

เออนไชม์และยืนของเจอราโนลเจอราโนล ไดฟอสเฟต ฟอสฟາเตส
ในวิธีชีวสังเคราะห์ของเปลาโนกอลในเปล่าน้อย

นางนาภุศจิ นวลแก้ว

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรคุณวีปัณฑิต

สาขาวิชาเคมีและผลิตภัณฑ์ธรรมชาติ

คณะเภสัชศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย

ปีการศึกษา 2547

ISBN 974-53-2091-9

ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

GERANYLGERANYL DIPHOSPHATE PHOSPHATASE ENZYME AND
GENE OF PLAUNOTOL BIOSYNTHETIC PATHWAY
IN *CROTON STELLATOPILOSUS* OHBA

Mrs. Natsajee Nualkaew

A Dissertation Submitted in Partial Fulfillment of the Requirements

For the Degree of Doctor of Philosophy in Pharmaceutical Chemistry and Natural Products

Faculty of Pharmaceutical Sciences

Chulalongkorn University

Academic Year 2004

ISBN 974-53-2091-9

Accepted by the Faculty of Pharmaceutical Sciences, Chulalongkorn University in Partial Fulfillment of the Requirements for the Doctor's Degree.

Boony Tantisira

Bonny J. Ramsden Dean of the Faculty of
Pharmaceutical Sciences

(Associate Professor Boonyong Tantisira, Ph.D.)

THESIS COMMITTEE

S. Wongscriptipatana

(Associate Professor Sumphan Wongseripipatana, Ph.D.)

O. Mc-Sheral

(Associate Professor Wanchai De-Eknamkul, Ph.D.)

Nonglukasue Sriwongsas, Member

(Associate Professor Nongluksna Sriubolmas, Ph.D.)

Khart Savonbe . Member

(Khanit Suwanborirux, Ph.D.)

Maree Chanama Member

(Assistant Professor Manee Chanama, Ph.D.)

นาฏศิลป์ นวลแก้ว : เอนไซม์และยีนของเจอรานิลไดฟอสเฟต ฟอสฟาเตส ในวิถีชีวสังเคราะห์ของเปลาโนทอลในเปลือกน้ำข้อ (GERANYLGERANYL DIPHOSPHATE PHOSPHATASE ENZYME AND GENE OF PLAUNOTOL BIOSYNTHETIC PATHWAY IN CROTON STELLATOPILOSUS OHBA) อาจารย์ที่ปรึกษา : รศ.ดร. วันชัย ดีอกนามกุล, อาจารย์ที่ปรึกษาร่วม : Professor Dr. Meinhart H. Zenk 147 หน้า 1. ISBN 974-53-2091-9.

