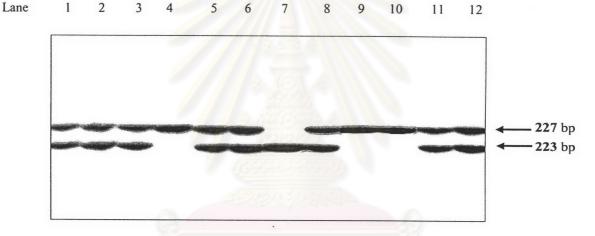
CHAPTER V

RESULTS

1. 5' – end labeling primer (kinase reaction) of TGF- β 2 gene

1.1 5' – end labeling primer analysis of TGF- β 2; an insertion ACAA in the 5' - untranslated region (5' UTR) at position + 71_72

Polymorphism of a 4-bp insertion $+71_{72}$ ACAA in the 5' – untranslated region (5' UTR) of the transforming growth factor beta2 (TGF- β 2) gene were identified by the 5' – end labeling primer method. The patterns are indicated by arrows (FigureA).



FigureA. The representative of 5' – end labeling primer from samples with homozygous 4-bp insertion (ACAA), heterozygous of 4-bp insertion (ACAA) and homozygous for the common allele

Lane 1 is heterozygous positive control for the +71_72 insACAA

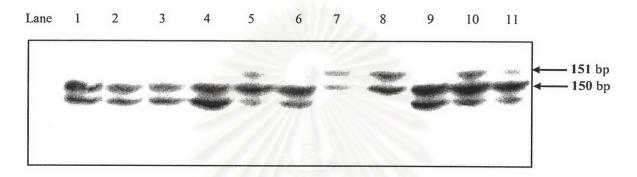
Lenes 2, 3, 5, 6, 8, 11, and 12 are heterozygous for the +71_72 insACAA

Lanes 7 is homozygous for the common allele

Lanes 4, 9 and 10 are homozygous for the +71 72insACAA

1.12 5' – end labeling primer analysis of TGF- β 2; an insertion A in the intron6 region at position + 94400_94401

Polymorphism of a 1-bp insertion+ 94400_94401 insA in the intron6 region of the transforming growth factor beta2 (TGF- β 2) gene were identified by the 5' – end labeling primer method. The patterns are indicated by arrows (FigureB).



FigureB. The representative of 5' – end labeling primer from samples with homozygous 1-bp insertion (A), heterozygous of 1-bp insertion (A) and homozygous for the common allele.

Lanes 5, 10 and 11 are heterozygous for the insertion A.

Lanes 1, 2, 3, 4, 6 and 9 are homozygous for the common allele.

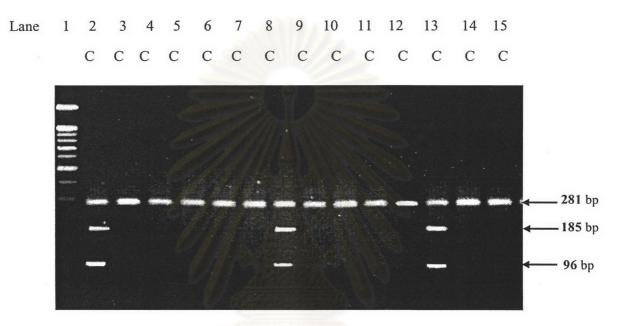
Lanes 7 and 8 are homozygous for the insertion A.

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

2. Polymerase Chain Reaction - Restriction Fragment Length Polymorphism analysis of TGF-\$\beta2\$ gene

2.1 PCR-RFLP analysis of TGF-β2 at position + 720

Polymorphism at +720T/G in the intron1 region of the TGF- β 2 were identified by the PCR-RFLP method. If a G was present at this position, the TaaI (Tsp4CI) restriction enzyme would cut the 281 bp PCR product into two fragments; 185 and 96 bp. No digestion would occur if a T was present. (FigureC).



FigureC. The representative of PCR-RFLP results from samples with homozygous of + 720T and heterozygous + 720T/G.

Lane 1 is 100 bp molecular marker.

