteran stored-product insects. Journal of Stored Products Research 39: 333-342.
- Rees, D. 2004. Insects of stored products. Australia: CSIRO Publishing.
- Rowlands, D. G. 1970. The metabolic fate of dichlorvos on stored wheat grains. Journal of Stored Products Research 6: 19-32.
- Shaaya, E., Kostjukovski, M., Eilberg, J. and Sukprakarn, C. 1997. Plant oils as fumigants and contact insecticides for the control of stored-product insects.
 Journal of Stored Products Research 33: 7-15.
- Silverstein, R. M., Bassler, G. C. and Morrill, T. C. 1981. Spectrometric identification of organic compounds. 4th edition. Canada: John Wiley & Sons.
- Singh, M., Srivastava, R. P. and Chauhan, S. S. 1995. Effect of Japanese mint (*Mentha arvensis*) oil as fumigant on nutritional quality of stored sorghum.
 Plant Foods for Human Nutrition 47: 109-114.
- Sukprakarn, C. 1989. Insect pests of stored products. In R. L. Semple, A. S. Frio, P. A. Hicks, and J. V. Lozare (eds.), Mycotoxin prevention and control in foodgrains. Bangkok: RAP Publication.
- Sukprakarn, C., Naunwat, K., Nilpanich, P., Visarathanonth, P., Chankaewmanee, B., Uraichuen, C. and Kengkanpanich, R. 1996. Stored product insect pests and management. 2nd edition. Bangkok: Funny publishing. (Printed in Thai).

- Varma, J. and Dubey, N. K. 2001. Efficacy of essential oils of *Caesulia axillaria* and *Mentha arvensis* against some storage pests causing biodeterioration of food commodities. International Journal of Food Microbiology 68: 207-210.
- Vogel, A. I. 1980. **Textbook of practical organic chemistry**. 14th edition. London: English Language Book Society.
- World Meteorological Organization (WMO). 1995. Methyl bromide. Environmental Health Criteria 166. WMO, Geneva.
- Yuenyongsawad, S. and Kummee, S. 2004. Composition and antimicrobial evaluation of volatile oil from *Cinnamomum porrectum* stem. Available from: <u>http://www.scisoc.or.th/stt/28/web/content/J_10/J04.htm[</u>2006, November 10].

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย APPENDIX

สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย

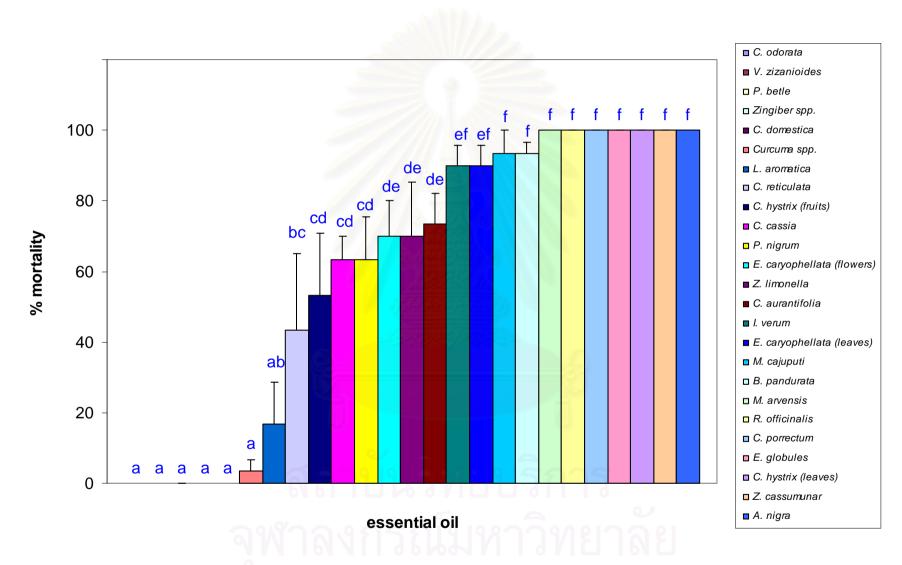


Figure A1 The fumigant activity of essential oils against *Sitophilus oryzae* adults at concentration of 1000 µL/L air after 24 h treatment.

