

REFERENCES

1. Goosen, M.F.A., *Application of Chitin and Chitosan*, Pennsylvania: Technomic Publishing, **1997**, pp. 79-86.
2. Zakaria, Z., *Lactic Acid Purification of Chitin from Prawn Waste Using a Horizontal Rotating Bioreactor*, Ph.D. Thesis, Loughborough University, **1997**.
3. Tracey, M. V., "Chitin", *Rev. Pure Appl. Chem.*, **1957**, 7, pp. 1-14.
4. Blackwell, J., "Physical Methods for The Determination of Chitin Structure and Conformation", *Method Enzymol*, **1988**, 161, pp. 435-442.
5. Simpson, B. K., Gagne, N., and Simpson, M. V., "Bioprocessing of Chitin and Chitosan", in *Fisheries Processing: Biotechnological Applications*, (ed. Martin, A. M.), Chapman & Hall, London, **1994**, pp. 155-173.
6. No, H. K., and Meyers, S. P., "Preparation of Chitin and Chitosan", in *Chitin Handbook*, (eds. Muzzarelli, R. A. A., and Peter, M. G.), European Chitin Society, 1997, pp. 475-489.
7. Shahidi, F., and Synowiecki, J., "Isolation and Characterization of Nutrients and Value-added Products from Snow Crab (*Chionoecetes opilio*) and Shrimp (*Pandalus borealis*) Processing Discards", *J. Agric. Food. Chem.*, **1991**, 39, pp. 1527-1532.
8. Carroad, P. A., and Tom, R. A., "Bioconversion of The Chitin Wastes: Process Conception and Selection of Microorganism", *J. Food Sci.*, **1978**, 43, pp. 1158-1161.
9. Muzzarelli, R. A. A.; Jeuniaux, C., and Gooday, G. W., *Chitin in Nature and Technology*, New York: Plenum Press, **1986**, pp. 287-293.
10. Shahidi, F.; Kamil, J.; Arachchi, V., and Jeon, Y. J., "Food Application of Chitin and Chitosans", *Trends in Food Sci. & Technol.*, **1999**, 10, pp. 37-51.
11. Hirano, S., and Nagao, N., "Effects of Chitosan, Pectic Acid, Lysozyme and Chitinase on The Growth of Several Phytopathogens", *Agric. Biol. Chem.*, **1989**, 53, pp. 3065-3066.
12. Akiyama, K.; Kawazu, K., and Kobayashi, A., "A Novel Method for Chemo-Enzymatic Synthesis of Elicitor-Active Chitosan Oligomers and Partially *N*-Deacetylated Chitin Oligomers Using *N*-Acetylated Chitotrioses as

- Substrates in A Lysozyme-Catalyzed Transglycosylation Reaction System”, *Carbohydr. Res.*, **1995**, pp. 279, 151.
13. Brodelius, P.; Funk, C.; Haner, A., and Villegas, M., “A Procedure for The Determination of Optimal Chitosan Concentrations for Elicitation of Cultured Plant Cells”, *Phytochemistry*, **1989**, 28, pp. 2651-2654.
 14. Yamada, A.; Shibuya, N.; Kodama, O., and Akatsuka, T., “Induction of Phytoalexin Formation in Suspension-Cultured Rice Cells by *N*-Acetylchitooligosaccharides”, *Biosci. Biotech. Biochem.*, **1993**, 57, pp. 405-409.
 15. Vander, P.; Vrum, K. M.; Domard, A.; Eddine, N.; Gueddari, E., and Moerschbacher, B. M., “Comparison of The Ability of Partially *N*-Acetylated Chitosans and Chitooligosaccharides to Elicit Resistance Reactions in Wheat Leaves”, *Plant Physiol.*, **1998**, 118, pp. 1353-1359.
 16. Suzuki, S., “Studies on Biological Effects of Water Soluble Lower Homologous Oligosaccharides of Chitin and Chitosan”, *Fragrance J.*, **1996**, 15, pp. 61-68.
 17. Suzuki, K.; Tokoro, A.; Okawa, Y.; Suzuki, S., and Suzuki, M., “Enhancing Effects of *N*-Acetylchitooligosaccharide on The Active Oxygen-Generating and Microbicidal Activity of Peritoneal Exudate Cells in Mice”, *Chem. Pharma. Bull.*, **1985**, 33, pp. 886-888.
 18. Tokoro, A.; Tatewaki, N.; Suzuki, K.; Mikami, T.; Suzuki, S., and Suzuki, M., “Growth-Inhibitory Effect on Hexa-*N*-Acetylchitohexoase and Chitohexoase Against Meth-A Solid Tumor”, *Chem. Pharma. Bull.*, **1988**, 36, pp. 784-790.
 19. Suzuki, K.; Tokoro, A.; Okawa, Y.; Suzuki, S., and Suzuki, M., “Effect of *N*-Acetylchitooligosaccharides on Activation of Phagocytes”, *Microbiol. Immunol.*, **1986**, 30, pp. 777-787.
 20. Tokoro, A.; Kobayashi, M.; Tatewaki, N.; Suzuki, K.; Okawa, Y.; Mikami, T.; Suzuki, S., and Suzuki, M., “Protective Effect of *N*-Acetyl Chitohexoase on *Listeria Monocytogens* Infection in Mice”, *Microbiol. Immunol.*, **1989**, 33, pp. 357.
 21. Tsukada, K.; Matsumoto, T.; Aizawa, K.; Tokoro, A.; Naruse, R.; Suzuki, S, and Suzuki, M., “Antimetastatic and Growth Inhibitory Effect of *N*-Acetylchitohexoase in Mice Bearing Lewis Lung Carcinoma”, *Jpn. J. Cancer Res.*, **1990**, 81, pp. 259-265.