Lane 2, 8 and 13 are heterozygous of + 720T/G.

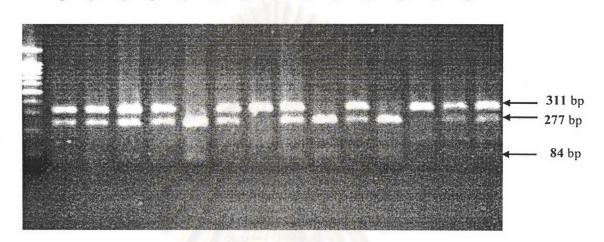
Lane 3, 4, 5, 6, 7, 9, 10, 11, 12, 14 and 15 are homozygous of + 720T.

No homozygous of G in this position.

2.2 PCR-RFLP analysis of TGF-β2 at position + 89835

Polymorphism at + 89835A/G in the intron5 region of the TGF- β 2 were identified by the PCR-RFLP method. If a G was present at this position, the FspBI (MaeI) restriction enzyme would cut the 311 bp PCR product into two fragments; 227 and 84 bp. No digestion would occur if an A was present. (FigureD).

Lane 1 10 11 12 13 15 2 3 5 8 9 14 4 6 7 С С С С С С С С С С С С С С



FigureD. The representative of PCR-RFLP results from samples with homozygous

+ 89835A, heterozygous+ 89835 A/G and homozygous + 89835G

Lane 1 is 100 bp molecular marker.

Lane 5 and 13 are homozygous of + 89835 A.

Lane 2, 3, 4, 5, 7, 9, 11, 14 and 15 are heterozygous of + 89835 A/G.

Lane 6, 10 and 12 are homozygous of + 89835 G.

3. The association results of TGF- $\beta 2$ gene polymorphisms with SLE

We assessed the quality of the genotype data by testing for Hardy-Weinberg equilibrium in the sample, using Fisher's exact test (p < 0.05). There are three significant deviations from Hardy-Weinberg equilibrium in all SNPs in this study.

3.1 TGF-β2 gene polymorphism at position +71_72insACAA

Genotype and allele frequencies for $+71_72$ insACAA at the 5' UTR of TGF- β 2 gene in healthy controls and SLE patients were shown in table 9 and 10. Thirty-nine of 133 healthy controls (29.3%) were homozygous for the common _/_ genotype, sixty-nine (51.9%) were heterozygous for the _/insACAA and twenty-five (18.8%) were homozygous for the insACAA/insACAA genotype. The allele frequencies were 55.3% for common _ allele and 44.7% for ACAA allele. In comparison, sixty-six of 155 SLE patients (43.1%) were homozygous for the common _/_ genotype, sixty-five (42.5%) were heterozygous for the _/insACAA and twenty-two (14.4%) were homozygous for the insACAA/insACAA genotype. The allele frequencies were 64.4% for common _ allele and 35.6% for ACAA allele. The +71_72 allele at the 5' UTR of TGF- β 2 gene was found to be significantly increased in SLE patients compared to healthy controls (p = 0.048, OR = 1.43, 95%CI = 1.00-2.03). The effect of +71_72 allele at the 5' UTR of TGF- β 2 gene was similar to autosomal recessive mode of inheritance. The presence of one common _/_ genotype conferred with the significant OR of 1.83 (p = 0.02, OR = 1.83, 95%CI = 1.09-3.08).

3.2 TGF-β2 gene polymorphism at position +720 (T/G)

Genotype and allele frequencies for +720 (T/G) at the intron1 of TGF- β 2 gene in healthy controls and SLE patients were shown in table 11 and 12. Xero of 133 healthy controls (0.0%) were homologous for the G/G genotype, twenty-nine (21.8%) were heterozygous for the G/T and one-hundred and four (78.2%) were homozygous for the T/T genotype. The allele frequencies were 10.9% for G allele and 89.1% for T allele. In comparison, xero of 155 SLE patients (0.0%) were homozygous for the G/G genotype, fifteen (9.8%) were heterozygous for the G/T and one-hundred and thirty- eight (90.2%) were homozygous for the T/T genotype. The allele frequencies were homozygous for the T/T genotype. The allele frequencies were 4.9% for G allele and 95.1% for T allele. The +720 T allele at the intron1 of TGF- β 2 gene was found to be significantly increased in SLE patients compared to healthy controls (p = 0.01, OR = 2.37,

95%CI = 1.19-4.77). The effect of +720 T allele at the intron1 of TGF- β 2 gene was similar to autosomal recessive mode of inheritance. The presence of one genotype (T/T) conferred with the significant OR of 2.57 (*p* =0.008, OR = 2.57, 95%CI = 1.25-5.32).