การป้อนสารติดnakของเจอรานิลเจอรานิลออล [1^{-3}H]GGOH เข้าสู่ไปและก้อนคลัลสของเปลือกน้ำข้อ พบว่าสารติดnakถูกนำไปใชในการสร้างเปลาโนทอล จากการศึกษาด้านเอนไซม์ทำให้สามารถตรวจพบกิจกรรมของ เอนไซม์เจอรานิลเจอรานิลไดฟอสเฟต ฟอสฟาเตส (GGDP phosphatase) ได้ทั้งในสารสกัดเอนไซม์จากใบและในคลอโรพลาส เมื่อทำการแยกเอนไซม์เบื้องต้น โดยวิธีเจลฟิลเตชัน โครโนโตกราฟ ทำให้พบว่า GGDP phosphatase ในสารสกัดเอนไซม์ มีอยู่สองชนิดคือ PI และ PII การศึกษาคุณสมบัติของเอนไซม์ทำให้พบว่า PI มีน้ำหนักโมเลกุล 232 kD ประกอบด้วย 4 หน่วยย่อยของโปรตีนที่มีขนาด 58 kD โดยมีค่า K_m 0.2 mM ค่า V_{max} 277.8 pkat/mg protein และมีสภาวะการทำงานเหมะสมที่ pH 6.0-6.5 ในขณะที่เอนไซม์ PII เป็นโปรตีนเดียวที่มีน้ำหนักโมเลกุล 30-34 kD โดยมีค่า K_m 0.1 mM ค่า V_{max} 7.5 nkat/mg มีสภาวะการทำงานเหมะสมที่ pH 6.5-7.0 และถูกยับยั้งการเร่งปฏิริยาโดยสารตั้งต้นที่ความเข้มข้นสูง นอกจากนี้ยังพบว่า PI และ PII เป็นเอนไซม์ที่เกิดติดกันบนแข็ง โดยกิจกรรมเอนไซม์สามารถยับยั้งได้โดยโมลิบเดตอ่อนเอนไซม์ทั้งสองมีความจำเพาะของการใช้เจอรานิลเจอรานิลไดฟอสเฟตสูงที่สุด เมื่อเปรียบเทียบกับการใช้สารตั้งต้น เจอรานิลฟอสเฟต และฟานเนซิลไดฟอสเฟต การศึกษาระบวนการเร่งปฏิริยาของเอนไซม์พบว่า การตัดหมู่ไดฟอสเฟตออกจากเจอรานิลเจอรานิลไดฟอสเฟต โดยเอนไซม์ PI และ PII เกิดขึ้นโดยผ่านปฏิริยา 2 ขั้นตอนคือ สารตั้งต้นเจอรานิลเจอรานิลไดฟอสเฟต เปลี่ยนไปเป็นเจอรานิลเจอรานิลโนโนฟอสเฟต และเจอรานิลเจอรานิลออล ตามลำดับ การโคลนยืนจากใบเปลือกน้ำข้อ โดยอาศัยข้อมูลของลำดับกรดอะมิโนของพรีนิลไดฟอสเฟต ฟอสฟาเตส จากฐานข้อมูล Swiss-Prot ทำให้ได้ยืนที่สามารถครองคราฟอสฟาเตส 33.6 kD และยังคงแสดงกิจกรรมเอนไซม์เจอรานิลเจอรานิลไดฟอสเฟต ฟอสฟาเตส เอนไซม์ฟอสฟาเตส ที่แสดงออกนี้ มีกระบวนการเร่งปฏิริยาการเปลี่ยนจาก GGDP ไปเป็น GGOH ในแบบ 2 ขั้นตอน เช่นเดียวกับเอนไซม์ GGDP phosphatase ที่แยกได้จากส่วนใน ก้อน GGDP ผ่าน GGMP ไปเป็น GGOH ในที่สุด

สาขาวิชา เภสัชเคมีและผลิตภัณฑ์ธรรมชาติ
ปีการศึกษา 2547

ลายมือชื่อนิสิต..... นพ. ใจดี
ลายมือชื่ออาจารย์ที่ปรึกษา.....

4376955333 : MAJOR PHARMACEUTICAL CHEMISTRY AND NATURAL PRODUCTS
 KEY WORD: CROTON STELLATOPILOSUS / GERANYLGERANIOL / GERANYLGERANYL DIPHOSPHATE / GERANYLGERANYL DIPHOSPHATE PHOSPHATASE / PLAUNOTOL BIOSYNTHESIS / PRENYL DIPHOSPHATE PHOSPHATASE

NATSAJEE NUALKAEW : GERANYLGERANYL DIPHOSPHATE PHOSPHATASE ENZYME AND GENE OF PLAUNOTOL BIOSYNTHETIC PATHWAY IN *CROTON STELLATOPILOSUS* OHBA. THESIS ADVISOR : ASSOC. PROF. WANCHAI DE-EKNAMKUL, Ph.D., THESIS COADVISOR : PROF. MEINHART H. ZENK, Ph.D., 147 pp. ISBN 974-53-2091-9.

