

REFERENCES

- Abbt-Braun, G., Frimmel, F. H., and Schulten, H.-R., (1989). Structural investigations of aquatic humic substances by pyrolysis-field ionization mass spectrometry and pyrolysis-gas chromatography/mass spectrometry. Water Research. 23: 1579-1591.
- Aieta, M., (1998). The impacts of the Strontia Springs/Bi-City water exchange practice by Denver water on Thornton source water quality and the Columbine treatment plant performance. pp. 26+ Appendix D.
- Almendros, G., Dorado, J., Gonzalez-Vila, F. J., and Martin, F. (1997). Pyrolysis of carbohydrate-derived macromolecules: Its potential in monitoring the carbohydrate signature of geopolymers. Journal of Analytical and Applied Pyrolysis. 40: 599–610.
- Amirtharajah, A., Dennett, K.E., and Stustill A. (1993). Ferric chloride coagulation for removals of dissolved organic matter and trihalomethane precursors. Water Science and Technology. 27 (11): 113-121.
- Amy, G.L., Collins M.R., Kuo C.J., and King P.H. (1987). Comparing gel permeation chromatography and ultrafiltration for the molecular weight characterization of aquatic organic matter. Journal of American Water Works Association. 79(1): 43-49.
- Amy G.L. (1994). Using NOM characterization for the evaluation of treatment. In Natural organic matter in drinking water: origin, characterization, and removal. Workshop Proceedings, AWWRF, September 19-22, (1993) Chamonix, France, 19-26.
- AWWA (1993). Characterization of natural organic matter and its relationship to treatability, 1st Ed, AWWARF & AWWA, USA
- AWWA (1998). Removal of DBP Precursors by GAC Adsorption. Subject Area: Water treatment, published by the AWWA research foundation and AWWA, pp. 49
- AWWA (2000). Characterization of natural organic matter in drinking water. 1st Ed, AWWARF & AWWA, USA
- Baker A. (2001). Fluorescence excitation-emission matrix characterization of some sewage-impacted rivers. Environmental Science and Technology. 35: 948-953.
- Baker A. (2002). Fluorescence excitation–emission matrix characterization of river waters impacted by a tissue mill effluent. Environmental Science and Technology. 36: 1377-1382.
- Baker A. and Genty D. (1999). Fluorescence wavelength and intensity variations of cave waters. Journal of Hydrology. 217: 19-34.

- Barber, L.B., Leenheer, J.A., Noyes, T.I., and Stiles, E.A. (2001). Nature and transformation of dissolved organic matter in treatment wetlands. Environmental Science and Technology. 35: 4805-4816.
- Bellar, T.A., Lichtenberg, J.J., and Kroner, R.C. (1974). The occurrence of organohalides in chlorinated drinking water. Journal of American Water Works Association. 66 (12): 703-706.
- Biber, M. V., Gulacar, F. O., and Buffle, J. (1996). Seasonal Variations in Principal Groups of Organic Matter in a Eutrophic Lake Using Pyrolysis/GC/MS. Environmental Science and Technology. 30: 3501-3507.
- Bolto, B., Dixon D., Eldridge, R., King, S., and Linge, K. (2002). Removal of natural organic matter by ion exchange. Water Research. 36:5057-5065.
- Bruchet A. Application de la technique de pyrolyse]CG]SM a l'etude des matieres organiques non volatiles de l'eau. 'PhD. Thesis. Universite de Poitiers. Poitiers, France, 1985:78.
- Bruchet A., Anselme C., Duguet J.P., and Mallevialle L. (1987). THM formation potential and organic content: a new approach in water chlorination chemistry. Environmental Impact Health Effects. 6: 633-647.
- Bruchet, A., Rousseau, C., and Mallevialle, J. (1990). Pyrolysis-GC/MS for investigating high-molecular weight THM precursors and other refractory organics. Journal of American Water Works Association. 82 (9): 66-74.
- Chang, C-Y., Hsieh, Y-H., Lin, Y-M., Hu, P-Y., Liu, C-C., and Wang, K-H. (2001). The organic precursors affecting the formation of disinfection by-products with chlorine dioxide. Chemosphere. 44: 1153-1158.
- Chen, W., Westerhoff, P., Leenheer, J.A., and Booksh, K. (2003). Fluorescence excitation - emission matrix regional integration to quantify spectra for dissolved organic matter. Environmental Science and Technology. 37: 5701-5710.
- Christman, R.F., Norwood, D.L., Seo, Y., and Frimmel, F.H. 1989. Oxidative Degradation of Humic Substances from Freshwater Environments: Humic Substances II. New York: Wiley and Sons.
- Christy, A. A., Bruchet, A., and Rybacki, D. (1999). Characterization of natural organic matter by Pyrolysis/GC/MS. Environment International. 25 (2/3): 181-189.
- Coble, P.G. (1996). Characterization of marine and terrestrial DOM in seawater using excitation-emission matrix spectrometry. Marine Chemistry. 51: 325-346.

- Coble, P. G., Green, S. A., and Blough, N. V. (1990). Characterization of dissolved organic matter in Black Sea by fluorescence spectroscopy. Nature. 48: 432-435.
- Coble, P.G., Schultz, C.A., and Mopper, K. (1993). Fluorescence contouring analysis of DOC intercalibration experiment samples: a comparison of techniques. Marine Chemistry. 41: 173-178.
- Collins, M., Amy G., and Steelink C. (1986). Molecular weight distribution, carboxylic acidity, and humic substances content of aquatic organic matter: Implications for removal during water treatment. Environmental Science and Technology. 20(10): 1028-1032.
- Croue, J.P, Martin, B. Simon, P., and Legube, B. (1993). Hydrophobic and hydrophilic matters of retention-extraction, characterization and quantification of water. Water Supply. 11(1): 79-90.
- Croue, J.P., Korshin, G.V., and Benjamin, M. 2000. Characterization of Natural Organic Matter in Drinking Water: United State of America, American Water Works Association: 324.
- Cunha, L.C., Serve, L., Gadel, F., and Blazi, J.L. (2000). Characterization of riverine particulate organic matter by pyrolysis GC/MS. The Science of the Total Environment. 256: 191-204.
- Day, G.M., Beckett, R., Hart, B.T., and Mckelvie, I.D. (1991). Characterization of natural organic matter from four victorian freshwater systems. Australian Journal of Marine and Freshwater Research. 42(6): 675-687.
- Debroux J.-F. (1998). The physical-chemical and oxidantreactive properties of effluent organic matter (EfOM) intended for potable reuse. Ph.D. Thesis, University of Colorado, Boulder
- Department of Water Affairs and Forestry (1993). South African Water Quality Guidelines. Volume 2: Recreational Use. Pretoria.
- Dignac, M.F., Ginestet, P., Rybacki, D., Bruchet, A., Urbain, V., and Scribe P. (2000). Fate of wastewater organic pollution during activated sludge treatment: nature of residual organic matter. Water Research. 34 (17): 4185-4194.
- Eaton A. (1995). Measuring UV-absorbing organic: a standard method. Journal of American Water Works Association. 2: 86-90.
- Eckenfelder, W.W., Jr., and Ford, D.L. (1970). Water Pollution Control. Austin, Tex.: Pemberton Press.

- Eckenfelder, W.W., Jr. (1970). Water Quality Engineering for Practicing Engineers. New York: Barnes & Noble.
- Edzwald, J. K. (1993). Coagulation in drinking water treatment: particles, organics and coagulants. Water Science and Technology. 27(11): 21–35.
- Edzwald, J. K., Becker, W. C., and Wattier, K. (1985). Surrogate parameters for monitoring organic matter and THM precursors. Journal of American Water Works Association. 77(4): 122.
- Fabbri, D., Mongardi, M., Montanari, L., Galletti, G. C., Chiavari, G., and Scotti, R. (1998). Comparison between CP/MAS ^{13}C -NMR and pyrolysis-GC/MS in the structural characterization of humins and humic acids of soil and sediments. Fresenius' Journal of Analytical Chemistry. 362: 299–306.
- Fan, L., Harris, J., Roddick, F., Booker, N. (2001). Influence of the characteristics of natural organic matter on the fouling of microfiltration membranes. Water Research. 35(18): 4455-4463.
- Faure, P., Schlepp, L., Mansuy-Hault, L., Elie, M., Jarde, E., and Pelletier M. (2006). Aromatization of organic matter induced by the presence of clays during flash pyrolysis-gas chromatography-mass spectrometry artifact. Journal of Analytical and Applied Pyrolysis. 75: 1-10.
- Fukushima, T., Park, J., Imai, A., and Matsushige, K. (1995). Dissolved organic carbon in a eutrophic lake: dynamics, biodegradation and origin. Aquatic Sciences. 58: 139-57.
- Gadel, F. and Bruchet, A. (1987). Application of pyrolysis-gas chromatography-mass spectrometry to the characterization of humic substances resulting from decay of aquatic plants in sediments and waters. Water Research. 21: 1195–1206.
- Galapate, R.P., Baes, A.U., and Okada, M. (2001). Transformation of dissolved organic matter during ozonation: effects on trihalomethane formation potential. Water Research. 35: 2201-2206.
- Gehr, R., Startz, C., and Offringa, G. (1993). Removal of trihalomethanes precursors from eutrophic water by dissolved air floatation. Water Research. 27: 41–49.
- Gloyna, E.F. (1976). Facultative Waste Stabilization Pond Design. In Ponds as a Wastewater Treatment Alternative, edited by E.F.Gloyna, J.F. Malina, Jr., and E.M. Davis Austin, Tex.: University of Texas Press.

- Gloyna, E.F. (1968). Basis for Waste Stabilization Ponds Designs. In Advances in Water Quality Improvement, edited by E.F. Gloyna and W.W. Eckenfelder, Jr. Austin, Tex: University of Texas Press.
- Goslan, E.H., Gurses, F., Banks, J., and Parsons, S.A. (2006). An investigation into reservoir NOM reduction by UV photolysis and advanced oxidation processes. Chemosphere. 65: 1113-1119
- Goslan, E. H., Voros, S., and Banks, J. (2004). A model for predicting dissolved organic carbon distribution in a reservoir water using fluorescence spectroscopy. Water Research. 38(3): 783-791.
- Gray, K.A., McAuliffe, K.S., Bornick, R., Horne, A.J., and Bachand, P. (1996). Evaluation of organic quality in Prado Wetlands and Santa Ana River by Pyrolysis-GC/MS (Final Report). Orange County Water District, CA.
- Harrington, G., Bruchet, A., Rybacki, D., and Singer, P. (1996). Characterization of natural organic matter and its reactivity with chlorine. In: Water disinfection and natural organic matter characterization and control, Minear R., Amy G., (editors). ACS symposium series 649. Washington, DC: ACS, 138–58.
- Hatcher, P. G., Dria, K. J., Kim, S., and Frazier, S. W. (2001). Modern analytical studies of humic substances. Soil Science. 166: 770–794.
- Her, N., Amy, G., Sohn, J., and Yoon Y. (2001). Characterization of DOM as a function of MW by HPLC-SEC using UVA, DOC and fluorescence detection. Proceeding of the Water Quality Technology Conference of American Water Work Association. Nashville, November, 11-15.
- Homklin, S. 2004. Removal of hydrophobic and hydrophilic dissolved organic matters in natural water by alum coagulation. Master thesis. Inter-Department Program in Environmental Management, Chulalongkorn University, Thailand.
- Hu, J.Y., Ong, J.H., Shan, J.B., Kang., and Ng, W.J. (2003). Treatability of organic fractions derived from secondary effluent by reverse osmosis membrane. Water Research. 37:4801-4809.
- Imai, A., Fukushima, T., Matsushige, K., and Kim, Y-H. (2001). Fractionation and characterization of dissolved organic matter in a shallow eutrophic lake, its flowing rivers, and other organic matter sources. Water Research. 35: 4019-4028.

- Imai, A., Fukushima, T., Matsushige, K., Kim, Y-H., and Choi, K. (2002). Characterization of dissolved organic matter in effluent from wastewater treatment plant. Water Research. 36: 859-870.
- Imai, A., Matsushige, K., and Nagai, T. (2003). Trihalomethane formation potential of dissolved organic matter in a shallow eutrophic lake. Water Research. 37: 4284-4294.
- Janhom T. 2004. Reduction of trihalomethane created from six fractions of dissolved organic matters in raw water supply by alum coagulation. Master thesis. Inter-Department Program in Environmental Management, Chularongkorn University.
- Jiarsirikul V. 2003. Relationships between trihalomethane formation potential and natural organic matter surrogates in raw water and coagulated water of shallow wells near a closed unsanitary solid waste dumping site. Master thesis. Inter-Department Program in Environmental Management, Chulalongkorn University, Thailand
- Kasuga, I., Nakajima, F., and Furumai, H. (2003). Analysis of dissolved organic matter and bacterial community in degradation of algal bloom by EEMs and PCR-DGGE. Journal of Japan Society on Water Environment. 26 (3): 171- 174.
- Kavanaugh, M.C., Trussell, A.R., Cromer, J., and Trussell, R.R. (1980). An empirical kinetic model of trihalomethane formation: applications to meet the proposed trihalomethane standard. Journal of American Water Works Association. 10: 578-582.
- Kennedy, M.D., Chun, H.K., Yangali, V.A.Q., Heijman, B.G.J., and Schippers, J.C. (2005). Natural organic matter (NOM) fouling of ultrafiltration membranes: fractionation of NOM in surface water and characterization by LC-OCD. Desalination. 178: 73-83.
- Kim, H-C. and Yu, M-J. (2005). Characterization of natural organic matter in conventional water treatment processes for selection of treatment processes focused on DBPs control. Water Research. 39: 4779-4789.
- Kimura, K., Hane, Y., Watanabe, Y., Amy, G., and Ohkuma, N. (2004). Irreversible membrane fouling during ultrafiltration of surface water. Water Research. 38: 3431-3441.
- Kitis, M., Karanfil, T., Wigton, A., and Kilduff, J.E. (2002). Probing reactivity of dissolved organic matter for disinfection by-product formation using XAD-8 resin adsorption and ultrafiltration fractionation. Water Research. 36: 3834-3848.
- Kogel-Knabner, I. (2000). Analytical approaches for characterizing soil organic matter. Organic Geochemistry. 31, 609-625.

- Komatsu, K., Nakajima, F., Furumai, H., and Miki, O. (2005). Characterization of dissolved organic matter (DOM) removed by iron coagulation using spectrofluorimetry and pyrolysis GC/MS analysis. Journal of Water Supply: Research and Technology-AQUA. 54 (3): 157-163.
- Korshin, G.V., Benjamin, M.M., and Sletten R.S. (1997). Adsorption of natural organic matter (NOM) on iron oxide: effects on NOM composition and formation of organo-halide compounds during chlorination. Water Research. 31(7): 1643-1650.
- Krasner, S. W., Croue, J. P., and Buffle, J. (1996). Three approaches for characterizing NOM. Journal of American Water Works Association. 88(6): 66-79.
- Lee, N., Amy, G., Croué, J-P., and Buisson, H. (2004). Identification and understanding of fouling in low-pressure membrane (MF/UF) filtration by natural organic matter (NOM). Water Research. 38: 4511-4523.
- Lee S. and Ahn K.-H. (2004). Monitoring of COD as an organic indicator in wastewater and treated effluent by fluorescence excitation-emission (FEEM) matrix characterization. Water Science and Technology. 50 (8): 57-63.
- Leenheer, J.A. (1981). Comprehensive approach to preparative isolation and fractionation of dissolved organic carbon from natural waters and wastewaters. Environmental Science and Technology. 15(5): 578-587.
- Leenheer, J.A., and Croué J.P. (2003). Characterizing dissolved aquatic organic matter. Environmental Science and Technology. January 1: 19A-26A.
- Liang, L. and Singer, P.C. (2003). Factors influencing the formation and relative distribution of haloacetic acids and trihalomethanes in drinking water. Environmental Science and Technology. 37: 2920-2928.
- Lu, X. Q., Hanna, J. V., and Johnson W. D. (2000). Source indicators of humic substances: an elemental composition, solid state ^{13}C CP/MASNMR and Py-GCIMS study. Applied Geochemistry. 15: 1019-1033.
- Pirnie, M., Inc. 1993. Guidance Manual for Enhanced Coagulation and Enhanced Softening. Manwah, NJ: USEPA.
- Mancini, J.L., and Barnhart, E.L. (1968). Industrial Waste Treatment in Aerated Lagoons. In Advances in Water Quality Improvement, edited by E.F.Gloyna and W.W. Eckenfelder, Jr. Austin, Tex.: University of Texas Press.
- Marhaba, T.F. and Kochar, I.H. (2000). Rapid prediction of disinfection by-product formation potential by fluorescence. Environmental Engineering and Policy. 2: 29-36.

- Marhaba, T. F. and Pipada, N. S. (2000). Coagulation: effectiveness in removing dissolved organic matter fractions. Environmental Engineering Science. 17(2): 107-115.
- Marhaba, T.F. and Pu, Y. (2000). Rapid delineation of humic and non-humic organic matter fractions in water. Journal of Hazardous Materials. A73: 221-234.
- Marhaba, T. F. and Van, D. (1999). Chlorinated Disinfection By-product formation potential of dissolved organic matter fractions at an ozonation water treatment plant. Environmental Research. 3(3): 255-268.
- Marhaba, T. F. and Van, D. (2000). The variation of mass and disinfection by-product formation potential of dissolved organic matter fractions along a conventional surface water treatment plant. Journal of Hazardous Materials. 73(3): 133-147.
- Marhaba, T.F., Pu, Y., and Bengraine, K., (2003). Modified dissolved organic matter fractionation technique for natural water. Water Research. 34 (14): 3543-3550.
- Marhaba T.F. and Washington M.B. (1998). Drinking water disinfection byproducts: history and current practice. Advance Environmental Research. 2 (1): 103-115.
- Martin, J. W. (1995). Characterization of Neuropathological Shape Deformations. PhD thesis, Massachusetts Institute of Technology.
- McKnight, D.M., Boyer, E., Westerhoff, O., Doran, P., Kulbe, T., and Anderson, D. (2001). Spectrofluorometer characterization of dissolved organic matter for identification of precursor organic material and aromaticity. Limnology and Oceanography. 46(1): 38-48.
- Metcalf & Eddy, Inc. (1979). Wastewater Engineering Treatment, Disposal and Reuse. 2nd ed. New York: McGraw-Hill.
- Miller, J. W., and Uden, P.C. (1983). Characterization of Nonvolatile Aqueous Chlorination Products of Humic Substances. Environment Engineering Science. 17: 150-159.
- Murruy, C.A. and Parson, S.A. (2004). Advance oxidation processes: flowsheet options for bulk natural organic matter removal. Water Science and Technology. 4 (4): 113-119.
- Musikavong, C., Wattanachira, S., Marhaba, T.F., and Pavasant, P. (2005). Reduction of organic matter and trihalomethane formation potential in reclaimed water from treated industrial estate wastewater. Journal of hazardous material. B127: 48-57.
- Muttamara S. and Puetpaiboon U. (1996). Nitrogen removal in baffled waste stabilization ponds. Water Science and Technology. 33(7):173-181.

- Muttamara S. and Puetpaiboon U. (1997). Roles of baffles in waste stabilization ponds. Water Science and Technology. 35(8): 275-284.
- Nakajima F., Hanabusa M., and Furumai H. (2002). Excitation-emission fluorescence spectra and trihalomethane formation potential in the Tama River, Japan. Water Science and Technology: Water Supply. 2: (5-6), 481-486.
- Nameche, TH. and Vasel, J.L. (1998). Hydrodynamic studies and modelization for aerated lagoons and waste stabilization ponds. Water Research. 32 (10): 3039-3045.
- National Research Council, 1998. Issues in Potable Reuse: The Viability of Augmenting Drinking Water Supplies with Reclaimed Water. National Academy Press, Washington, DC.
- Owen, D. M., Pirnie, M. and Amy, G. L.(1993). Characterization of natural organic matter and its relationship to treatability. AWWARF order number: 90631.
- Owen, D. M., Amy, G. L., and Chowdhury, Z. K. (1995). NOM characterization and treatability. Journal of American Water Works Association. 87(1): 46-63.
- Page, D.W., Van Leeuwen, J.A., Spark, K.M., and Mulcahy D.E. (2002). Pyrolysis characterization of plant, humus and soil extracts from Australian catchments. Journal of Analytical and Applied Pyrolysis. 65: 269–285.
- Page, D.W., Van Leeuwen, J.A., Spark, K.M., and Mulcahy, D.E. (2003). Application of pyrolysis-gas chromatography/mass spectrometry for characterization of dissolved organic matter before and after alum treatment. Journal of Analytical and Applied Pyrolysis. 67: 247–262.
- Panyapinyopol, B., Marhaba, T.F., Kanokkantapong, V. and Pavasant, P. (2005). Characterization of precursors to trihalomethanes formation in Bangkok source water. Journal of Hazardous Material. 120 (1-3):229-236.
- Peschel, G. and Wildt, T., (1988). Humic substances of natural and anthropogeneous origin. Water Research. 22: 105–108.
- Pettersson, C., Bishop, K. H., and Lee, Y. H. (1995). Relations between Organic Carbon and Methylmercury in Humic Rich Surface Waters from Svartberget Catchment in Northern Sweden. Water Air and Soil Pollution. 80:971-979.
- Phumpaisanchai A. 2005. Removal of Hydrophobic and Hydrophilic Natural Organic Matters in Reservoirs by Alum Coagulation Master Thesis. Graduated School, Chiang Mai University. Thailand

- Poerschmann, J., Kopinke, F.D., Balcke, G., and Mothes, S. (1998). Pyrolysis pattern of anthropogenic and natural humic organic matter. Journal of Microcolumn Separations. 10: 401–411.
- Pouwels, A. D., Eijkel, G. B., and Boon, J. J. (1989). Curie-point pyrolysis-capillary gas chromatographyhigh- resolution mass spectrometry of microcrystalline cellulose. Journal of Analytical and Applied Pyrolysis. 14: 237-280.
- Ralph, J., and Hatfield, R. D. (1991). Pyrolysis-GC-MS characterization of forage materials. Journal of Agricultural and Food Chemistry. 39: 1426-1437.
- Reckhow, D.A., S., P.C., and Malcolm, R.L. (1990). Chlorination of Humic Materials By- Product Formation and Chemical Interpretations. Environmental Engineering Science. 24: 1655-1664.
- Reynolds, D.T. and Richards, A.P. (1996). Unit Operation and Process in Environmental Engineering 2nd ed., U.S.PWS publishing Company,
- Reynolds, T.D. and Yang, J.T. (1966). Model of Completely Mixed Activated Sludge Process. Proceeding of the 21st Annual Purdue Industrial Waste Conference. Part 2
- Ritter, A.V., Masion, A., Boulange, T., Rybacki, D., and Bottero J-Y. (1999). Removal of natural organic matter by coagulation-flocculation: A pyrolysis-GC-MS study. Environmental Engineering Science. 33: 3027-3032.
- Rodriguez, M.J., Vinette, Y., Serodes, J., and Bouchard, C. (2003). Trihalomethanes in drinking water of Greater Quebec Region (Canada): Occurrence, variations and modeling, Environmental Monitoring and Assessment. 89:69-93.
- Roek J.J. (1974). Formation of haloform during chlorination of natural waters. Journal of the Society for Treatment Examination. 23: 234-243
- Roek J.J. (1977). Chlorination reactions of fulvic acids in natural waters. Environmental Engineering Science. 11(5): 478-482.
- Sanz-Jimenez, C. (1994). Analytical pyrolysis of humic substances: pitfalls, limitations, and possible solutions. Environmental Engineering Science. 28: 1773-1780.
- Smally, Jr., F.E., Howell, G.D., Kravitz, R., Jewell, J.T. (1988). Protein in natural waters and their relation to the formation of chlorinated organics during water disinfection. Environmental Engineering Science. 22: 537–542.
- Sharp, E.L., Jarvis, P., Parson, S.A., and Jefferson B. (2006). Impact of fractional character on coagulation of NOM. Colloids and Surfaces A: Physicochemistry Engineering Aspect. 286: 104–111.

- Sierra, M.M.D., Giovanelo, M., Parlanti, E. and Soriano-Sierra E.J. (2005). Fluorescent fingerprint of fulvic and humic acids from varied origins as viewed by single-scan and excitation/emission matrix techniques, *Chemosphere*. 58: 715-733.
- Sinsabaugh, III, R.L., Hoehn, R.C., Knocke, W.R. and Linkins, A.E. (1986). Removal of dissolved organic carbon by coagulation with iron sulfate. *Journal of American Water Works Association*. 78 (5): 74-82.
- Sirivedhin, T. and Gray, A. (2005). Part I. Identifying anthropogenic markers in surface waters influenced by treated effluents: a tool in potable water reuse. *Water Research*. 39: 1154-1164.
- Smart, P.L., Finlayson, B.L., Rylands, W.D. and Ball, C.M. (1976). The relation of fluorescence to dissolved organic carbon in surface water. *Water Research*. 31: 2012-2018.
- Standard Methods for the Examination of Water and Wastewater* 1995 19th ed, American Public Health Association/American Water Works Association/Water Environment Federation, Washington, DC.
- Stevenson, F. J. 1982. *Spectroscopic Approaches, Humus Chemistry, Genesis, Composition, Reactions*; New York; Wiley and Sons.
- Swietlik, J., Dabrowska, A., Raczyk-Stanislawiak, U., and Nawrocki, J. (2004). Reactivity of natural organic matter fractions with chlorine dioxide and ozone. *Water Research*. 38:547-558
- Tambo, N. (1989). Evaluation of extent of humic substance removal by coagulation. *Aquatic Humic Substances*. Edited by Suffet. Washington, D.C.: American Chemical Society.
- Thacker, P.N., Kaur, P. and Rudra, A. (2002). Trihalomethane formation potential and concentration changes during water treatment at Mumbai (India). *Environmental Monitoring and Assessment*. 73: 253-262.
- Thapa, P.B. 2002. *Characterization of natural organic matter in reservoirs by multi-analytical approaches*. Ph.D. dissertation, Graduate School of Engineering, The University of Tokyo.
- Thaveemaitree Y. 2005. *Adsorption of di-(2-ethylhexyl) phthalate (DEHP) on solid generated in coagulation and flocculation process in drinking water treatment plants*. Ph.D. dissertation, Graduate School of Engineering, The University of Tokyo

- Thurman, E.M. Organic geochemistry of natural waters. Martinus Nijhoff/Junk, Boston, MA, 1985, pp 497.
- Thurman, E.M. and Malcolm, R.L. (1981). Preparative isolation of aquatic humic substances. Environmental Science and Technology. 15 (4): 463–466.
- Traganza, E.D., (1969). Fluorescence excitation and emission spectra of dissolved organic matter in sea water. Bulletin of Marine Science. 19: 897-904.
- USEPA (1998). National Primary Drinking Water Regulation: Disinfectants And Disinfection By-Product (D/DBP), Final Rule. Fed Register. 63:69389-69476.
- USEPA (1999). Enhanced coagulation and enhanced precipitative softening guidance manual: Office of water (4607)[Online]. Available online from: <http://www.epa.gov> [2001, November 10]
- Wattanachira, S., Permsuk, O., and Marhaba, T.F. (2003). Trihalomethane Species and NOM Parameters in Water Supply of a Small Rural Waterworks. The proceeding of the International Symposium on Southeast Asian Water Environment. pp. 395-402, October 23-25, AIT, Bangkok, Thailand.
- Wattanachira, S., Musikavong, C., Permsuk, O., and Pavasant, P. (2004). Removal of Surrogates for Natural Organic Matter and Probability of Finding Trihalomethanes in the Produced Water Supply of Small Waterworks in Chiang Mai, Thailand. Songklanakarin Journal of Science and Technology. 26 (Suppl.1): 25-37.
- White D.M., Garland D.S., Beyer L., Yoshikawa K. (2004). Pyrolysis-GC/MS fingerprinting of environmental samples. Journal of Analytical and Applied Pyrolysis. 71:107-118.
- White D. M., Garland D. S., Narr J., and Woolard, C.R. (2003). Natural organic matter and DBP formation potential in Alaskan water supplies. Water Research. 37: 939-947
- Widrig, D.L., Gray, K.A., and McAuliffe, K.S., (1996). Removal of algal-derived organic material by preozonation and coagulation: monitoring changes in organic quality by pyrolysis-GC-MS. Water Research. 30: 2621–2632.
- Wolfbeis, O.S. (1985). Fluorescence of organic natural products. In S.G. Schulman (Editor) Molecular luminescence spectroscopy: Methods and application. Part I. John Wiley & Sons, New York., p.167.
- World Health Organization, Guidelines for Drinking Water Quality: (1994), 2nd ed., Vol 1, Recommendation, WHO, Geneva.

- World Health Organization, Guidelines for Drinking Water Quality: (1996), Health Criteria and Other Supporting Information, 2nd ed., Vol 2, Geneva, p 940.
- Wu, Y.C., and Kao, D.F. (1976). Yeast Plant Wastewater Treatment. Journal SED 102, no.EE5:969.
- Yamashita, Y. and Tanoue, E. (2003). Chemical characterization of protein-like fluorophores in DOM in relation to aromatic amino acids. Marine Chemistry. 82: 255-271.
- Young, M.S. and Uden, P.C. (1994). Byproducts of aqueous chlorination of purines and pyrimidines. Environmental Science and Technology. 28: 1755–1758.
- Zepp, R.G., Sheldon, W.M., and Moran, M.A. (2004). Dissolved organic fluorophores in southeastern US coastal waters: correction methods for eliminating Rayleigh and Raman scattering peaks in excitation-emission matrices. Marine Chemistry. 89: 15-36
- Zouboulis, A.I., Jun, W., and Katsoyiannis, I.A. (2003). Removal of humic acids by flotation, Colloids and Surface A: Physicochemistry Engineering Aspect. 231: 181-193.