3.3 TGF-\beta2 gene polymorphism at position +89835 A/G

Genotype and allele frequencies for +89835 A/G at the intron5 of TGF- β 2 gene in healthy controls and SLE patients were shown in table 13 and 14. Thirteen of 133 healthy controls (9.8%) were homozygous for the A/A genotype, sixty-nine (51.9%) were heterozygous for the A/G and fifty-one (38.3%) were homozygous for the G/G genotype. The allele frequencies were 35.7% for A allele and 64.3% for G allele. In comparison, twenty-eight of 155 SLE patients (18.3%) were homozygous for the A/A genotype, sixty-nine (45.1%) were heterozygous for the A/A genotype, sixty-nine (45.1%) were heterozygous for the A/G and fifty-six (36.6%) were homozygous for the G/G genotype. The allele frequencies were 40.9% for A allele and 59.1% for G allele. There were no significant differences in allele and genotype frequency of the +89835 (A/G) polymorphism at the intron5 of TGF- β 2 gene between patients with SLE and healthy controls.

3.4 TGF-B2 gene polymorphism at position +94400 944001insA

Genotype and allele frequencies for +94400_944001insA at the intron6 of TGF- β 2 gene in healthy controls and SLE patients were shown in table 15 and 16. Eighty-eight of 133 healthy controls (66.2%) were homologous for the common _/_ genotype, fourty-one (30.8%) were heterozygous for the _/insA and four (3.0%) were homozygous for the insA/insA genotype. The allele frequencies were 81.6% for common _ allele and 18.4% for A allele. In comparison, one-hundred and thirty-two of 155 SLE patients (86.2%) were homozygous for the common _/_ genotype, twenty (13.1%) were heterozygous for the _/insA and one (0.7%) were homozygous for the insA/insA genotype. The allele frequencies were 92.8% for wild type _ allele and 7.2% for A allele. The +94400_944001 _(common) allele at the intron6 of TGF- β 2 gene was found to be significantly increased in SLE patients compared to healthy controls (p = 0.00008, OR = 2.91, 95%CI = 1.66-5.15). The effect +94400_944001 _ allele at the intron6 of TGF- β 2 gene was similar to autosomal recessive mode of inheritance. The presence of one common (_/_) genotype conferred with the significant OR of 1.83 (p = 0.0001, OR = 3.21, 95%CI = 1.73-6.02). 4. Haplotype analysis of TGF-β2 gene at position (+71_72insACAA, +720T/G, +89835A/G, +94400_94401insA), respectively

The haplotype frequencies of the TGF- β 2 gene polymorphism were also calculated by PHASE program. The haplotype frequencies in patients with SLE and normal controls were shown in table 17. In haplotype analysis of 4 positions (+71_72insACAA, +720T/G, +89835A/G, +94400_94401insA) of TGF- β 2 gene, we found 11 haplotypes; _/T/G/_, insACAA/T/A/_, insACAA/T/G/_, _/T/G/insA, _/T/A/_, insACAA/G/A/_, insACAA/G/G/_, insACAA/T/G/insA, insACAA/T/A/_insA, _/G/G/_ and _/T/A/insA in patients with SLE and normal concrols were shown in table 18. After comparing haplotype frequencies of the 4 positions of TGF- β 2 gene polymorphism between patients with SLE and normal controls, the _/T/G/insA, _/T/A/_ and insACAA/G/A/_ were found to be significantly increased in patient with SLE compared to normal controls (*p* = 0.01, OR = 0.49, 95%CI = 0.27-0.89, *p* = 0.001, OR = 2.64, 95%CI = 1.58-4.42 and *p* = 0.00005, OR = 0.05, 95%CI = 0.02-0.32), respectively.