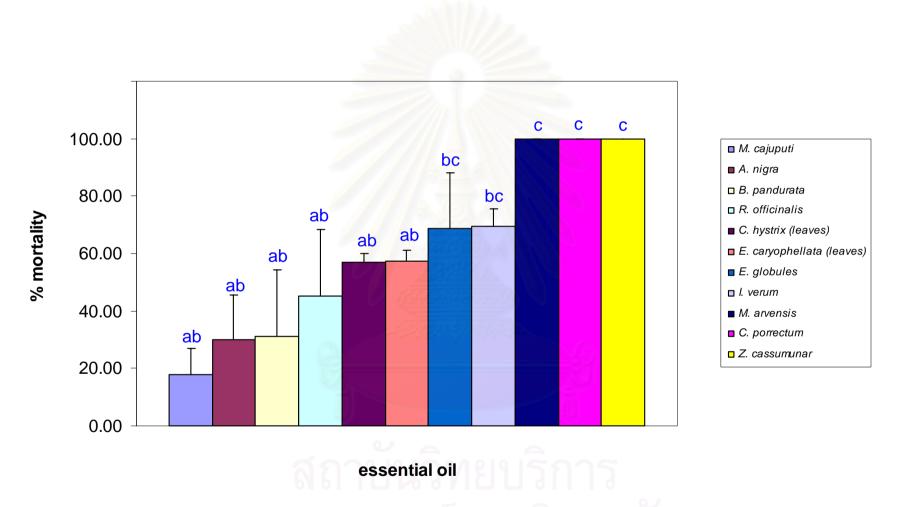


Figure A2 The fumigant activity of essential oils against *Sitophilus oryzae* adults at concentration of 400 µL/L air after 24 h treatment.

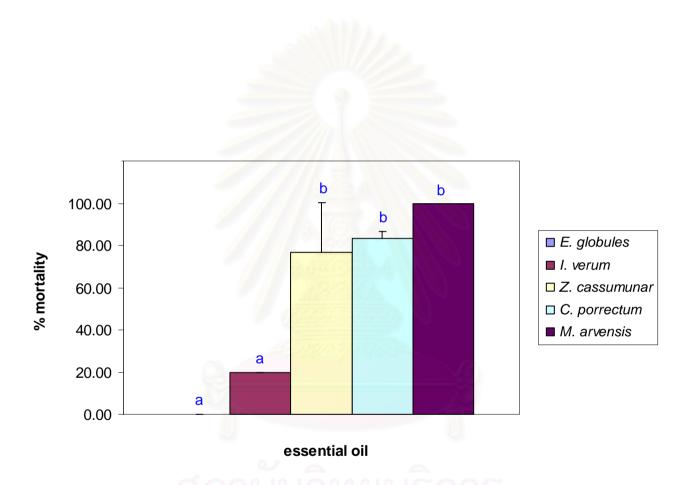


Figure A3 The fumigant activity of essential oils against *Sitophilus oryzae* adults at concentration of 200 µL/L air after treatment for 24 h.





Table A1 The LC₅₀ of three effective essential oils against *S. oryzae*.

nlant			Mean of %Mortality (±SE), n=3						LC ₅₀ ^a	
plant	50	100	120	140	160	220	280	340	400	(µL/L air)
M. arvensis	0.0 ± 0.0	13.3 ± 6.6	30 ± 5.8	53.33 ± 3.3	60.0 ± 20.0	100.0 ± 0.0	100.0 ± 0.0	100.0 ± 0.0	100.0 ± 0.0	138
Z. cassumunar	0.0 ± 0.0	0.0 ± 0.0	-	-	0.0 ± 0.0	10.0 ± 5.8	73.3 ± 12.0	96.7 ± 3.3	100.0 ± 0.0	260
C. porrectum	33.3 ± 8.8	60.0 ± 11.5	-	-	83.3 ± 3.3	83.3 ± 8.8	93.3 ± 6.6	96.7 ± 3.3	96.7 ± 3.3	79

^a Mortality percentage of each concentration were calculated LC₅₀ using Probit analysis Program





Table A2 Residues of menthol and safrole (mg/kg) on fumigated rice comparison between groups of pre-aeration and 24 h aeration.

	Residue in rice (mg/kg)						
Cround	men	thol	safrole				
Groups	Pre-	24 h	Pre-	24 h			
	aeration	aeration	aeration	aeration			
1. Positive control	3873.48	1660.28	9650.76	1592.83			
2. Negative control	0.00	0.00	0.00	0.00			
3. Fumigated rice	489.33	0.00	0.00	0.00			



VITAE

Miss Wachiraporn Phoonan was born on April 15, 1982 in Saraburi province, Thailand. She graduated a Bachelor Degree of Science in Biology, from the Department of Zoology, Kasetsart University, Bangkok, Thailand in 2003. She graduated in Master of Science in Biotechnology in 2006 from the Program of Biotechnology, Faculty of Science, Chulalongkorn University, Bangkok, Thailand. During the course of study, she obtained financial support from Graduate School Chulalongkorn University and TRF-Master Thesis Grants, Thailand.



สถาบันวิทยบริการ จุฬาลงกรณ์มหาวิทยาลัย