22. Suzuki, K.; Mikami, T.; Okawa, Y.; Suzuki, S., and Suzuki, M., "Antitumor Effect of Hexa-*N*-Acetylchitohexaose and Chitohexaose", *Carbohydr. Res.*, **1986**, *151*, pp. 403-408.
23. Shikhman, A.R.; Kuhn, K.; Alaaeddine, N., and Lotz, M., "*N*-Acetylglucosamine Prevents IL-1 Beta-Mediated Activation of Human Chondrocytes", *J. Immunology*, **2001**, *166*, pp. 5155-5160.
24. Hiroshi, H., "Disaccharide as Starting Materials for The Synthesis of Valuable Compounds", *Trends in Glycoscience and Glycotechnology*, **2000**, *12*, pp. 185-190.
25. Takahashi, S.; Terayama, H.; Koshino, H., and Kuzuhara, H., "Synthesis of Novel Azapseudodisaccharide Related to Allosamidin Employing *N,N'*-Diacetylchitobiose as A Key Material", *Tetrahedron*, **1999**, *55*, pp. 14871-14884.
26. Richardson, S. C. W.; Kolbe, H. V. J., and Duncan, R., "Potential of Low Molecular Mass Chitosan as A DNA Delivery System: Biocompatibility, Body Distribution and Ability to Complex and Protect DNA", *Int. J. Pharm.*, **1999**, *178*, pp. 231-243.
27. Rupley, S. "The Hydrolysis of Chitin by Concentrated Hydrochloric Acid, and The Preparation of Low-Molecular-Weight Substrates For Lysozyme A", *Biochim. Biophys. Acta*, **1964**, *83*, pp. 245-255.
28. Takahashi, Y.; Miki, F., and Nagase, K., "Effect of Sonolysis on Acid Degradation of Chitin to Form Oligosaccharides", *Bull. Chem. Soc. Jpn.*, **1995**, *68*, pp. 1851-1857.
29. Bosso, C.; Defaye, J.; Domard, A.; Gadelle, A., and Pedersen, C., "The Behavior of Chitin Towards Anhydrous Hydrogen Fluoride. Preparation of β -(1 \rightarrow 4)-Linked 2-Acetamido-2-Deoxy-D-Glucopyranosyl Oligosaccharides", *Carbohydr. Res.*, **1986**, *156*, pp. 57-68.
30. Inaba, T.; Ohguchi, T.; Iga, Y., and Hasegawa, E., "Synthesis of 4-Methylcoumarin-7-yloxy Tetra-*N*-Acetyl- β -Chitotetraoside, A Novel Synthetic Substrate for The Fluorometric Assay of Lysozyme", *Chem. Pharm. Bull.*, **1984**, *32*, pp. 1597-1603.

31. Kurita, K.; Tomita, K.; Ishii, S.; Nishimura, S., and Shimoda, K., "Beta-Chitin as A Convenient Starting Material for Acetolysis for Efficient Preparation of *N*-Acetylchitooligosaccharides", *J. Poly. Sci.: Part A: Poly. Chem.*, **1993**, *31*, pp. 2393-2395.
32. Defaye, J., Gadelle, A., and Pedersen, C., *Chitin and Chitosan*, London: Elsevier, **1989**, pp. 415-429.
33. Horowitz, S.; Roseman, S., and Blumenthal, H. J., "The Preparation of Glucosamine Oligosaccharides. I. Separation", *J. Am. Chem. Soc.*, **1957**, *79*, pp. 5046.
34. Uchida, Y., Izume, M., and Ohtakara, A., *Chitin and Chitosan*, London: Elsevier, **1989**, pp. 373-382.
35. Mitsutomi, M., Uchiyama, A., Yamagami, T., Watanabe, T., "Mode of Action of Family 19 Chitinase", In : A Domard, GAF Roberts, KM Varum (eds), *Advances in Chitin Science*, **1997**, *2*, Jacques Andre, Lyon, pp. 250-255.
36. Kramer, K. J., Koda, D., "Insect Chitin : Physical State, Synthesis Degradation and Metabolic Regulation", *Insect Biochem.*, **1986**, *16*, pp. 851-877.
37. Koda, D., Hoshika, H., Matsushita, M., Tanaka, A., Ida, A., Kono, M., "Purification and Characterization of β -N-acetylhexosaminidase from The Liver of a Prawn", *Penaeus japonicus. Biosci. Biotech. Biochem.*, **1996**, *60*, pp. 194-199.
38. Takiguchi, Y., and Shimahara, K., "Isolation and Identification of a Thermophilic Bacterium Producing *N,N'*-Diacetylchitobiose from Chitin", *Agric. Biol. Chem.*, **1989**, *53*, pp. 1537.
39. Takayanagi, T.; Ajisaka, K.; Takiguchi, Y., and Shimahara, K., "Isolation and Characterization of Thermostable Chitinases from *Bacillus licheniformis* X-7u", *Biochim. Biophys. Acta*, **1991**, *1078*, pp. 404-410.
40. Aiba, S., "Preparation of *N*-acetylchitooligosaccharides by Hydrolysis of Chitosan with Chitinase Followed by *N*-Acetylation", *Carbohydr. Res.*, **1994**, *265*, pp. 323-328.
41. Kobayashi, S.; Kiyosada, T., and Shoda, S. I., "A Novel Method for Synthesis of Chitobiose *via* Enzymatic Glycosylation Using A Sugar Oxazoline as Glycosyl Donor", *Tetrahedron Letters*, **1997**, *38*, pp. 2111-2112.