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

	SLE patients	Healthy controls
	n = 153	n =133
Genotype frequencies		
/	66(43.1%)	39(29.3%)
_/ACAA(ins)	65(42.5%)	69(51.9%)
ACAA/ACAA(ins)	22(14.4%)	25(18.8%)
Allele frequencies		
- 2	197(64.4%) ^a	147(55.3%)
ACAA(ins)	109(35.6%)	116(44.7%)

Table9. Genotype and allele frequencies for TGF- β 2 polymorphism at position +71_72insACAA in healthy controls and SLE patients.

 $p^{a} p = 0.048, OR = 1.43, 95\%CI = 1.00-2.03$

Table10. Risk of SLE associated with TGF- β 2 (insACAA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	. n = 153	n =133
140	T	
_dominance, A(ins) wild type		
/ or _/ACAA(ins)	131(85.6%) ^a	108(81.2%)
ACAA/ACAA(ins)	22(14.4%)	25(18.8%)
_recessive, ACAA(ins) wild type		
1.911081058	66(43.1%) ^b	39(29.3%)
/ACAA or ACAA/ACAA(ins)	87(56.9%)	94(70.7%)

^a *p* =0.40

^b p = 0.02, OR = 1.83, 95%CI = 1.09-3.08

Table11. Genotype and allele frequencies for TGF- β 2 polymorphism at position +720 (T/G) in healthy controls and SLE patients.

	SLE patients	Healthy controls
	n = 153	n =133
Genotype frequencies		
G/G	0(0.0%)	0(0.0%)
G/T	15(9.8%)	29(21.8%)
T/T	138(90.2%)	104(78.2%)
Allele frequencies		
т	291(95.1%) ^a	237(89.1%)
G	15(4.9%)	29(10.9%)

^a *p* = 0.01, OR = 2.37, 95%CI = 1.19-4.77

Table12. Risk of SLE associated with TGF- β 2 (T/G) genotype according to different models of inheritance.

	SLE patients n = 153	Healthy controls n =133
T dominance, G wild type	Å	1
T/T or G/T	153(100.0%)	133(100.0%)
G/G	0(0.0%)	0(0.0%)
T recessive, G wild type		
T/T	138(90.2%) ^b	104(78.2%)
G/G or G/T	15(9.8%)	29(21.8%)

 b p =0.008, OR = 2.57, 95%CI = 1.25-5.32

	SLE patients	Healthy controls
	n = 153	n =133
Genotype frequencies		
A/A	28(18.3%)	13(9.8%)
A/G	69(45.1%)	69(51.9%)
G/G	56(36.6%)	51(38.3%)
Allele frequencies		
A	125(40.9%) ^a	95(35.7%)
G	181(59.1%)	171(64.3%)

Table13. Genotype and allele frequencies for TGF- β 2 polymorphism at position +89835 (A/G) in healthy controls and SLE patients.

Table 14. Risk of SLE associated with TGF- $\beta 2$ (A/G) genotype according to different models of inheritance.

	SLE patients n = 153	Healthy controls n =133
A dominance, G wild type	Ă	
AA or A/G	97(63.4%) ^a	82(61.7%)
G/G	56(36.6%)	51(38.3%)
A recessive, G wild type		
A/A	28(18.3%) ^b	13(9.8%)
A/G or G/G	125(81.7%)	120(90.2%)

^b p =0.06

	SLE patients	Healthy controls	
	n = 153	n =133	
enotype frequencies			
/	132(86.2%)	88(66.2%)	
_/A(ins)	20(13.1%)	41(30.8%)	
A/A(ins)	1(0.7%)	4(3.0%)	
llele frequencies			
-	284(92.8%) ^a	217(81.6%)	
A(ins)	22(7.2%)	49(18.4%)	

Table15. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in healthy controls and SLE patients.