The whole leaves and callus cultures of *Croton stellatopilosus* Ohba could incorporate [1^3H]geranylgeraniol into plaunotol. Enzymological studies showed that geranylgeranyl diphosphate phosphatase (GGDP phosphatase) activity was present in cell-free extracts of the leaves and of chloroplasts. The crude enzyme extract could be separated into two peaks of GGDP phosphatase activities (PI and PII) by gel filtration. Purification and characterization of both enzyme peaks revealed that PI was a tetrameric enzyme of 232 kD with its subunit size of 58 kD. It showed optimum pH of 6.0-6.5, an apparent K_m of 0.2 mM, and a V_{max} of 277.8 pkat/mg. PII was a monomeric enzyme of 30-34 kD with an apparent K_m value of 0.1 mM, V_{max} of 7.5 nkat/mg and exhibited substrate inhibition. Both PI and PII appeared to be membrane-bound enzymes and their activities could be inhibited by Mo^{2+} . Both preferred GGDP as their substrate rather than geranyl diphosphate or farnesyl diphosphate. Study on the catalysis of the reaction revealed that GGOH was formed from GGDP by two successive monodephosphorylations, rather than a one-step diphosphate dephosphorylation. Cloning of the phosphatase genes from *C. stellatopilosus* leaves, based on the available information of prenyl diphosphate phosphatase in Swiss-Prot database, was also performed using *E. coli* as expression system. The results showed that the encoded protein had its molecular weight of 33.6 kD and could exhibit GGDP phosphatase activity. The expressed phosphatase also showed its process of catalytic dephosphorylation by using two sequential steps of monophosphate dephosphorylation from GGDP to GGMP and to GGOH.

Field of study Pharmaceutical Chemistry
 and Natural Products

Academic year 2004

Student's signature.....

Advisor's signature.....

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude and deep appreciation to Associate Professor Dr. Wanchai De-Eknamkul, my major advisor for his guidance, valuable advice, encouragement and understanding throughout my work.

I especially thank Professor Dr. Meinhart H. Zenk, my co-advisor for his valuable advice, supervision and understanding throughout my work especially in the part of enzymology. I am also greatful thank Professor Toni M. Kuchan, for her guidance, valuable advice in the field of molecular biology.

I would like to thank Mr. Nils Guennewich, Dr. Karin Springob, and Ms Maja Raschke, for their kindly help and suggestion for my work in Germany.

I am grateful to laboratory assistants, colleagues and friends of the Faculty of Pharmaceutical Sciences, Chulalongkorn University and also the staffs, postgraduates, and friends of Biocenter and those of Institute of Plant Biochemistry, Halle/Saale, Germany, for their assistance and friendship.

I would like to thank the Royal Golden Jubilee Ph.D. Program, The Thailand Research Fund (TRF) for financial support, the German Academic Exchange (DAAD) for partial research grant in Germany and Faculty of Pharmaceutical Sciences, Chulalongkorn University, Biocenter, and Institute of Plant Biochemistry, Germany for providing laboratories facilities, equipments and chemical compounds.

I would like to thank The Institute of Biotechnology and Genetic Engineering, Chulalongkorn University; Siri Ruckhachati Medicinal Plant Garden, Mahidol University; and Faculty of Pharmaceutical Sciences, Srinakharinwirot University for their support of plant materials.

I am much indebted to Khon-Kaen University for permission enabling me to undertake this higher study.

Finally, I am grateful to my parents, my husband and my family for their love, encouragement, support and entirely care. The usefulness of this dissertation, I dedicate to my parents and all of my teachers.

CONTENTS

	Page
ABSTRACT (Thai).....	iv
ABSTRACT (English)	v
ACKNOWLEDGEMENT.....	vi
CONTENTS.....	vii
LIST OF TABLES.....	ix
LIST OF FIGURES.....	x
ABBREVIATIONS.....	xiv
CHAPTER	
I INTRODUCTION.....	1
II HISTORICAL.....	4
1. Botanical description of <i>Croton stellatopilosus</i> Ohba	4
2. Thai Folk medicinal uses.....	6
3. Bioactive compounds in <i>Croton stellatopilosus</i>	6
4. Various techniques used for obtaining plaunitol.....	7
5. Methods used in biosynthetic study of secondary metabolites.....	8
6. Previous biosynthetic study in <i>Croton stellatopilosus</i>	9
7. Compartmentalization of diterpene biosynthesis in plants and in <i>Croton stellatopilosus</i>	10
8. Phosphatases: Clarification.....	12
9. Phosphatases involving in terpenoid pathways.....	12
10. Amino acid sequences of phosphatase enzymes utilizing GGDP.....	15
11. N-terminal targeting sequences.....	25