42. Domard, A.; Jeuniaux, C; Muzzarelli, R. A. A., and Roberts, G. A. F., *Advances in Chitin Science* Lyon: Andre Publ., **1996**, pp. 192-197.
43. Shashiwa, H.; Fujishima, S.; Yamana, N.; Kawasaki, N.; Nakayama, A.; Muraki, E., and Aiba, S., "Production of *N*-Acetyl-D-Glucosamine from β -Chitin by Enzymatic Hydrolysis", *Chem. Lett.*, **2001**, pp. 308-309.
44. Sukwattanasinitt, M.; Zhu, H.; Shashiwa, H., and Aiba, S., "Utilization of Commercial Non-Chitinase Enzymes from Fungi for Preparation of 2-Acetamido-2-Deoxy-D-Glucose from β -Chitin", *Carbohydr. Res.*, **2002**, *337*, pp. 133-137.
45. Sashiwa, H.; Fujishima, S.; Yamano, N.; Kawasaki, N.; Nakayama, A.; Muraki, E.; Hiraga, K.; Oda, K., and Aiba, S., "Production of *N*-Acetyl-D-Glucosamine from α -Chitin by Crude Enzymes from *Aeromonas hydrophila* H-2330", *Carbohydr. Res.*, **2002**, *337*, pp. 761-763.
46. Pichyangkura, R.; Kudan, S.; Kuttiyawong, K.; Sukwattanasinitt, M., and Aiba, S., "Quantitative Production of 2-Acetamido-2-Deoxy-D-Glucose from Crystalline Chitin by Bacterial Chitinase", *Carbohydr. Res.*, **2002**, *337*, pp. 557-559.
47. Matsuoka, K.; Matsuzawa, Y.; Kusano, K.; Terunuma D., and Kuzuhara, H., "An Improved Preparation of *N,N'*-Diacetylchitobiose by Continuous Enzymatic Degradation of Colloidal Chitin Using Dialysis Tubing as A Convenient Separator", *Biomacromolecules*, **2000**, *1*, pp. 798-800.
48. Jolles, P., and Muzzarelli, R. A. A., "Chitin and Chitinases", Birkhäuser Vorlag, Basel. **1999**.
49. Boller, T., "Cellular Molecular and Molecular Biology of Plant Stress", Alan R. Liss Inc., New York, **1985**.
50. Henrissat, B., Bairoch, A. "New Families in The Classification of Glycosyl Hydrolases Based on Amino Acid Sequence Similarities", *Biochem. J.*, **1993**, *293*, pp. 781-788.
51. Perrakis, A., Tews, I., Dauter, Z., Oppenheim, A. B., Chet, J., Wilson, K. S., and Vorgias, C. E., "Crystal Structure of a Bacteria Chitinase at 2.3 Å Resolution", *Structure*, 1994, *2*, pp. 1169-1180.
52. Terwisscha van Scheltinga, A.C., Kalk, K. H., Beintemo, J. J., and Dijkstra, B. W., "Crystal Structure of Hevamine, a Plant Defence Protein with Chitinase

- and Lysozyme Activity and Its Complex with an Inhibitor”, *Structure*, **1994**, *2*, pp. 1181-1189.
53. Roey, P. V., Rao, V., Plummer, T. H., and Tarentino, A., “Crystal Structure of Endo- β -N-acetylglucosaminidase F1, an α/β -barrel Enzyme Adapted for a Complex Substrate”, *Biochemistry*, **1994**, *33*, pp. 13989-13996.
 54. Rao, V., Guan, C., and Roey, P. V., “Crystal Structure of Endo- β -N-acetylglucosaminidase H at 1.9 Å Resolution: Active-site Geometry and Substrate Recognition”, *Structure*, **1995**, *3*, pp. 449-457.
 55. Ohno, T., Armand, S., Hata, T., Nikaidou, N., Henrissat, B., Mitsutomi, M., and Watanabe, T. “A Molecular Family 19 Chitinase Found in The Prokaryotic Organism *Streptomyces griseus* HUT 6037”, *J. Bacteriol.*, **1996**, *178*, pp. 5056-5057.
 56. Tews, I., Wilson, K. S., and Vorgias, C. E., “Enzymatic Mechanism of N-acetylglucosaminidase Revealed by Structural Studies on Enzyme Substrate-Inhibitor Complexes”, *Adv. Chitin. Sci.*, **1996**, *1*, pp. 23-26.
 57. Davies, G. J.; Wilson, K. S., and Henrissat, B., “Nomenclature for Sugar-Binding Subsites in Glycosyl Hydrolases”, *Biochem. J.*, **1997**, *321*, pp. 557-559.
 58. Honda, Y., and Fukamizo, T., “Substrate Binding Subsites of Chitinase from Barley Seeds and Lysozyme from Goose Egg White” *BBA-Protein Struct. M*, **1998**, *1388*, pp. 53-65.
 59. Tews, I, Terwisscha van Scheltinga, A.C., Perrakis, A., Wilson, K.S., and Dijkstra, B.W., “Substrate-Assisted Catalysis Unifies Two Families of Chitinolytic Enzymes”, *J. Am. Chem. Soc.*, **1997**, *119*, pp. 7954-7959.
 60. Brameld, K.A., and Goddard III, W.A., “Substrate Distortion to A Boat Conformation at Subsite -1 Is Critical in The Mechanism of Family 18 Chitinase”, *J. Am. Chem. Soc.*, **1998**, *120*, pp. 3571- 3580.
 61. Kuttiyawong, K., “Cloning and nucleotide sequencing of chitinase gene from *Burkholderia cepacia* TU09”, MSC. Thesis, Department of Biochemistry, Faculty of Science, Chulalongkorn University, **2001**.
 62. Nishimura, S.; Kuzuhara, H.; Takiguchi, Y., and Shimahara, K, “Peracetylated Chitobiose: Preparation by Specific Degradations of Chitin, and Chemical Manipulations”, *Carbohydr. Res.*, **1989**, *194*, pp. 223-231.