^a *p* = 0.00008, OR = 2.91, 95%CI = 1.66-5.15

Table 16. Risk of SLE associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
Aug.	n = 153	n =133
dominance, A(ins) wild type		
/ or _/A(ins)	152(99.3%) ^ª	129(97.0%)
A/A(ins)	1(0.7%)	4(3.0%)
recessive, A(ins) wild type		
/	132(86.3%) ^b	88(66.2%)
/A or A/A(ins)	21(13.7%)	45(33.8%)

° *p* =0.29

^b p = 0.0001, OR = 3.21, 95%CI = 1.73-6.02

Haplotype frequencies	SLE patients (n=306)	Healthy controls (n=266)
/T/G/	107 (35.0%)	79 (29.7%)
insACAA/T/A/_	55 (18.0%)	44 (16.5%)
insACAA/T/G/_	36 (11.8%)	39 (14.7%)
_/T/G/insA	22 (7.2%)	36 (13.5%)
_/T/A/ _	68 (22.2%)	26 (9.8%)
insACAA/G/A/_	1 (0.3%)	18 (6.8%)
insACAA/G/G/_	13 (4.2%)	9 (3.4%)
insACAA/T/G/insA	2 (0.7%)	6 (2.3%)
insACAA/T/A/insA	1 (0.3%)	3 (1.1%)

Table17. Haplotype frequencies of the TGF- β 2 polymorphism (+71_72insACAA, +720T/G, +89835 A/G, +94400_94401insA respectively) between normal controls and SLE patients.

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

1 (0.3%)

0 (0%)

3 (1.1%)

3 (1.1%)

/G/G/

_/T/A/insA

Table 18. Association of the TGF- β 2 polymorphism (+71_72insACAA, +720T/G, +89835 A/G, +94400_94401insA respectively) between normal controls and SLE patients.

Aplotype frequencies	SLE patients	Healthy controls	p-value
	(n=306)	(n=266)	
/T/G/	107 (35.0%)	79 (29.7%)	0.21
other haplotype	199 (65.0(%)	187 (70.3%)	
insACAA/T/A/_			
other haplotype	55 (18.0%)	44 (16.5%)	0.73
	251(82.0%)	222(83.5%)	
insACAA/T/G/_	36 (11.8%)	39 (14.7%)	0.37
other haplotype	270(88.2%)	227(85.3%)	
_/T/G/insA	22 (7.2%) ^a	36 (13.5%)	0.02
other haplotype	284(92.8%)	230(86.5%)	
_/T/A/ _	68 (22.2%) ^b	26 (9.8%)	0.0001
other haplotype	238(77.8%)	240(%)	
		233/4	
insACAA/G/A/_	1 (0.3%) ^c	18 (6.8%)	0.00005
other haplotype	305(99.7%)	248(90.2%)	
insACAA/G/G/_	13 (4.2%)	9 (3.4%)	0.75
other haplotype	293(95.8%)	257(%)	0.75
insACAA/T/G/insA	2 (0.7%)	6 (2.3%)	0.20
other haplotype	304(99.3%)	260(96.6%)	5
insACAA/T/A/insA	1 (0.3%)	3 (1.1%)	0.52
other haplotype	305(99.7%)	263(%)	
	61 V 1 1 6 6 16	A PITO PIC	1915
/G/G/	1 (0.3%)	3 (1.1%)	0.52
other haplotype	305(99.7%)	263(98.9%)	
/T/A/insA	0 (0%)	3 (1.1%)	0.20
other haplotype			0.20
1 .71-	306(100.0%)	263(98.9%)	

^a p = 0.01, OR = 0.49, 95%CI = 0.27-0.89 ^b p = 0.0001, OR = 2.64, 95%CI = 1.58-4.42 ^c p = 0.00005, OR = 0.05, 95%CI = 0.02-0.32

5. Linkage Disequilibrium (LD)

Linkage Disequilibrium coefficients (|D'| and r^2) among TGFB2 SNP at position + 71_72 ins(ACAA), + 720 (T/G), + 89835 (A/G) and + 94400_94401ins(A). Data was shown in table 19 (see appendix F).