APPENDICES

APPENDIX A
DOC, UV-254, SUVA, THMFP AND SPECIFIC THMFP VALUES

Table A-1: DOC, UV-254, SUVA, THMFP and specific THMFP of the effluent water from detention pond collected in June 2004 and those of its DOM fractions

Parameter	Eff ¹ . Water from detention pond	DOM fractions					
		HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	6.81±0.2	0.43±0.02	0.27±0.02	2.16±0.04	0.73±0.08	1.86±0.09	1.60±0.10
UV-254 (cm ⁻¹)	0.208 ±0.004	0.006 ±0.0001	0.010 ±0.001	0.035 ±0.0006	0.015 ±0.0001	0.045 ±0.0002	0.014 ±0.0004
SUVA (L/mg-m)	3.05±0.06	1.47±0.02	3.88±0.37	1.62±0.03	2.05±0.01	2.43±0.01	0.84±0.02
THMFP (µg/L)	625±4	54±2	50±6	306±6	112±2	243±6	178±9
Specific THMFP (µg THMFP/mg DOC)	90±6.7	125±5.2	190±22	142±2.9	153±2.3	130±8.8	112±5.6

Remark:¹Eff = Effluent**Table A-2:** DOC, UV-254, SUVA, THMFP and specific THMFP of industrial estate wastewater and the effluent water from aeration, facultative, oxidation and detention pond collected in September 2004

Parameter	Industrial Estate Wastewater	Eff ¹ . Water from aeration ponds	Eff ¹ . Water from facultative ponds	Eff ¹ . Water from oxidation pond	Eff ¹ . Water from detention pond
DOC (mg/L)	14.0±0.2	6.52±0.05	5.96±0.1	5.63±0.1	NA ²
UV-254 (cm ⁻¹)	0.182±0.006	0.113±0.002	0.110±0.001	0.109±0.002	NA
SUVA (L/mg-m)	1.28±0.02	1.71±0.01	1.83±0.04	1.94±0.04	NA
THMFP (µg/L)	1245±44	563±33	553±35	491±29	NA
Specific THMFP (µg THMFP/mg DOC)	89±3.2	86±5.1	93±5.8	87±5.2	NA

Remark:¹Eff = Effluent and ²NA = Not available**Table A-3:** DOC, UV-254, SUVA, THMFP and specific THMFP of industrial estate wastewater and the effluent water from aeration, facultative, oxidation and detention pond collected in October 2004

Parameter	Industrial Estate Wastewater	Eff ¹ . Water from aeration ponds	Eff ¹ . Water from facultative ponds	Eff ¹ . Water from oxidation pond	Eff ¹ . Water from detention pond
DOC (mg/L)	12.2±0.07	5.99±0.02	6.07±0.13	6.11±0.02	NA ²
UV-254 (cm ⁻¹)	0.143±0.0004	0.112±0.0003	0.108±0.0001	0.106±0.0003	NA
SUVA (L/mg-m)	1.13±0.009	1.90±0.002	1.75±0.034	1.75±0.001	NA
THMFP (µg/L)	1109±39	497±12	435±13	373±21	NA
Specific THMFP (µg THMFP/mg DOC)	91±3.2	83±1.9	72±2.0	61±3.5	NA

Remark:¹Eff = Effluent and ²NA = Not available

Table A-4: DOC, UV-254, SUVA, THMFP and specific THMFP of industrial estate wastewater and the effluent water from aeration, facultative, oxidation and detention pond collected in February 2005

Parameter	Industrial Estate Wastewater	Eff ^l . Water from aeration ponds	Eff ^l . Water from facultative ponds	Eff ^l . Water from oxidation pond	Eff ^l . Water from detention pond
DOC (mg/L)	10.3±0.25	5.55±0.02	6.01±0.47	5.60±0.11	6.14±0.29
UV-254 (cm ⁻¹)	0.163±0.002	0.098±0.0003	0.112±0.007	0.100±0.003	0.112±0.005
SUVA (L/mg-m)	1.58±0.05	1.78±0.01	1.84±0.03	1.75±0.05	1.82±0.07
THMFP (µg/L)	1214±3	530±16	564±11	572±7	588±5
Specific THMFP (µg THMFP/mg DOC)	118±0.3	96±1.4	93±0.5	100±0.7	95±0.6

Remark: ^lEff = Effluent

Table A-5: DOC, UV-254, SUVA, THMFP and specific THMFP of DOM fractions of industrial estate wastewater collected in February 2005

Parameter	DOM fractions					
	HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	1.51±0.09	0.21±0.02	2.58±0.05	1.72±0.10	1.65±0.10	2.25±0.03
UV-254 (cm ⁻¹)	0.021 ±0.0004	0.007 ±0.0007	0.029 ±0.0016	0.026 ±0.0015	0.027 ±0.0023	0.019 ±0.0007
SUVA (L/mg-m)	1.36±0.03	3.38±0.33	1.11±0.06	1.51±0.09	1.63±0.14	0.83±0.03
THMFP (µg/L)	141±11	18±5	371±18	115±18	148±8	95±1
Specific THMFP (µg THMFP/mg DOC)	93±8.5	86±14.7	144±5.9	67±9.0	90±4.2	42±0.2

Table A-6: DOC, UV-254, SUVA, THMFP and specific THMFP of DOM fractions of the effluent water from aeration ponds collected in February 2005

Parameter	DOM fractions					
	HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	0.59±0.09	0.12±0.01	1.72±0.03	0.91±0.01	1.14±0.05	1.63±0.05
UV-254 (cm ⁻¹)	0.011 ±0.0007	0.002 ±0.0007	0.021 ±0.0003	0.012 ±0.0004	0.014 ±0.0011	0.005 ±0.0010
SUVA (L/mg-m)	1.87±0.12	2.02±0.30	1.22±0.02	1.29±0.04	1.23±0.10	0.29±0.06
THMFP (µg/L)	70±6	15±3	218±13	60±4	108±13	66±6
Specific THMFP (µg THMFP/mg DOC)	119±9.0	124±48.2	127±1.6	65±2.6	95±7.8	40±3.9

Table A-7: DOC, UV-254, SUVA, THMFP and specific THMFP of DOM fractions of the effluent water from facultative ponds collected in February 2005

Parameter	DOM fractions					
	HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	0.43±0.03	0.10±0.01	2.80±0.05	0.63±0.04	1.27±0.05	1.24±0.06
UV-254 (cm ⁻¹)	0.010 ±0.0009	0.002 ±0.0004	0.046 ±0.0002	0.013 ±0.0005	0.023 ±0.0033	0.005 ±0.0005
SUVA (L/mg-m)	2.35±0.20	2.87±0.58	1.63±0.01	2.04±0.07	1.80±0.26	0.41±0.02
THMFP (µg/L)	54±8	10±2	262±15	38±3	171±24	48±4
Specific THMFP (µg THMFP/mg DOC)	125±17.0	142±42.2	93±6.3	60±3.2	141±16.9	39±3.3

Table A-8: DOC, UV-254, SUVA, THMFP and specific THMFP of DOM fractions of the effluent water from oxidation ponds collected in February 2005

Parameter	DOM fractions					
	HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	0.41±0.06	0.10±0.01	2.10±0.05	0.71±0.05	1.46±0.03	1.24±0.05
UV-254 (cm ⁻¹)	0.009 ±0.0002	0.002 ±0.0008	0.034 ±0.0013	0.012 ±0.0013	0.018 ±0.0002	0.004 ±0.0003
SUVA (L/mg-m)	2.09±0.06	2.08±0.35	1.61±0.06	1.74±0.19	1.23±0.01	0.29±0.02
THMFP (µg/L)	50±8	15±2	244±24	50±6	174±4	59±3
Specific THMFP (µg THMFP/mg DOC)	122±18.3	142±34.2	116±13.9	71±19.4	119±2.4	47±2.0

Table A-9: DOC, UV-254, SUVA, THMFP and specific THMFP of DOM fractions of the effluent water from detention pond collected in February 2005

Parameter	DOM fractions					
	HPON	HPOB	HPOA	HPIB	HPIA	HPIN
DOC (mg/L)	0.45±0.06	0.10±0.03	2.62±0.12	0.74±0.10	1.52±0.07	1.11±0.05
UV-254 (cm ⁻¹)	0.007 ±0.0044	0.001 ±0.00005	0.043 ±0.0001	0.011 ±0.00005	0.018 ±0.0007	0.003 ±0.0002
SUVA (L/mg-m)	1.64±0.05	1.40±0.05	1.62±0.005	1.48±0.01	1.17±0.04	0.27±0.01
THMFP (µg/L)	45±7	12±1	273±3	48±6	192±6	50±5
Specific THMFP (µg THMFP/mg DOC)	96±8.9	120±4.7	104±0.9	63±4.8	131±1.5	47±3.1

Table A-10: DOC, UV-254, SUVA, THMFP and specific THMFP of industrial estate wastewater and the effluent water from aeration, facultative, oxidation and detention pond collected in July 2005

Parameter	Industrial Estate Wastewater	Eff ¹ . Water from aeration ponds	Eff ¹ . Water from facultative ponds	Eff ¹ . Water from oxidation pond	Eff ¹ . Water from detention pond
DOC (mg/L)	14.9±0.7	5.64±0.13	5.78±0.15	5.71±0.25	5.37±0.05
UV-254 (cm ⁻¹)	0.171±0.002	0.119±0.001	0.120±0.004	0.113±0.002	0.124±0.017
SUVA (L/mg-m)	1.16±0.06	2.09±0.05	2.12±0.07	1.97±0.05	2.31±0.018
THMFP (µg/L)	1351±21	² NA	NA	468±3.5	NA
Specific THMFP (µg THMFP/mg DOC)	91±1.4	NA	NA	82±0.6	NA

Remark:¹Eff = Effluent and ²NA = Not available

APPENDIX B
IDENTIFICATION OF PYROLYSIS FRAGMENTS,
DETAIL OF PYROLYSIS FRAGMENTS AND PYROCHROMATOGRAM OF ALL
WATER SAMPLES

Identification of Pyrolysis Fragments

1. Open the data file obtained from the pyrolysis GC/MS using the GC/MS Postrun analysis program. Figure B-1 shows the output of pyrochromatograms of the HPOA fraction of the aeration pond effluent.

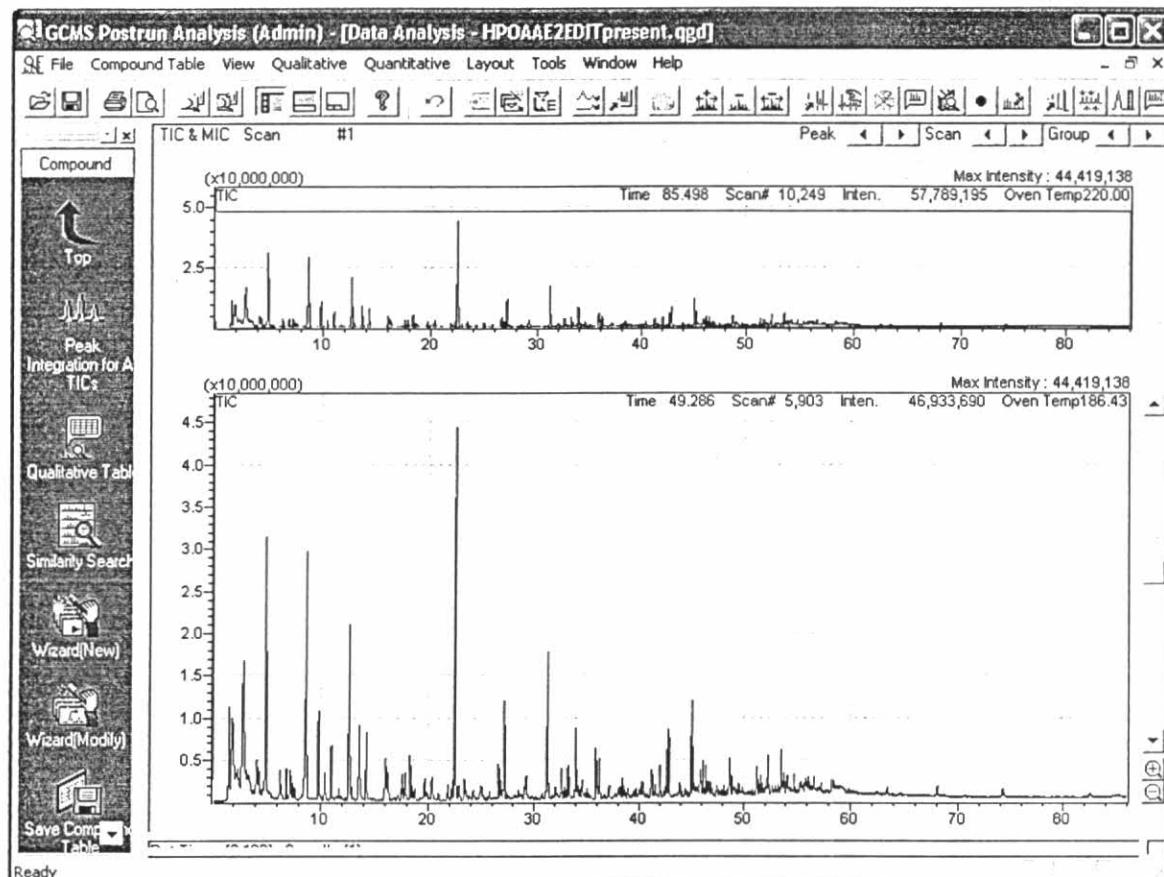


Figure B-1 The output of pyrochromatograms of the HPOA fraction of the aeration pond effluent

2. Integrate the pyrochromatograms of the HPOA fraction of the aeration pond effluent. The result is shown in Figure B-2.

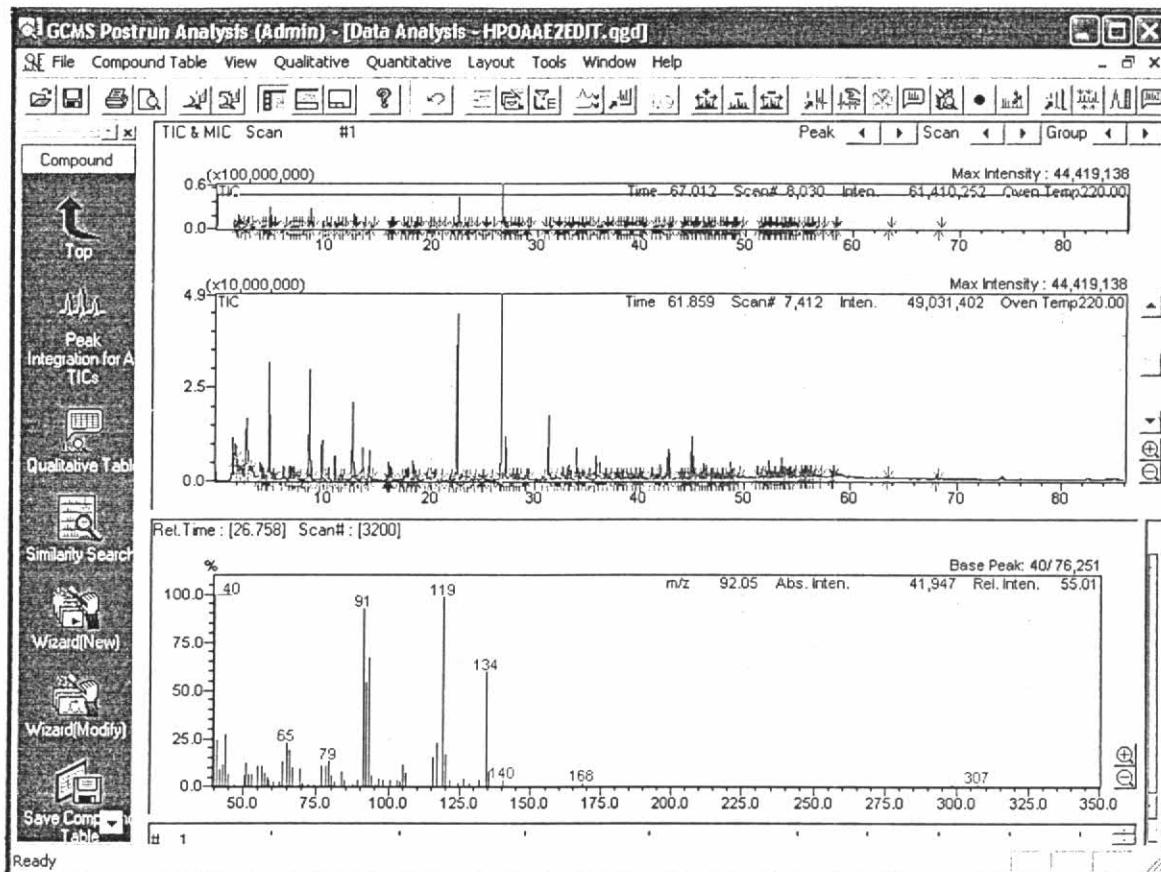


Figure B-2 The output of integrated pyrochromatograms of the HPOA fraction of the aeration pond effluent

3. Open the qualitative table and select the pyrolysis peak (pyrolysis fragment). Figure B-3 shows the outcome of pyrochromatograms of the HPOA fraction of the aeration pond effluent after selecting the pyrolysis peak

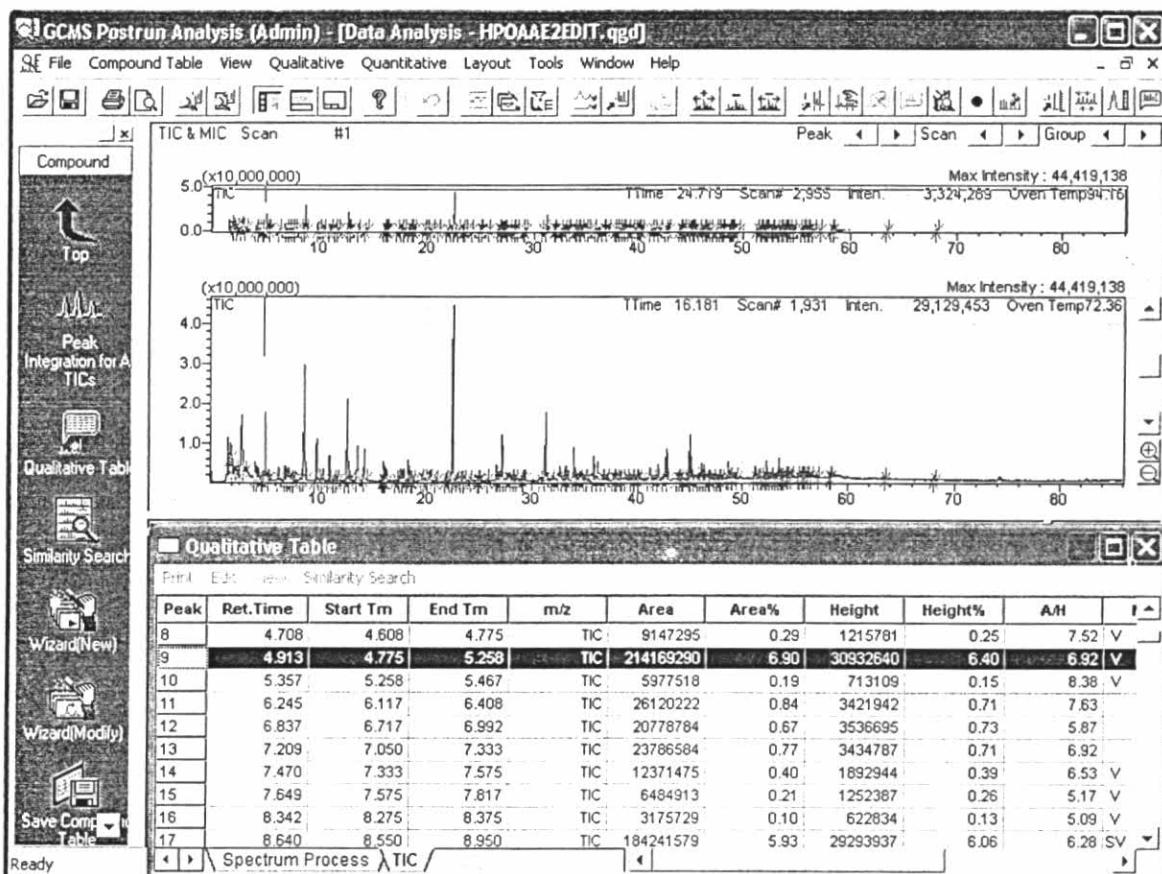


Figure B-3 The outcome of pyrochromatograms of the HPOA fraction of the aeration pond effluent after selecting the pyrolysis peak

4. Choose the similarity search icon. Figure B-4 shows the similarity search results of the selected pyrolysis peak (pyrolysis fragment). As can be seen, the results provide the list of possible compound names with their matching percentage from high to low, molecular weight, mass spectra and chemical structure. When the matching percentage was less than 85%, the pyrolysis fragment was defined as an unknown fragment. In the case of a $85\% \leq$ matching percentage of a pyrolysis fragment $\leq 90\%$, the pyrolysis fragment was defined as an acceptable match fragment, whereas, in the case of a matching percentage of a pyrolysis fragment $> 90\%$, the pyrolysis fragment was defined as a satisfactory match fragment.

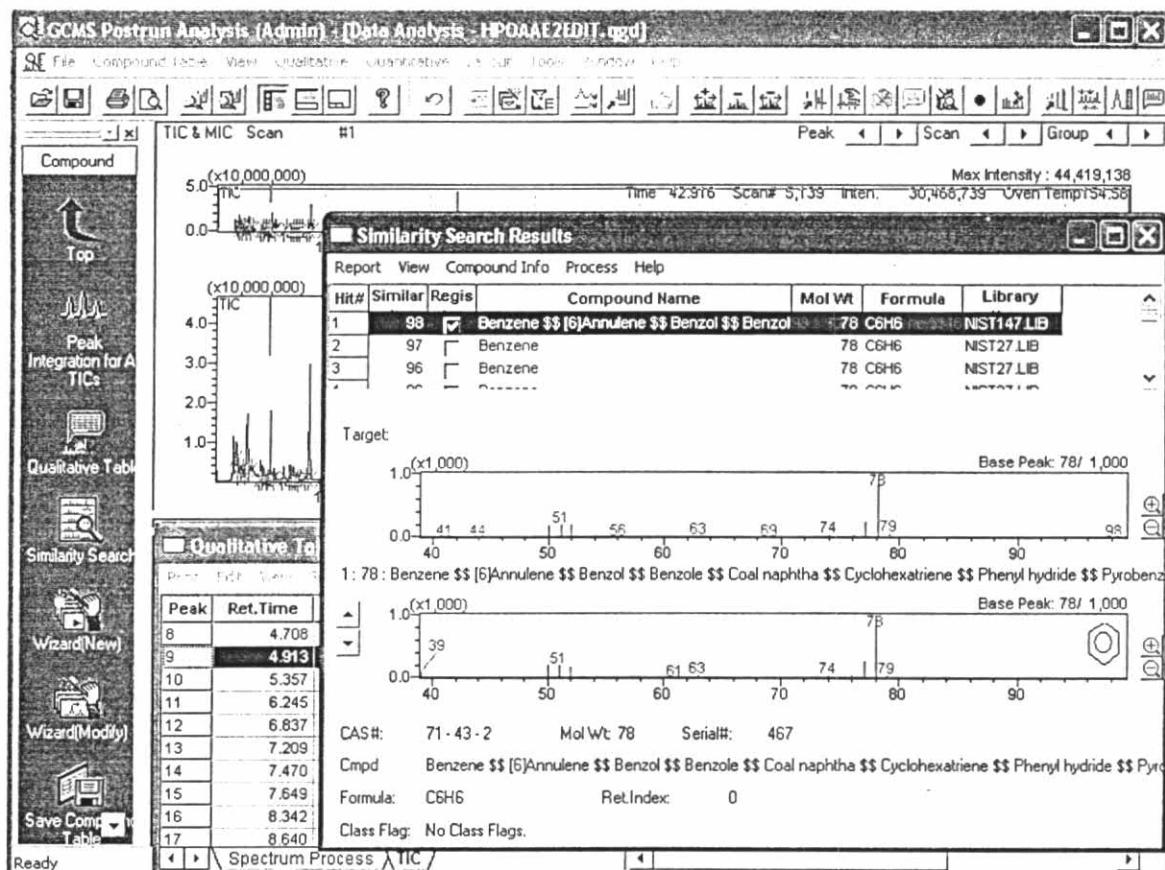


Figure B-4 The similarity search results of the selected pyrolysis peak of the HPOA fraction of the aeration pond effluent

Detail of Pyrolysis Fragment

Table B-1: Average relative ratios of the pyrolysis fragment in influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL ³	*Propene	2.45				
AL	*2-Butene	2.55	1.70			
AL	*1-Pentene	2.47	4.19			1.79
AL	*1,3-Pentadiene	0.29				
AL	1-Pentene, 4-methyl				4.82	3.04
AL	*1-Hexene	3.48	4.28			
AL	Cyclohexene	0.73	0.56			
AL	2,4-Hexadiyne					5.61
AL	*1-Heptene	3.52	5.03	2.93	3.62	
AL	*1-Octene	2.38	2.40	1.86	3.10	1.60
AL	1,3-Octadiene					0.10
AL	*1-Nonene	2.16	2.57	2.68	2.51	1.42
AL	*1-Decene	2.06	2.01	1.33	2.11	1.03
AL	5-Decyne					0.11
AL	*1-Undecene	1.85	1.72	1.21	1.84	0.75
AL	1,4 Undecadiene	0.20	0.26			0.12
AL	*1-Dodecene	1.81	1.76	0.79		0.62
AL	2-Dodecene	0.20			1.52	
AL	3-Dodecene	0.51				
AL	6-Dodecyne	0.37	0.43			0.11
AL	Undecene, 5-methyl-	0.19				
AL	2,4-Dodecadiene, (E,Z)	0.22				
AL	2-Butyl-1-decene	0.23				
AL	*1-Tridecene	2.04	1.70		1.58	0.79
AL	*Z-1,6 Tridecadiene	0.44	0.38			0.14
AL	*1-Tetradecene	1.95	2.05	0.67	1.86	0.81
AL	*Cyclotetradecane	0.12				
AL	*7-Tetradecyne	0.28				
AL	*1-Pentadecene	2.02	2.25	0.81	2.14	1.45
AL	*3-Hexadecene	0.14				
AL	Cyclopentene, 1-hexy		0.39			
AL	3-Hexadecyne	0.37				0.27
AL	Cyclopentene, 1-octyl	0.38	0.53			0.38
AL	Cyclohexene, 3-nonyl	0.19				0.17
AL	Cyclotetradecane		0.79			
AL	*1-Heptadecene	0.51	0.89			0.46
AL	*Cyclohexadecane	0.55				
AL	*1-Nonadecene		0.99			0.29
AL	10-Methyl-octadec-1-ene	0.26			1.43	
AL	Octadecane, 2-methyl					
AL	*9-Eicosene, (E)	0.69				

Table B-1: Average relative ratios of the pyrolysis fragment in influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL	*Nonadecane, 2-methyl			2.73		
AL	*Eicosane			2.82	3.01	
AL	*Eicosane 10 methyl			2.74	4.00	
AL	*Tricosane			2.21		
AR ⁴	*Toluene	1.00	1.00	1.00	1.00	1.00
AR	*p-Xylene	0.54				
AR	*Styrene	0.53	0.72	0.81	0.80	0.40
AR	*Benzene, propyl	0.13				0.11
AR	*Benzene, butyl					
AR	*Benzene, pentyl-					0.11
AR	*Naphthalene	0.13	0.19			0.19
AR	*Benzene, heptyl					0.10
AR	*Benzene, octyl-					0.20
AR	*Benzene, nonyl	0.11	0.23			0.29
PN ⁵	*Phenol	0.34		1.93	0.89	0.45
PN	*Phenol, 4-methyl	0.41			0.23	
PN	.alpha.,.alpha.'-Dihydroxy-m-diisopropylbenzene			2.46		
PN	Benzeneethanol, .alpha.-methyl-3-(1-methylethyl)			7.08		
PN	*Phenol, 4-(1,1,3,3-tetramethylbutyl)	0.29				
ON ⁶	Ethylamine				1.47	
ON	*Acetonitrile			0.82		
ON	*Isobutyronitrile			2.12		
ON	Butanenitrile	0.13	0.25	0.51		
ON	*Butanenitrile, 2-methyl	0.10	0.62	0.40	0.26	0.15
ON	*Butanenitrile, 3-methyl	0.23	0.46	0.85	0.42	0.34
ON	*Pyrrole	0.33		1.27	0.75	0.19
ON	1H-Pyrazole, 3-methyl			0.87		
ON	*1H-Pyrrole, 2-methyl			0.38		
ON	*1H-Pyrrole, 3-methyl			0.37		
ON	*Pantanenitrile, 4-methyl	0.23		0.68		0.13
ON	*Acetamide			4.20		
ON	*Benzonitrile			2.31		0.46
ON	Octanenitrile		0.32	0.31		
ON	Benzenepropanenitrile		0.19			
ON	Nonanenitrile		0.23			
ON	*Indole	0.49				
ON	*Decanenitrile		0.42			
ON	*Undecanenitrile		0.42			
ON	*Dodecanenitrile		2.16	0.62		
ON	*Tridecanenitrile		1.89		0.91	
ON	Tetradecanenitrile			0.74		0.20

Table B-1: Average relative ratios of the pyrolysis fragment in influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ON	*Pentadecanenitrile		8.48		1.34	
ON	Hexadecanenitrile		0.47	1.82	1.53	0.40
ON	*Heptadecanenitrile	0.57	2.01	1.25	1.42	0.59
ON	Octadecanenitrile		4.61			
AH ⁷	*Acetaldehyde					2.80
AH	*Furfural			0.66		
AH	Heptanal					0.15
AH	Benzaldehyde					0.29
AH	*Octanal					0.12
AH	Nonanal					0.11
AH	Decanal			0.39		0.18
AH	*E-14-Hexadecenal					0.43
AH	E-15-Heptadecenal					0.33
AH	Octadecanal					0.55
KT ⁸	*2-Pentadecanone	0.21				
KT	*2-Hexadecanone	0.11				
KT	*2-Heptadecanone	0.31				
AC ⁹	*1-Hexanol, 2-ethyl	1.21	3.28		1.53	
AC	1-Decanol, 2-methyl	0.15				
AC	2-Methyl-1-undecanol	0.18				
AC	3-Dodecen-1-ol	0.24				
AC	* 1-Octanol, 2-butyl	0.13				
AC	*1-Decanol, 2-hexyl	0.53				
AC	*9-Tetradecen-1-ol	0.27	1.12			0.44
AC	1-Hexadecanol, 2-methyl	0.45				
CA ¹⁰	Acetic acid					1.81
CA	*Tetradecanoic acid			4.18		0.66
CA	*n-Hexadecanoic acid					2.75
OT ¹¹	*Cyclotrisiloxane, hexamethyl	0.82	1.39	3.77	1.38	0.20
OT	Cyclotetrasiloxane, octamethyl		0.79			
OT	Silane, tetramethyl-			9.84		
	Sum of relative ratio of AL	37.64	36.88	24.21	32.11	21.06
	Sum of relative ratio of AR	2.44	2.14	1.81	1.80	2.41
	Sum of relative ratio of PN	1.04	0.00	11.46	1.12	0.45
	Sum of relative ratio of ON	2.08	22.53	19.52	8.08	2.46
	Sum of relative ratio of AH and KT	0.63	0.00	1.05	0.00	4.97
	Sum of relative ratio of OT	3.98	6.58	17.79	2.91	5.86
	Total unknown	6.19	17.62	21.44	9.6	7.13
	Sum of relative ratio	53.99	85.75	97.28	55.62	44.34

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones, ⁹AC = alcohol, ¹⁰CA = carboxylic acids and ¹²OT = others. The pyrolysis fragments that have a average relative ratio more than 0.2 were identified as the major pyrolysis fragments in the samples, Common fragments = Italic bold letter.