Chapter	Page
III MATERIALS AND METHODS.....	30
1. Chemicals.....	30
2. Feeding experiments and detection of enzyme activity.....	33
3. Purification and characterization of GGDP phosphatase.....	36
4. Gene cloning of <i>Croton stellatopilosus</i> prenyl diphosphate phosphatase.....	42
IV RESULTS.....	68
1. Preparation of compounds for biosynthetic studies.....	68
2. Biosynthetic study of <i>C. stellatopilosus</i>	74
3. Purification and characterization of GGDP phosphatase from <i>C. stellatopilosus</i>	83
4. Gene cloning of prenyl diphosphate phosphatase from <i>C. stellatopilosus</i>	100
V DISCUSSION.....	115
VI CONCLUSION.....	133
REFERENCES.....	136
VITA.....	147

LIST OF TABLES

Table	Page
1. Some properties of bornyl pyrophosphate phosphohydrolases.....	13
2. Phosphatase activities in an enzyme preparation from orange flavedo.....	14
3. Peptide targeting domains for transport to different organelles.....	26
4. Microorganisms used in this study.....	43
5. Reaction mixture of first strand DNA synthesis using for RACE PCR.....	58
6. Reaction mixtures of 3'- and 5'-RACE PCR.....	59
7. Summary of GGDP phosphatase purification from <i>C. stellatopilosus</i> leaves.....	88
8. Rf value of proteins from SDS-PAGE gel.....	89
9. Vo/Ve value of proteins from Superose 6 gel filtration column.....	91
10. The influence of metal ions on the enzyme activity.....	95
11. Substrate specificity of PI and PII.....	96
12. Degenerated primers designed from prenyldiphosphate phosphatase.....	100
13. Comparison of some properties of prenyl diphosphate phosphatase.....	121
14. Percent (%) homology of prenyl diphosphate phosphatase and phosphatidic acid to prenyl diphosphate phosphatase from <i>C. stellatopilosus</i>	125
15. Percent (%) homology of <i>C. stellatopilosus</i> prenyl diphosphate phosphatase to terpene synthases and terpene cyclases.....	128
16. Properties of PI and PII from <i>C. stellatopilosus</i>	134

LIST OF FIGURES

Figure		Page
1	Proposed biosynthetic pathway for the formation of plaunotol.....	3
2	Various parts of <i>Croton stellatopilosus</i> Ohba.....	5
3	Bioactive compounds in <i>Croton stellatopilosus</i>	7
4	Compartmentation of terpenoid biosynthesis.....	11
5	Electron micrograph of the palisade mesophyll cell x 3,800 time.....	11
6	Pathway of camphor biosynthesis from geranyl diphosphate in <i>Salvia officinalis</i>	13
7	Prenylation by <i>E</i> -isoprenyl diphosphate synthase.....	16
8	Structures of <i>E</i> - Isoprenyl diphosphate synthases.....	17
9	Schematic representation of the active site of FPPS with FDP (product) and IDP bound.....	18
10	Prenylation by <i>Z</i> -isoprenyl diphosphate synthase	19
11	Schematic representation of the active site of UPPS with FDP chain elongation and IDP bound.....	19
12	Comparison of amino acid sequences between terpene synthase.....	20
13	Transmembrane model of mung bean vacuolar pyrophosphatase.....	22
14	Consensus sequence of various phosphatases with phosphatase sequence motif.....	23
15	Comparison of amino acid sequence between the deduced AtLpp1p, AtLpp2p, and AtLpp3p proteins.....	24
16	Dephosphorylation of diacylglycerol pyrophosphate phosphatase from diacylglycerol pyrophosphate.....	25
17	Pathways for protein targeting into thylakoids.....	27
18	Transit peptides of luminal proteins from <i>Arabidopsis thaliana</i>	29
19	Diagram of chemical synthesis of [1- ³ H]GGOH.....	68