Table19. Linkage disequilibrium coefficients (D and r²) among TGFB2 SNPs

	+ 71_72 ins	+ 720	+ 89835	+ 94400_94401ins
+ 71_72 ins	- /	0.8467	0.0788	0.4610
+ 720	0.0649	11 strait	0.6597	1.0000
+ 89835	0.0051	0.0155	-	0.7360
+94400_94401ins	0.0094	0.0042	0.0290	-

6. The association results of TGF-\$2 gene polymorphisms with clinical manifestation SLE

We analyze the association between clinical manifestation in patients with SLE and polymorphism of the position $+71_{72insACAA}$, +720T/G, +89835 A/G and $+94400_{94401insA}$ of TGF- β 2 gene by using chi-square test and odds ratio.

6.1 Clinical manifestation of SLE patients

The clinical expression of SLE is tremendously varied among individuals. In this study, we obtained clinical data of 127 patients, as shown in table20.

6.2 The 4 positions (+71_72insACAA, +720T/G, +89835 A/G and +94400_94401insA) of TGF-β2 gene polymorphisms and clinical presentation of SLE

There is one significant association between insA allele at position $+94400_{94401insA}$. The association is with cellular cast (p = 0.025, OR = 3.34, 95%CI = 1.05-10.46).

Clinical manifestation	No. of patients with SLE (%)
1. Malar rash	72 (56.7%)
2. Discoid rash	36 (28.3%)
3. Photosensitivity	47 (37.0%)
4. Oral ulcers	51 (40.1%)
5. Arthritis	84 (66.1%)
6. Proteinuria	82 (64.6%)
7. Cellular cast	26 (20.5%)
8. Anemia	64 (50.4%)
9. Leukopenia	44 (34.6%)
10. Lymphopenia	47 (37.0%)
11. Thrombocytopenia	6 (4.7%)
12. Anti-DNA antibodies	29 (22.8%)

Table20. Clinical manifestation of patients with SLE in this study

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย

	SLE patients with malar rash	Healthy controls without malar rash
	n = 72	n =55
Genotype frequencies		
/	65(90.3%)	47(85.5%)
_/A(ins)	7(9.7%)	7(12.7%)
A/A(ins)	0(0.0%)	1(1.8%)
Allele frequencies		
-	137(95.1%)	101(91.8%)
A(ins)	7(4.9%)	9(8.2%)

Table21. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with malar rash.

Table22. Risk of malar rash associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with malar rash	without malar rash
	n = 72	n =55
	โกิทยุทรัพเ	ยากร
dominance, A(ins) wild type		
/ or _/A(ins)	72(100.0%) ^a	54(98.2%)
A/A(ins)	0(0.0%)	1(1.8%)
recessive, A(ins) wild type		
1	65(90.3%) ^b	47(85.5%)

 $^{a} p = 0.43$

 $^{b} p = 0.58$

	SLE patients with malar rash	Healthy controls without malar rash
	n = 36	n =91
Genotype frequencies		
/	32(88.9%)	80(87.9%)
_/A(ins)	4(11.1%)	10(11.0%)
A/A(ins)	0(0.0%)	1(1.1%)
Allele frequencies		
-	68(94.4%)	170(93.4%)
A(ins)	4(5.6%)	12(6.6%)

Table23. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with discoid rash.

Table24. Risk of malar rash associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with malar rash $n = 36$	without malar rash n =91
dominance, A(ins) wild type	ไวิทยทรัพร	ากร
/ or _/A(ins)	36(100.0%) ^a	90(98.9%)
A/A(ins)	0(0.0%)	1(1.1%)
· · · · · · · · ·		
recessive, A(ins) wild type		
recessive, A(ins) wild type	32(88.9%) ^b	80(87.9%)

 $p^{a} p = 0.72$ $p^{b} p = 0.57$ Table25. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with photosensitivity.