Table B-2: Average relative ratio of the pyrolysis fragment in HPON of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPON				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL3	* <i>Propene</i>	1.03	2.31	0.83	0.21	0.63
AL	* <i>2-Butene</i>	0.62	1.08	0.95	0.30	0.50
AL	*1-Pentene	0.95	0.66	0.44		0.98
AL	*1,3-Cyclopentadiene					0.88
AL	Cyclobutane			1.28		
AL	*1-Hexene	0.89			0.32	1.32
AL	Cyclohexene		1.73			0.12
AL	2,4-Hexadiyne					
AL	*1-Heptene	1.74				
AL	*1,3-Pentadiene, 2,4-dimethyl				0.11	
AL	2,4-Hexadiene, 2-methyl	0.12	0.16			
AL	1,3,5-Heptatriene					0.13
AL	* <i>I-Octene</i>	0.49	0.34	0.28	0.26	1.24
AL	* <i>I-Nonene</i>					1.34
AL	* <i>I-Decene</i>	0.32	0.39	0.38	0.33	1.13
AL	* <i>I-Undecene</i>	0.28	0.34	0.24	0.47	0.89
AL	2,6-Octadiene, 2,7-dimethyl			0.16		
AL	1,4 Undecadiene					0.12
AL	*1-Dodecene		0.36	0.24		0.75
AL	2-Dodecene	0.26				
AL	6-Dodecyne					0.20
AL	2,4-Dodecadiene, (E,Z)					0.11
AL	*1-Tridecene					0.84
AL	*Z-1,6 Tridecadiene					0.19
AL	*1-Tetradecene					0.89
AL	2,6-Octadiene, 4-methy			0.10		
AL	*7-Tetradecyne					0.22
AL	*1-Pentadecene	0.42				1.04
AL	3-Hexadecyne					0.29
AL	*1-Heptadecene					0.55
AR ⁴	*Benzene			1.42	0.88	2.60
AR	* <i>Toluene</i>	1.00	1.00	1.00	1.00	1.00
AR	*Ethylbenzene	0.94	0.82	0.67	0.65	
AR	* <i>p-Xylene</i>	0.61	0.31	0.31	0.30	0.24
AR	Benzene, 1,3-dimethyl			0.12		
AR	o-xylene		0.24		0.12	0.13
AR	* <i>Styrene</i>	0.47	0.48	0.28	0.27	0.29
AR	* <i>Benzene, (1-methylethyl)</i>	0.25	0.22	0.17	0.16	0.10
AR	* <i>Benzene, propyl</i>	0.58	0.33	0.20	0.28	0.15
AR	Benzene, 1-ethyl-2-methyl					0.10
AR	* <i>alpha.-Methylstyrene</i>	0.60	0.43	0.32	0.38	0.22

Table B-2: Average relative ratio of the pyrolysis fragment in HPON of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPON				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AR	*Benzene, 1-ethenyl-2-methyl		0.10			
AR	*Indane	0.16				
AR	*Benzene, butyl	0.82	0.23			
AR	*Benzene, (1-methylenepropyl)	0.31				
AR	*Indene		0.22			
AR	*Benzene, (2-methyl-1-propenyl)	0.11				
AR	*Benzene, 1-ethenyl-3-ethyl	0.15	0.14			
AR	*Benzene, (1-methylenebutyl)	0.39	0.25			
AR	* Benzene, pentyl	0.76	0.30	0.13	0.14	0.10
AR	Benzene, (1-methylpentyl)	0.36	0.14			
AR	* Naphthalene	0.11	0.23	0.15	0.13	0.11
AR	*Benzene, (1-methylenepentyl)	0.67	0.15			
AR	*Benzene, hexyl	1.03				
AR	Benzene, (1-methylhexyl)	0.27				
AR	Benzene, (3-methyl-1-methylenepentyl)	0.55				
AR	Hex-1-enylbenzene	0.20				
AR	Naphthalene, 1-methyl	0.26	0.27			
AR	Naphthalene, 2-methyl		0.32		0.17	
AR	*Benzene, heptyl	0.73				0.11
AR	*Benzene, (1-methylheptyl)	0.19				
AR	*Biphenyl	0.36	0.50	0.13		0.17
AR	Benzene, (1-methylenepentyl)	0.65				
AR	Benzene, 2-heptenyl	0.21				
AR	*Indole	0.70	0.70			
AR	*Benzene, octyl-	0.58				0.26
AR	*Benzene, (1-butylhexyl)	1.74				
AR	*Benzene, (1-propylheptyl)	1.36				
AR	*Benzene, (1-ethyloctyl)	1.12				
AR	*Benzene, nonyl	0.43				0.28
AR	*Benzene, (1-methylnonyl)	0.51				
AR	Naphthalene, 2-(1-methylethyl)		0.37			
AR	Naphthalene, 2-methyl-1-propyl		0.95	0.63	0.42	0.19
AR	*Benzene, (1-pentylhexyl)	1.96	0.52	0.14	0.21	
AR	Benzene, (1-butylheptyl)	3.48	0.89	0.34	0.47	
AR	*Benzene, (1-propyloctyl)	2.43	0.77	0.27	0.33	
AR	*Benzene, (1-ethylnonyl)	1.67	1.30	0.56	0.69	
AR	*Benzene, 1,1'-(1,1,2,2-tetramethyl-1,2-ethanediyl)bis	0.50				
AR	Benzene, (1-methyldecyl)	0.53	1.34		0.52	
AR	* Benzene, (1-pentylheptyl)	2.54	1.24	0.26	0.47	0.10
AR	* Benzene, (1-butyloctyl)	2.08	1.09	0.26	0.52	0.32
AR	* Benzene, (1-propylnonyl)	1.24	1.29	0.89	0.51	0.31

Table B-2: Average relative ratio of the pyrolysis fragment in HPON of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPON				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AR	Benzene, (1-pentyloctyl)	1.20	1.31		0.66	0.62
AR	Benzene, (1-butynonyl)	0.82			0.70	0.32
AR	Benzene, (1-propyldecyl)	0.42			0.56	0.33
PN ⁵	* <i>Phenol</i>	0.47	1.76	0.86	0.89	0.36
PN	* <i>Phenol, 2-methyl</i>	0.15	0.77	0.27	0.39	0.11
PN	Phenol, 2,5-dimethyl-		0.13			
PN	* <i>Phenol, 4-methyl</i>	0.40	0.53	0.24	0.34	0.23
PN	* <i>Phenol, 2-ethyl</i>		0.14			
PN	* <i>Phenol, 2,4-dimethyl</i>	0.21	0.47	0.14	0.27	0.18
PN	* <i>Phenol, 4-ethyl</i>	0.11	0.34		0.17	
PN	* <i>Phenol, 4-(1-methylethyl)</i>		2.19	0.59	0.71	
PN	* <i>Phenol, 2-methyl-5-(1-methylethyl)</i>		0.36			
PN	Phenol, p-tert-butyl		0.31			
PN	* <i>Phenol, 4-(1-methylpropyl)</i>		0.59	0.29	0.32	
PN	p-Isopropenylphenol		2.55			
PN	2-Allyl-4-methylphenol			0.26		
PN	Phenol, 2-methyl-6-(2-propenyl)		0.75			
PN	Phenol, 2-(2-methyl-2-propenyl)				0.36	
PN	.alpha.,.alpha.'-Dihydroxy-m-diisopropylbenzene		0.56			
PN	* <i>Phenol, 4-(1,1,3,3-tetramethylbutyl)</i>	1.03	3.68	0.79	0.78	0.38
PN	* <i>Phenol, 4-(1-methyl-1-phenylethyl)</i>	1.54	3.62	0.57		
ON ⁶	*Acetonitrile	0.35				0.82
ON	2-Propenenitrile					0.74
ON	*Butanenitrile, 2-methyl-	0.12				
ON	*Butanenitrile, 3-methyl	0.17	0.16	0.19		0.29
ON	* <i>Pyrrole</i>	0.98	0.26	0.30	0.24	0.33
ON	1H-Pyrazole, 3-methyl-	0.15		0.17	0.16	
ON	*1H-Pyrrole, 2-methyl-	0.15				
ON	*1H-Pyrrole, 3-methyl-	0.30				
ON	*Pantanenitrile, 4-methyl	0.11				0.13
ON	*Acetamide	0.13	0.34	0.15	0.38	
ON	1H-Pyrrole, 3-ethyl	0.26				
ON	*Aniline			1.08	0.71	0.63
ON	*Pyridine, 3-methyl	0.14	0.70			
ON	*Benzonitrile		0.32	0.24		
ON	*2H-Benzotriazole, 2-methyl		0.58	0.21	0.31	0.10
ON	6-Quinoxalinamine		0.22			0.50
ON	*Pantanenitrile					0.36
ON	*Pantanenitrile, 2-methyl					0.18
ON	Benzene propanenitrile	0.14				0.10
ON	*1H-Pyrrole-2-carbonitrile	0.26				

Table B-2: Average relative ratio of the pyrolysis fragment in HPON of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPON				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ON	1H-Benzotriazole, 1-ethenyl-		0.77		0.33	
ON	*1H-Benzotriazole, 1-methyl		3.10	1.02	1.33	
ON	*Undecanenitrile	0.91				
ON	*Dodecanenitrile	1.34				
ON	*Benzonitrile, 2-hydroxy		0.89			
ON	Dodecanamide		1.93			
ON	*Heptadecanenitrile			0.11		0.25
AH7	*Butanal, 2-methylene-				0.45	0.12
AH	*2-Pentenal, (E)	0.15	1.30	0.42		
KT8	Acetone	0.71				
KT	*2-Butanone	0.49				
KT	*2-Pantanone	0.18				
KT	*2-Propanone, 1-hydroxy				0.13	
KT	2,3-Hexanedione			0.22	0.14	
KT	3-Penten-2-one, (E)-			0.14		
KT	*Cyclopentanone					0.11
KT	*2-Cyclopenten-1-one, 2-methyl-				0.21	
KT	*2-Octanone	0.15				
KT	*Methanone, dicyclopropyl-				0.72	
KT	2-Cyclopenten-1-one, 3,4-dimethyl	0.29				
KT	Acetophenone			0.18		0.1
KT	2-Cyclohexen-1-one, 3,5,5-trimethyl			0.17		0.12
KT	2,6,6-Trimethyl-2-cyclohexene-1,4-dione	0.18		0.23	0.26	0.12
KT	*2-Heptadecanone					0.23
AC ⁹	2-Hexen-1-ol, (E)		0.27			0.16
AC	*1-Hexanol, 2-ethyl	0.57				1.63
AC	*Cyclopentanol		0.44			
AC	3-Dodecen-1-ol					0.25
AC	* 1-Octanol, 2-butyl		0.53	0.29	0.51	
AC	*9-Tetradecen-1-ol					0.23
AC	1-Naphthalenol		1.00			
CA ¹⁰	*Tetradecanoic acid		1.16			
CA	*n-Hexadecanoic acid		1.61			

Table B-2: Average relative ratio of the pyrolysis fragment in HPON of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPON				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ES ¹¹	2-Propenoic acid, 2-methyl ester		7.40	6.20	4.43	1.92
ES	2-Propenoic acid, 2-methyl-, oxiranylmethyl ester		1.84		0.50	
ES	2-Propenoic acid, 2-methyl-, 2-ethyl-2-[(2-methyl-1-oxo-2-propenyl)oxy]methyl]-1,3-propanediyl ester		8.48	2.50	3.02	
OT ¹²	*Cyclotrisiloxane, hexamethyl	1.17	1.99	1.48	0.91	1.28
OT	Cyclotetrasiloxane, octamethyl-	0.34	0.75	0.33	0.26	
OT	Benzofuran, 2,3-dihydro-2-methyl			0.71		
	Sum of relative ratio of AL	7.11	7.37	4.90	2.01	14.34
	Sum of relative ratio of AR	38.06	18.47	8.27	10.55	8.06
	Sum of relative ratio of PN	3.91	18.75	4.01	4.23	1.26
	Sum of relative ratio of ON	5.51	9.28	3.44	3.45	4.42
	Sum of relative ratio of AH and KT	2.16	1.30	1.35	1.91	0.79
	Sum of relative ratio of OT	2.07	27.57	11.51	9.62	5.47
	Total unknown	10.3	16.37	7.54	7.9	6.44
	Sum of relative ratio	69.14	99.12	41.01	39.68	40.77

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones, ⁹AC = alcohol, ¹⁰CA = carboxylic acids , ¹¹ES = Ester and ¹²OT = others, Common fragments = Italic bold letter.

Table B-3: Average relative ratio of the pyrolysis fragment in the HPOB fraction of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPOB				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL ³	*1-Pentene	0.63				
AL	*1-Hexene	0.60				
AL	2,4-Hexadiene	1.10	1.48		2.95	
AL	2,4-Hexadiyne					0.78
AL	*1-Heptene	0.98				
AL	Hexane, 2-methyl-4-methylene	0.25				
AL	*1-Octene	0.36		0.52	0.57	0.36
AL	*1-Nonene	0.36		0.68	0.82	0.41
AL	*1-Decene	0.34		0.60	1.23	0.29
AL	*1-Undecene	0.25		0.31	0.57	0.15
AL	*1-Dodecene	0.20		0.32	0.59	0.16
AL	*1-Tridecene	0.25			0.49	0.15
AL	*1-Tetradecene			0.29	0.54	0.20
AL	*1-Pentadecene	0.38		0.28	0.51	
AL	3-Hexadecyne					0.14
AL	*1-Heptadecene	0.52				
AL	*1-Nonadecene	0.13	0.33		0.39	
AL	*Nonadecane, 2-methyl		0.56			
AL	*Eicosane	0.27	1.39			
AL	*Eicosane, 2-methyl-	0.28				
AL	Heneicosane		0.42			
AR ⁴	*Benzene			3.71		
AR	*Toluene	1.00	1.00	1.00	1.00	1.00
AR	*Ethylbenzene					
AR	*p-Xylene	0.12	0.13	0.30	0.12	0.12
AR	Benzene, 1,3-dimethyl					
AR	*Styrene	0.22	0.26	1.23	0.63	0.27
AR	*.alpha.-Methylstyrene			0.30		
AR	*Naphthalene	0.10	0.24	0.16	0.18	0.10
AR	Naphthalene, 1-methyl		0.12			
AR	*Biphenyl		0.15			
PN ⁵	*Phenol	0.57	1.51	0.77	1.93	0.94
PN	*Phenol, 2-methyl	0.10	0.31		0.45	0.23
PN	*Phenol, 4-methyl	0.40	0.53		0.32	
ON ⁶	Ethylamine			1.56		
ON	*Acetonitrile	0.66	0.58	1.31		
ON	*Pyridine	0.20				
ON	*Butanenitrile, 3-methyl	0.15		0.35	0.21	0.16
ON	*Pyrrole	0.49		0.45		

Table B-3: Average relative ratio of the pyrolysis fragment in the HPOB fraction of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPOB				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ON	*1H-Pyrrole, 3-methyl	0.22		0.20		
ON	*Pantanenitrile, 4-methyl	0.14				
ON	*Acetamide	0.89				
ON	1H-Pyrrole, 3-ethyl	0.21				
ON	*Benzonitrile		0.19		0.26	
ON	Benzeneopropanenitrile		0.16			
ON	1H-Pyrrole-2-carbonitrile		0.13			
ON	*Indole	0.17				
ON	1H-Indole, 2,3-dimethyl	0.10				
ON	1H-Indole, 1,2,3-trimethyl-	0.25				
ON	*1H-Isoindole-1,3(2H)-dione		0.49			
ON	*1H-Benzotriazole		1.62		1.63	
ON	*Tridecanenitrile			0.20		
ON	*Pentadecanenitrile			0.28		
ON	*Heptadecanenitrile	0.23	0.35	0.93	0.25	
AH ⁷	Butanal, 3-methyl			0.11		
AH	*Furfural	0.46	1.34	0.46	0.20	0.31
AH	*Octanal		3.17	0.78		
AH	*2-Furancarboxaldehyde, 5-methyl	1.29			0.62	0.51
AH	*E-14-Hexadecenal	0.17				0.22
AH	E-15-Heptadecenal				0.42	0.19
KT ⁸	*2,3-Butanedione			1.27		
KT	*Cyclopentanone	0.12	0.17	0.38	0.86	0.24
KT	*2-Cyclopenten-1-one	0.29			0.36	0.13
KT	*2-Cyclopenten-1-one, 2-methyl-		0.16			
KT	*Ethanone, 1-(2-furanyl)		0.44			
KT	Cycloheptanone				0.18	
KT	1H-Isoindole-1,3(2H)-dione			0.16		
KT	*2-Heptadecanone	0.13				
AC ⁹	*1-Butanol			0.90		
AC	*1-Hexanol, 2-ethyl	0.32				
CA ¹⁰	*Formic acid		0.61			
CA	*Acetic acid		3.30		1.70	1.58
CA	*Propanoic acid		0.26			0.16
CA	*Hexanoic acid				0.18	
CA	*Benzoic Acid		0.60		0.42	0.52
CA	Nonanoic acid				0.15	
CA	*n-Decanoic acid				0.29	
CA	*1,2-Benzenedicarboxylic acid		0.25			
CA	*Benzoylformic acid					0.20
CA	Dodecanoic acid		0.38	0.21	0.62	0.36

Table B-3: Average relative ratio of the pyrolysis fragment in the HPOB fraction of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPOB				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
CA	*Tetradecanoic acid		1.92	2.63	2.11	4.73
CA	*Pentadecanoic acid		0.83	1.27	0.51	3.60
CA	*n-Hexadecanoic acid		3.64	6.81	6.42	7.25
CA	*Heptadecanoic acid					22.44
OT	*Methane, chloro	1.33				
OT ¹¹	<i>*Cyclotrisiloxane, hexamethyl</i>	0.46	5.49	0.34	0.52	0.81
OT	Cyclotetrasiloxane, octamethyl	0.12	0.43			
OT	*Dodecane, 1-chloro-					0.15
OT	*Phthalic anhydride				0.82	0.11
OT	Tetradecane, 1-chloro					0.20
	Sum of relative ratio of AL	6.90	4.19	3.00	8.67	2.65
	Sum of relative ratio of AR	1.44	1.90	6.69	1.93	1.49
	Sum of relative ratio of PN	1.07	2.36	0.77	2.71	1.17
	Sum of relative ratio of ON	3.70	3.52	5.28	2.35	0.16
	Sum of relative ratio of AH and KT	2.46	5.29	3.17	2.65	1.61
	Sum of relative ratio of OT	2.24	17.12	12.15	13.75	42.12
	Total unknown	1.33	7.31	7.35	7.80	5.61
	Sum of relative ratio	19.14	42.29	38.42	37.75	50.07

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones, ⁹AC = alcohol, ¹⁰CA = carboxylic acids , ¹¹ES = Ester and ¹²OT = others, Common fragments = italic bold letter.

Table B-4: Average relative ratios of the pyrolysis fragment in HPOA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPOA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL ³	* <i>Propene</i>	0.51	0.36	0.56	0.24	0.30
AL	* <i>2-Butene</i>	0.62	0.22	0.24	0.28	0.18
AL	*1-Pentene	0.53			0.53	0.28
AL	* <i>1,3-Cyclopentadiene</i>	1.01	0.86	2.81	0.96	1.10
AL	*1-Hexene	0.40			0.32	0.38
AL	*1,3,5-Hexatriene, (Z)	0.51	0.30	0.70		
AL	* <i>1,3-Cyclohexadiene</i>	0.43	0.23	0.50	0.14	0.21
AL	*Cyclopentene, 3-ethenyl	0.14	0.11	0.10		0.11
AL	1,3,5-Heptatriene,			0.10		
AL	*1,3-Cyclopentadiene, 5,5-dimethyl	0.16	0.13			0.11
AL	*1,3,5-Hexatriene, 3-methyl-, (Z)	0.10	0.10			
AL	*1-Octene	0.27		0.10	0.50	0.29
AL	*Cyclopentene, 3-ethylidene-1-methyl	0.10			0.60	0.26
AL	*1-Nonene				0.43	0.21
AL	*1-Decene	0.36			0.25	0.10
AL	Cyclobutane, 3-hexyl-1,1,2-trimethyl		0.10		0.11	
AL	*1-Undecene	0.19			0.38	0.19
AL	1,4 Undecadiene				0.13	
AL	*1-Dodecene	0.25	0.10		0.10	0.22
AL	6-Dodecyne	0.10			0.10	
AL	2,4-Dodecadiene, (E,Z)				0.11	
AL	*1-Tridecene	0.30			0.10	
AL	*6-Tridecene, (E)				0.10	
AL	*Z-1,6 Tridecadiene	0.10		0.20		
AL	*Benzocycloheptatriene					
AL	*1-Tetradecene	0.30			0.59	0.20
AL	*7-Tetradecyne	0.10			0.24	0.17
AL	*1-Pentadecene	0.34			0.08	
AL	3-Hexadecyne	0.10			0.14	0.10
AL	Cyclopentene, 1-octyl				0.10	
AL	*1-Heptadecene	0.10			0.62	0.11
AL	*1-Nonadecene	0.10			0.23	0.16
AL	*Tridecane 7-hexyl				0.12	
AL	*9-Eicosene, (E)				0.21	0.11
AL	*Eicosane		0.10	0.11	0.30	0.17
AL	*Eicosane, 2-methyl-			0.12	0.17	
AL	Heneicosane		0.10			0.34
AL	*Eicosane 10 methyl					0.19
AR ⁴	* <i>Benzene</i>	1.52	1.52	3.13	2.35	2.18
AR	* <i>Toluene</i>	1.00	1.00	1.00	1.00	1.00
AR	*Ethylbenzene	0.53	0.35	0.16		0.10

Table B-4: Average relative ratios of the pyrolysis fragment in HPOA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPOA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AR	*Styrene	0.12	0.19		0.10	0.10
AR	*Benzene, (1-methylethyl)	0.13	0.11			
AR	*Benzene, propyl	0.17	0.10			
AR	Benzene, 1-ethyl-2-methyl	0.10	0.10			
AR	*.alpha.-Methylstyrene	0.10	0.10			
AR	Benzene, 1-ethyl-3-methyl	0.10	0.10			
AR	Benzene, 1,2,3-trimethyl		0.17			
AR	*Benzene, 1-ethenyl-4-methyl		0.13			
AR	Benzene, 1-ethyl-4-methyl		0.10			
AR	*Benzene, 1-ethenyl-2-methyl	0.17				
AR	*Benzene, butyl	0.26	0.13			
AR	<i>Indene</i>	0.26	0.45	0.26	0.41	0.30
AR	*Benzene, (1-methylenebutyl)		0.12			
AR	*Benzene, pentyl	0.14	0.12		0.10	
AR	*1H-Indene, 1-methyl	0.10	0.12			
AR	*1H-Indene, 2-methyl	0.10	0.10			
AR	Benzene, (1-methylpentyl)		0.10			
AR	*Naphthalene	0.10	0.16	0.10	0.11	
AR	*Benzene, (1-methylenepentyl)		0.10			
AR	Naphthalene, 1-methyl	0.12	0.11	0.10		
AR	Naphthalene, 2-methyl	0.10	0.16		0.11	
AR	*Benzene, heptyl				0.10	
AR	*Biphenyl	0.14	0.27		0.19	0.10
AR	*Naphthalene, 1-ethyl		0.12			
AR	Naphthalene, 1,3-dimethyl		0.11			
AR	*Benzene, octyl-	0.10			0.22	0.10
AR	1,1'-Biphenyl, 4-methyl		0.12	0.11		
AR	*Benzene, nonyl				0.14	0.10
AR	Fluorene		0.10	0.11	0.12	0.10
AR	*Benzene, decyl				0.16	
AR	Benzene, (1-methyldecyl)		0.10			
AR	*Benzene, (1-pentylheptyl)		0.10		0.10	
AR	*Benzene, (1-butyoctyl)				0.10	
AR	*Benzene, (1-propynonyl)		0.10			
AR	Benzene, (1-ethyldecyl)		0.11			
AR	*Benzene, undecyl-				0.10	
AR	Benzene, (1-methylundecyl)		0.10		0.10	
AR	9H-Fluorene, 2-methyl		0.11			
AR	Benzene, (1-pentyloctyl)		0.10		0.10	

Table B-4: Average relative ratios of the pyrolysis fragment in HPOA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPOA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AR	Benzene, tetradecyl				0.10	
AR	Benzene, (1-methyldodecyl)		0.10			
PN ⁵	*Phenol	0.13	0.45	0.28	0.12	
PN	*Phenol, 2-methyl	0.10	0.28	0.24		
PN	*Phenol, 4-methyl	0.10	0.18	0.12	0.11	
PN	*Phenol, 2-ethyl	0.10				
PN	*Phenol, 2,4-dimethyl		0.10	0.11		
PN	*Phenol, 4-ethyl		0.10			
PN	*Phenol, 4-(1-methylethyl)		0.28	0.17		
PN	*1-Naphthalenol		0.13	0.11		
ON ⁶	*Acetronitrile		0.10			
ON	*Isobutyronitrile	0.12	0.14		0.10	0.12
ON	*Butanenitrile, 2-methyl-	0.10	0.10		0.10	0.10
ON	*Butanenitrile, 3-methyl	0.24	0.26	0.10	0.15	0.22
ON	*Pyrrole	0.26	0.80	0.50	0.43	0.48
ON	*1H-Pyrrole, 2-methyl	0.11	0.13	0.10		0.10
ON	*1H-Pyrrole, 3-methyl	0.10	0.22	0.13		0.10
ON	*Pantanenitrile, 4-methyl	0.10				
ON	*1H-Pyrrole, 2,3-dimethyl-	0.10	0.10			
ON	*Pyridine, 3-methyl					0.10
ON	*Indole	0.16	0.15	0.10	0.22	0.13
ON	*Pentadecanenitrile				0.15	
ON	*Heptadecanenitrile	0.10			0.17	
AH ⁷	*E-14-Hexadecenal	0.11				0.11
KT ⁸	*Methyl Isobutyl Ketone	0.27	0.43	0.27	0.21	0.35
KT	*Cyclopentanone, 2-methyl		0.15			
KT	*Cyclopentanone, 2,5-dimethyl		0.10			
KT	*Cyclopentanone, 2,4,4-trimethyl-	0.82	1.47	0.98	0.47	0.75
KT	*2-Cyclopenten-1-one,2 3,4-dimethyl	0.10				
KT	*2-Pentadecanone				0.10	
KT	*2-Heptadecanone	0.10			0.15	
AC ⁹	*1-Hexanol, 2-ethyl	0.18				0.20
AC	*1-Octanol, 2-butyl				0.56	
AC	*2-Hexyl-1-octanol		0.10			
AC	1-Decanol, 2-methyl				1.27	
AC	*2-Ethyl-1-dodecanol					0.35
AC	*1-Decanol, 2-ethyl				0.30	
AC	*1-Decanol, 2-hexyl				0.36	0.14
AC	*9-Tetradecen-1-ol	0.10			0.10	
CA ¹⁰	Heptanoic acid, anhydride		0.11			
CA	Dodecanoic acid				0.30	

Table B-4: Average relative ratios of the pyrolysis fragment in HPOA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPOA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ES ¹¹	Allyl heptanoate			0.18		
OT ¹²	Cyclotetrasiloxane, octamethyl	0.10				
OT	Silane, tetramethyl-					0.14
	Sum of relative ratio of AL	7.12	2.92	5.34	7.95	5.47
	Sum of relative ratio of AR	5.33	7.17	4.97	5.64	4.07
	Sum of relative ratio of PN	0.42	1.52	1.03	0.23	0.00
	Sum of relative ratio of ON	1.40	2.01	0.93	1.32	1.34
	Sum of relative ratio of AH and KT	1.40	2.15	1.25	0.93	1.21
	Sum of relative ratio of OT	0.38	0.21	0.18	2.87	0.82
	Total unknown	0.99	1.31	0.85	2.86	1.58
	Sum of relative ratio	17.04	17.27	14.54	21.80	14.49

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones, ⁹AC = alcohol, ¹⁰CA = carboxylic acids, and ¹²OT = others, Common fragments = Italic bold letter.