63. Bradford, M. M., "A Rapid and Sensitive Method for the Quantitatively of Microgram Quantities of Protein Utilizing the Principle of Protein-dye Binding", *Anal. Biochem.*, **1976**, 72, pp. 248-254.
64. Imoto, T., and Yagishita, K., "A Simple Activity Measurement of Lysozyme", *Agr. Biol. Chem.* **1971**, 35, pp. 1154-1156.



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย



APPENDICES

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย



APPENDIX A

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

1. Reagent preparation

1.1 Reagent for protein assaying

- Bradford Stock solution
 - 100 mL 95% ethanol
 - 200 mL 85% phosphoric acid
 - 350 mg Serva Blue G
 - stable indefinitely at room temperature.
- Bradford working Buffer
 - 425 mL distilled water
 - 15 mL 95% ethanol
 - 30 mL 85% phosphoric acid
 - 30 mL Bradford Stock Solution
 - Filter through Whatman No. 1 paper, store at room temperature in brown glass bottle. Usable for several weeks, but may need to be refiltered.

1.2 Reagent for chitinase activity assaying

- Color reagent
 - Potassium hexaferrocyanate ($K_3Fe(CN)_6$) (0.5 g) and Na_2CO_3 (45.34 g) were dissolved in DI-water 500 mL.

2. Preparation the calibration curve of *N*-acetyl-D-glucosamine for HPLC analysis

Calibration curve of GlcNAc was made by varying the concentration and measuring the peak area by HPLC.

Table A1 The concentration of standard solution of GlcNAc and peak area.

Standard No.	Conc. GlcNAc (mg/mL)	Conc. GlcNAc (mM)	Peak area (mV*Sec)
1	0.0093	0.0420	14.983
2	0.0186	0.0841	27.667
3	0.0620	0.2803	98.046
4	0.1860	0.8408	308.345
5	0.1210	0.5470	195.536
6	0.2420	1.0940	387.937
7	0.4840	2.1880	749.393
8	1.2400	5.6055	1987.448

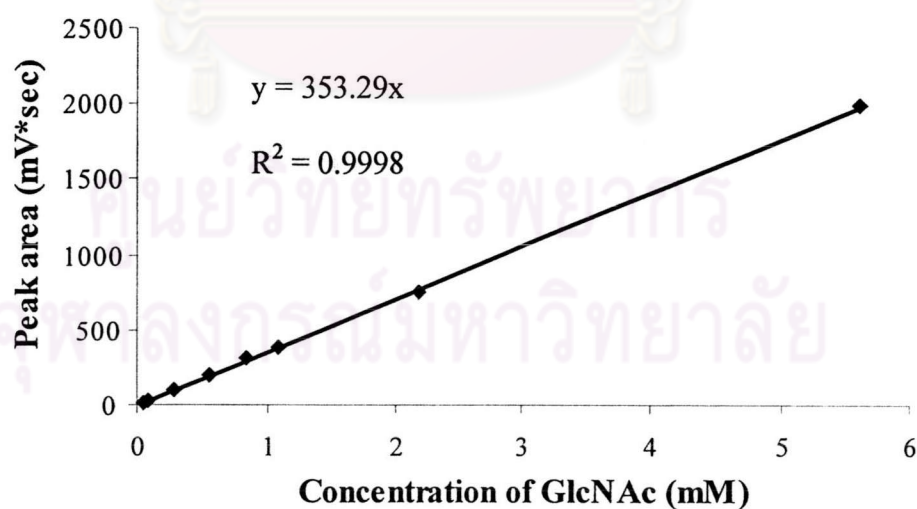


Figure A1 Correlation between concentration of standard *N*-acetyl-D-glucosamine and peak area by HPLC.

3. Preparation the calibration curve of N,N' -diacetylchitobiose for HPLC analysis

Calibration curve of $(\text{GlcNAc})_2$ was made by varying the concentration and measuring the peak area by HPLC.

Table A2 The concentration of standard solution of $(\text{GlcNAc})_2$ and peak area.

