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APPENDICES

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APPENDIX A

RT-PCR AND SPECIFIC BUFFER

1. Preparation of the reaction mix for cDNA synthesis

Before starting the experiment thaw all reagents except of the polymerase, mix them thoroughly and centrifuge briefly. Use a sterile microcentrifuge tube to add the reagents to the reaction mix in a fixed order as outlined in the table

Table 2 Preparation of the reaction mix for cDNA synthesis

RT Reacion	Volume/1 sample (μ l)	Volume/2 sample (μ l)	Volume/3 sample (μ l)	Volume/4 sample (μ l)
10X Reaction Buffer	2	4	6	8
25mM MgCl ₂	4	8	12	16
dNTP	2	4	6	8
Random Primer	2	4	6	8
RNase Inhiitor	1	2	3	4
AMV Reverstranscriptase	1	2	3	4
Master Mix	12	24	36	48
Master Mix Aliquot	12	2x12	3x12	4x12
2 ug of RNA (Xi)	Xi	2xXi	3xXi	4xXi
RNase Free Water (20-12-Xi)	(20-12-Xi)	(20-12-Xi)	(20-12-Xi)	(20-12-Xi)
Total Volume	20	2x20	3x20	4x20

i = The number labeled in each sample

2. Preparation of the reaction mix for PCR

Before starting the experiment thaw all reagents except of the polymerase, mix them thoroughly and centrifuge briefly. Use a sterile microcentrifuge tube to add the reagents to the reaction mix in a fixed order as outlined in table 3.

Table 3 Preparation of the reaction mix for PCR

PCR Reaction	Volume/1 sample (μ l)	Volume/2 sample (μ l)	Volume/3 sample (μ l)	Volume/4 sample (μ l)
10X Reaction Buffer	2.5	5	7.5	10
25mM MgCl ₂	1.5	3	4.5	6
dNTP	0.5	1	1.5	2
Taq DNA Polymerase	0.2	0.4	0.6	0.8
Forward Primer	0.7	1.4	2.1	2.8
Reward Primer	0.7	1.4	2.1	2.8
Master Mix	6.1	12.2	18.3	24.4
Master Mix Aliquot	6.1	2x6.1	3x6.1	4x6.1
cDNA	5	2x5	3x5	4x5
RNase Free Water (25-6.1-5)	13.9	2x13.9	3x13.9	4x13.9
Total Volume/sample	25	2x25	3x25	4x25

3. Buffers preparations for RT-PCR

10x Ficoll loading buffer 10 ml

Ficoll	25	g
Bromphenol blue	0.025	g
0.5 M EDTA (pH 8.0)	0.2	ml

Adjust volume to 10 ml with dH₂O, Store at -20^oc

1.5% Agarose gel (w/v) 100 ml

Agarose	1.5	g
1x TBE	100	ml

Dissolved by heating and occasional mixing until no granules of agarose are visible.

Add ethidium bromide (stock 10 mg/ml) 10 μ l (final concentration 1 μ g/ml)

10x TBE buffer (pH 8.0) 1 liter

Tris base	108	g
EDTA 2H ₂ O (pH 8.0)	40	ml
dH ₂ O	800	ml
Slowly add the boric acid, anhydrous	55	g

Adjust the pH to 8.0 with conc.HCl

Adjust the volume to 1 liter with dH₂O

1x TBE buffer 1 liter

10x TBE buffer	100	ml
dH ₂ O	900	ml

Adjust the pH to 7.4 with conc. HCl

Adjust the volume to 1 liter with dH₂O

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APPENDIX B

ZYMOGRAM AND SPECIFIC BUFFER

Table 4 Preparing the solutions for Tris/Glycine SDS-Polyacrylamide Gel Electrophoresis for zymogram

Reagents	10%Separating gel (ml)	4%Stacking gel (ml)
H ₂ O	4.1	3
1.5 M Tris-HCl (pH 8.8)	2.5	-
0.5 M Tris-HCl (pH 6.8)	-	2.5
Gelatin/Casein	0.01 (g)	0.01 (g)
10% SDS	0.1	0.05
30% Acrylamide mix	3.25	0.7
10% APS	0.05	0.025
TEMED	0.004	0.004
Total Volume	10	5

1. Renaturing protein and developing protein

After running, washed the gel with 3 changes of 1x renaturing buffer for 15 min each. Decant the zymogram renaturing buffer and replace with 1x zymogram developing buffer. Equilibrate the gel for 30 min at room temperature with gentle agitation then replace with fresh 1x zymogram developing buffer and incubate at 37^oc overnight for maximum sensitivity. Stain gel with Coomassie Blue R-250 for 30 min. Areas of protease activity will appear as clear bands against a dark blue background where the protease has digested the substrate.

2. Buffer preparations for zymogram

2x non reducing sample buffer 10 ml

0.5 M Tris-HCl, pH 6.8	1.5	ml
Glycerol	2.5	ml
10% (w/v) SDS	4	ml
1% Bromophenol blue	0.1	ml
dH ₂ O	2.15	ml

5x Zymogram running buffer (pH 8.3) 600 ml

Tris base	9	g
Glycine	43.2	g
SDS	3	g
dH ₂ O	500	ml

Adjust the pH to 8.3 with conc. HCl and conc. NaOH

Adjust the volume to 600 sml with dH₂O

10x Renaturing buffer 100 ml

Triton X-100	25	ml
dH ₂ O	100	ml

10x developing buffer 1 liter

1 M Tris base	12.1	g
Tris-HCl	63	g
NaCl	117	g
50 mM CaCl ₂	7.4	g
Brij-35	2	g
dH ₂ O	1000	ml

Stain 0.5% coomassei blue 500 ml

Methanol	200	ml
Acetic acid	50	ml
Coomassei blue	2.5	g

Adjust the volume to 500 ml with dH₂O

Destain 500 ml

Methanol	25	ml
Acetic acid	37.5	ml

Adjust the volume to 500 ml with dH₂O



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BIOGRAPHY

Name	Miss Krongkan Nasee	Sex	Female
Birth date	Oct 28, 1979	Age	26
Nationality	Thai		
Place of birth	Chaing rai, Thailand		
Home address	1012/14, moo 12, Thumbon Weang, Meung, Chaing rai, 57000 Thailand		
Education	2001: Bachelor of Science (Biology) Kasetsat University		



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