Table B-5: Average relative ratios of the pyrolysis fragment in HPIB of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPIB				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL ³	*Propene					1.07
AL	*2-Butene		0.43			
AL	*1-Pentene					1.20
AL	*1,3-Cyclopentadiene	2.02		0.65		
AL	*1,3-Pentadiene					0.54
AL	*1-Hexene		0.46	0.97		1.25
AL	*1,3,5-Hexatriene, (Z)		1.99			
AL	2,4-Hexadiene			4.37		
AL	2,4-Hexadiyne	3.89			7.44	3.83
AL	*1-Octene	0.71	0.41	1.02	1.70	0.46
AL	*1-Nonene	0.80	0.43	1.04	1.63	0.40
AL	*1-Decene	0.65	0.40	0.90	1.11	0.28
AL	*1-Undecene	0.79	0.35	0.78		0.23
AL	*2-Undecene				1.03	
AL	*1-Dodecene	0.51	0.28	0.61	0.81	0.26
AL	Cyclododecene, (E)	0.16				
AL	*1-Tridecene	0.64	0.32	0.50	0.78	
AL	*1-Tetradecene	0.58	0.24	0.60	0.73	0.22
AL	*7-Tetradecyne	0.27	0.17			0.10
AL	*1-Pentadecene	0.91		1.15		0.31
AL	*1-Nonadecene		0.22			
AR ⁴	*Toluene	1.00	1.00	1.00	1.00	1.00
AR	p-Xylene		0.11			
AR	o-xylene		0.11			
AR	*Styrene	0.20	0.14	0.37	0.29	0.24
AR	*Benzene, octyl-	0.16				
AR	*Benzene, (1-butylOctyl)					0.13
AR	Benzene, (1-ethylDecyl)					0.11
PN ⁵	*Phenol	0.14	0.56			0.11
ON ⁶	*Acetonitrile	4.50				2.52
ON	1,2-Propanediamine				5.35	
ON	*2-Propenenitrile	2.51				0.94
ON	*Isobutyronitrile					1.25
ON	*2-Propenenitrile, 2-methyl-	0.26				
ON	Butanenitrile					0.22
ON	*2-Methyl-2-butenenitrile	0.19				
ON	*Butanenitrile, 2-methyl-	0.10	0.20	0.27		0.46
ON	2-Pentenenitrile	0.10				
ON	*Pyrrole	0.18	0.19	0.51	0.52	0.27
ON	*Butanenitrile, 3-methyl	0.33	0.25	0.60	0.35	1.69
ON	*2,4-Pentadienenitrile	0.22				0.30

Table B-5: Average relative ratios of the pyrolysis fragment in HPIB of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPIB				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
ON	*Pyridine, 3-methyl					0.39
ON	*Benzonitrile	0.30				
ON	*Benzonitrile, 2-methyl	0.15				
ON	*Indole					0.12
AH ⁷	*Acetaldehyde			1.17		
AH	*E-14-Hexadecenal		0.12			
KT ⁸	*Cyclopentanone			0.19		
KT	*Cyclopentanone, 2,4,4-trimethyl-		0.52			
AC ⁹	*1-Hexanol, 2-ethyl					0.96
AC	*1-Decanol, 2-methyl		0.54			
AC	*1-Decanol, 2-ethyl				0.91	
AC	*9-Tetradecen-1-ol		0.14			
	Sum of relative ratio of AL	11.93	5.71	12.61	15.23	10.14
	Sum of relative ratio of AR	1.36	1.36	1.37	1.29	1.49
	Sum of relative ratio of PN	0.14	0.56	0.00	0.00	0.11
	Sum of relative ratio of ON	8.84	0.64	1.38	6.22	8.16
	Sum of relative ratio of AH and KT	0.00	0.64	1.37	0.00	0.00
	Sum of relative ratio of OT	0.00	0.67	0.00	0.91	0.96
	Total unknown	2.04	1.57	2.29	2.53	1.59
	Sum of relative ratio	24.31	11.14	19.01	26.18	22.46

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones and ⁹AC = alcohol, Common fragment = Italic bold letter

Table B-6: Average relative ratios of the pyrolysis fragment in HPIA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPIA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AL ³	*Propene	1.78	2.21			2.16
AL	*2-Butene	0.97	2.05			
AL	*1-Pentene	1.31	2.26			
AL	Cyclopentene			2.26		
AL	*1-Hexene	3.44	3.02			5.02
AL	2,4-Hexadiene	0.61	3.91	4.35		
AL	*1-Heptene	3.52			6.09	5.64
AL	*1-Octene	1.86	1.34	1.87	2.60	2.22
AL	*1-Nonene	1.75	1.16	2.13	2.19	1.61
AL	*1-Decene	1.20	0.93	2.96	2.03	1.21
AL	*1-Undecene	1.06	0.85	2.58	1.06	0.98
AL	1,4 Undecadiene		0.14			
AL	*1-Dodecene	0.96	0.75	2.43	0.87	0.77
AL	Cyclododecene, (E)				0.19	
AL	6-Dodecyne	0.20				
AL	Undecane, 2,10-dimethyl			4.05		
AL	2,4-Dodecadiene, (E,Z)		0.21			
AL	*1-Tridecene	0.86	0.75	17.18	0.90	0.77
AL	*Z-1,6 Tridecadiene	0.22	0.14			
AL	Tridecane, 3-methyl			1.97		
AL	*1-Tetradecene	0.93	0.82	1.49	0.86	0.82
AL	*Dodecane, 2,6,11-trimethyl			2.51		
AL	*7-Tetradecyne	0.20	0.25			
AL	Tetradecane, 2-methyl-			6.29		
AL	*1-Pentadecene	1.00	1.12	12.20	1.00	0.84
AL	3-Hexadecyne		0.16			
AL	Cyclopentene, 1-octyl	0.31		1.41		
AL	Cyclohexene, 3-nonyl	0.14				
AL	Cyclotetradecane				0.44	
AL	*1-Heptadecene	0.55	0.43	3.51	0.36	
AL	*1-Nonadecene		0.48			
AL	10-Methyl-octadec-1-ene	0.69				
AL	*Nonadecane, 2-methyl	1.28				
AL	*Eicosane	1.85				
AL	*Eicosane 10 methyl	1.26				
AL	*Tricosane	0.95				
AR ⁴	*Toluene	1.00	1.00	1.00	1.00	1.00
AR	o-xylene	0.19				
AR	*Styrene	0.30	0.22	1.07	0.53	2.07
AR	*Naphthalene		0.16			
AR	*Biphenyl		0.16			

Table B-6: Average relative ratios of the pyrolysis fragment in HPIA of influent wastewater and effluent water from the aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPIA				
		Inf. ¹ wastewater	Eff. ² Water from aeration ponds	Eff. Water from facultative ponds	Eff. Water from oxidation ponds	Eff. Water from detention pond
AR	Benzene, (1-methyldecyl)	0.27				
PN ⁵	*Phenol	0.16				
PN	*Benzenemethanol, .alpha.,.alpha.-dimethyl-	0.55				
PN	*Phenol, 4-methyl	0.19				
ON ⁶	*1,2-Propanediamine				2.51	
ON	*Butanenitrile, 2-methyl-	0.16				
ON	*Butanenitrile, 3-methyl	0.42	0.49			0.43
ON	*Pyrrole	0.21	0.30			0.23
ON	*Pantanenitrile, 4-methyl	0.29	0.33			0.29
ON	*Indole	0.53	0.34			
ON	*Tridecanenitrile		0.30			
ON	Tetradecanenitrile	0.49	0.52			
ON	*Pentadecanenitrile			0.33		
ON	Hexadecanenitrile		0.37			0.40
ON	Dodecanamide					
ON	*Heptadecanenitrile	0.94				
AH ⁷	*Acetaldehyde				0.47	2.37
AH	*Furfural					
AH	*E-14-Hexadecenal	0.42				0.27
AH	E-15-Heptadecenal	0.50				
KT ⁸	*Cyclopentanone			0.99	0.89	
KT	*2-Heptadecanone	0.48			0.48	
AC ⁹	*1-Hexanol, 2-ethyl	1.99		1.29	1.17	0.62
AC	*1-Octanol, 2-butyl			4.12		
AC	10-Undecen-1-ol	0.19		0.69		
AC	*1-Decanol, 2-hexyl			1.33		
AC	*9-Tetradecen-1-ol	0.18		2.47		
OT ¹²	*Cyclotrisiloxane, hexamethyl				1.97	
	Sum of relative ratio of AL	28.91	22.99	69.20	18.58	22.03
	Sum of relative ratio of AR	1.76	1.54	2.07	1.53	3.07
	Sum of relative ratio of PN	0.90	0.00	0.00	0.00	0.00
	Sum of relative ratio of ON	3.04	2.66	0.00	2.84	1.35
	Sum of relative ratio of AH and KT	1.40	0.00	0.99	1.84	2.64
	Sum of relative ratio of OT	2.36	0.00	9.91	3.14	0.62
	Total unknown	7.12	7.82	19.01	8.57	1.32
	Sum of relative ratio	45.49	35.00	101.18	36.50	31.03

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones ⁹AC = alcohol and ¹²OT = others
Common fragments = Italic bold letter

Table B-7: Average relative ratio of the pyrolysis fragment in HPIN of influent wastewater, and effluent water from aeration, facultative, oxidation and detention ponds

Chemical Classes	Important fragment	Average relative ratio of HPIN				
		Inf. ¹ wastewater	Eff. ² Water after aeration ponds	Eff. Water after facultative ponds	Eff. Water after oxidation ponds	Eff. Water after detention pond
AL ³	*Propene	1.72	0.61			
AL	*2-Butene	2.80	0.73	4.81		1.37
AL	*1-Pentene	1.52				1.08
AL	*1-Hexene	3.44				2.95
AL	2,4-Hexadiene		2.39	3.28		
AL	*1-Heptene	5.44				4.33
AL	1-Pentene, 2,4,4-trimethyl				0.12	
AL	*1-Octene	1.31	0.67	1.26	0.45	0.95
AL	*Cyclopentene, 3-ethylidene-1-methyl				0.53	
AL	*1-Nonene	1.33	0.78	1.66		1.14
AL	*1-Decene	1.29	0.51	1.46	0.34	1.14
AL	*1-Undecene	1.08	0.38	1.29	0.30	0.92
AL	1,4 Undecadiene	0.17				
AL	*1-Dodecene	0.83	0.42	1.74	0.24	0.83
AL	6-Dodecyne	0.26				0.17
AL	*1-Tridecene	0.74	0.46	2.07	0.23	1.10
AL	*Z-1,6 Tridecadiene	0.22				
AL	*1-Tetradecene	0.69	0.35	1.78	0.18	0.69
AL	*1-Pentadecene		0.44		0.18	1.17
AL	3-Hexadecyne	1.00				
AL	*1-Heptadecene	0.63		3.79	0.11	
AL	*1-Nonadecene		0.43	6.98		
AL	*Tridecane 7-hexyl		0.87		0.25	
AL	Octadecane, 2-methyl	1.06				1.15
AL	*9-Eicosene, (E)		1.78		0.74	
AL	*Nonadecane, 2-methyl	2.49		16.90		2.80
AL	*Nonadecane, 9-methyl		3.25			
AL	*Eicosane	3.58		27.80	1.58	5.85
AL	*Eicosane, 2-methyl-		3.35		2.71	
AL	Heneicosane	3.45		26.17		6.75
AL	*Eicosane 10 methyl		2.69		2.90	
AL	*Tricosane	2.21				4.73
AL	*Tetracosane				1.93	
AR ⁴	*Benzene				3.04	
AR	*Toluene	1.00	1.00	1.00	1.00	1.00
AR	*p-Xylene	0.28	0.40	0.49	0.15	0.20
AR	o-xylene		0.23			0.49
AR	*Styrene	0.29	0.25	0.59	0.26	0.36
AR	*Indene		0.25			
AR	*Naphthalene	0.14	0.13			0.15
AR	*Benzene, hexyl			0.87		

Table B-7: Average relative ratio of the pyrolysis fragment in HPIN of influent wastewater, and effluent water from aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPIN				
		Inf. ¹ wastewater	Eff. ² Water after aeration ponds	Eff. Water after facultative ponds	Eff. Water after oxidation ponds	Eff. Water after detention pond
AR	*Benzene, (1-butyloctyl)	0.13				
AR	*Benzene, undecyl-			0.90		
PN ⁵	*Phenol	1.06		2.63	0.71	1.87
PN	*Phenol, 2-methyl	0.31		0.93	0.16	0.32
PN	*Phenol, 4-methyl	0.45		0.75	0.18	0.49
PN	Phenol, p-tert-butyl			1.22		
PN	*Phenol, 4-(1,1,3,3-tetramethylbutyl)	0.72		7.70	0.86	1.70
ON ⁶	*Acetonitrile (SI:85)			2.07		
ON	*Butanenitrile, 3-methyl	0.20	0.35		0.11	0.12
ON	*Pyrrole	0.79	0.58	1.18	0.46	0.45
ON	*1H-Pyrrole, 2-methyl	0.24	0.21	0.39		0.24
ON	*1H-Pyrrole, 3-methyl	0.39	0.25	0.32		0.20
ON	*Pantanenitrile, 4-methyl	0.23				0.54
ON	*Acetamide	0.76		3.34		0.64
ON	1H-Pyrrole, 3-ethyl	0.45	0.19			
ON	Caprolactam			3.97		1.12
ON	*Benzonitrile, 2-hydroxy					0.56
ON	Tetradecanenitrile					0.19
ON	*Heptadecanenitrile	0.41				0.64
AH ⁷	*Acetaldehyde				0.65	
AH	*2-Pentenal, (E)	0.33				
AH	Butanedia					0.17
AH	*Furfural					0.94
AH	*2-Furancarboxaldehyde, 5-methyl					0.48
AH	*E-14-Hexadecenal	0.32				0.32
KT ⁸	Acetone		1.70			
KT	*2-Butanone	3.51	0.84			
KT	*2-Butanone, 3-methyl-			0.60		0.34
KT	*2,3-Butanedione					0.23
KT	*2-Pentanone				0.12	
KT	*2,3-Pentanedione			0.56		
KT	*2-Propanone, 1-hydroxy	0.40				0.34
KT	2,3-Pentanedione, 4-methy					0.33
KT	*Cyclopantanone	0.50		0.49	0.25	0.22
KT	*Cyclopantanone, 2-methyl		0.17			
KT	*2-Cyclopenten-1-one	2.22		3.91	0.66	0.81
KT	*2-Cyclopenten-1-one, 2-methyl-	1.13		1.67	0.33	0.52
KT	*Ethanone, 1-(2-furanyl)	0.28				
KT	2-Cyclopenten-1-one, 2,3-dimethyl-	0.20				
KT	*Butyrolactone	0.26		0.71		0.24
KT	2-Cyclopenten-1-one, 2,3,4-dimethyl	0.17				

Table B-7: Average relative ratio of the pyrolysis fragment in HPIN of influent wastewater, and effluent water from aeration, facultative, oxidation and detention ponds (*continued*)

Chemical Classes	Important fragment	Average relative ratio of HPIN				
		Inf. ¹ wastewater	Eff. ² Water after aeration ponds	Eff. Water after facultative ponds	Eff. Water after oxidation ponds	Eff. Water after detention pond
AC ⁹	*2-Propanol, 2-methyl					
AC	*1-Hexanol, 2-ethyl	1.57	0.25	4.41	0.66	1.88
AC	* 1-Octanol, 2-butyl			8.94		
CA ¹⁰	*Acetic acid					0.72
CA	*Benzoic Acid					0.24
CA	Isopropyl Myristate					0.34
OT ¹²	*Cyclotrisiloxane, hexamethyl	1.26			0.90	1.37
OT	Cyclotetrasiloxane, octamethyl				0.13	
	Sum of relative ratio of AL	37.29	20.10	100.97	12.76	39.14
	Sum of relative ratio of AR	1.84	2.26	3.85	4.45	2.20
	Sum of relative ratio of PN	2.54	0.00	13.24	1.90	4.38
	Sum of relative ratio of ON	3.47	1.57	11.27	0.57	4.71
	Sum of relative ratio of AH and KT	9.31	2.72	7.93	2.00	4.94
	Sum of relative ratio of OT	2.83	0.25	13.35	1.69	4.54
	Total unknown	12.42	4.32	48.76	5.16	15.74
	Sum of relative ratio	69.69	31.22	199.36	28.53	75.64

Remark: ¹Inf = Influent and ²Eff. = Effluent, ³AL= aliphatic hydrocarbon, ⁴AR = aromatic hydrocarbon, ⁵PN =phenol, ⁶ON = organic nitrogen, ⁷AH = aldehydes, ⁸KT = ketones, ⁹AC = alcohol, ¹⁰CA = carboxylic acids , and ¹²OT = others, Common fragment = Italic bold letter

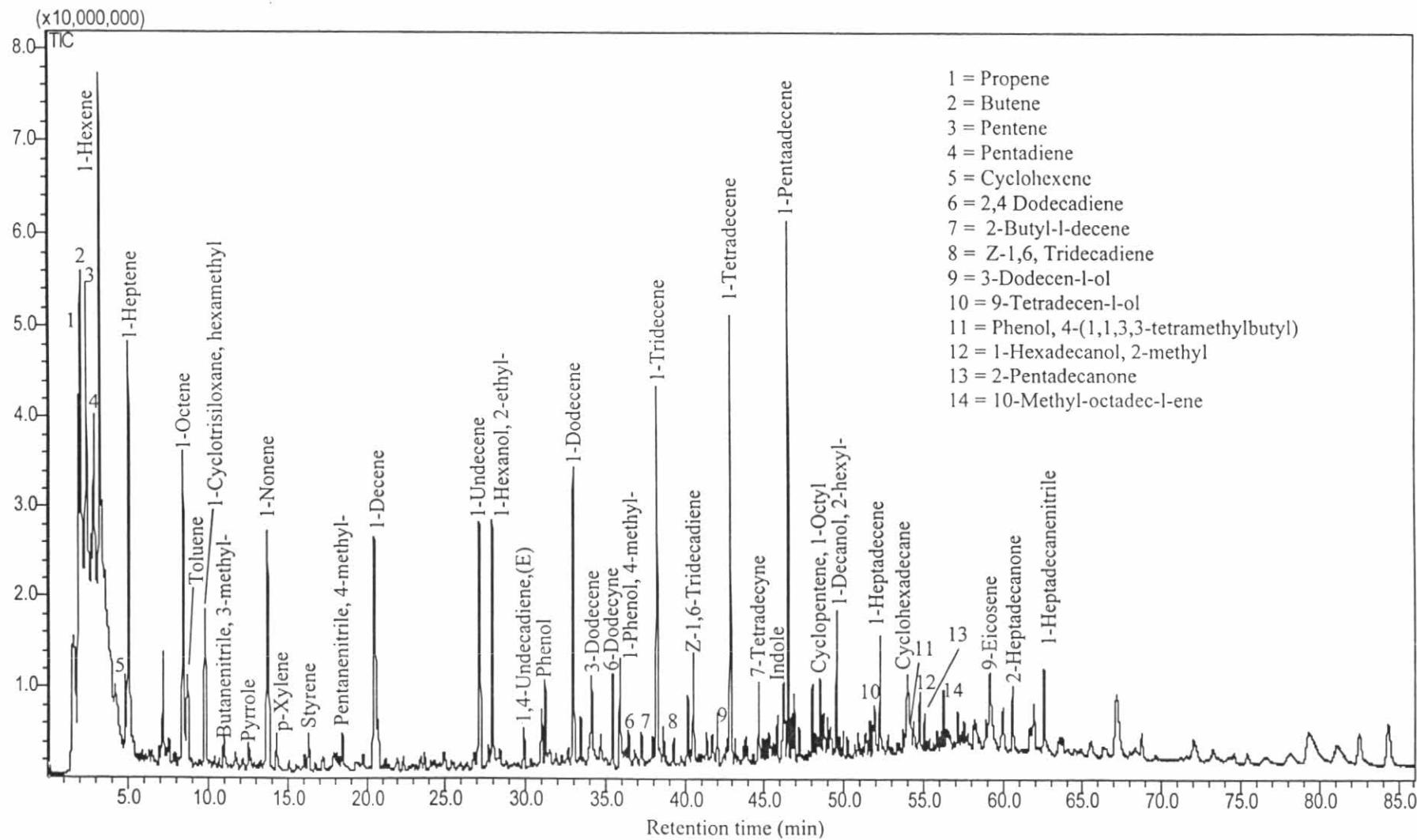


Figure B-5: Pyrochromatogram of major fragments of influent wastewater

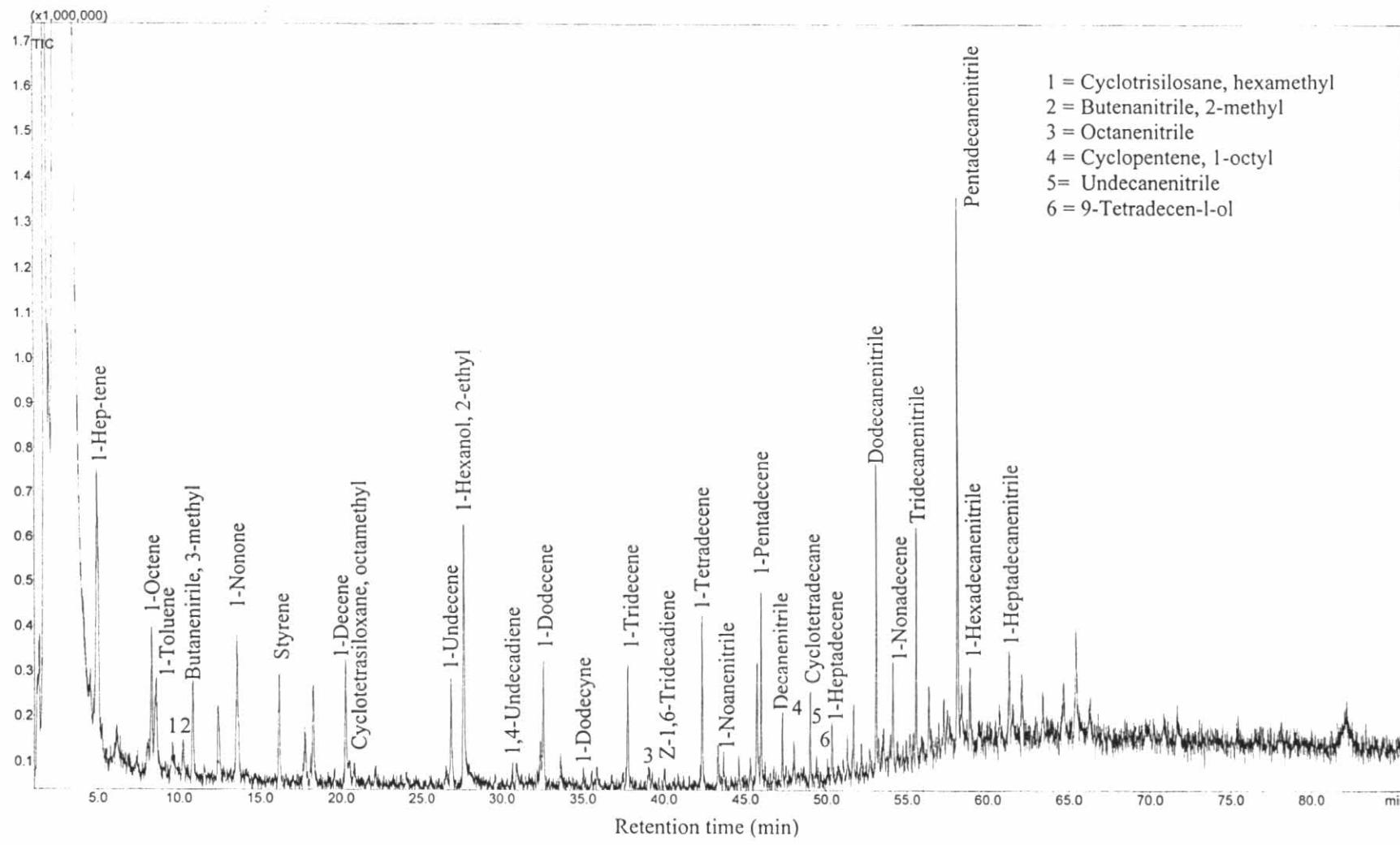


Figure B-6: Pyrochromatogram of major fragments of the aeration pond effluent

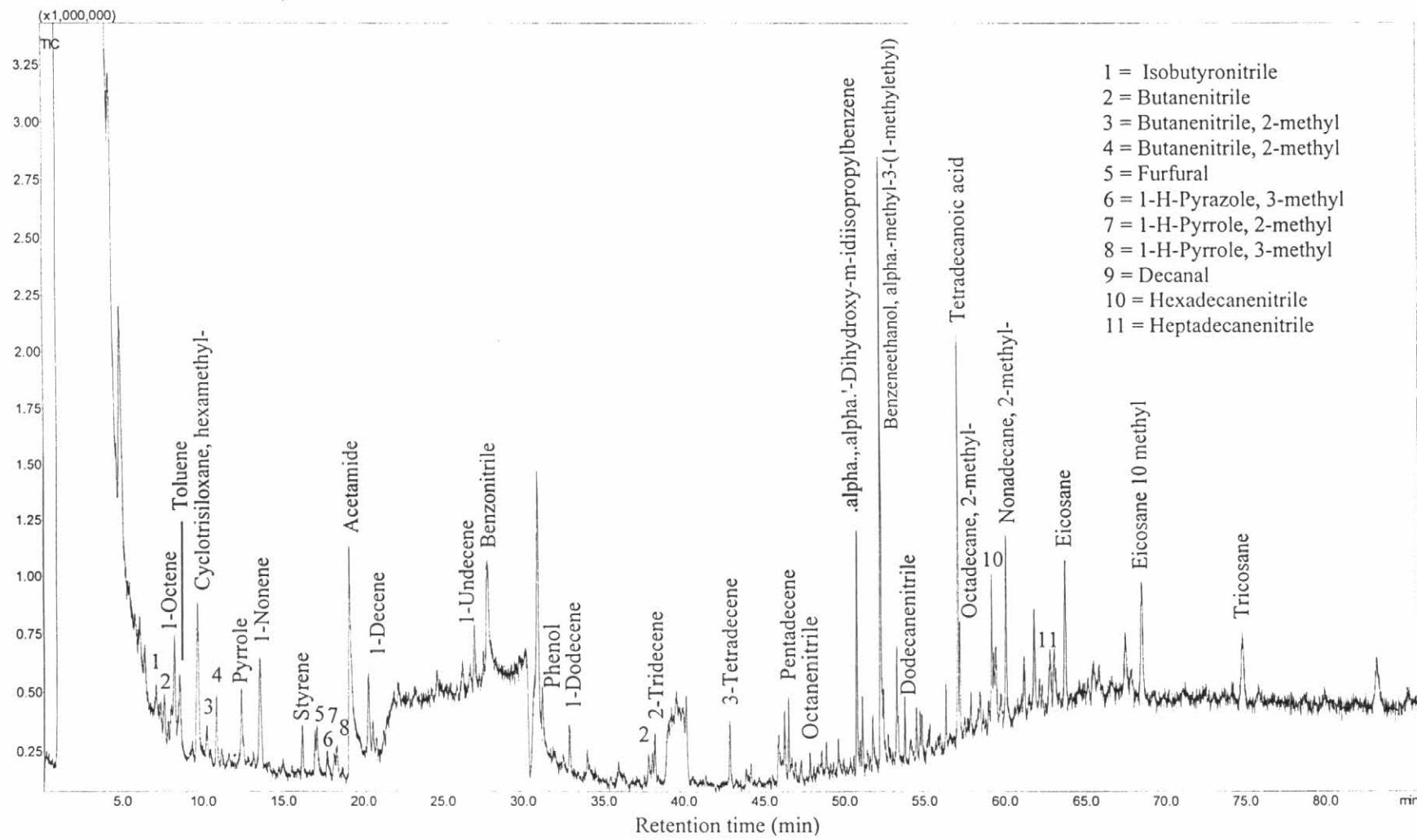


Figure B-7: Pyrochromatogram of major fragments of the facultative pond effluent

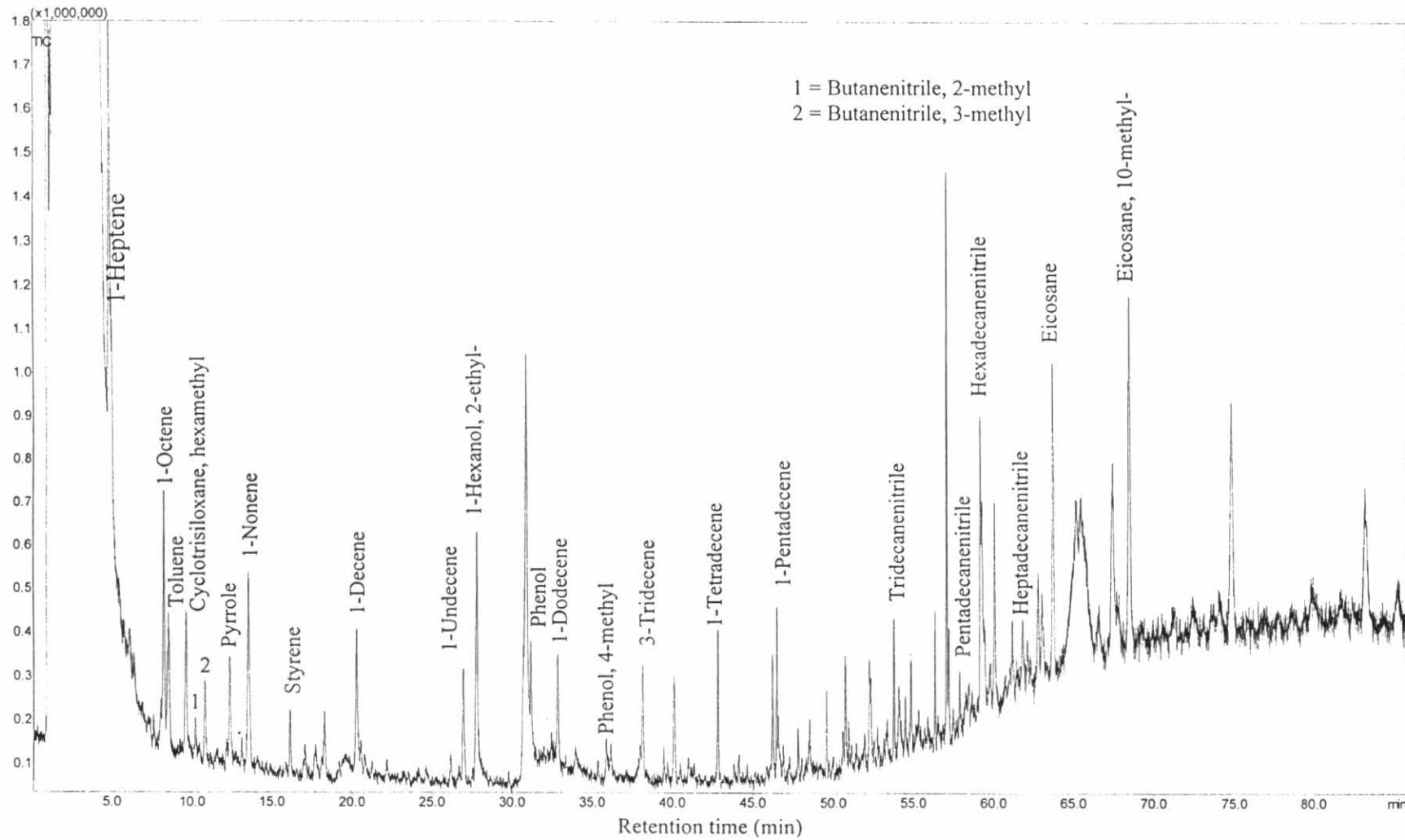


Figure B-8: Pyrochromatogram of major fragments of the oxidation pond effluent

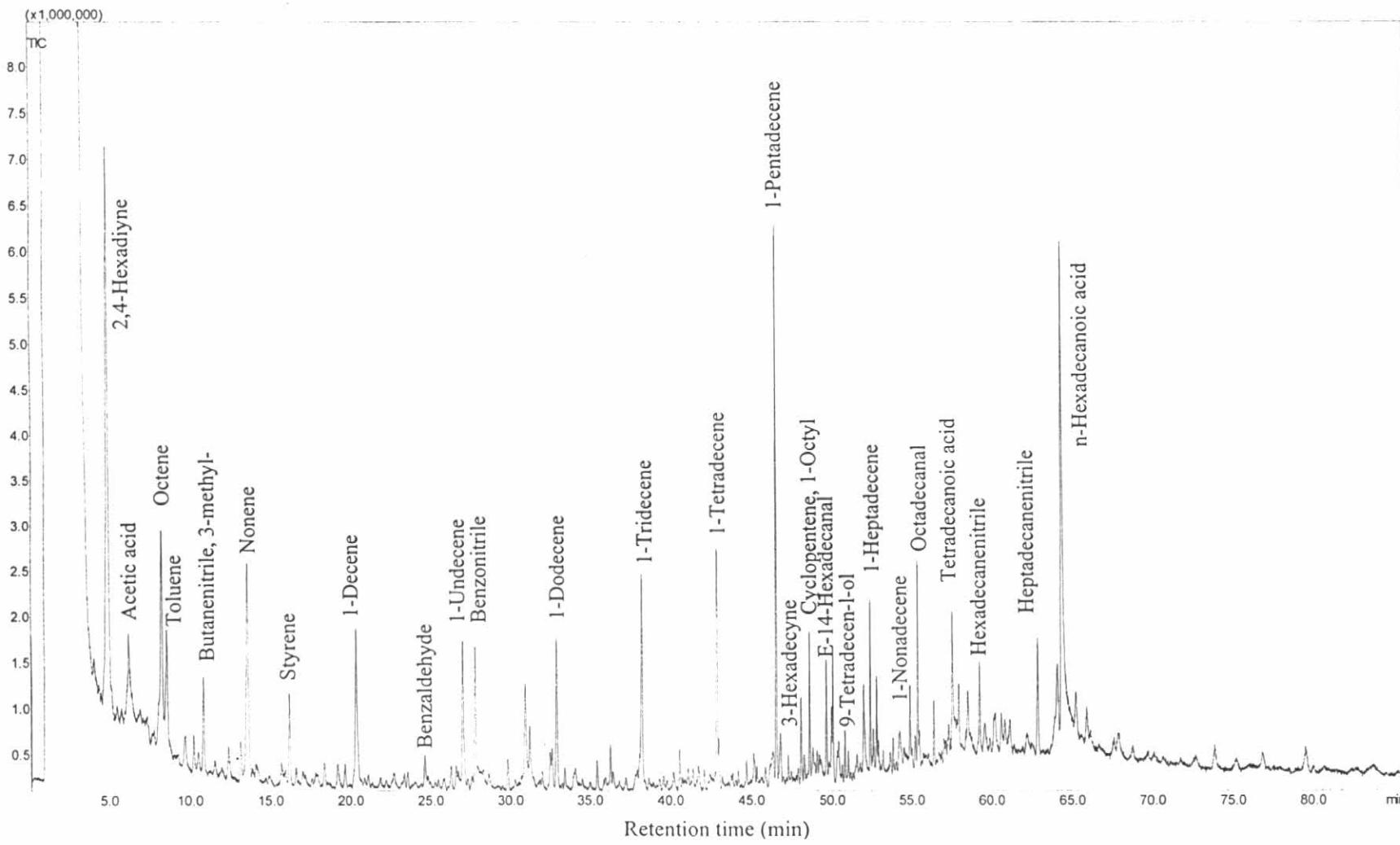


Figure B-9: Pyrochromatogram of major fragments of the detention pond effluent

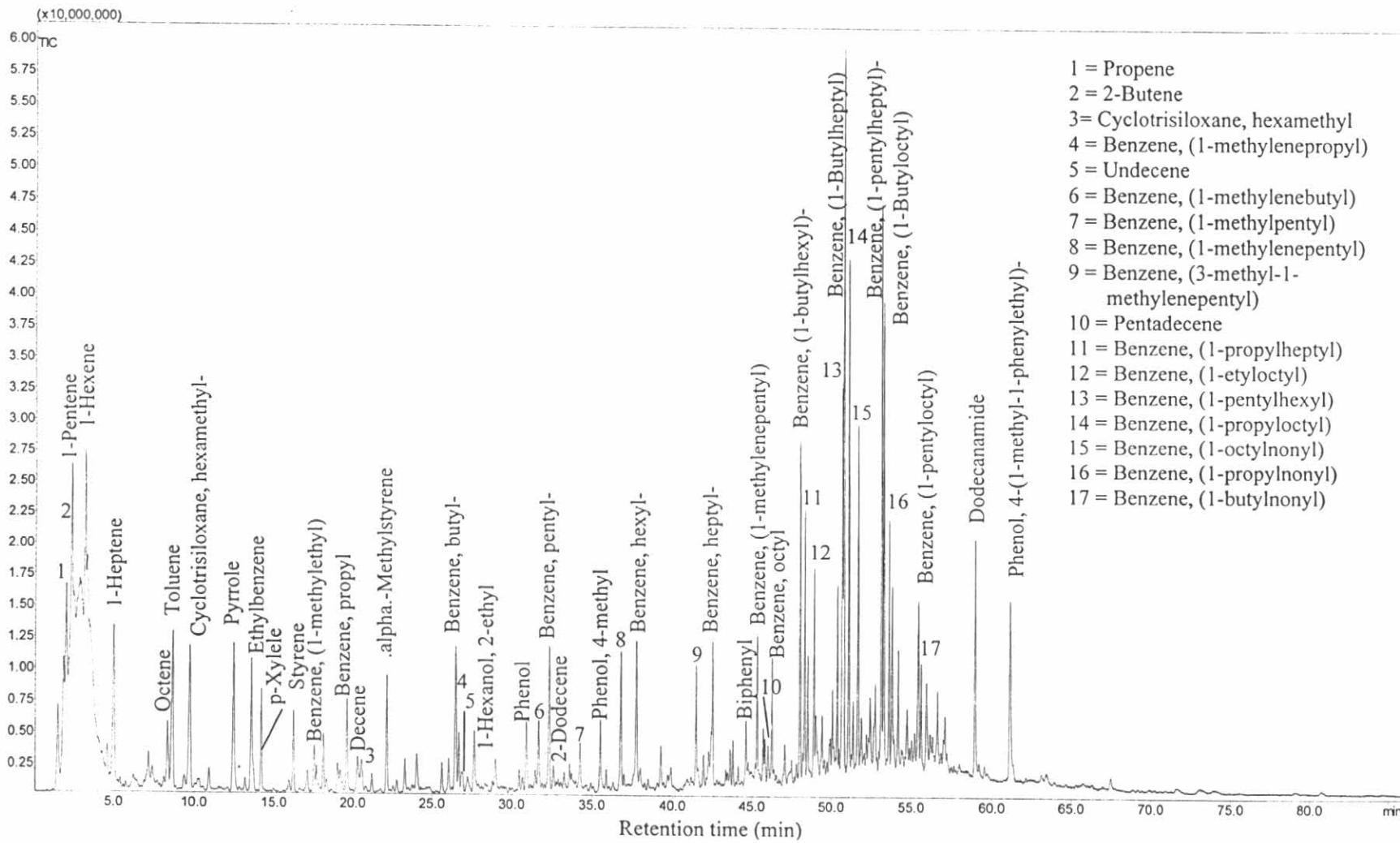


Figure B-10: Pyrochromatogram of major fragments of the HPON of influent wastewater