Standard No.	Conc. $(\text{GlcNAc})_2$ (mg/mL)	Conc. $(\text{GlcNAc})_2$ (mM)	Peak area (mV*Sec)
1	0.05	0.1178	57.786
2	0.10	0.2356	111.022
3	0.20	0.4712	235.395
4	0.32	0.7540	380.000
5	0.60	1.4137	676.788
6	1.00	2.3562	1128.094
7	1.28	3.0159	1494.989
8	1.60	3.7699	1900.128

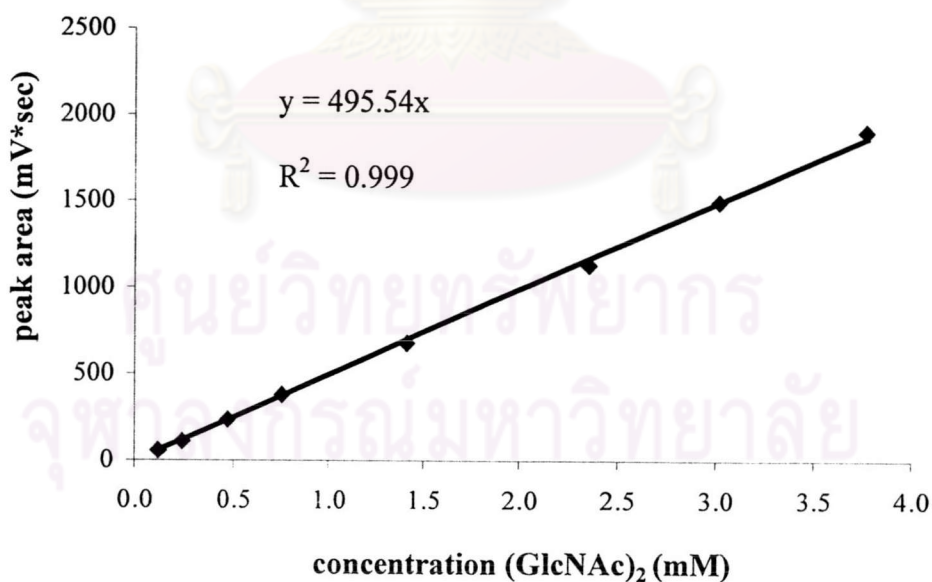


Figure A2 Correlation between concentration of standard N,N' -diacetylchitobiose and peak area by HPLC.

4. Preparation calibration curve of protein concentration by Bradford's colorimetric method

Calibration curve for BSA was made by determining the absorbance value at 595 nm of standard BSA according to the method of Bradford.

Table A3 The concentration of standard BSA and absorbance

Standard No.	$\mu\text{g BSA}$	Absorbance
1	0.0	0.000
2	2.5	0.097
3	5.0	0.150
4	7.5	0.208
5	10.0	0.249
6	12.5	0.291
7	15.0	0.367
8	17.5	0.379
9	20.0	0.427

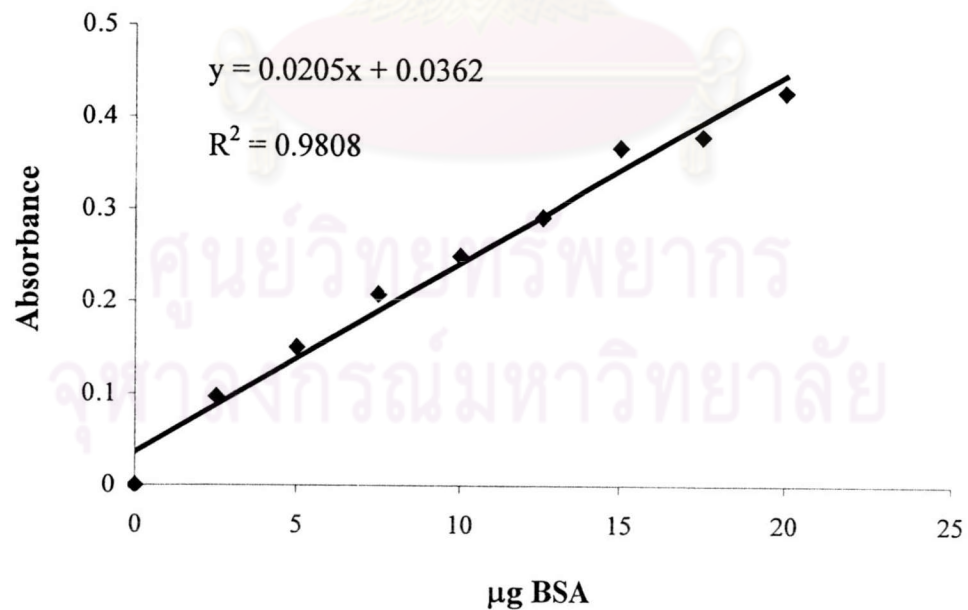


Figure A3 Relationship between standard protein (BSA) concentration and optical density (absorbance) at 595 nm.

5. Preparation calibration curve of N-acetyl-D-glucosamine for chitinolytic enzyme assay by colorimetric method

Calibration curve for GlcNAc was made by determining the absorbance value at 420 nm of standard GlcNAc according to the method of Schales.