	SLE patients with photosensitivity	Healthy controls without photosensitivity
	n = 47	n =80
Genotype frequencies		
/	41(87.2%)	71(88.8%)
_/A(ins)	6(12.8%)	8(10.0%)
A/A(ins)	0(0.0%)	1(1.2%)
Allele frequencies		
_	88(93.6%)	150(93.8%)
A(ins)	6(6.4%)	10(6.2%)

Table26. Risk of photosensitivity associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with photosensitivity	without photosensitivity
×	n = 47	n =80
dominance, A(ins) wild type	วิทยทรัพเ	ยากร 🐳
/ or _/A(ins)	47(100.0%) ^a	79(98.8%)
A/A(ins)	0(0.0%)	1(1.2%)
recessive, A(ins) wild type		
/	41(87.2%)	71(88.8%)
/A or A/A(ins)	6(12.8%) ^b	9(11.2%)

° *p* =0.63

 $^{b} p = 0.98$

	SLE patients with oral ulcers n = 51	Healthy controls without oral ulcers n = 76
Genotype frequencies		
_/	45(88.2%)	67(88.2%)
_/A(ins)	6(11.8%)	8(10.5%)
A/A(ins)	0(0.0%)	1(1.3%)
Allele frequencies		
_	96(94.1%)	142(93.4%)
A(ins)	6(5.9%)	10(6.6%)

Table27. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with oral ulcers.

Table28. Risk of oral ulcers associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with oral ulcers	without oral ulcers
	n = 51	n =76
	วิทยุงวัพย	ากร
dominance, A(ins) wild type		
/ or _/A(ins)	51(00.0%) ^a	75(98.7%)
A/A(ins)	0(0.0%)	1(1.3%)
recessive, A(ins) wild type		
/	45(86.3%) ^b	67 (88.2%)
/A or A/A(ins)	6(13.7%)	9 (11.8%)

 $^{a} p = 0.60$

 $^{b} p = 0.79$

Table29. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with persistent proteinurie.

	SLE patients	Healthy controls
	with persistent proteinurie	without persistent proteinurie
	n = 82	n =45
Genotype frequencies		
/	73(89.0%)	39(86.7%)
_/A(ins)	8(9.8%)	6(13.3%)
A/A(ins)	1(1.2%)	0(0.0%)
Allele frequencies		
-	154(93.9%)	84(93.3%)
A(ins)	10(6.1%)	6(6.7%)

Table230. Risk of persistent proteinurie associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with persistent proteinurie	without persistent proteinurie
	n = 82	n =45
dominance, A(ins) wild type		
/ or _/A(ins)	81(98.8%) ^a	45(100.0%)
A/A(ins)	1(1.2%)	0(0.0%)
recessive, A(ins) wild type		
/	73(89.0%) ^b	39(86.7%)
/A or A/A(ins)	9(11.0%)	6(13.3%)

^a *p* =0.65

 $^{b}p = 0.92$

	SLE patients with cellular cast	Healthy controls without cellular cast
	n = 26	n =101
Genotype frequencies		
/	20(76.9%)	92(91.1%)
_/A(ins)	5(19.2%)	9(8.9%)
A/A(ins)	1(3.8%)	0(0.0%)
Allele frequencies		
	45(86.5%)	193(95.5%)
A(ins)	7(13.5%)	9(4.5%)

Table31. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with cellular cast.

Table32. Risk of cellular cast associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE pațients	Healthy controls
	with cellular cast	without cellular cast
	n = 26	n =101
dominance, A(ins) wild type	โกิญยาตรั้งแย	ากร
/ or _/A(ins)	25(96.2%)	101(97.0%)
A/A(ins)	1(3.8%) ^a	0(3.0%)
	1(3.8%) ^a	0(3.0%)
A/A(ins)	1(3.8%) ^a 20(76.9%)	0(3.0%) 92(91.1%)

^a *p* =0.20

^b p = 0.08, OR = 3.07, 95%CI = 0.85-10.92

	SLE patients with arthritis	Healthy controls	
		without arthritis	
	n = 84	n =43	
Genotype frequencies			
/	76(90.5%)	36(83.7%)	
_/A(ins)	8(9.5%)	6(14.0%)	
A/A(ins)	0(0.0%)	1(2.3%)	
Allele frequencies			
-	160(95.2%)	78(90.7%)	
A(ins)	8(4.8%)	8(9.3%)	

Table33. Genotype and allele frequencies for TGF- β 2 polymorphism at position +94400_94401insA in SLE patients with arthritis.

Table34. Risk of arthritis associated with TGF- β 2 (insA) genotype according to different models of inheritance.

	SLE patients	Healthy controls
	with arthritis	without arthritis
	n = 84	n =43
dominance ((inc) wild the		- W
dominance, A(ins) wild type		
/ or _/A(ins)	84(100.0%) ^a	42(97.0%)
A/A(ins)	0(0.0%)	1(3.0%)
recessive, A(ins) wild type		
/	76(90.5%) ^b	36(83.7%)
/A or A/A(ins)	8(9.5%)	7(16.3%)

^w p =0.34

 $^{b} p = 0.41$

7. The distribution of 5'UTR and Introns polymorphisms (+71_72insACAA, +720T/G, +89835 A/G and +94400_94401insA, respectively)

Besides association study of TGFB2 polymorphisms with SLE disease, this study also provides the basic knowledge of the frequencies of TGFB2 polymorphisms (position +71_72insACAA, +720T/G and +89835 A/G and, respectively) in Thai population. The distributions of the three positions polymorphisms between Thai population and Caucasian population previous reports were compared.

Pattern of TGFB2 gene polymorphisms (+71_72insACAA, +720T/G and +89835 A/G)

7.1 Pattern of TGFB2 at position +71_72insACAA

Allele frequencies for the polymorphism at +71_72insACAA in 5'UTR region of the TGFB2 gene were analyzed. The analyzed showed significant differences in allele frequencies between Thai and Caucasians population ($^{a}\chi^{2} = 19.35$, p<0.001).

7.2 Pattern of TGFB2 at position +720T/G

Allele frequencies for the polymorphism at +720T/G in intron1 region of the TGFB2 gene were analyzed. The analyzed showed no significant differences in allele frequencies between Thai and Caucasians population.

7.3 Pattern of TGFB2 at position 89835 A/G

Allele frequencies for the polymorphism at 89835 A/G in intron5 region of the TGFB2 gene were analyzed. The analyzed showed no significant differences in allele frequencies between Thai and Caucasians population. Table35. Allele and genotype frequencies of the TGFB2 promoter polymorphisms in healthy Thais individuals compared Caucasian population.

Gene/Allele/Genotype	This study (Thai)	Caucasian
+71_72insACAA ^a	N=133	N=187
allele	147 (55%)	273 (73%)
- ACAAins allele	116 (45%)	101 (27%)
/_ genotype	39 (29%)	102 (54%)
/ACAAins genotype	69 (52%)	16 (9%)
- ACAAins/ACAAins	25 (19%)	69 (37%)
genotype		
+720T/G ^b	N=133	N=12
- Tallele	237 (89%)	11 (91%)
- Gallele	29 (11%)	1 (9%)
- T/T genotype	104 (78%)	
- T/G genotype	29 (22%)	-
- G/G genotype	0 (0%)	
+89835 A/G [°]	N=133	N=60
- A allele	95 (36%)	97 (81%)
- G allele	171 (64%)	23 (19%)
- A/A genotype	13 (10%)	40 (67%)
- A/G genotype	69 (52%)	17 (28%)
- G/G genotype	51 (38%)	3 (5%)
+94400_94401insA	N=133	2
allele	217 (82%)	NON - REPORT
- Ains allele	48 (18%)	
 / genotype 	88 (66%)	
/Ains genotype	41 (31%)	
- Ains/Ains genotype	4 (3%)	

 ${}^{a}\chi^{2}$ =19.35, p<0.00001; compare between allele frequencies in Thai with Caucasian population (Alansari, Hajeer et al. 2001). ^bNot significant ; compare between genotype and allele frequencies in Thai with Caucasian population. ^c\chi^{2} =65.55, p<0.00000; compare between allele frequencies in Thai with Caucasian population.

ศูนย์วิทยทรัพยากร จุฬาลงกรณ์มหาวิทยาลัย