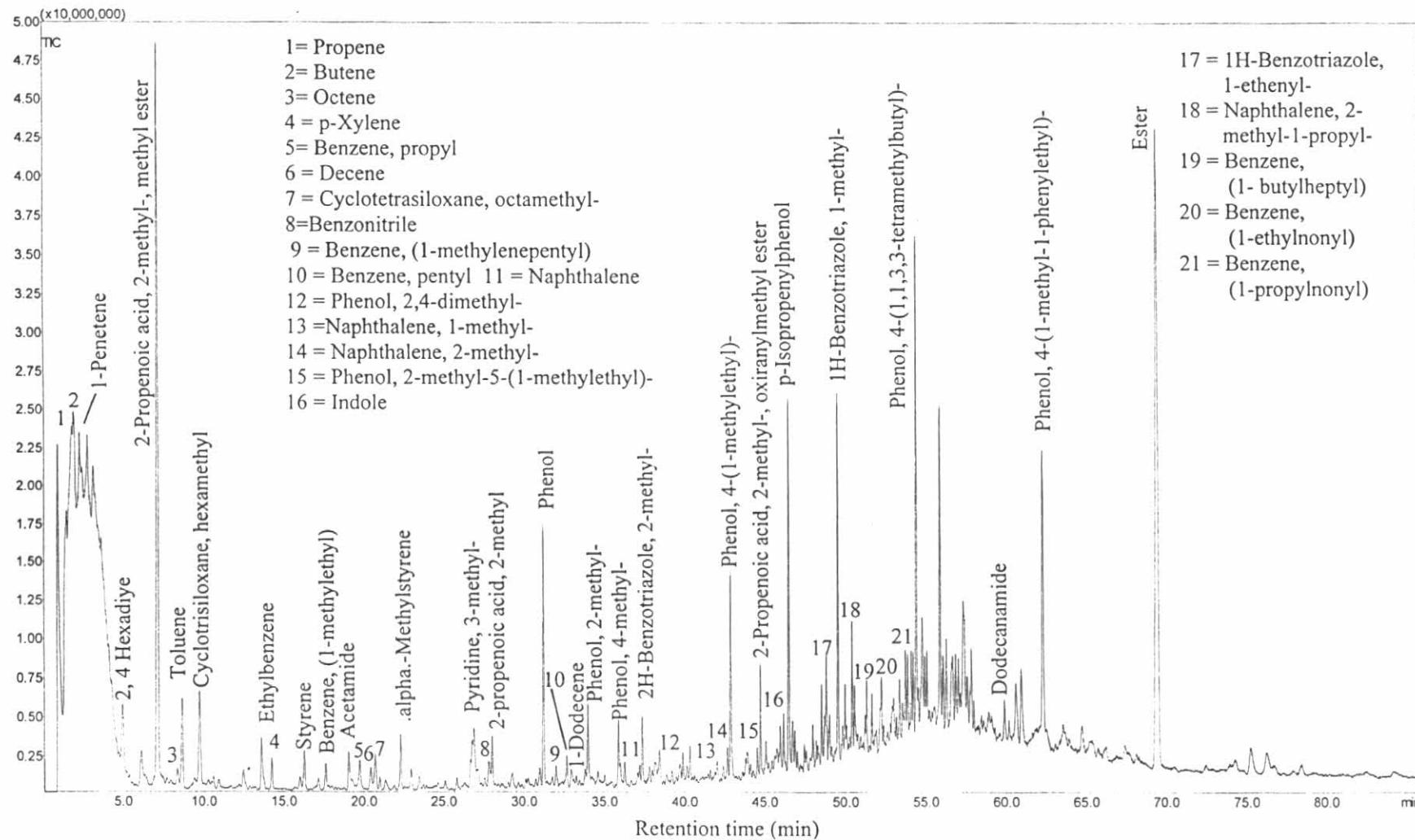


Figure B-11: Pyrochromatogram of major fragments of the HPON of the aeration pond effluent

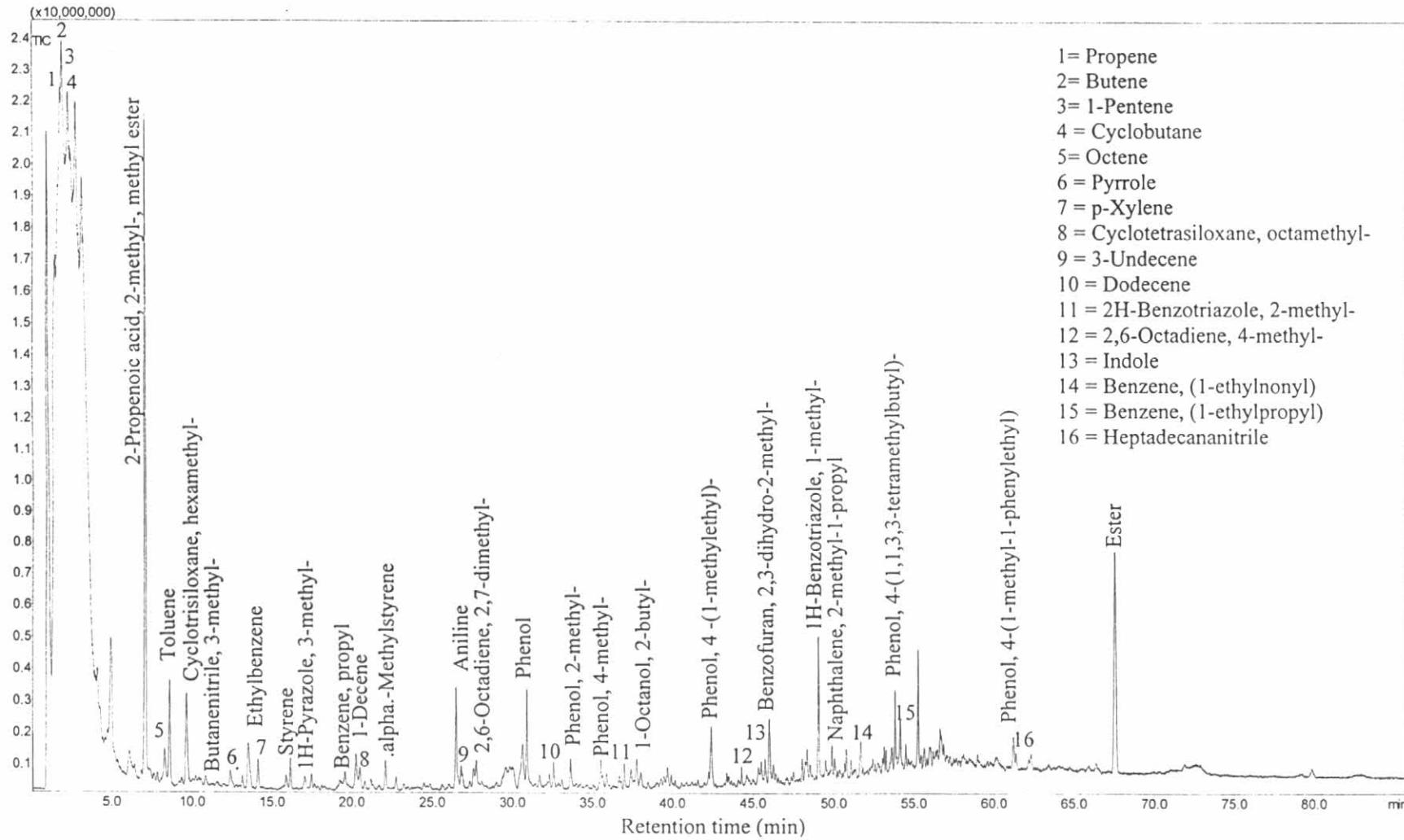


Figure B-12: Pyrochromatogram of major fragments of the HPON of the facultative pond effluent

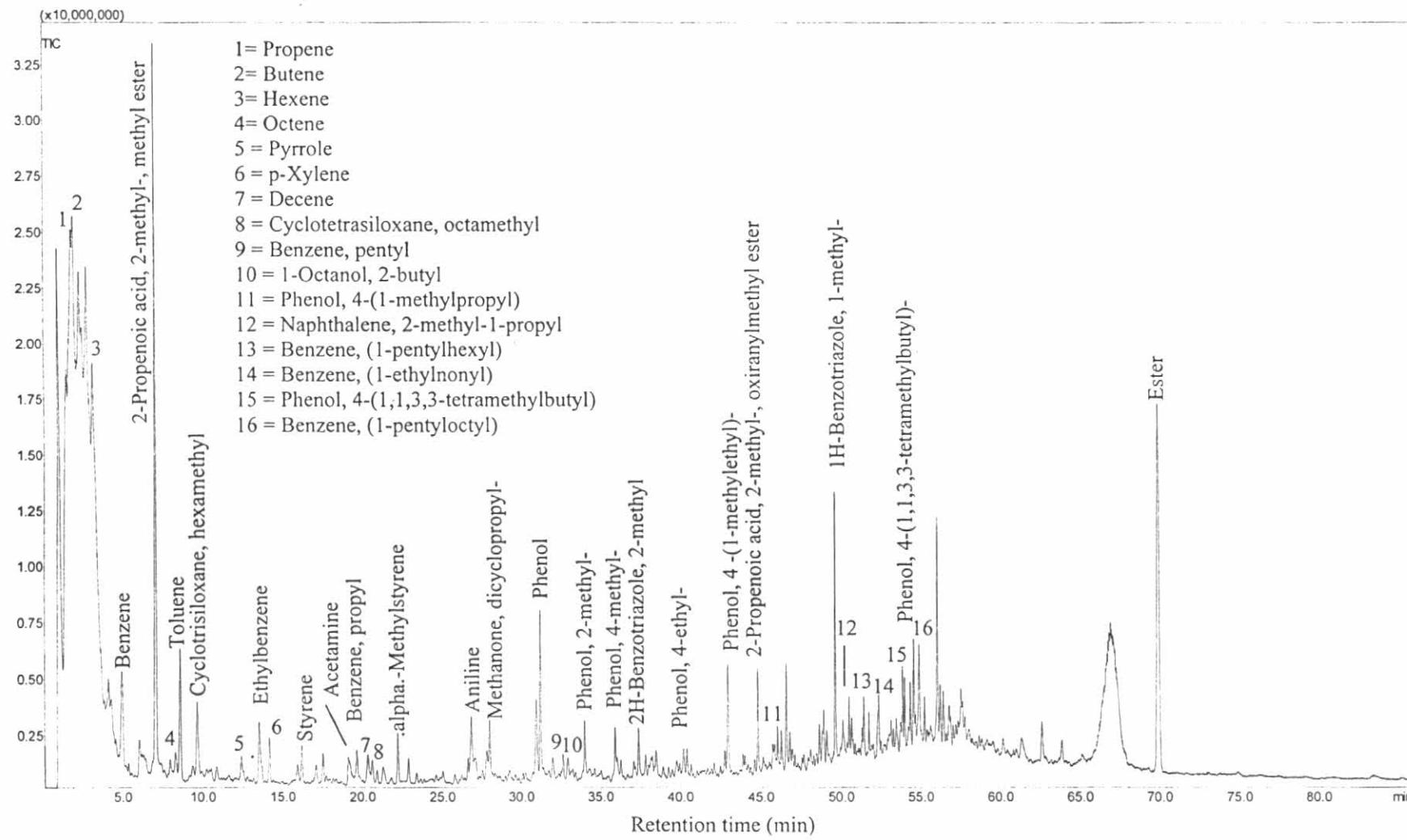


Figure B-13: Pyrochromatogram of major fragments of the HPON of the oxidation pond effluent

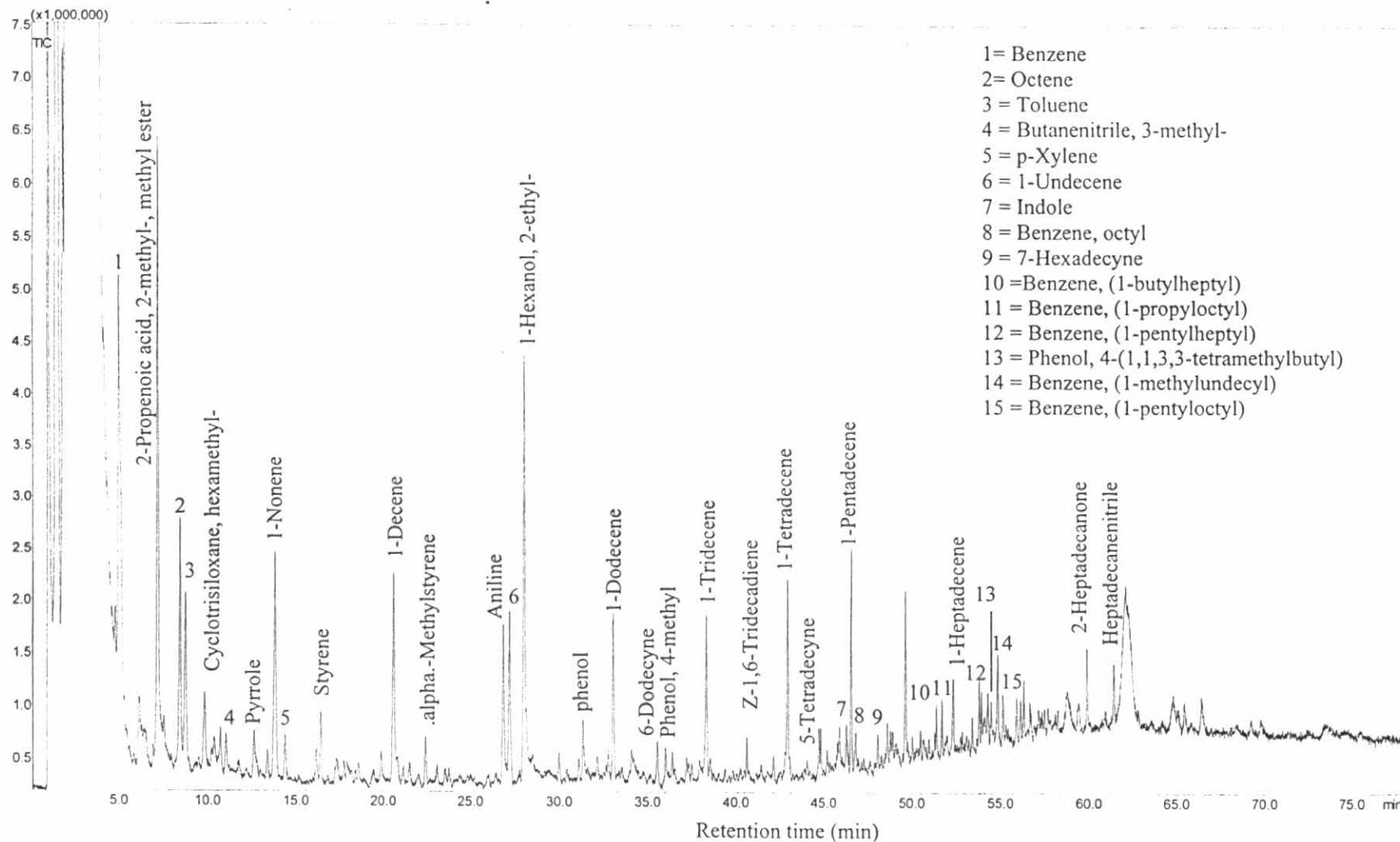


Figure B-14: Pyrochromatogram of major fragments of the HPON of the detention pond effluent

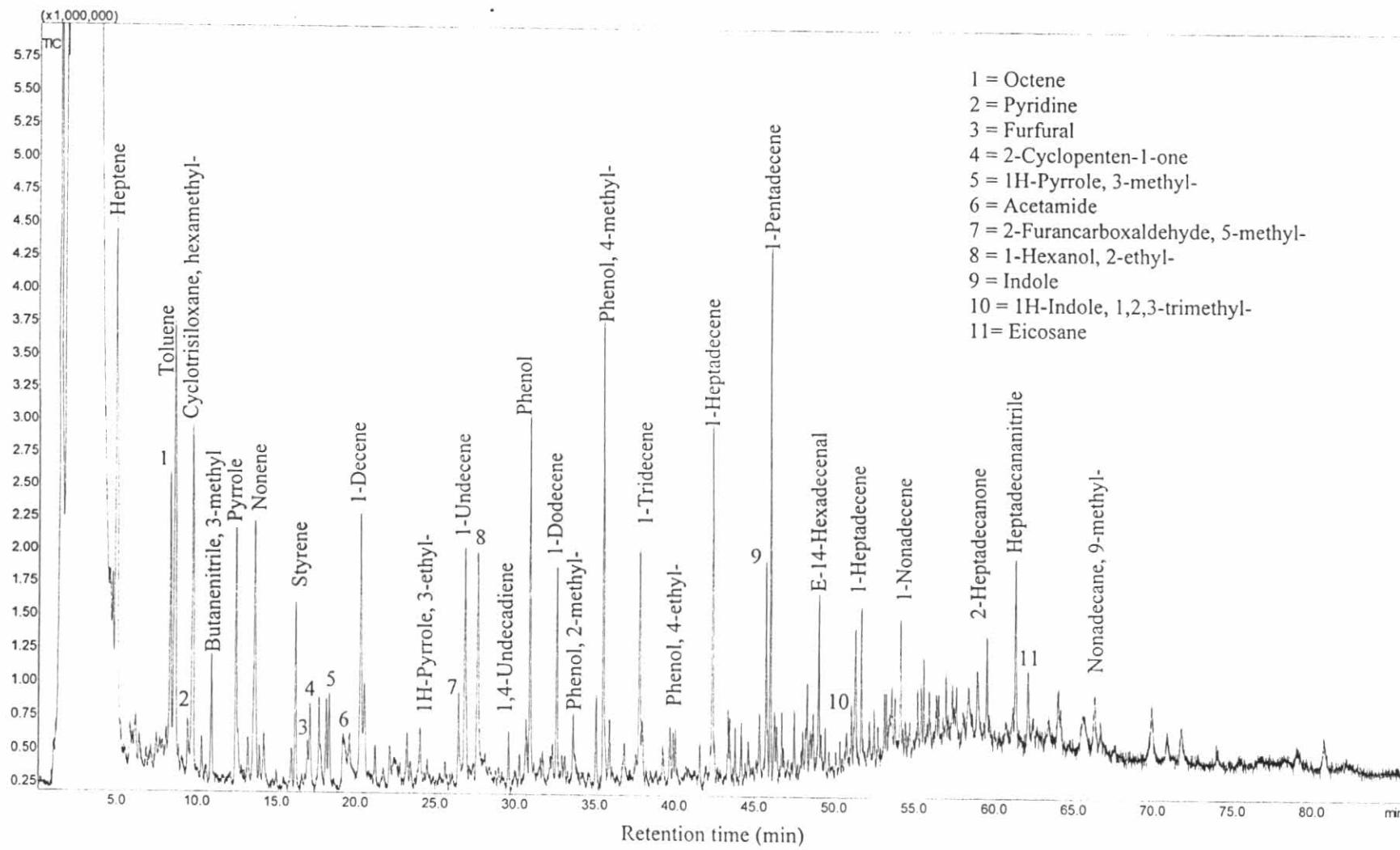


Figure B-15: Pyrochromatogram of major fragments of the HPOB of influent wastewater

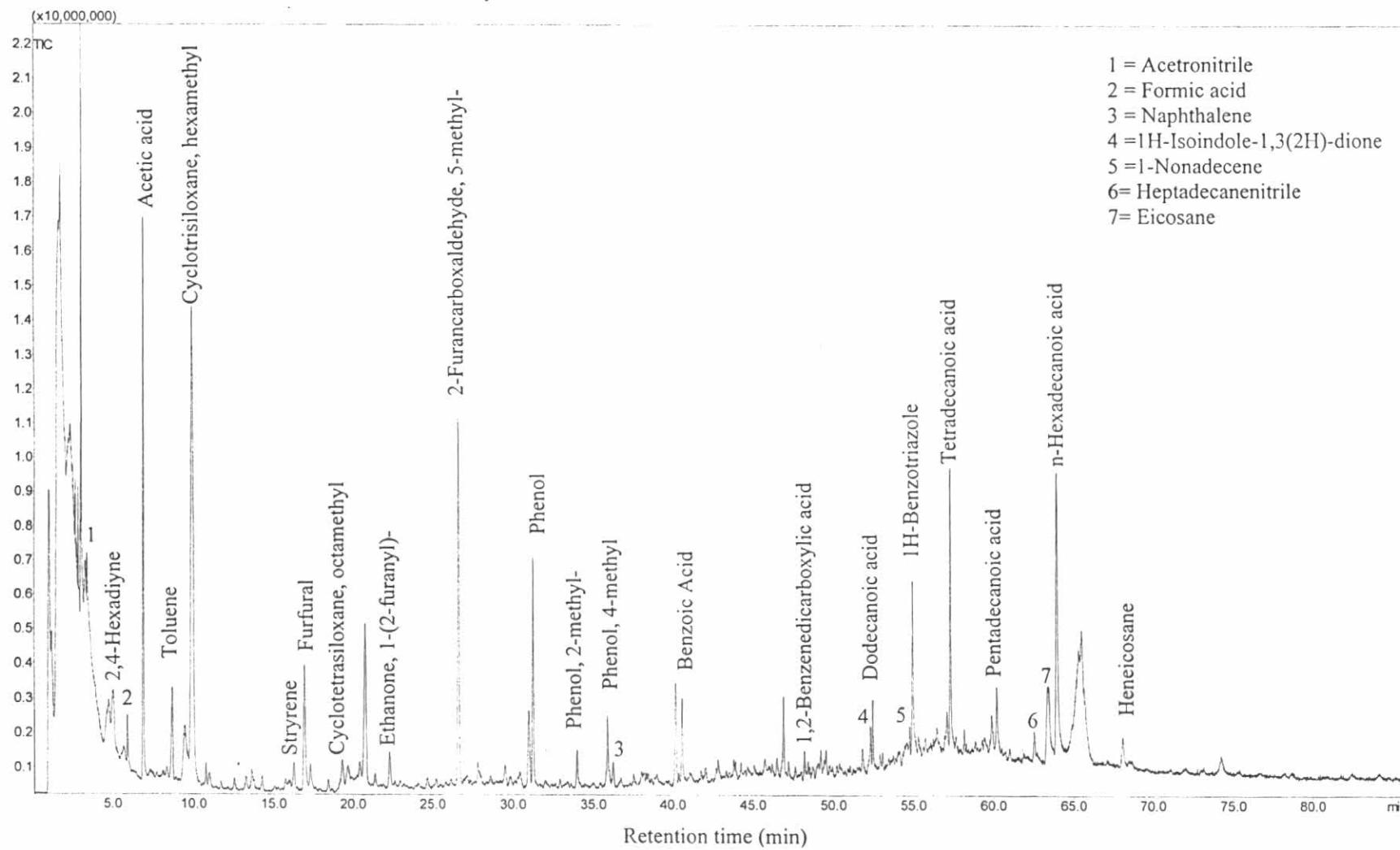


Figure B-16: Pyrochromatogram of major fragments of the HPOB of the aeration pond effluent

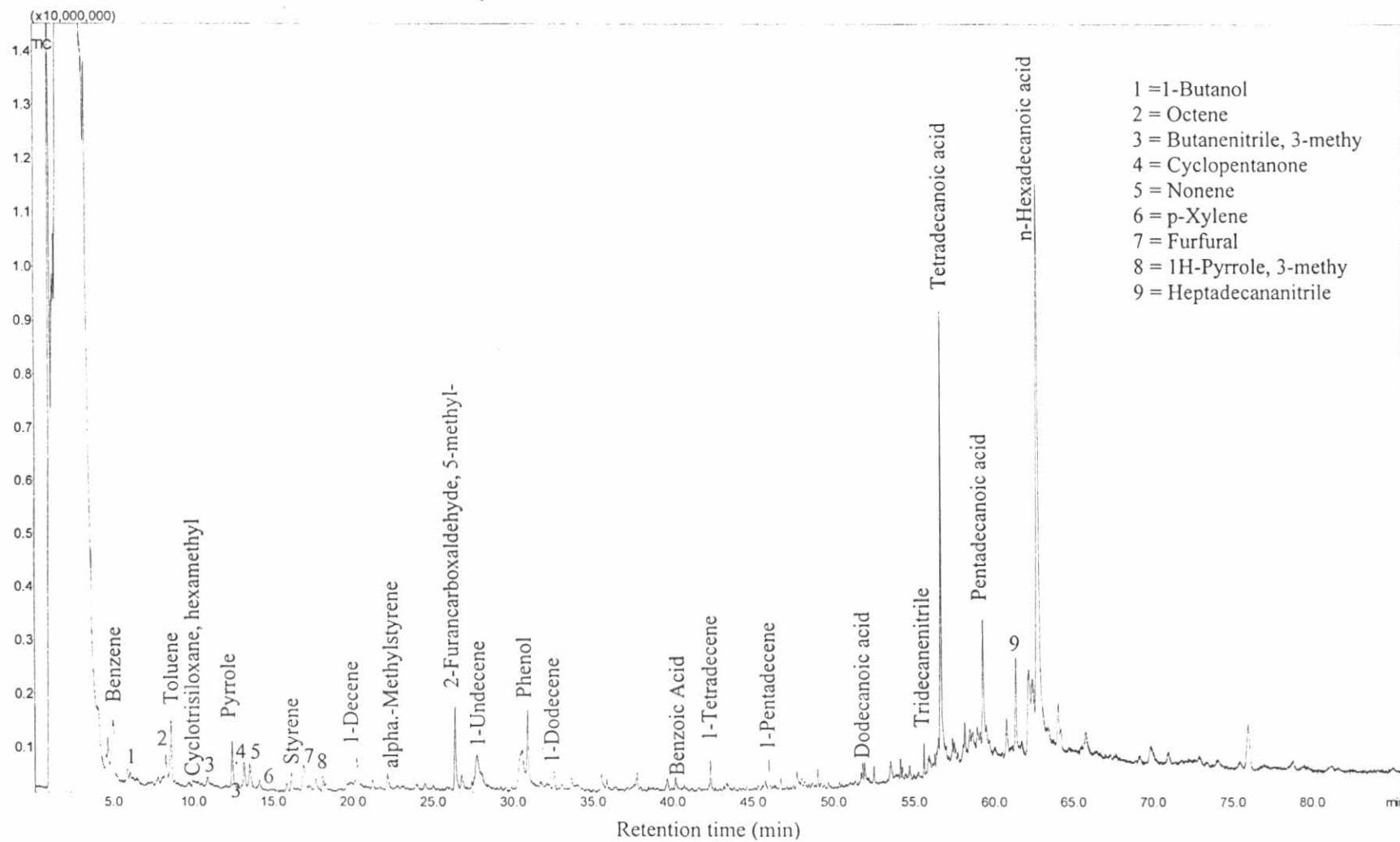


Figure B-17: Pyrochromatogram of major fragments of the HPOB of the facultative pond effluent