Table A4 The amount of standard solution of GlcNAc and Δ Absorbance

Standard No.	Amount of GlcNAc (μ mole)	Δ Absorbance
1	0.8029	0.886
2	0.7025	0.761
3	0.6021	0.650
4	0.5018	0.541
5	0.5018	0.516
6	0.4014	0.411
7	0.3011	0.311
8	0.2007	0.187
9	0.1004	0.094

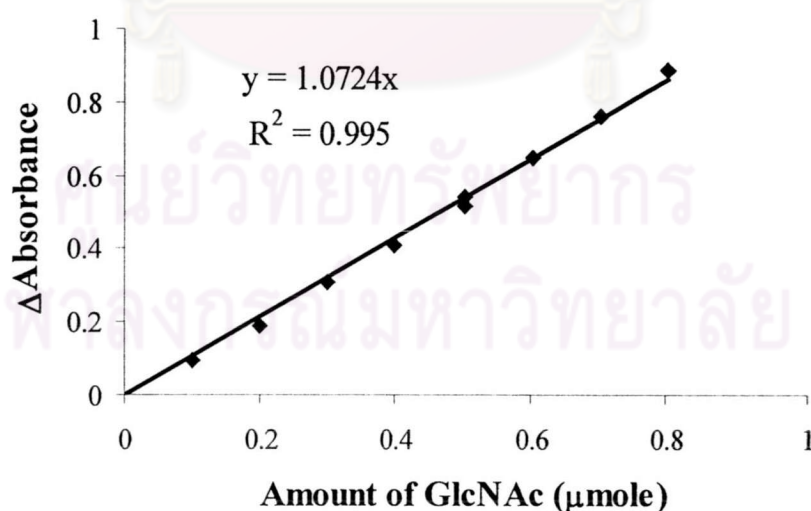


Figure A4 Correlation between amount of standard of N-acetyl-D-glucosamine and optical density (absorbance) at 420 nm.



APPENDIX B

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

Table B1 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of powder chitin on chitinolysis (The data in bold fonts were used in Figure 3.2).

[chitin] (mg/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
10.0	1	3.32	4.52	1.36
	2	6.06	6.98	1.15
	3	7.82	7.84	1.00
	4	10.23	8.85	0.87
20.0	1	3.35	6.84	2.04
	2	5.59	9.60	1.72
	3	7.39	11.02	1.49
	4	9.01	12.02	1.33
30.0	1	2.87	7.42	2.58
	2	4.95	10.93	2.21
	3	6.73	12.86	1.91
	4	8.34	14.64	1.76
40.0	1	2.83	7.80	2.76
	2	4.64	11.08	2.39
	3	8.11	14.90	1.84
	4	8.75	15.44	1.76
50.0	1	2.66	8.06	3.03
	2	4.24	11.28	2.66
	3	7.55	15.62	2.07
	4	9.0	17.88	1.99
60.0	1	2.38	7.80	3.27
	2	3.92	11.22	2.86
	3	7.01	15.54	2.22
	4	7.98	17.01	2.13
80.0	1	1.19	5.02	4.22
	2	3.56	10.95	3.07
	3	5.14	14.55	2.83
	4	6.10	16.49	2.70

Table B1 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of powder chitin on chitinolysis (continued).

[chitin] (mg/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
100.0	1	1.48	5.19	3.52
	2	2.46	7.72	3.14
	3	3.80	11.05	2.90
	4	4.85	13.67	2.82
200.0	1	0.90	3.06	3.39
	2	1.37	4.79	3.49



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

Table B2 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of fibrous chitin on chitinolysis (The data in bold fonts were used in Figure 3.3).

[chitin] (mg/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
10.0	1	3.39	3.53	1.04
	2	5.49	4.31	0.79
	3	7.55	4.68	0.62
	5	11.04	4.50	0.41
	20.0	1	3.13	3.94
2		5.01	5.96	1.19
3		7.04	7.37	1.05
5		10.25	8.69	0.85
30.0		1	3.28	5.60
	2	4.79	7.76	1.62
	3	6.57	9.62	1.46
	5	9.06	10.79	1.19
	60.0	1	2.68	5.81
2		3.98	8.99	2.26
3		5.25	10.89	2.07
5		8.92	15.97	1.79

Table B3 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of cellulase *Ac* on chitinolysis (The data in bold fonts were used in Figure 3.4).

[cellulase <i>Ac</i>] (mU/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
130.0	1	1.09	3.86	3.54
	3	3.09	10.01	3.24
	5	3.89	12.12	3.12
	9	5.22	14.43	2.76
520.0	1	4.68	10.56	2.26
	3	12.06	21.54	1.79
	5	17.96	29.66	1.65
	9	30.84	40.04	1.30
780.0	1	7.02	14.53	2.07
	3	17.48	29.04	1.66
	5	26.99	40.21	1.49
	9	44.93	49.40	1.10
1040.0	1	9.11	15.98	1.75
	3	21.86	30.37	1.39
	5	30.49	38.41	1.26
	9	52.16	47.48	0.91

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

Table B4 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of cellulase *Ac* on chitinolysis (The data in bold fonts were used in Figure 3.5).

[cellulase <i>Ac</i>] (mU/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
159.5	1	2.83	5.64	1.99
	3	5.58	8.92	1.60
	5	10.42	16.48	1.58
	9	7.94	11.92	1.50
319.0	1	4.98	8.71	1.75
	3	9.20	13.64	1.48
	5	19.06	28.24	1.48
	9	17.02	23.16	1.36
638.0	1	7.70	11.26	1.46
	3	18.30	21.14	1.16
	5	33.12	36.12	1.09
	9	28.62	26.64	0.93
957.0	1	10.41	13.81	1.33
	3	24.34	24.60	1.01
	5	51.74	48.72	0.94
	9	45.54	36.30	0.80
1276.0	1	12.55	17.17	1.37
	3	31.64	30.96	0.98
	5	47.80	42.10	0.88
	9	55.44	38.80	0.70

Table B5 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of pH of the reaction solution on chitinolysis (The data in bold fonts were used in Figure 3.6).