Figure	Page
20 Phosphoimage showed kinase activities of microsome from <i>C. stellatopilosus</i>	69
21 Diagram of enzymatic reaction of [1- ³ H]GGMP and [1- ³ H]GGDP formation from [1- ³ H]GGOH.....	70
22 TLC radiochromatograms of enzymatic products obtained from microsomal kinases	70
23 TLC radiochromatograms of dephosphorylation of [1- ³ H]GGDP and [1- ³ H]GGMP by alkaline phosphatase.....	71
24 TLC radiochromatograms of the fractions from MCI gel CHP20P column.....	72
25 Diagram of chemical synthesis of dibenzoyl plaunotol.....	73
26 FTICR of dibenzoyl plaunotol (C ₃₄ H ₄₂ O ₄).....	73
27 TLC radiochromatogram of CHCl ₃ extract obtained from the leaf fed with 10 µCi [1- ³ H]GGOH.....	74
28 TLC radiochromatogram showed trace amount of compound at the same Rf as dibenzoyl plaunotol.....	75
29 TLC phosphoimage of callus and cell suspension of <i>C. stellatopilosus</i> fed with [1- ³ H]GGOH.....	76
30 Sucrose gradient for chloroplast separation from <i>C. stellatopilosus</i> leaves.....	77
31 TLC radiochromatogram of GGDP phosphatase activity in chloroplast preparation of <i>C. stellatopilosus</i>	78
32 Phosphoimage of GGMP phosphatase activity in the chloroplast preparation of <i>C. stellatopilosus</i>	79
33 TLC patterns of products obtained from GGDP phosphatase activity in 20,000 x g fraction of <i>Croton stellatopilosus</i> leaves.....	80
34 A typical TLC-densitochromatogram of the reaction mixture.....	81
35 TLC-densitochromatogram showed the activity of GGDP phosphatase before and after membrane solubilization.....	82

Figure	Page
36 Optimization of GGDP phosphatase assay.....	84
37 Chromatogram obtained from BioGel A column chromatography of crude enzyme extract.....	86
38 Chromatogram of PI separation by Superose 6 column.....	86
39 Chromatogram of PII separation by Superose 6 column.....	87
40 Chromatogram of PII separation by UNO Q column.....	87
41 SDS-PAGE of various enzyme preparation obtained during enzyme purification of PI and PII from <i>C. stellatopilosus</i>	88
42 Standard calibration curve of Log molecular weight plotted against Rf values of standard proteins from 12% SDS-PAGE.....	90
43 Elution profile of Bio-Rad molecular weight standard on Superose 6 column.....	92
44 Standard calibration curve of Log molecular weight plotted against Vo/Ve of Superose 6 column.....	93
45 Lineweaver-Burk plot of GGDP phosphatase activity.....	94
46 Effect of pH on GGDP phosphatase activity.....	97
47 TLC radiochromatogram of dephosphorylation using [1- ³ H]GGDP as substrate.....	98
48 TLC radiochromatograms of dephosphorylation using [1- ³ H]GGMP as substrate.....	99
49 Comparison of amino acid sequences of various prenyl diphosphate phosphatases.....	101
50 Agarose gel of total RNA from fresh leaves of <i>C. stellatopilosus</i>	102
51 Agarose gel electrophoresis of Nested PCR product.....	103
52 Agarose gel of touchdown PCR product of 3'-RACE and 5'-RACE.....	104
53 Diagram of RACE PCR fragment and the connection into full-length gene.....	104

Figure	Page
54 Full-length gene of ORF1 and ORF2.....	105
55 Agarose gel of fusion gene with his-tag	106
56 Starting amino acid of 5 clones of <i>C. stellatopilosus</i> prenyl diphosphate phosphatase.....	107
57 SDS-PAGE of truncated protein ORF2.....	110
58 SDS-PAGE of protein ORF1 and ORF2.....	110
59 Western Blot of truncate ORF1 protein from HiTrap column by using His-Tag monoclonal antibody.....	111
60 Western Blot of recombinant His-tagged proteins from TALON resin column.....	112
61 TLC radiochromatograms of the dephosphorylation of recombinant prenyl diphosphate phosphatase I.....	113
62 TLC radiochromatograms of the dephosphorylation of recombinant prenyl diphosphate phosphatase I.....	114
63 Diagram of synthesizing 5'-end of gene by 5'-RACE PCR using SMART kit.....	123
64 Phylogenetic tree of prenyldiphosphate phosphatase, phosphatidic acid phosphatases, DGPP phosphatase and Dol-PP phosphatase.....	124
65 Alignment of the deduced amino acid sequence of ORF1 and ORF2 from <i>C. stellatopilosus</i> , prenyl diphosphate phosphatases and phosphatidic acid phosphatases.....	127
66 Transmembrane prediction of deduced amino acid sequences of ORF1 and ORF2.....	130

ABBREVIATIONS

%	=	percent (part per 100); percentage
β	=	beta
β -	=	beta-
δ	=	chemical shift
μCi	=	microCurie
μl	=	microliter (s)
μM	=	micro molar (s)
λ_{max}	=	wavelength at maximum absorption
μmol	=	micromole
/	=	per, or divided by
[1- ^{14}C]	=	carbon position 1 of the molecule is labeled with carbon 14
[1- ^3H]	=	hydrogen position 1 of the molecule is labeled with tritium
$^1\text{H-NMR}$	=	proton nuclear magnetic resonance
2,4-D	=	2,4-dichlorophenoxyacetic acid
Bis	=	N,N'-dimethyl bisacrylamide
bp	=	base pair
BSA	=	bovine serum albumin
cDNA	=	cloning DNA
cDNA	=	complementary deoxyribonucleic acid
CHCl_3	=	chloroform
Ci	=	Curie
cm	=	centimeter
cpm	=	counts per minute
CTP	=	cytidine 5'-triphosphate
Da	=	Dalton, unit of molecular mass (1/12 of C=1)
DNA	=	deoxyribonucleic acid
dpm	=	disintegrations per min
EDTA	=	ethylenediamine tetraacetic acid
e. g.	=	for example
EM	=	electron microscope

etc.	=	et cetera
Fig	=	Figure
g	=	gram
g	=	centrifugal force (relative to gravity)
GGPPS	=	geranylgeranyl pyrophosphate synthase
hr	=	hour
Hz	=	Hertz
IDP	=	isopentenyl diphosphate
kD	=	kilodalton, ($\times 10^3$ Da)
kg	=	kilogram
K_m	=	Michaelis constant = substrate concentration at which the rate of enzyme-catalysed reaction is half maximum rate
l	=	liter (s)
LS	=	Linsmair and Skoog (1965) medium
m	=	meter (s)
M	=	molar (s)
m/z	=	mass to charge ratio
mA	=	milliampere
MeOH	=	Methanol
mg	=	milligram
ml	=	milliliter
mm	=	millimeter
mM	=	millimolar
mmol	=	millimole
Mr	=	relative molecular mass
MS	=	Murishage and Skoog (1962) medium
MS	=	Mass spectrometry
MW	=	molecular weight
N ₂	=	Nitrogen atmosphere
NaOH	=	sodium hydroxide
nkat	=	nano katal

nm	=	nanometer (s)
no.	=	number
°C	=	degree Celsius
opt	=	optimum
PCR	=	polymerase chain reaction
pH	=	hydrogen ion concentration
pI	=	isoelectric point
pkat	=	pico katal
pmol	=	picomole (s)
ppm	=	part per million
q.s.	=	quantum suffices
R _f	=	distance spot moved/ distance solvent moved (TLC)
RNA	=	ribonucleic acid
rpm	=	revolutions per minute
SDS	=	sodium dodecyl sulfate
SDS-PAGE	=	sodium dodecyl sulfate-polyacrylamide gel electrophoresis
sec	=	second (s)
sp.act.	=	specific activity
TEMED	=	N,N,N',N'-tetramethylethylenediamine
TLC	=	thin layer chromatography
UV	=	ultraviolet light
v/v	=	volume/ volume
V _e	=	elution volume
V _{max}	=	maximum velocity of enzyme
V _o	=	void volume
w/v	=	weight/volume (concentration)
wt	=	weight