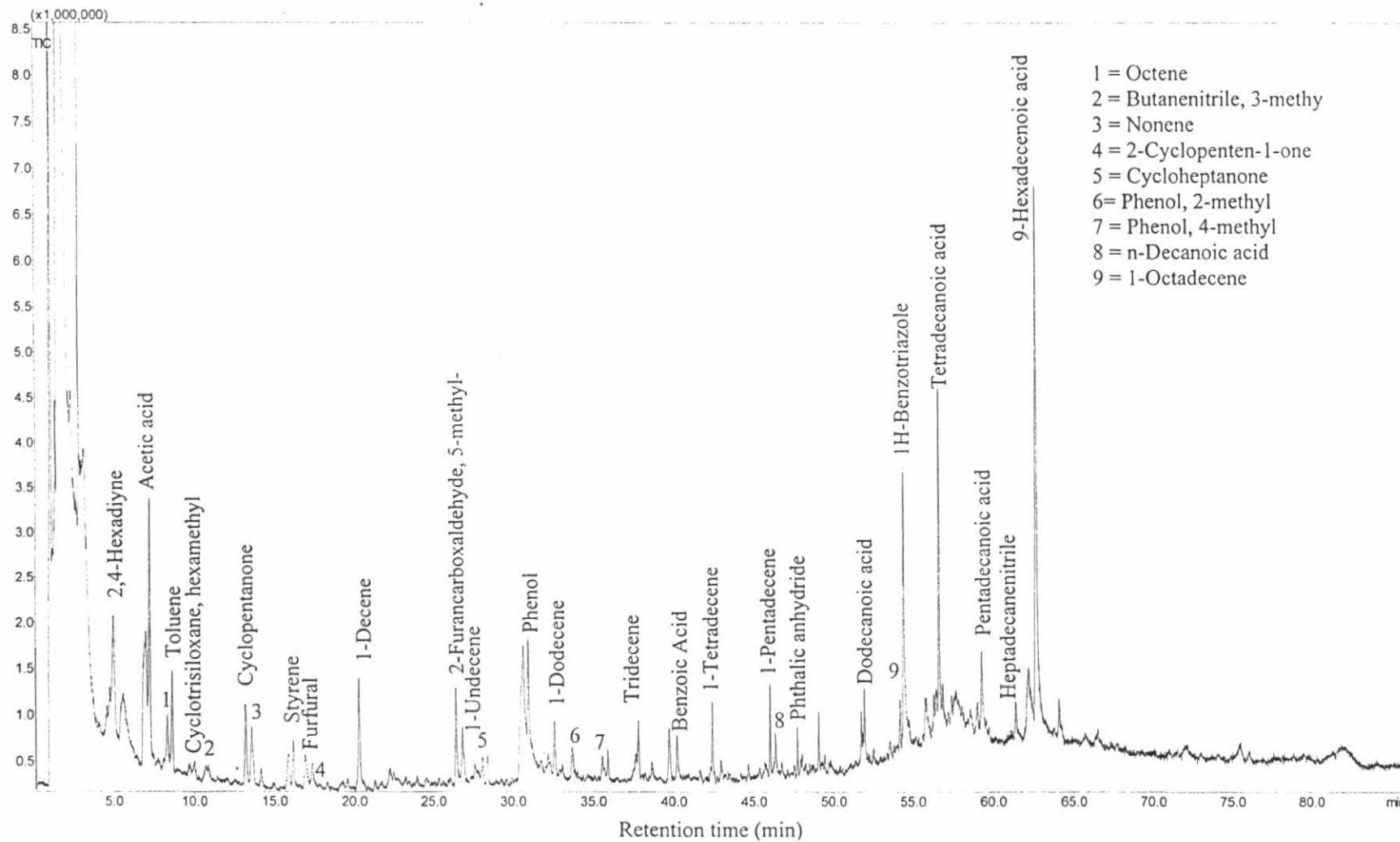


Figure B-18: Pyrochromatogram of major fragments of the HPOB of the oxidation pond effluent

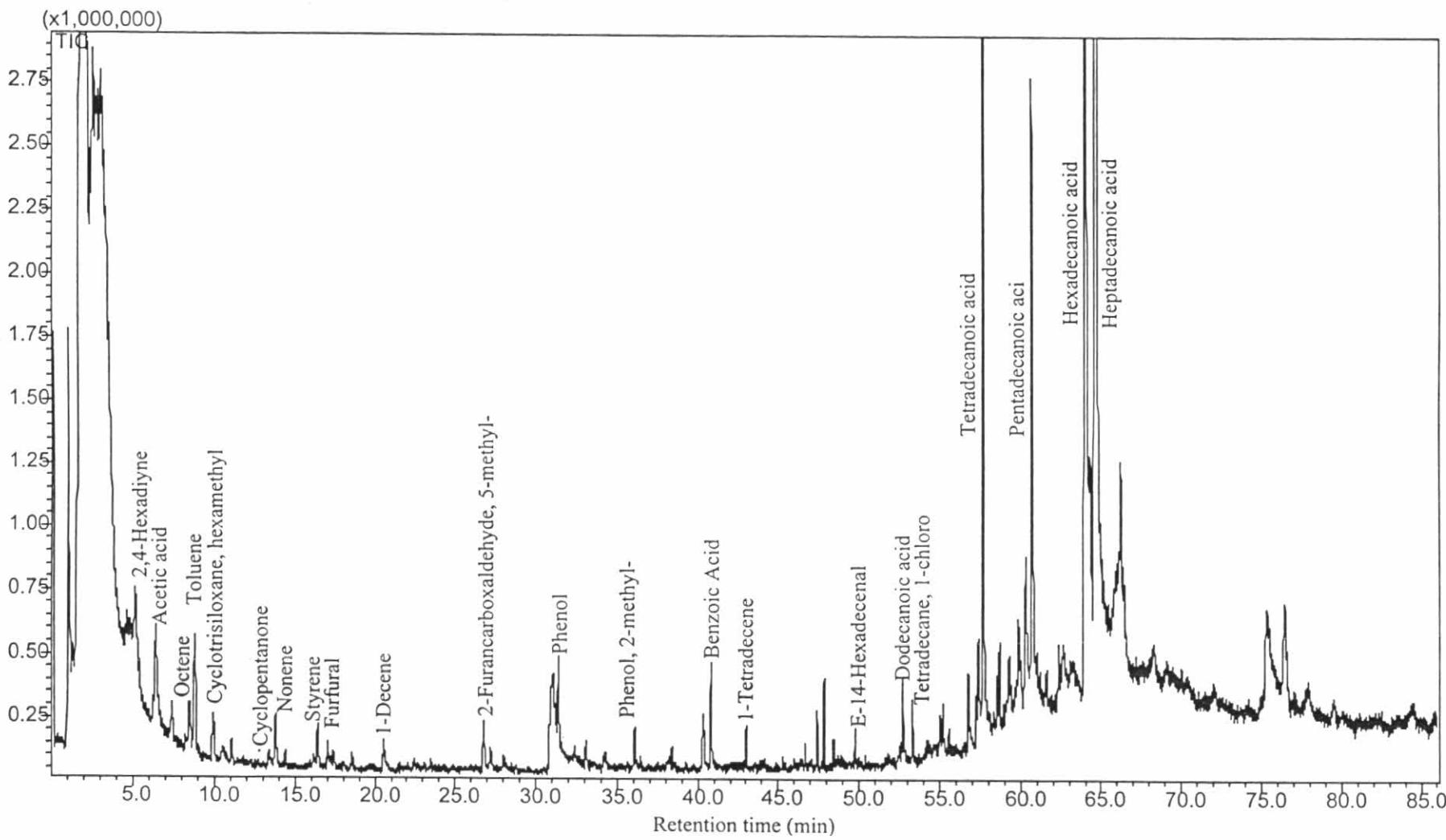


Figure B-19: Pyrochromatogram of major fragments of the HPOB of the detention pond effluent

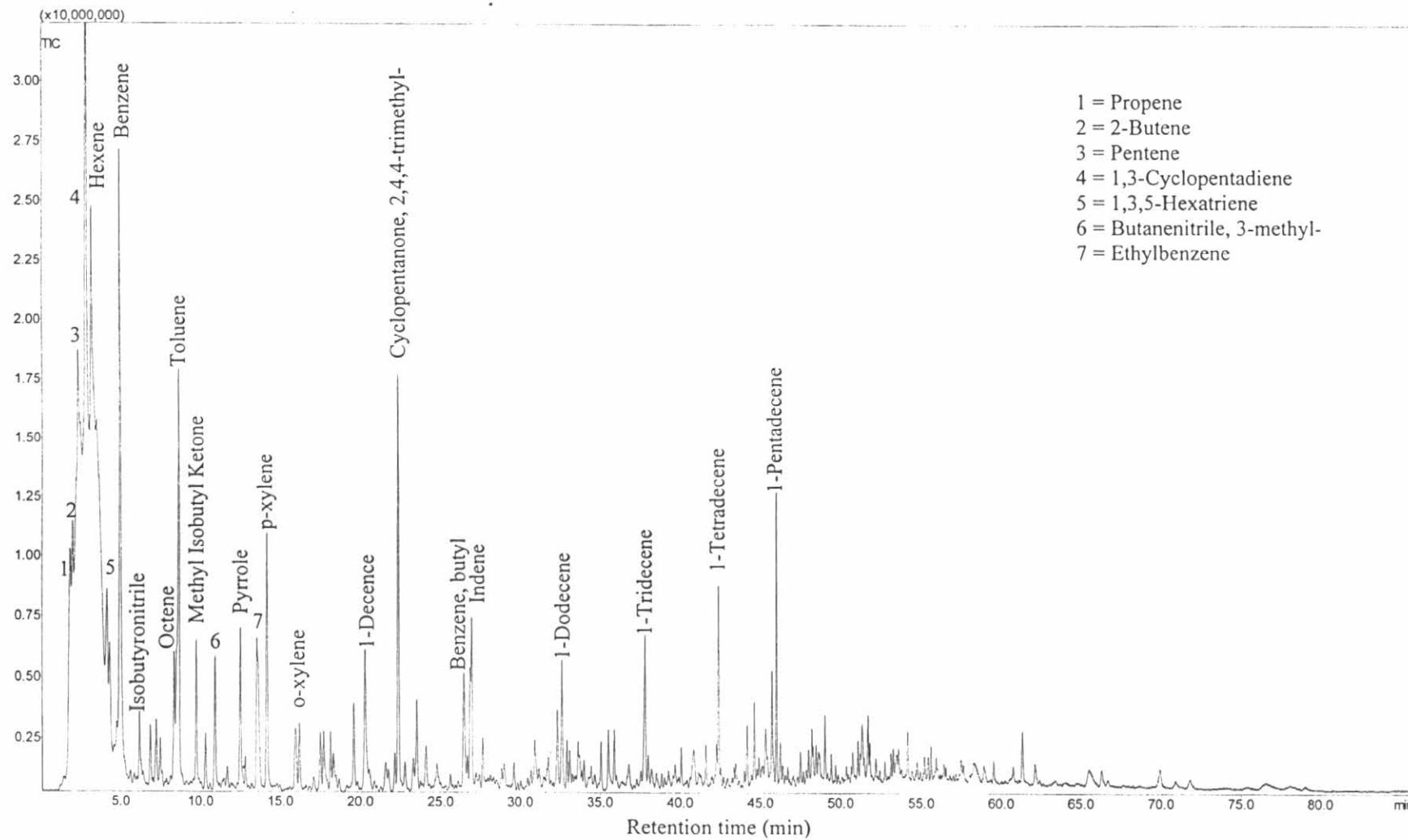


Figure B-20: Pyrochromatogram of major fragments of the HPOA of influent wastewater

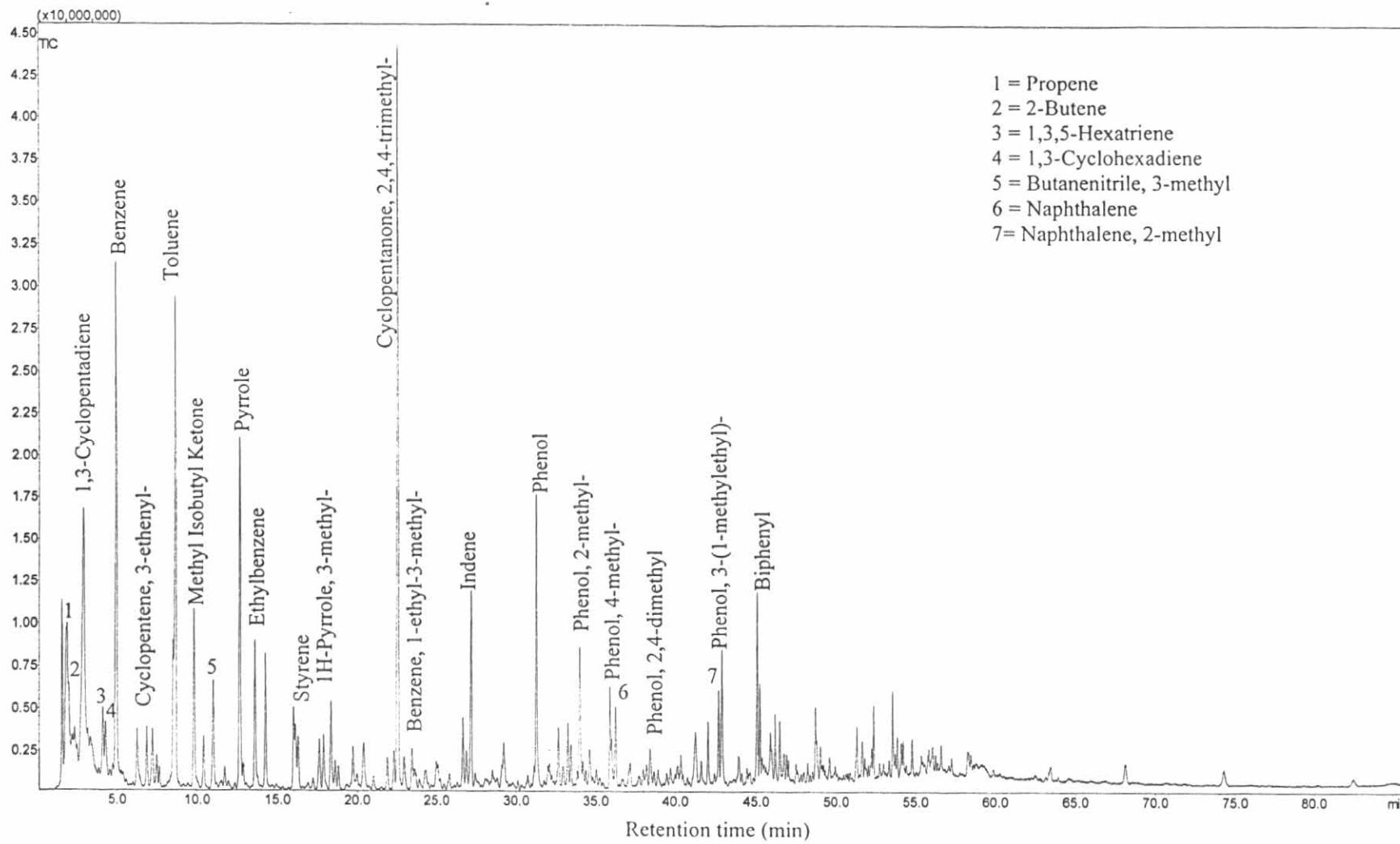


Figure B-21: Pyrochromatogram of major fragments of the HPOA of the aeration pond effluent

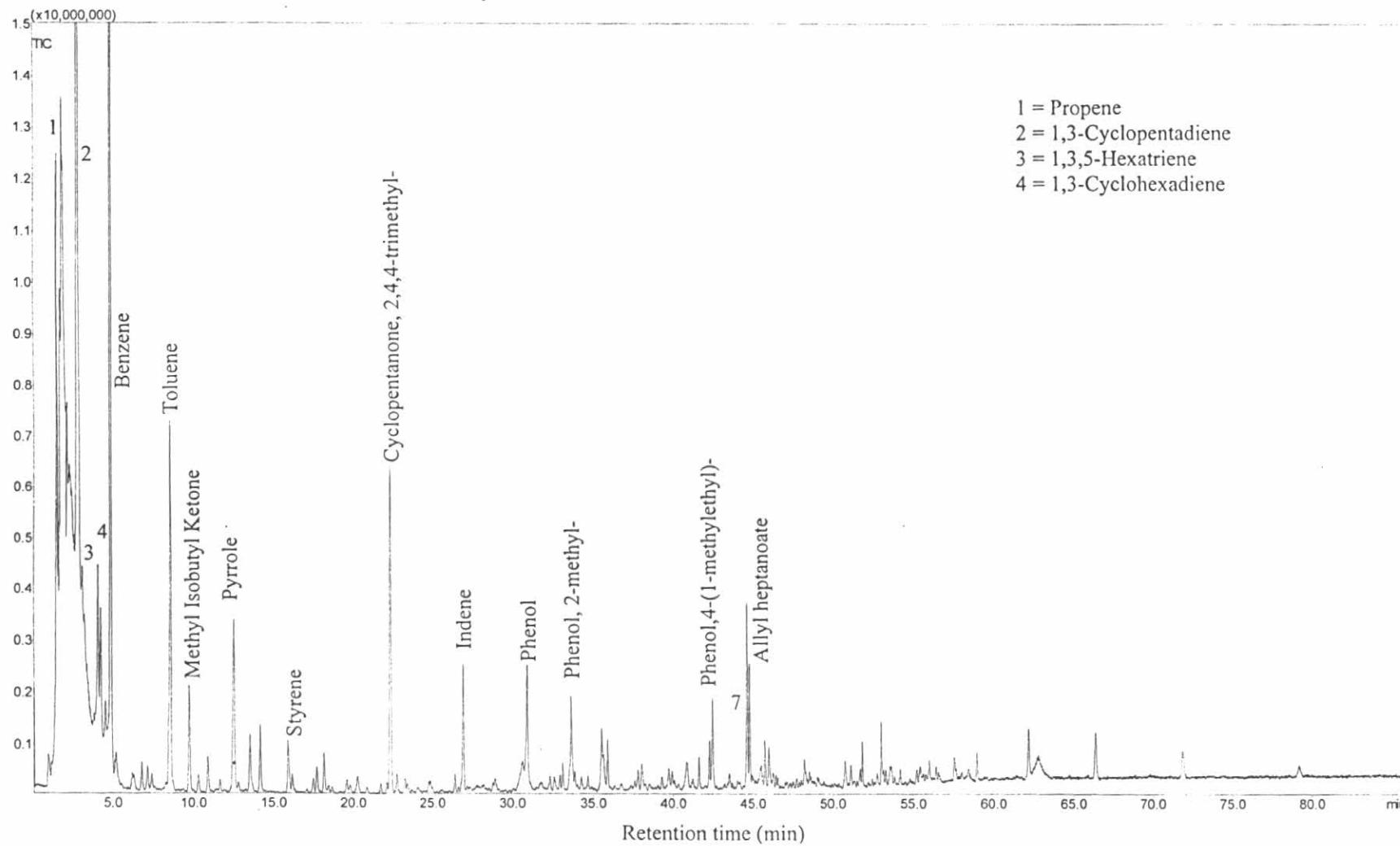


Figure B-22: Pyrochromatogram of major fragments of the HPOA of the facultative pond effluent

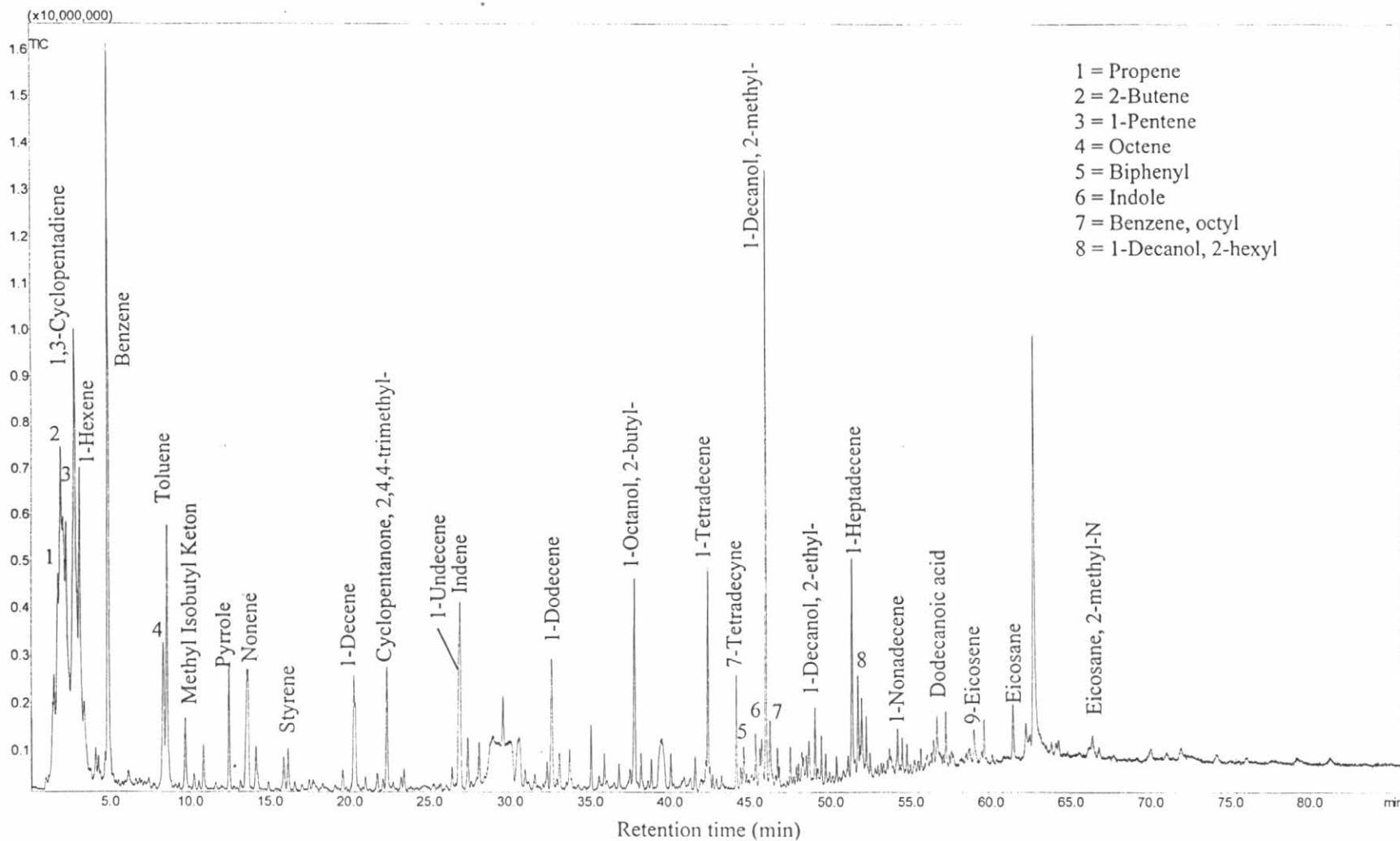


Figure B-23: Pyrochromatogram of major fragments of the HPOA of the oxidation pond effluent

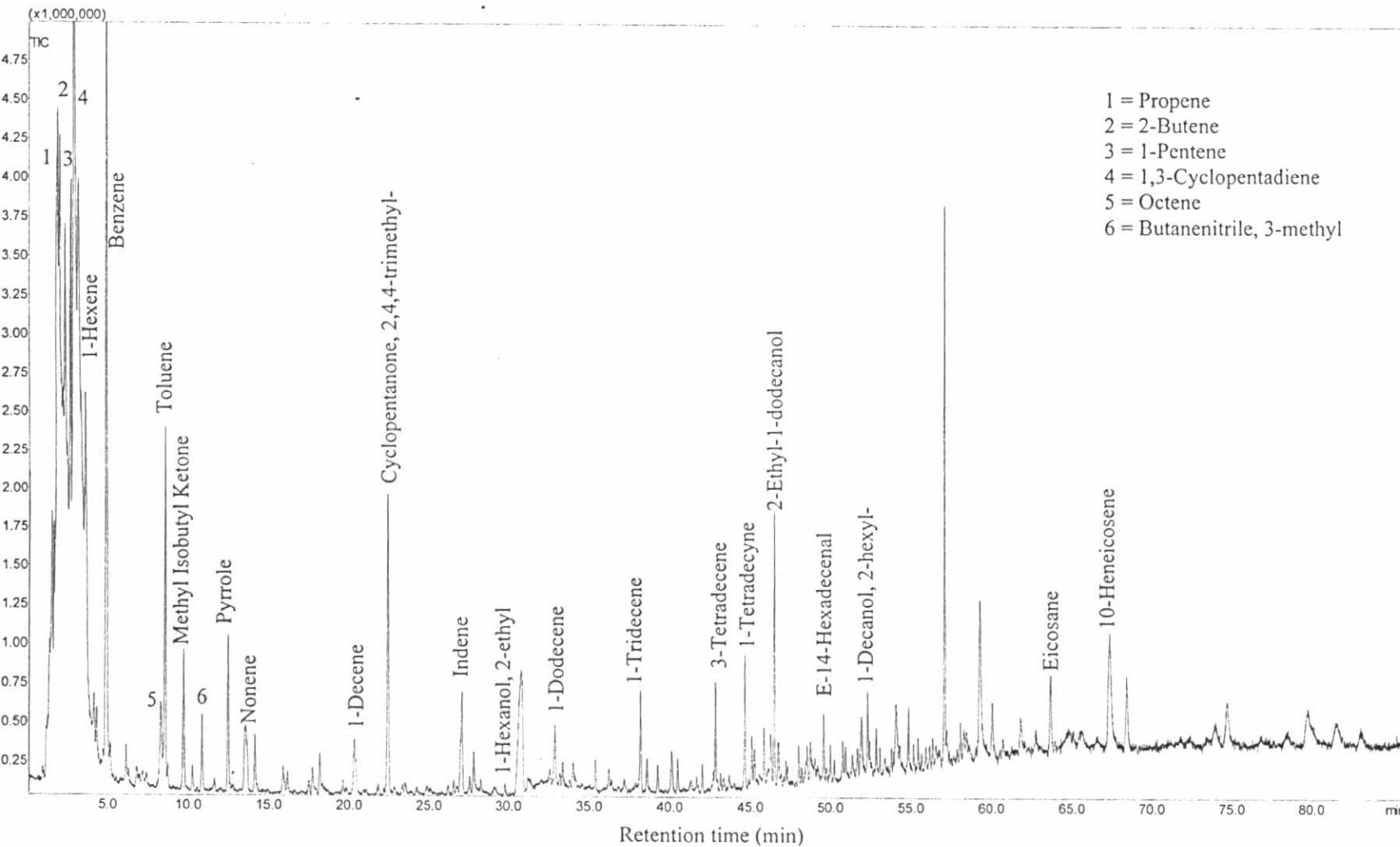


Figure B-24: Pyrochromatogram of major fragments of the HPOA of the detention pond effluent

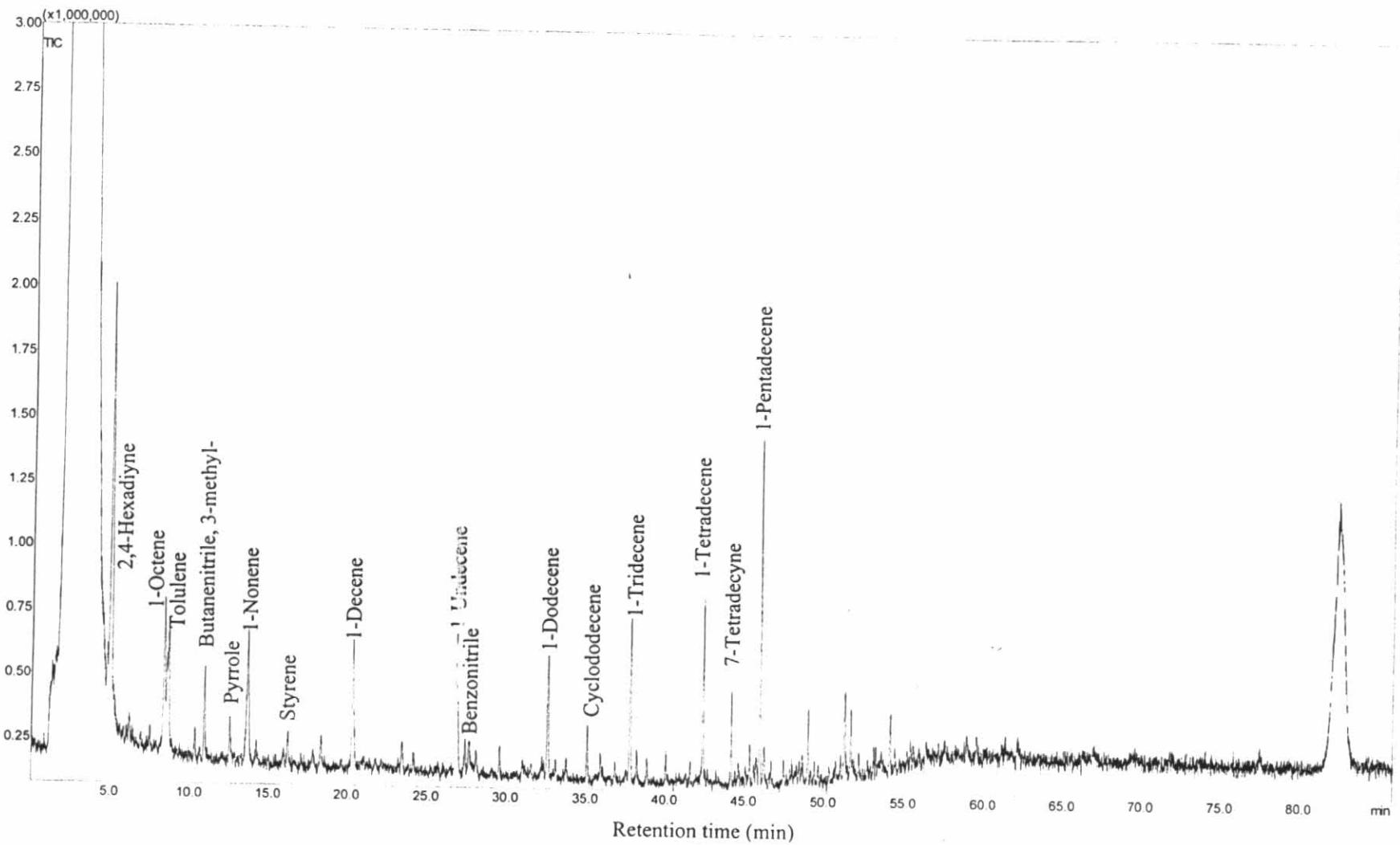


Figure B-25: Pyrochromatogram of major fragments of the HPIB of influent wastewater

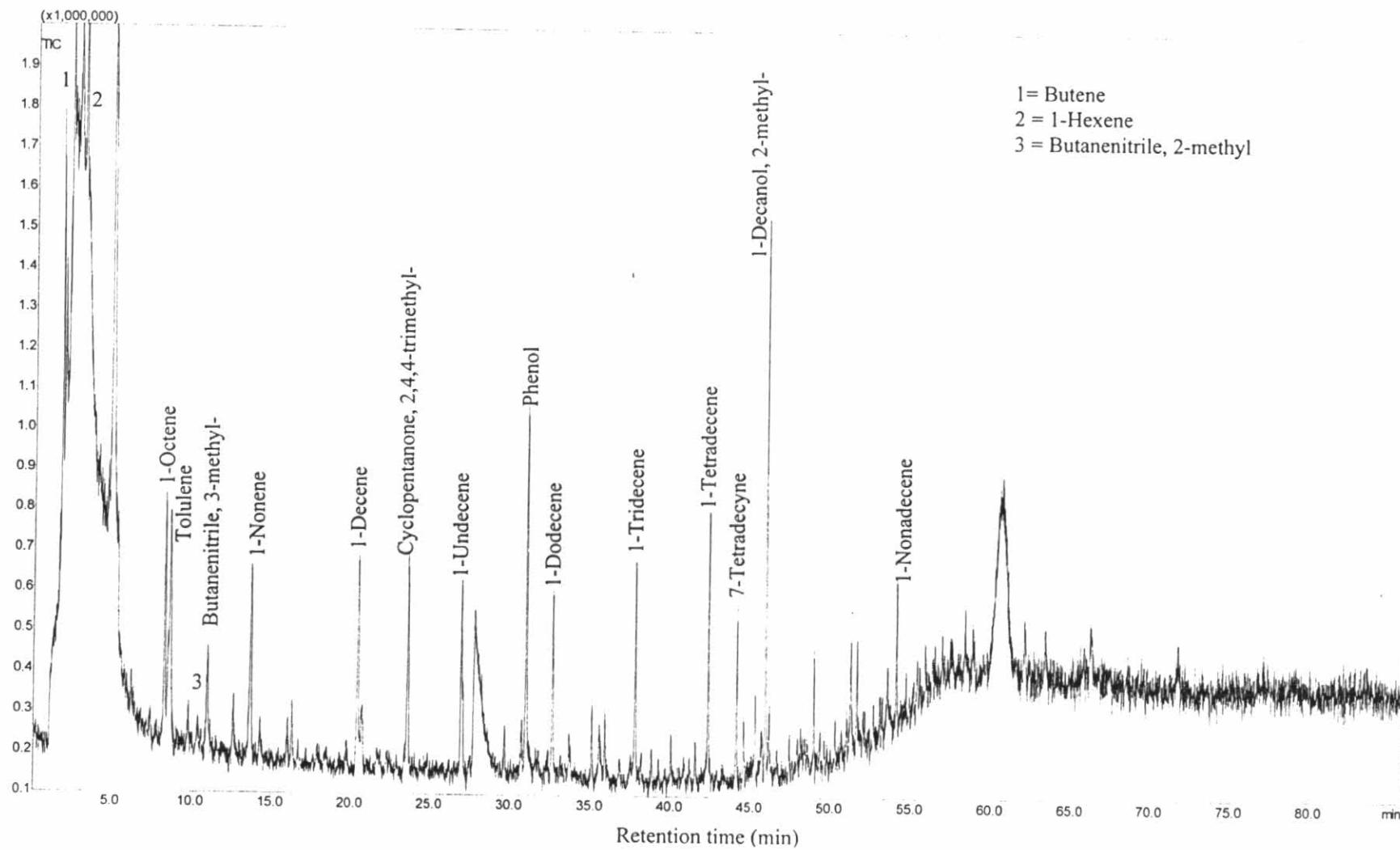


Figure B-26: Pyrochromatogram of major fragments of the HPIB of the aeration pond effluent

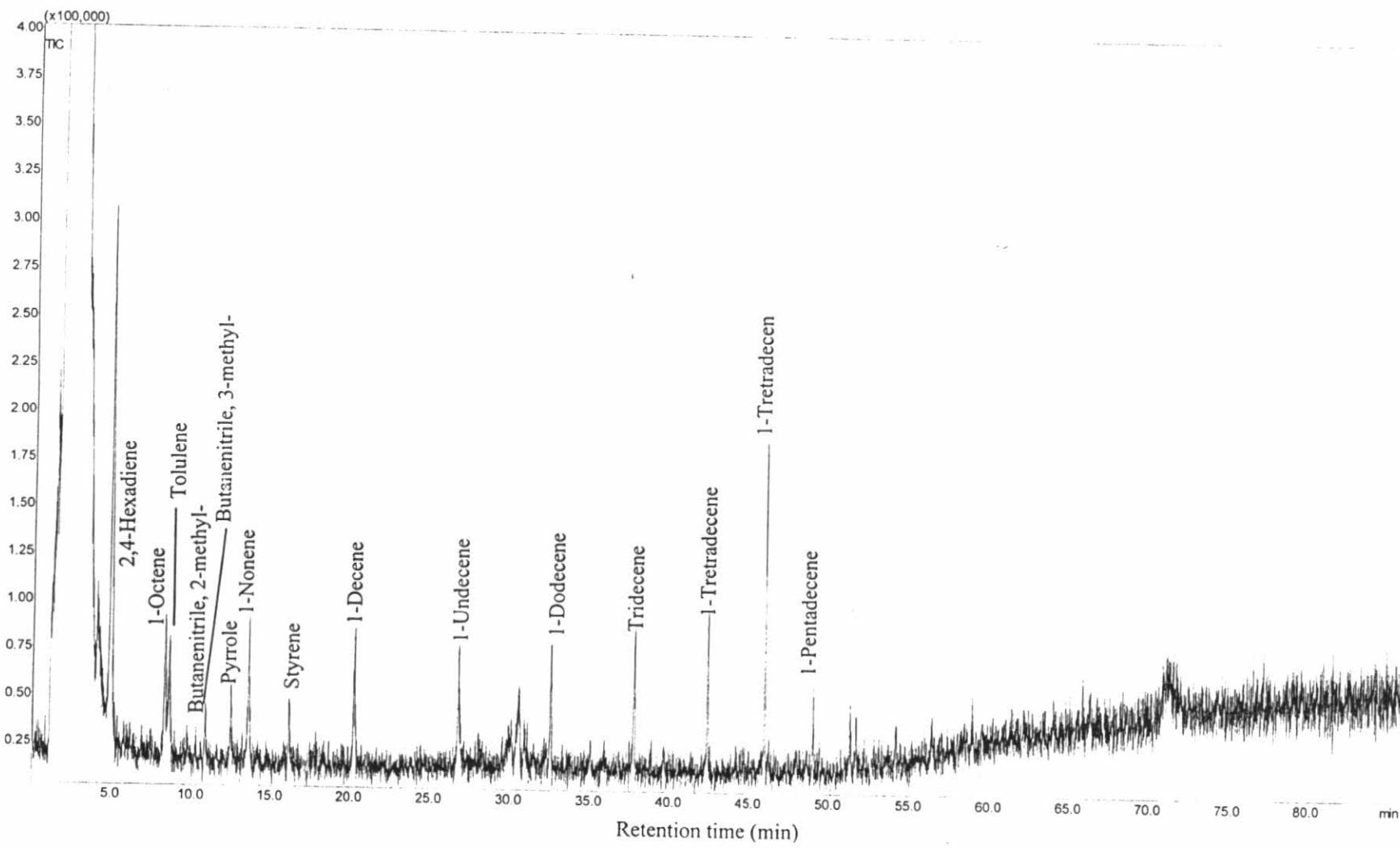


Figure B-27: Pyrochromatogram of major fragments of the HPIB of the facultative pond effluent

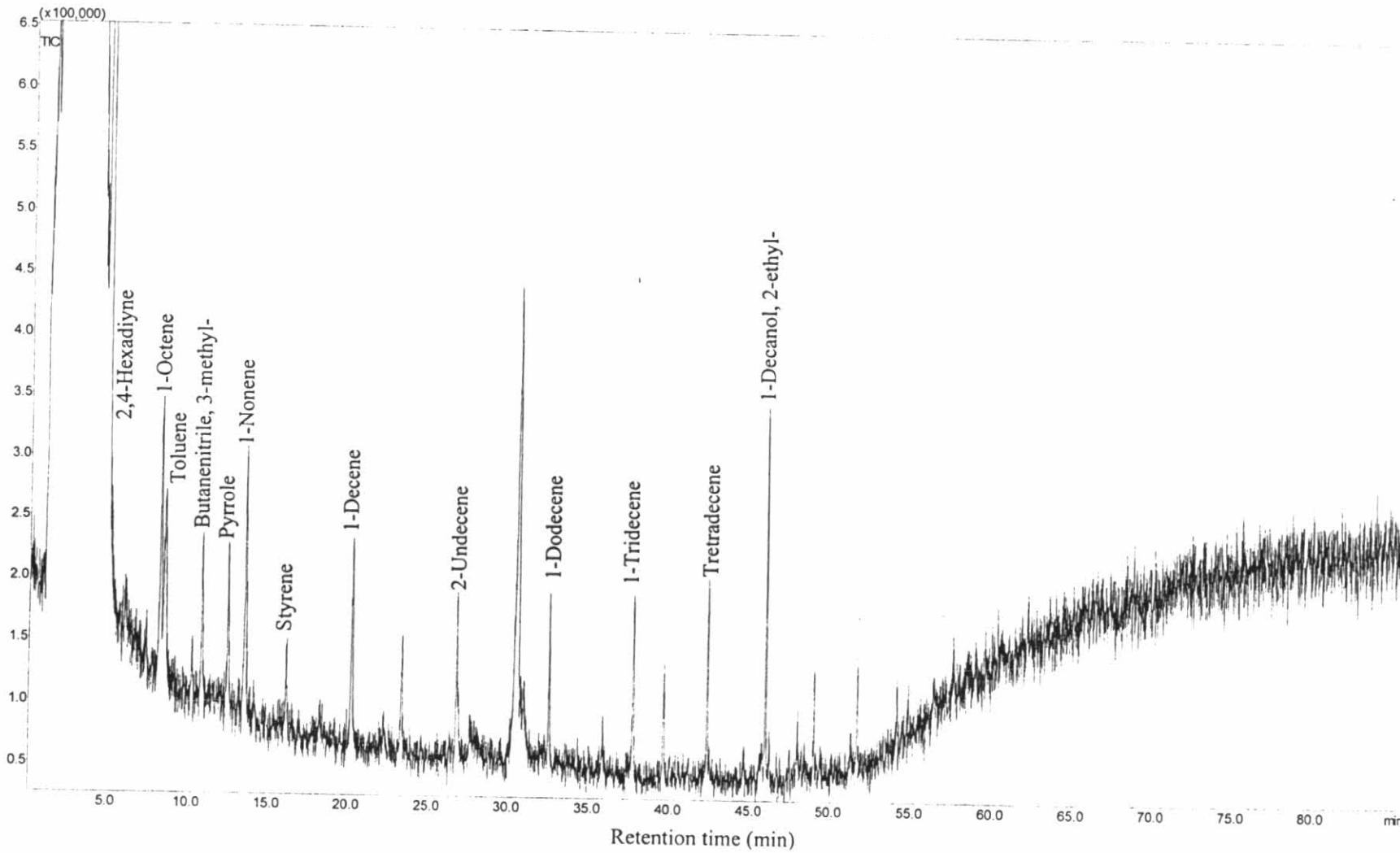


Figure B-28: Pyrochromatogram of major fragments of the HPIB of the oxidation pond effluent

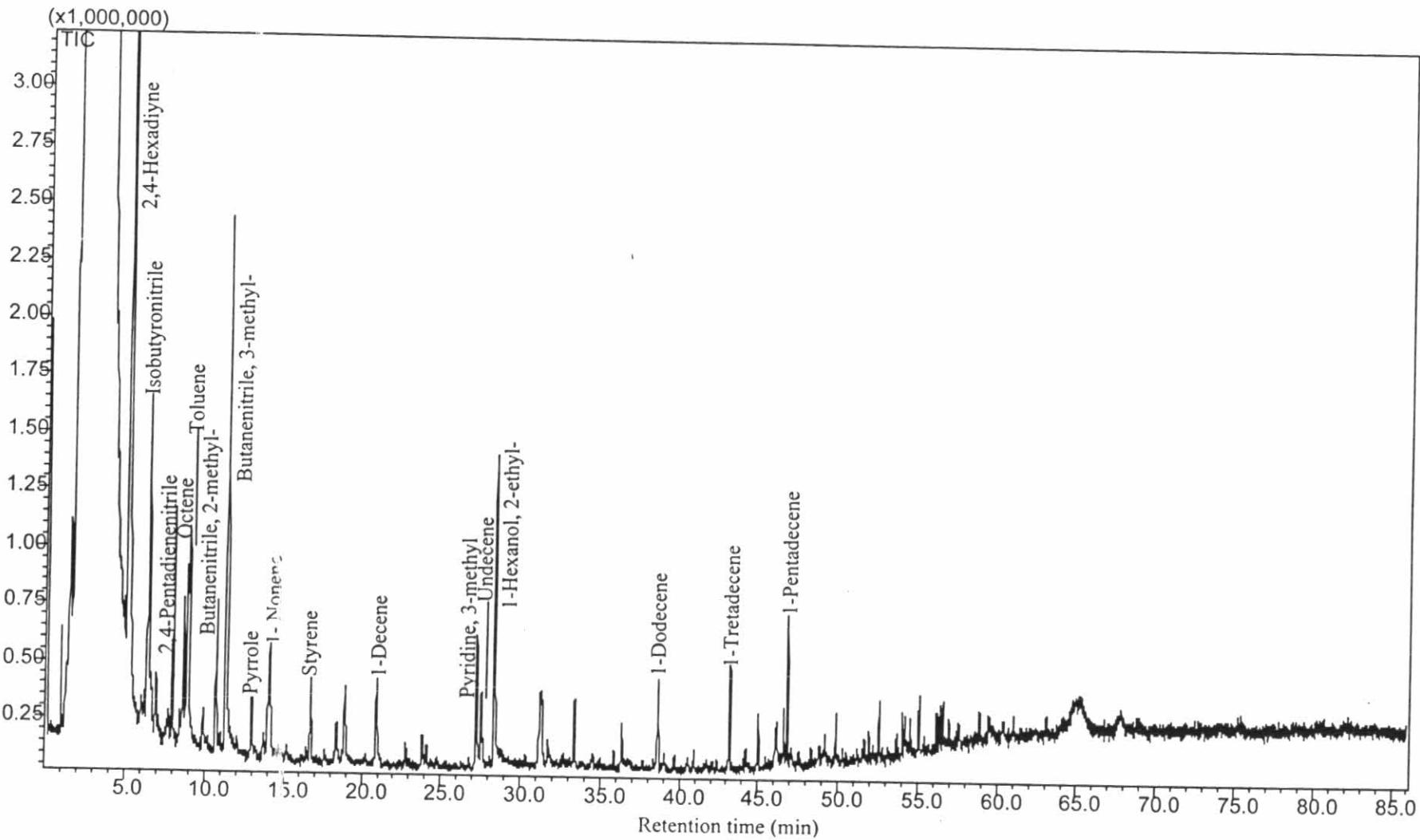


Figure B-29: Pyrochromatogram of major fragments of the HPIB of the detention pond effluent

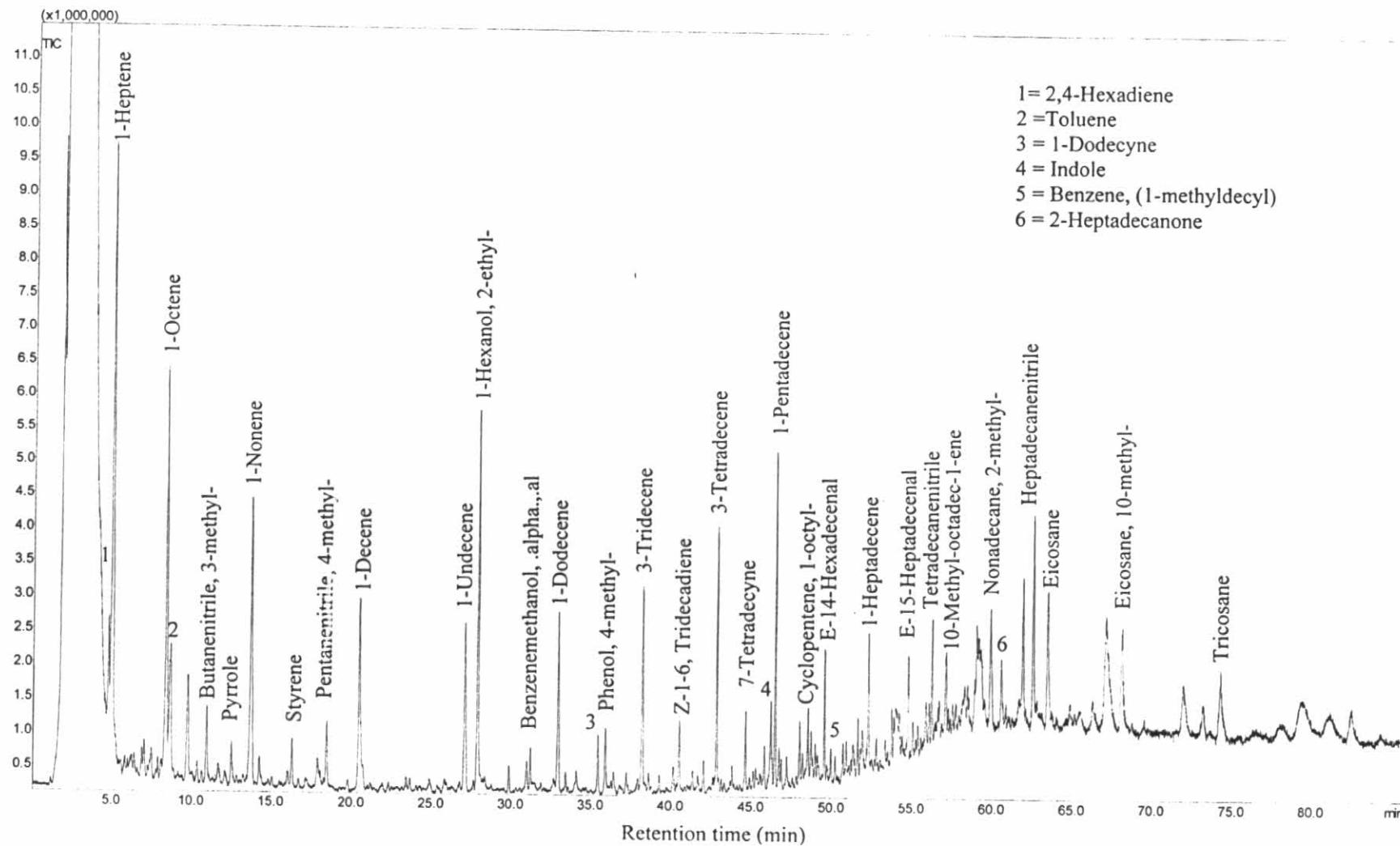


Figure B-30: Pyrochromatogram of major fragments of the HPIA of influent wastewater

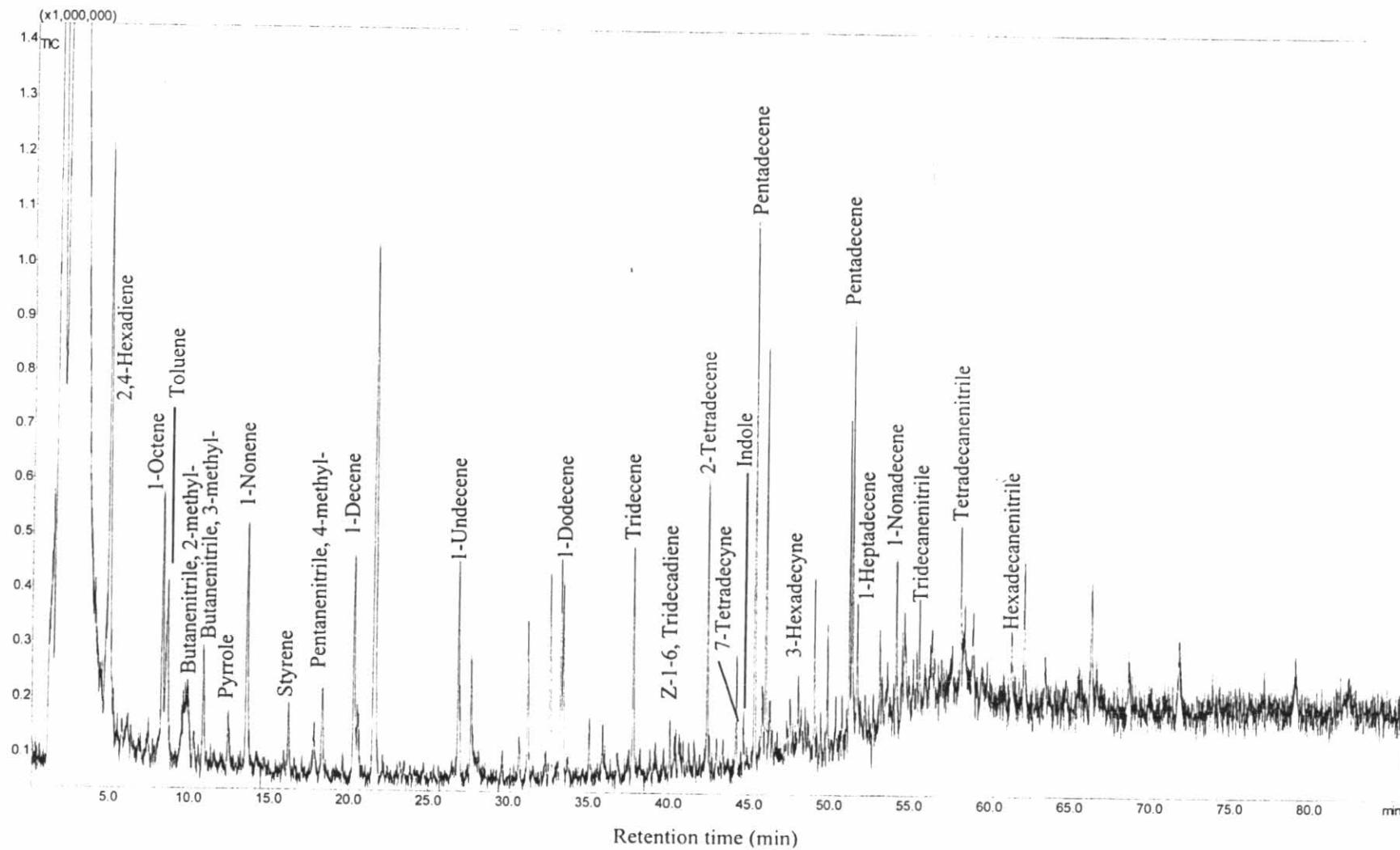


Figure B-31: Pyrochromatogram of major fragments of the HPIA of the aeration pond effluent

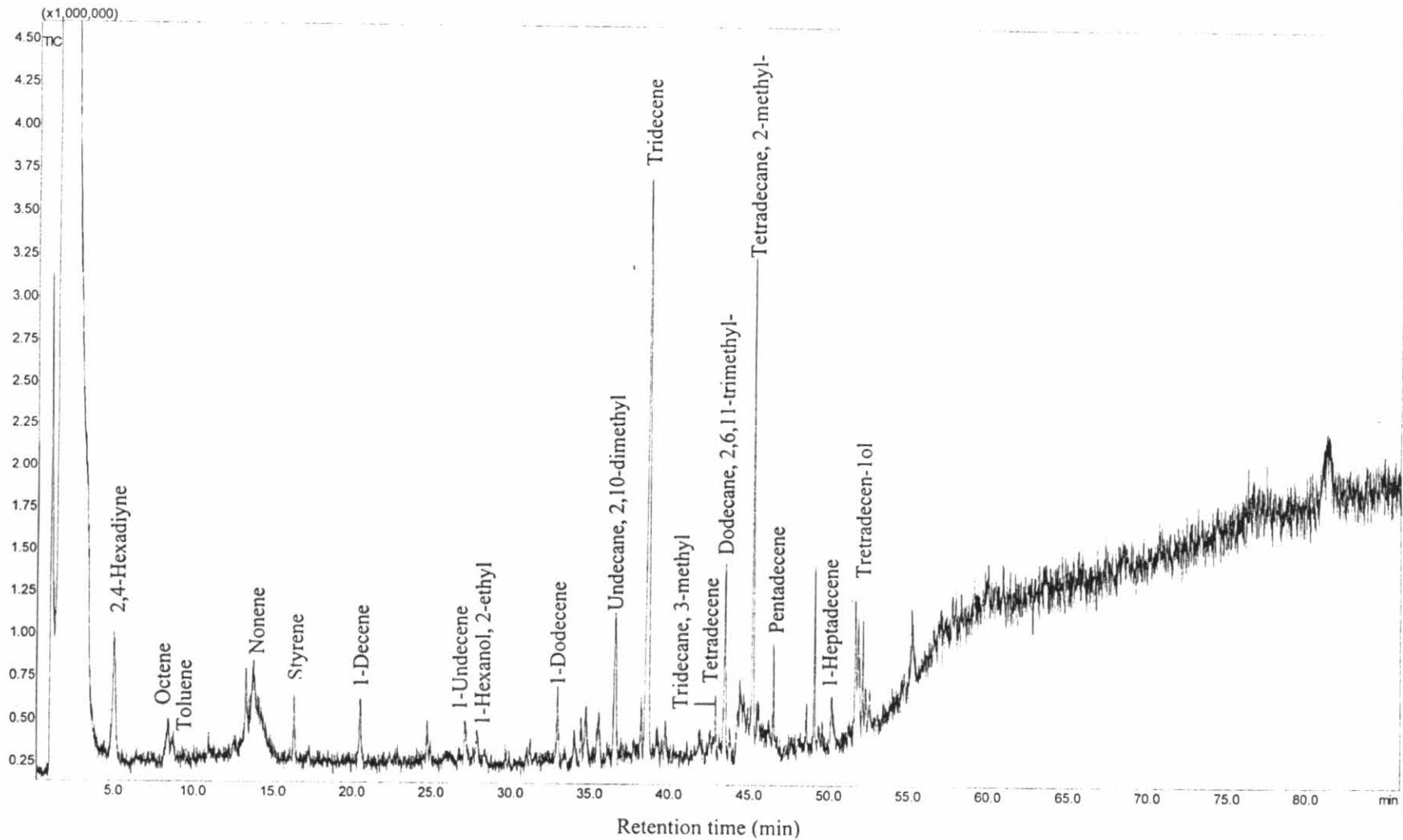


Figure B-32: Pyrochromatogram of major fragments of the HPIA of the facultative pond effluent

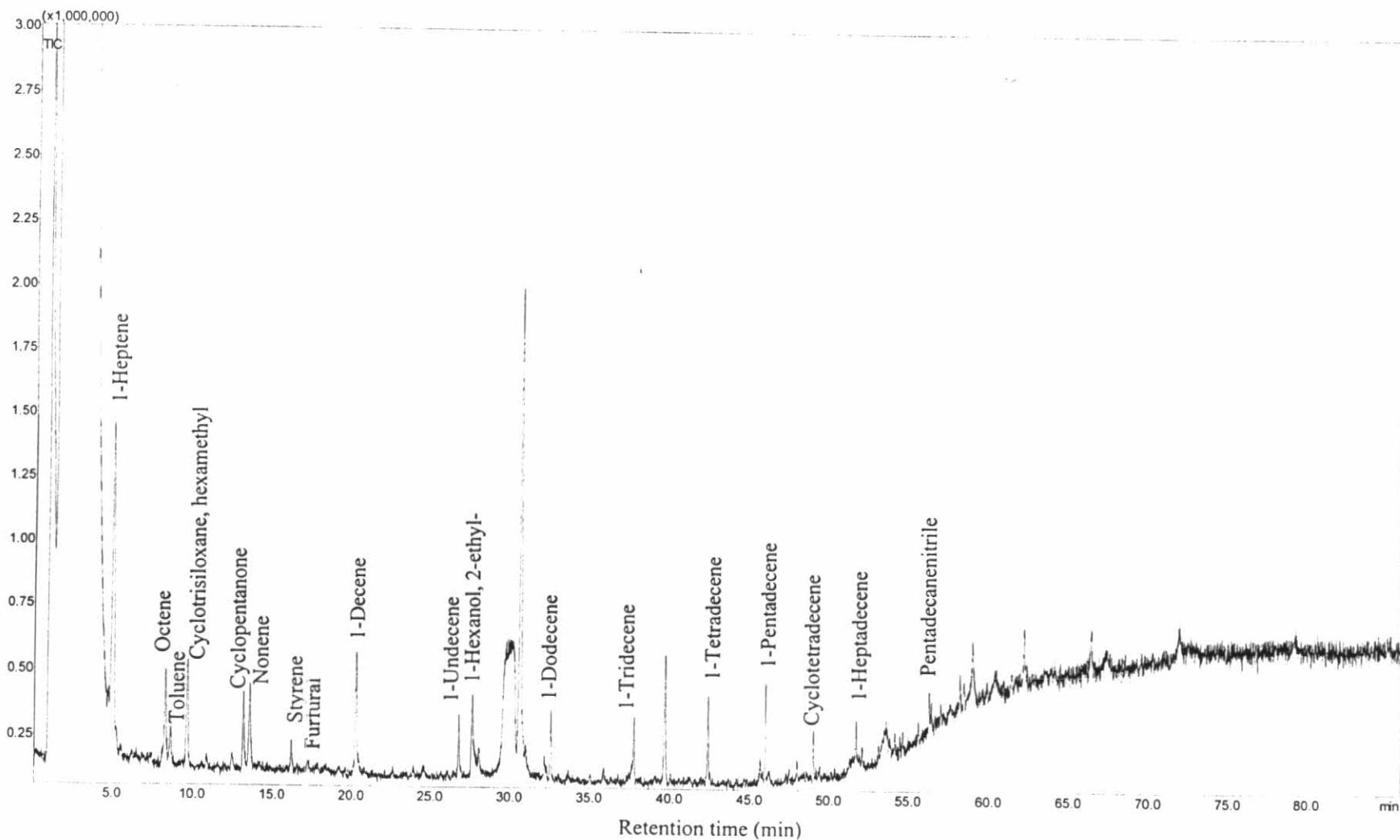


Figure B-33: Pyrochromatogram of major fragments of the HPIA of the oxidation pond effluent

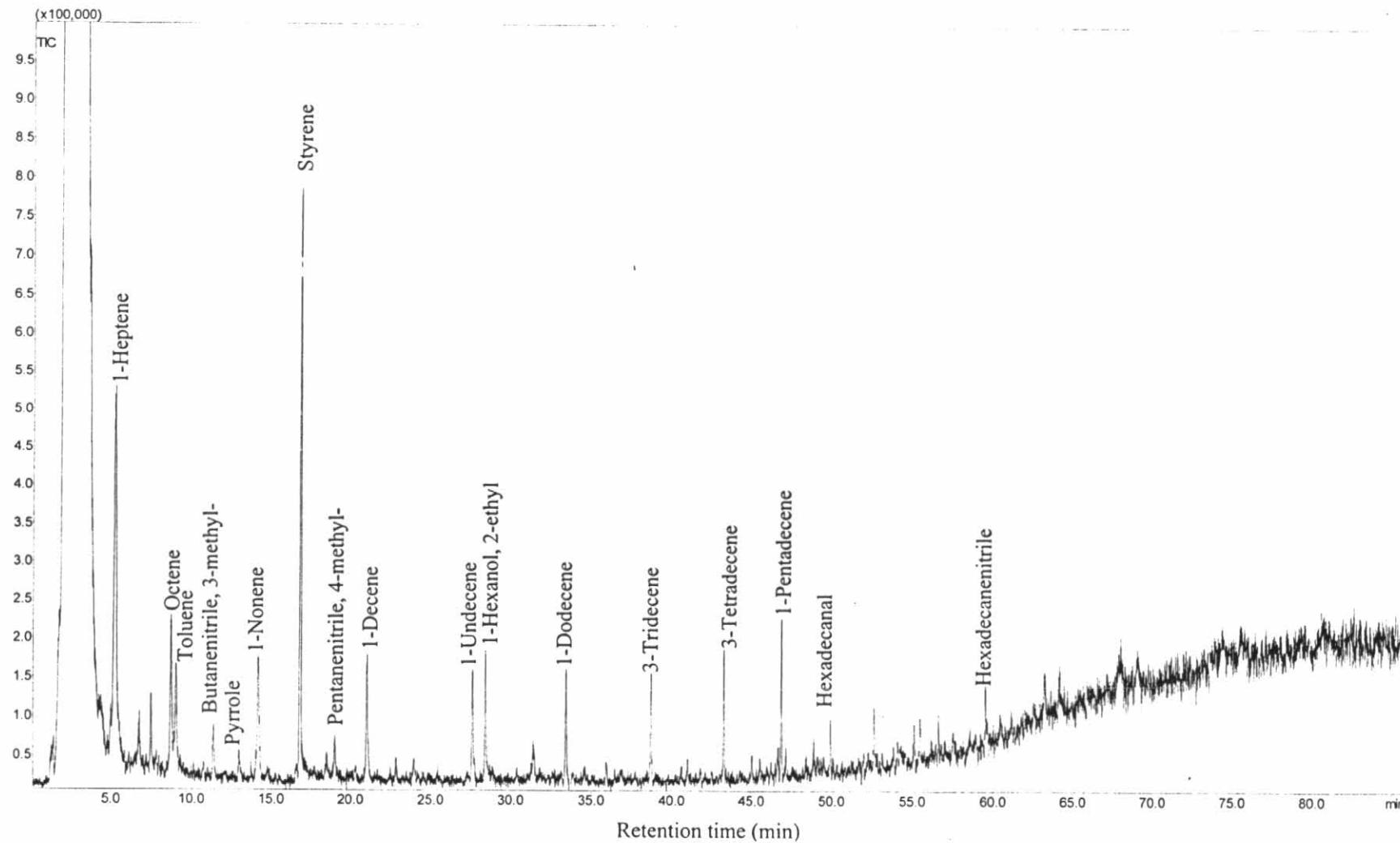


Figure B-34: Pyrochromatogram of major fragments of the HPIA of the detention pond effluent