McIlvaine buffer (pH)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
2.0	1	2.70	2.52	0.93
	3	6.96	5.56	0.80
	5	19.98	14.30	0.72
3.0	1	9.11	13.82	1.52
	3	17.74	21.91	1.24
	5	27.76	31.96	1.15
4.0	1	9.13	12.08	1.32
	3	16.45	17.17	1.04
	5	35.97	31.43	0.87
5.0	1	6.01	7.61	1.27
	3	12.47	11.83	0.95
	5	20.66	18.25	0.88

Table B6 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of buffer on chitinolysis (The data in bold fonts were used in Figure 3.7).

[McIlvaine buffer] (M)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
0.0	1	1.96	0.82	0.42
	3	1.29	1.37	1.06
	9	3.30	3.90	1.18
0.05	1	6.49	11.40	1.76
	3	13.56	18.29	1.35
	5	15.61	23.64	1.51
	9	33.81	30.60	0.91
0.1	1	8.25	15.61	1.89
	3	15.70	24.62	1.57
	5	24.75	32.78	1.32
	9	39.49	38.00	0.96
0.2	1	8.42	18.57	2.21
	3	19.90	31.36	1.58
	5	24.55	37.71	1.54
	9	37.93	42.10	1.11
0.4	1	6.42	15.97	2.49
	3	14.74	27.80	1.89
	9	26.65	36.20	1.36

Table B7 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of temperature on chitinolysis (The data in bold fonts were used in Figure 3.8).

Temperature (°C)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
30	1	7.52	15.47	2.06
	3	16.72	27.42	1.64
	5	22.85	32.35	1.42
	9	33.23	45.26	1.36
37	1	7.02	14.53	2.07
	3	17.48	29.04	1.66
	5	26.99	40.21	1.49
	9	44.93	49.40	1.10
45	1	11.81	17.95	1.52
	3	23.86	32.02	1.34
	5	32.62	37.89	1.16
	9	44.87	44.11	0.98
55	1	2.14	7.31	3.42
	3	16.67	5.50	0.33
	5	22.42	4.54	0.20

Table B8 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of enzyme affinity technique on chitinolysis (The data in bold fonts were used in Figure 3.9).

Chilling period (hrs)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
0	1	1.18	5.37	4.57
	2	2.21	8.23	3.73
	3	2.74	9.64	3.52
	4	3.28	10.87	3.31
1	1	1.36	5.60	4.11
	2	2.32	8.08	3.49
	3	3.00	9.82	3.27
	4	3.48	10.79	3.10
4	1	0.92	4.57	4.99
	2	1.58	6.59	4.16
	3	2.31	8.90	3.85
	4	2.78	10.20	3.67
24	1	1.18	5.37	4.55
	2	2.09	8.15	3.90
	3	2.65	9.71	3.66
	4	3.19	11.08	3.47

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

Table B9 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of fibrous chitin on chitinolysis (The data in bold fonts were used in Figure 3.10).

[chitin] (mg/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
10.0	1	0.16	4.67	28.84
	2	0.35	6.63	19.04
	3	0.57	8.06	14.19
	4	0.61	8.97	14.68
	7	0.79	9.73	12.27
20.0	1	0.12	5.24	44.42
	2	0.26	7.25	28.09
	3	0.43	9.28	21.74
	4	0.57	10.57	18.64
	7	0.76	11.61	15.33
30.0	1	trace	3.17	-
	2	trace	6.17	-
	3	trace	8.66	-
	4	0.18	7.92	43.75
	7	0.42	10.67	25.64
40.0	6	0.25	10.06	40.89
	7	0.25	7.82	31.41

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

Table B10 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of Chi 60 on chitinolysis (The data in bold fonts were used in Figure 3.11).

[cellulase <i>Ac</i>] (mU/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
4.0	1	Not observed	4.35	-
	2	Not observed	5.04	-
	3	Not observed	5.14	-
	6	Not observed	6.41	-
	8	Not observed	6.48	-
5.0	1	Not observed	5.30	-
	2	Not observed	6.97	-
	3	Not observed	7.05	-
	6	Not observed	9.06	-
	8	Not observed	9.08	-
6.55	1	Not observed	6.37	-
	2	Not observed	8.46	-
	3	Not observed	9.20	-
	6	Not observed	11.29	-
	8	Not observed	11.07	-
9.32	1	Not observed	7.70	-
	2	Not observed	9.95	-
	3	Not observed	10.63	-
	6	Not observed	14.61	-
	8	Not observed	17.10	-
11.0	1	Trace	9.22	-
	2	Trace	12.29	-
	3	Trace	13.14	-
	6	trace	16.98	-
	8	Trace	18.44	-

Table B10 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of Chi 60 on chitinolysis (continued).

[cellulase <i>Ac</i>] (mU/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
13.0	1	Trace	10.24	-
	2	Trace	12.59	-
	3	Trace	14.26	-
	6	Trace	19.72	-
	8	Trace	19.79	-
15.0	1	Trace	11.77	-
	2	Trace	14.39	-
	3	Trace	15.34	-
	6	Trace	22.19	-
	8	Trace	22.11	-
30.0	1	0.97	15.34	15.81
	2	1.41	19.21	13.62
	3	1.58	21.33	13.50
	6	1.89	23.60	12.49
	8	1.97	23.74	12.05
50.0	1	1.80	20.53	11.41
	2	2.54	24.99	9.84
	3	2.93	27.04	9.23
	6	3.61	31.53	8.73
	8	3.65	32.48	8.90
100.0	1	3.85	27.02	7.02
	2	5.32	32.86	6.18
	3	5.84	34.95	5.98
	6	6.29	37.99	6.04
	8	6.13	36.94	6.03

Table B10 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of Chi 60 on chitinolysis (continued).