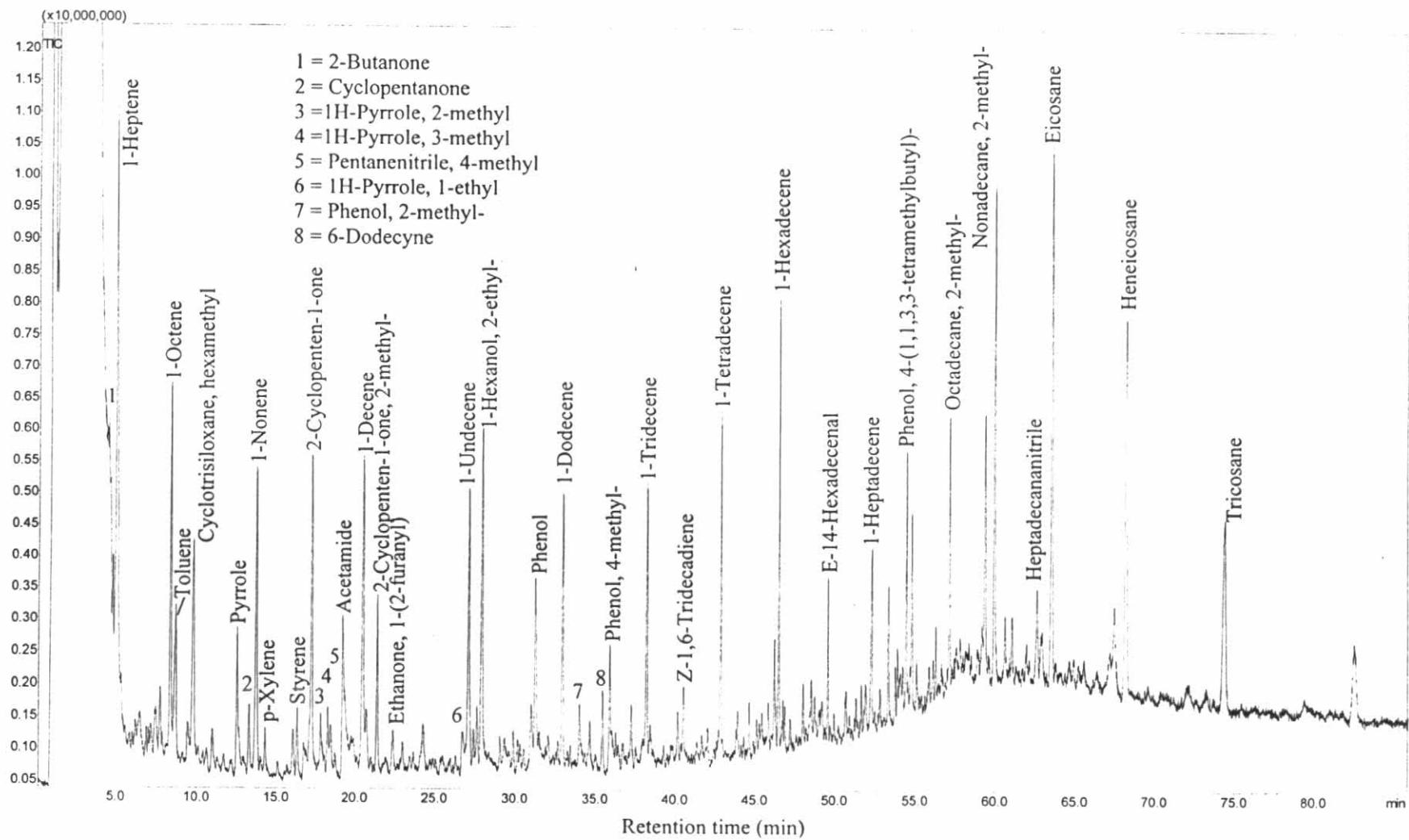


Figure B-35: Pyrochromatogram of major fragments of the HPIN of influent wastewater

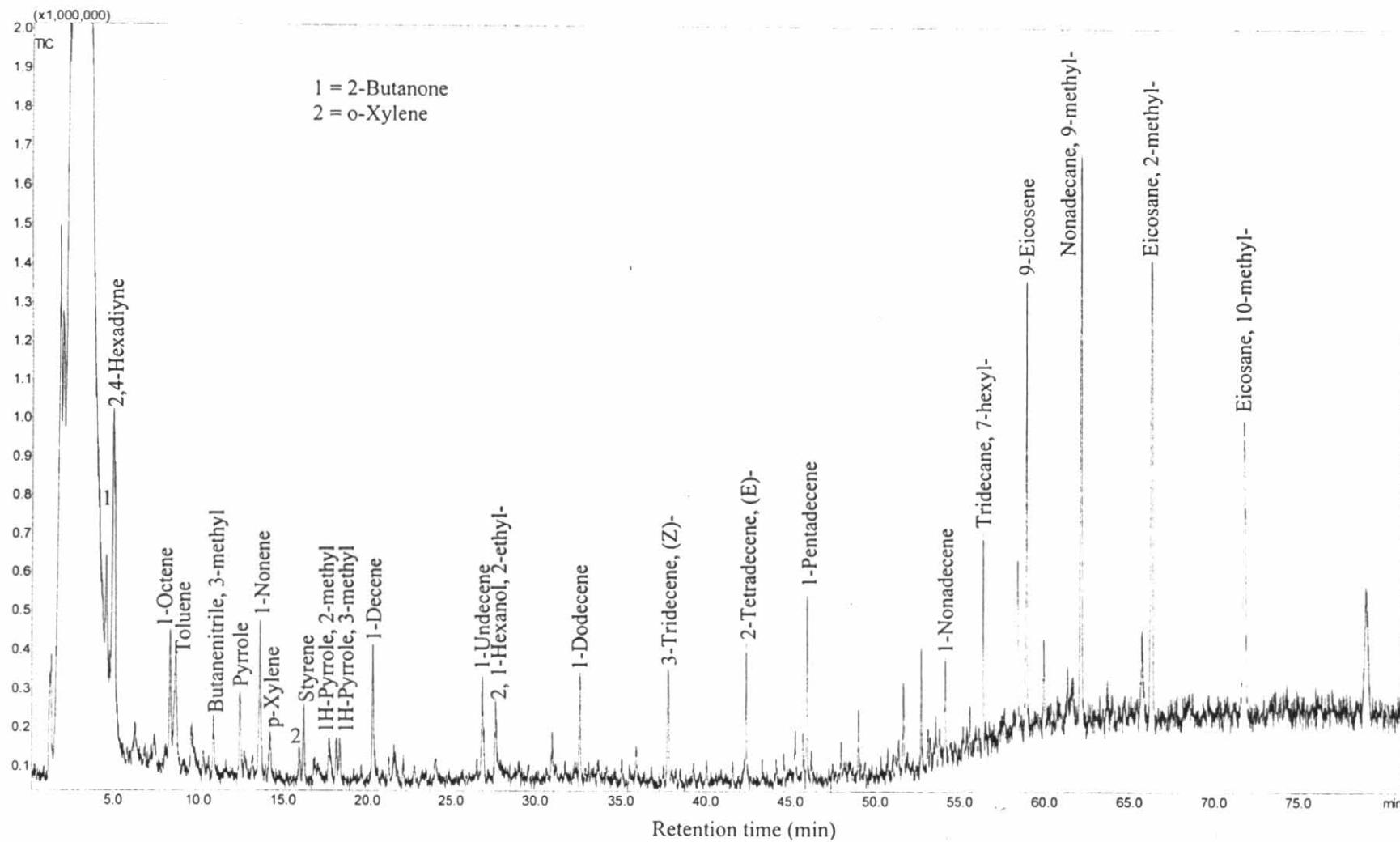


Figure B-36: Pyrochromatogram of major fragments of the HPIN of the aeration pond effluent

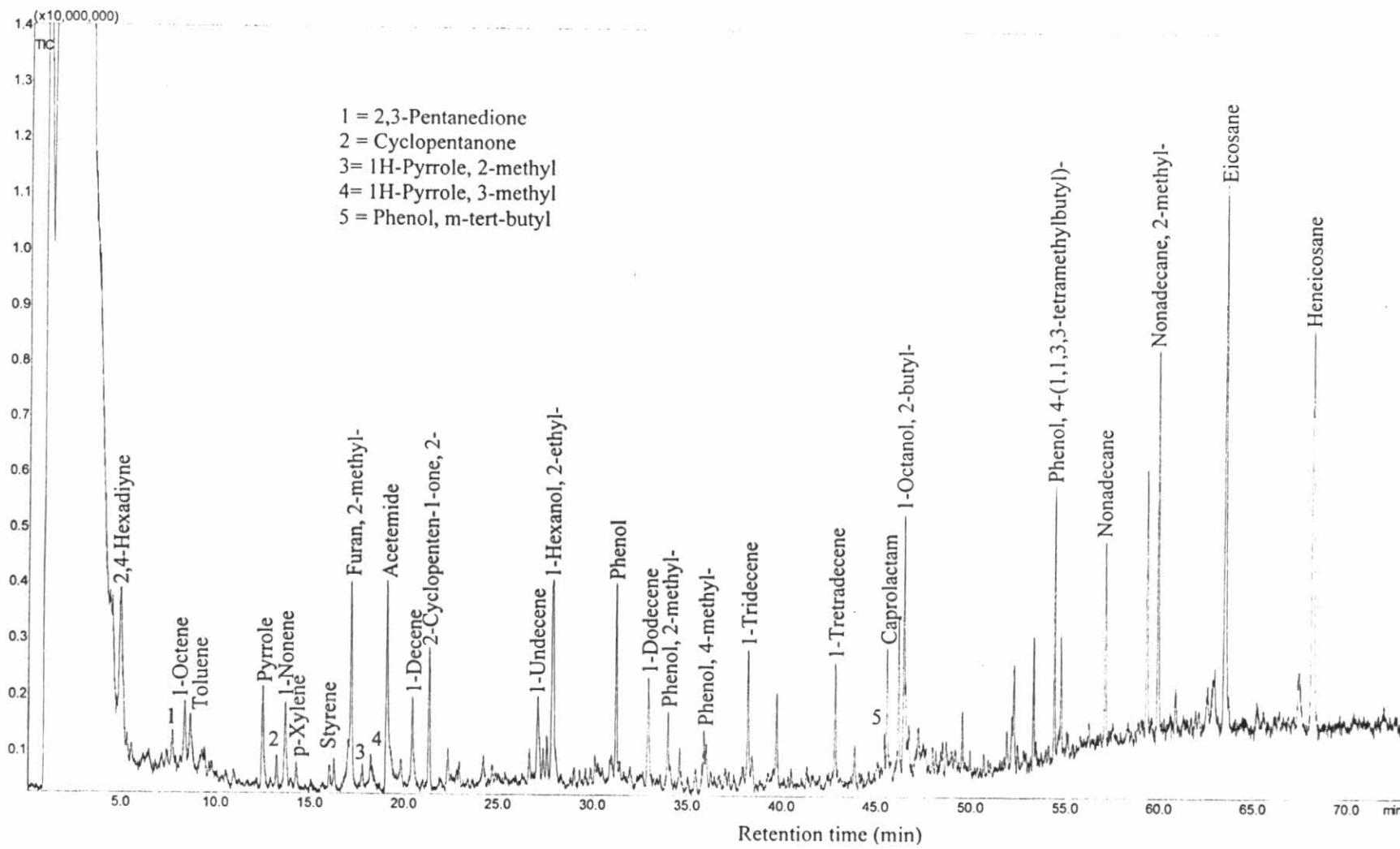


Figure B-37: Pyrochromatogram of major fragments of the HPIN of the facultative pond effluent

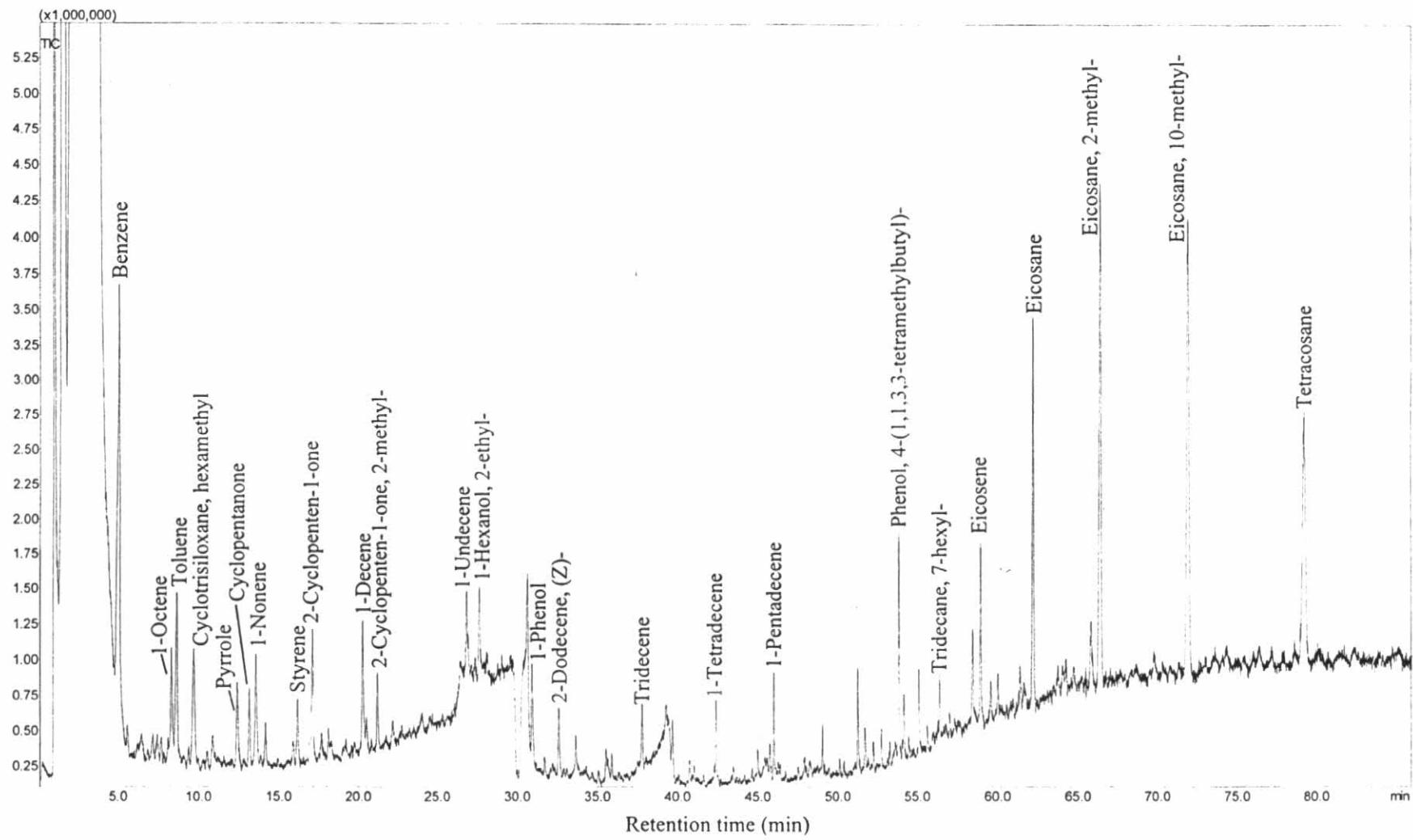


Figure B-38: Pyrochromatogram of major fragments of the HPIN of the oxidation pond effluent

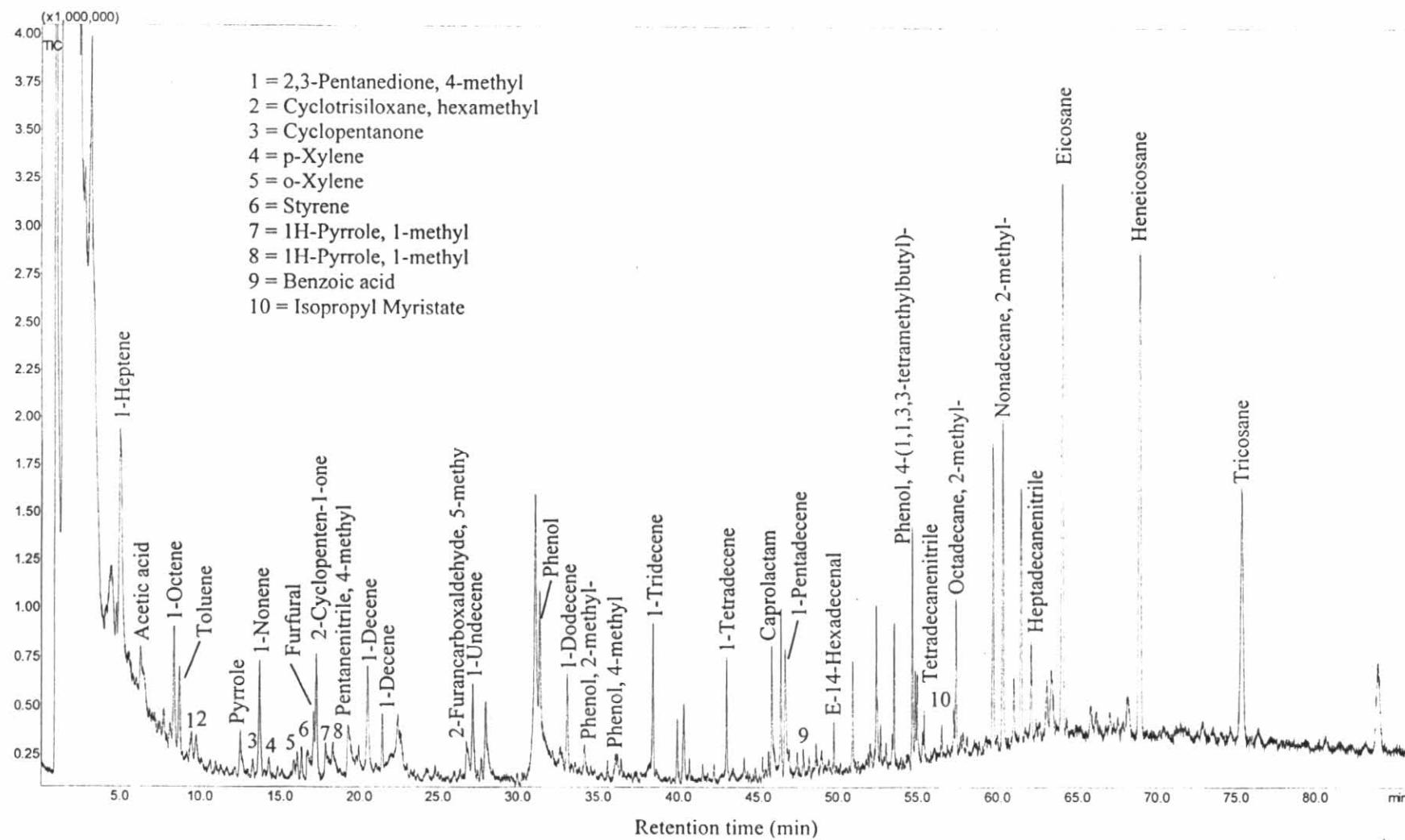


Figure B-39: Pyrochromatogram of major fragments of the HPIN of the detention pond effluent

APPENDIX C
ADVANTAGES, DISADVANTAGES AND LIMITATIONS OF DOM
CHARACTERIZATION USING RESIN FRACTIONATION, PYROLYSIS GC/MS
AND SPECTROFLUOROMETRY

Advantages, Disadvantages, and Limitations of DOM characterization Using Resin Fractionation, Pyrolysis GC/MS and Spectrofluorometry

The resin fractionation, pyrolysis gas chromatography mass spectrometer (GC/MS) and spectrofluorometry methods were utilized to characterize the dissolved organic matter (DOM) in industrial estate wastewater and effluent water from the aeration, facultative, oxidation and detention ponds of the Northern-Region Industrial Estate, Lamphun province, Thailand. The advantages, disadvantages and limitations using these three techniques to characterize DOM have been summarized according to the sample preparation, analytical and data interpretation processes of each technique.

Resin Fractionation

Resin fractionation of DOM in water is a technique that concentrates and categorizes the DOM in water into structurally more specific and physiochemically more analogous subgroups (Marhaba *et al.* 2003). It could be used to identify the major DOM fractions in water. By conducting trihalomethane formation potential (THMFP) tests on DOM fractions, the ability of each DOM fraction for reacting with chlorine to form carcinogenic substances such as trihalomethane (THMs) and haloacetic acids (HAAs) can be determined. The major THMs and HAAs precursors and the active DOM fractions that form THMs and HAAs can be determined. Sample fractionation provides opportunities to identify the complex positions of DOM fractions, especially the complex positions of the major DOM fractions and major THMs and HAAs precursors, by using other techniques such as pyrolysis GC/MS and spectrofluorometry analysis.

Nevertheless, resin fractionation also has disadvantages:

1. It consumes acetone, hexane and methanol in the resin preparation process and an acid and base solution in the sample preparation, resin preparation and elution processes.
2. It is time consuming and labor intensive.

The process has some limitations. The fractionation and isolation of organic matter only exhibits a good recovery of organic matter when the sample contains a low level of inorganic salt or salinity. A desalting step before fractionation might be needed prior to fractionation, even when the organic matter is only present in a small quantity (Leenheer,

2000). Specific conductance in a water sample should not exceed 2,000 $\mu\text{mhos}/\text{cm}$, as above this level the concentration of inorganic ionic salts would exceed the capacity of ion exchange resins. The nature of organic fractions is based on the operational definition. Finally, colloid clay could foul resin adsorbents.

Pyrolysis GC/MS Analysis

Pyrolysis is a method that thermally cleaves an organic molecule into volatile fragments, which are then separated by gas chromatography and identified by mass spectroscopy (MS). Pyrolysis GC/MS yields a reproducible fragmentation pattern or fingerprint, which is highly characteristic of the parent organic matter.

One advantage of the pyrolysis approach is that the natural biopolymers that represent the bulk of DOM precursors is able to be clearly identified because they yield very specific fragments with few interferences among the biopolymers (Leenheer and Croue, 2003). Saiz-Jimenez, (1994) proposed several advantages of pyrolysis GC/MS. First, it requires a small sample amount; only a few hundred micrograms of organic carbon was enough for analysis. Furthermore, an elaborate sample preparation such as derivatization and extraction is not required. Finally, it is capable to providing detailed molecular weight information. Since it does not require solvents for the extraction of the target chemical from a solid or liquid matrix, the target organic matter being studied is not partially oxidized or modified. The organic matrices in the environment that are composed of material too large to be volatilized at 300 °C can be analyzed by GC/MS, since, pyrolysis will thermally extract intact molecules or crack large molecules into fragments that can be separated and identified by GC/MS (White *et al.*, 2004). By using freeze-drying to concentrate DOM, there is the possibility of a high concentration factor, convenient for small sample volumes, easy operation, and a high recovery of DOC and UV-254.

While there are many advantages to utilizing pyrolysis GC/MS, there are also some disadvantages. After the freeze-drying process, a considerably high concentration of inorganic salts may be observed in the samples. It is also inconvenient to use with larger volumes (low DOC concentration) and takes longer in time. In addition, the overall effect in the DOM structure remains uncertain; while data interpretation can be time consuming

and complicated. Finally, it is difficult to make quantitative and provide the indirect information.

Pyrolysis' limitations are listed as follows:

1. In the sample preparation process, when a sample contains a low level of DOM, it is very difficult to receive a uniform fine powder after the freeze-drying process. The concentration of DOM with membranes or an evaporation technique is required.
2. Since, the pyrolysis technique is a destructive technique, fragments of the organic molecule can become involved with side reactions that form new compounds such as ring structures.
3. Pyrolysis results, especially of humic substances, are highly dependent on the temperature selected (Salz-Jimenez, 1994).
4. The library spectrum for some unknown compounds is still limited.
5. The specific biopolymer types of specific water sources such as wastewater and treated wastewater have not been widely established.

Spectrofluorometry Analysis

A fluorescent excitation-emission matrix (FEEM) is obtained by spectrofluorometry. An FEEM can provide information on the putative origin of fluorescent organic matter of DOM in water. It has the advantage of its simplicity due to its minimal sample amount, pretreatment, and analysis time requirements. It can be an alternative that allows for on-line processing and control at reduced operating costs. A fluorescence spectrophotometer can scan the entire usable band in a short period of time (in minutes) without sample pretreatment (Marhaba and Pu, 2000). However, conclusive results from past research established that it is difficult to quantify the level of fluorescent organic matter such as tyrosine-like, tryptophan-like, and humic and fulvic acids-like substances using FEEM analysis.

The limitations for spectrofluorometry analysis are as follows:

1. Since not every compound fluoresces, water samples that contain high levels of non-fluoresced compounds cannot be analyzed by this technique.
2. The fluorescent peak positions of target compounds have not been widely determined. The major putative origins of DOM in water have been at present classified into three groups: tyrosine-like, tryptophan-like, and humic and fulvic acids-like substances.
3. The metal-binding and subsequent fluorescent quenching could occur in water samples that have high pH values
4. The DOC level is significantly important to determining the fluorescent peaks in an FEEM. In the case of low DOC concentrations, the fluorescent peaks cannot be detected (under scale). The concentration of DOM with a membrane and evaporation technique, therefore, may be required. In the case of high DOC concentrations, the fluorescent intensity will be higher than the detection limit of spectrofluorometry (over scale) and the dilution of water samples using Milli-Q water would be required.

Finally the overall integration and application of DOM characterization by using resin fractionation, pyrolysis GC/MS and spectrofluorometry analysis is tabulated in Table C-1.

Table C-1: The overall integration and application of DOM characterization by using the resin fractionation, pyrolysis GC/MS and spectrofluorometry techniques

Techniques	Provided information			Applicability in water and wastewater treatment
	Mass	Physical chemical structure	Disinfection by-products (DBPs) /reactivity	
Resin fractionation	Yes (By conducting the DOC test)	Chemical	Yes (By conducting the DOC and THMFP test)	To monitor the major DOM fractions, major DBPs precursors and highly reactive DOM fractions that form DBPs.
Spectrofluorometry analysis	Only, tyrosine-like, tryptophan-like and humic and fulvic acids-like substances	Moderate both chemical and physical	Indirect	Online, easy monitoring, useful for checking alterations/additions of new substances, useful for checking the major DOM fractions, major DBPs precursors and highly reactive DOM fractions that form DBPs.
Pyrolysis GC/MS	Low	Chemical	Indirect	Identification of chemical class of DOM in unfractionated water and in the major DOM fractions, major DBPs precursors and highly reactive DOM fractions that form DBPs.

APPENDIX D
PUBLICATIONS

Peer Reviewed International Journals/Books

1. Musikavong C. and Wattanachira S. (2007) Reduction of Dissolved Organic Matter in Terms of DOC, UV-254, SUVA and THMFP in Industrial Estate Wastewater Treated by Stabilization Ponds. [Environmental Monitoring and Assessment](#) (Accepted: January 22, 2007)
2. Janhom T., Musikavong C., Wattanachira S., and Furumai H. (2006) Reactivity and Sensitivity of DOM Fractions to Form THMs in Raw Water Supply and Treated Wastewater Used for Reclaimed Water of The Northern-Region Industrial Estate, Thailand. [Southeast Asian Water Environment 2: Book published by International Water Association \(IWA\)](#). London, UK. (In press)

International Symposium/Conference Proceedings

1. Musikavong C., Wattanachira S., Nakajima F., and Furumai H. (2006) Three-Dimensional Fluorescent Spectroscopy Analysis for Evaluation of Organic Matter Removal from Industrial Estate Wastewater by Stabilization Ponds. [The seventh IWA Specialist Conference on Waste Stabilization Ponds: Advances in Pond Technology and Management](#). September 25-27, AIT, Bangkok, Thailand
2. Musikavong C., Wattanachira S. and Pavasant P. (2006) Relationships between Dissolved Organic Matter Surrogates and Area of Excitation Spectrum of Fluorescent Excitation-Emission Matrix in Industrial Estate Wastewater. [The proceeding of international conference on hazardous waste management for a sustainable future](#). January 10-12, Bangkok, Thailand.
3. Janhom T., Musikavong C., Wattanachira S., and Furumai H. (2005) Reactivity and Sensitivity of DOM Fractions to Form THMs in Raw Water Supply and in Treated Industrial Used for Reclaimed Water of the Northern-Region Industrial Estate. [The proceeding of the third international symposium on Southeast Asian Water Environment](#). pp. 201-208, December 6-9, AIT, Bangkok, Thailand.
4. Musikavong C., Wattanachira S. and Pavasant P. (2004) Dissolved Organic Matter in Treated Industrial Estate Wastewater Characterized by Resin Fractionation and Excitation-Emission Fluorescent Spectra. [The proceeding of the second international symposium on Southeast Asian Water Environment](#). pp. 158-165, December 1-3, Hanoi, Vietnam.

National Symposium/Conference Proceedings

1. Musikavong C., Wattanachira S. and Pavasant P. (2005) Application of Excitation-Emission Fluorescence Spectra to Quantify Trihalomethane Formation Potential of Wastewater and Treated Wastewater. [The proceeding of the fourth National Environmental Conferences](#). pp. 564-571, January 19-21, Chonburi, Thailand.

BIOGRAPHY

Name : Charongpun Musikavong
Date of Birth : May 1, 1977
Place of Birth : Nakornsithammarat, Thailand
Education : 1995, Certificate of Mathayomsuksa 6
Benjamarashutit School, Nakornsithammarat, Thailand
: 1999, B.Eng. (Environmental Engineering),
King Mongkut's University of Technology Thonburi,
Bangkok, Thailand
: 2003, M.Sc. (Environmental Management)
Chulalongkorn University,
Bangkok Thailand