[cellulase <i>Ac</i>] (mU/mL)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
150.0	1	9.31	26.66	2.86
	2	17.72	38.88	2.19
	3	20.46	41.30	2.02
	6	26.32	46.40	1.76
	8	26.60	44.98	1.69
250.0	1	7.82	38.26	4.89
	2	17.25	41.05	2.38
	3	17.36	40.68	2.34
	6	21.56	42.10	1.95
	8	21.70	41.50	1.91
350.0	1	15.12	36.64	2.42
	2	20.85	42.40	2.03
	3	20.40	40.00	1.96
	6	29.46	39.52	1.34
	8	34.90	38.88	1.11

Table B11 The yield of $(\text{GlcNAc})_2$ by the effect of pH of the reaction solution on chitinolysis (The data in bold fonts were used in Figure 3.12).

McIlvaine buffer (pH)	Time (day)	$[(\text{GlcNAc})_2]$ (mM)
3.0	1	0.10
	2	0.11
	3	0.13
	4	0.14
4.0	1	0.29
	2	0.26
	3	0.27
	4	0.28
5.0	1	0.88
	2	0.98
	3	1.16
	4	1.31
6.0	1	1.65
	2	2.03
	3	2.21
	4	2.40
7.0	1	0.93
	2	1.43
	3	1.70
	4	2.05

Table B12 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of concentration of buffer on chitinolysis (The data in bold fonts were used in Figure 3.13).

[McIlvaine buffer] (M)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
0.0	3	1.75	14.58	8.33
	5	4.93	34.94	7.09
	9	7.42	40.93	5.52
0.05	3	2.19	15.34	7.00
	5	5.46	34.39	6.30
	9	8.05	40.06	4.98
0.1	3	2.33	15.25	6.55
	5	5.93	34.45	5.81
	9	8.59	40.25	4.69
0.2	3	1.93	13.64	7.07
	5	5.01	31.32	6.25
	9	6.87	35.89	5.22
0.4	3	1.69	10.31	6.10
	5	-	26.00	-
	9	-	23.59	-

Table B13 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of temperature on chitinolysis (The data in bold fonts were used in Figure 3.14).

Temperature (°C)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc
32	1	8.26	28.33	3.43
	2	12.48	35.00	2.80
	3	13.63	37.52	2.75
	6	15.86	41.82	2.64
	8	15.68	36.16	2.31
37	1	9.31	26.66	2.86
	2	17.72	38.88	2.19
	3	20.46	41.30	2.02
	6	26.32	46.40	1.76
	8	26.60	44.98	1.69
45	1	18.59	38.35	2.06
	2	23.38	38.72	1.66
	3	25.20	39.12	1.55
	6	29.98	40.18	1.34
	8	30.00	36.14	1.20
50	1	13.61	32.49	2.39
	2	11.68	33.92	2.90
	3	18.56	36.56	1.97
	6	19.16	37.60	1.96
	8	21.78	39.46	1.81

Table B14 The yield of GlcNAc, (GlcNAc)₂ and mole ratio of (GlcNAc)₂/GlcNAc by the effect of enzyme affinity technique on chitinolysis (The data in bold fonts were used in Figure 3.15).

Chilling period (hrs)	Time (day)	[GlcNAc] (mM)	[(GlcNAc) ₂] (mM)	Mole ratio (GlcNAc) ₂ /GlcNAc	
Non adsorption	1	9.31	26.66	2.86	
	2	17.72	38.88	2.19	
	4	20.46	41.30	2.02	
	6	26.32	46.40	1.76	
	8	26.60	44.98	1.69	
	0	1	0.95	8.12	8.55
		2	3.24	11.10	3.43
		4	3.63	17.04	4.69
6		5.64	22.87	4.06	
8		5.61	23.19	4.14	
1	1	0.93	7.59	8.16	
	2	2.28	10.82	4.75	
	4	2.31	16.78	7.26	
	6	4.83	20.64	4.27	
	8	5.39	21.70	4.02	
3	1	0.85	7.04	8.28	
	2	2.84	9.95	3.50	
	4	4.46	16.30	3.65	
	6	4.92	21.93	4.46	
	8	6.05	23.08	3.81	
24	1	0.74	11.32	15.30	
	2	3.04	13.75	4.52	
	4	4.76	20.06	4.22	
	6	6.53	24.45	3.75	
	8	6.98	26.32	3.77	

VITAE

Miss Wasinee Prakobkij was born on February 15th, 1978 in Bangkok, Thailand. She received a Bachelor Degree of Science, majoring in Chemistry from Mahidol University, in 1999. Since 2000, she has been a graduate student studying Petrochemistry and polymer Science as her major course at Chulalongkorn University. During her studies towards the Master's Degree, she was supported by a research grant for her Master degree's thesis from the Graduate School, Chulalongkorn University.

Her present address is 161/11 Soi Watpracharabuethum, Rama V Rd., thanonnakhonchaisri, Dusit, Bangkok, Thailand 10300.



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย