

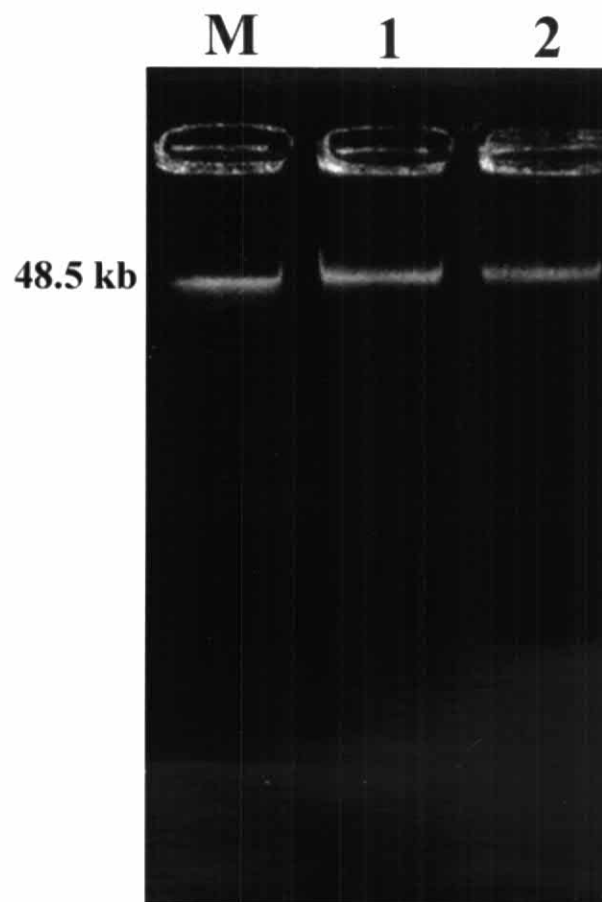
## CHAPTER IV

### RESULTS

#### 4.1 DNA Extraction

High molecular weight genomic DNA was located at approximately 48.5 kb, indicating good quality of extracted genomic DNA (Fig.4.1).

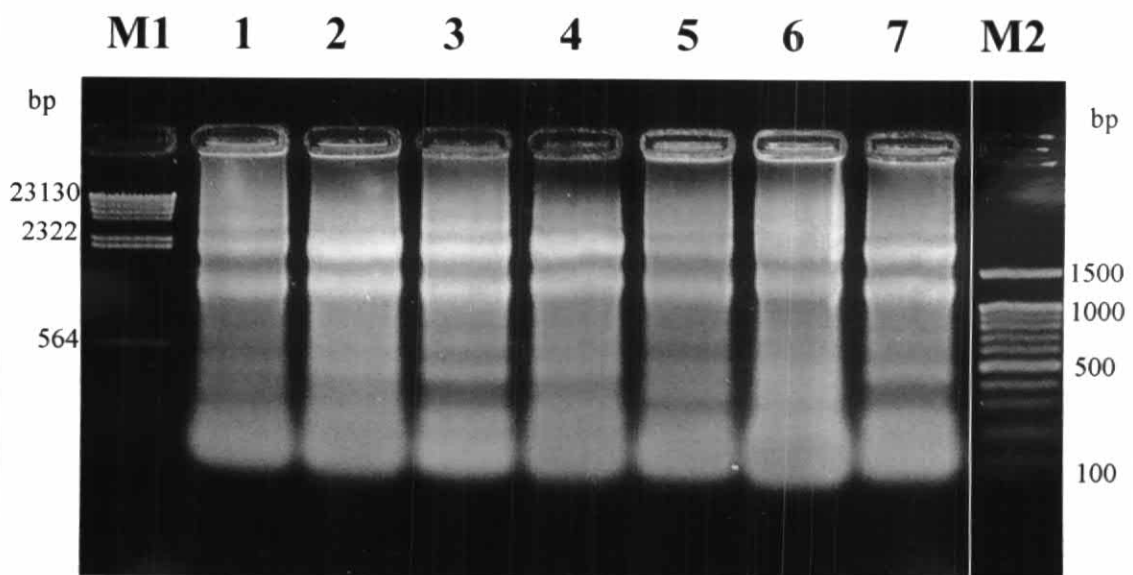
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**Figure 4.1** A 1.2% ethidium bromide stained agarose gel showing the quality of genomic DNA extracted from the muscle of *L. subviridis* (lane 1 and 2). Lane M is 100 ng of undigested lambda DNA marker.

## 4.2 RNA Extraction

The extracted RNA showed 28s and 18s rRNA indicating good quality of RNA (Fig. 4.2). RNA was used for mRNA purification for RACE cDNA synthesis and as the template for 1<sup>st</sup> strand cDNA synthesis, which was then used as the template for amplifying partial and full-length cDNA sequences as well as semiquantitative RT-PCR analysis.

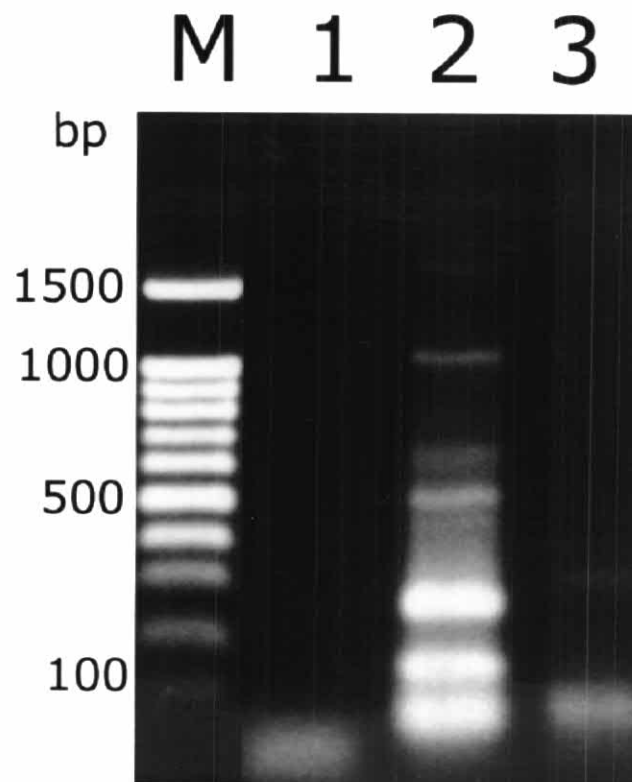


**Figure 4.2** A 1.2 % ethidium bromide stained agarose gel showing the quality of RNA extracted from the liver of *L. subviridis*. Lane 1 – 7 is total liver RNA of normal, 0, 0.05, 0.1, 0.5, 1 and 5 mg/kg E<sub>2</sub> treated (6 days) mullet, respectively. Lane M1 and M2 are  $\lambda$ HindIII and 100 bp DNA marker, respectively.

### 4.3 Cloning and characterization of estrogen receptor (ER) genes

#### 4.3.1 PCR amplification of estrogen receptor (ER) gene

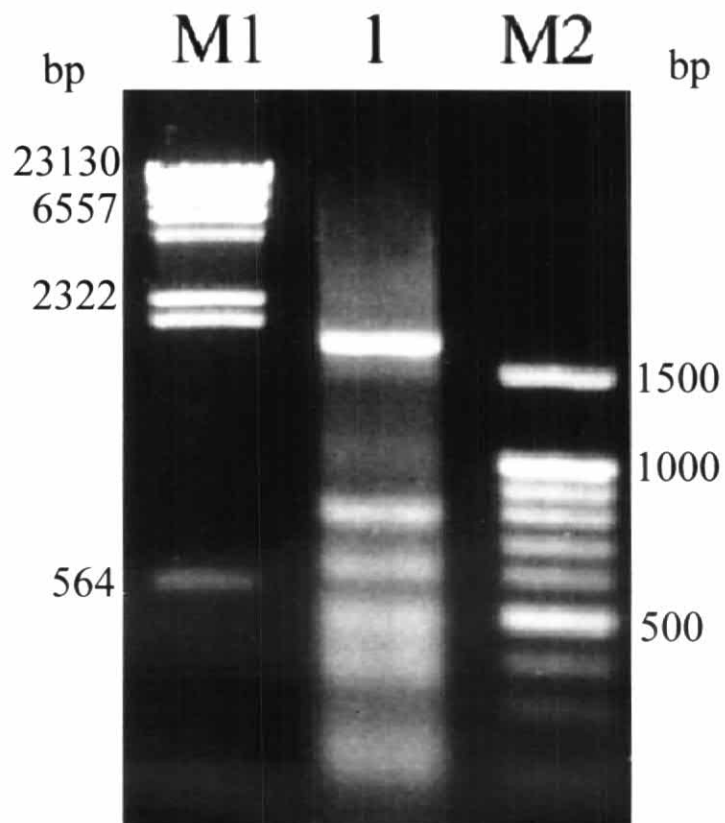
Partial coding sequence of ER gene was amplified from 1<sup>st</sup> strand cDNA template of the liver. DNA fragment at the size of 100 bp was obtained (Fig. 4.3). After sequencing (see in appendix B (*ER FR/59*) and BLAST analysis (see in appendix C), this PCR product was homologous with estrogen receptor alpha of flathead mullet (*Mugil cephalus*) with E-value of  $6e-36$ .



**Figure 4.3** PCR products of ER gene separated in 1.2% agarose gel. PCR products amplified from liver. Lane M is 100 bp markers, lane 1 is negative control, lane 2 is positive control, and lane 3 is PCR product of ER gene.

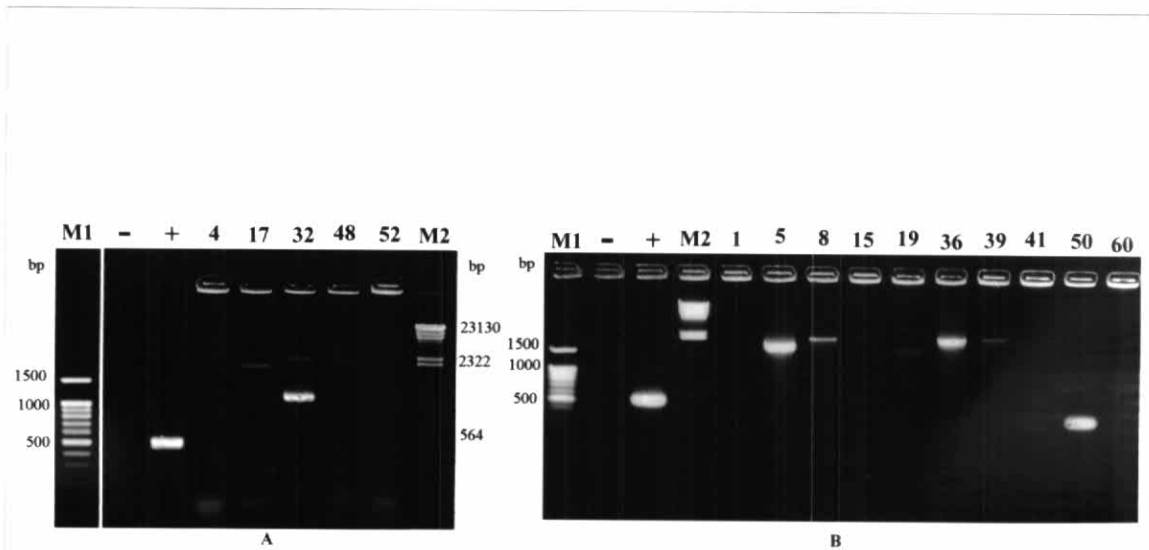
#### 4.3.2 Amplification of 5' and 3' cDNA ends of ER $\alpha$ gene by RACE-PCR

DNA fragment at the size of 1.76 kb was obtained from 3'RACE PCR amplification in liver (Fig. 4.4).

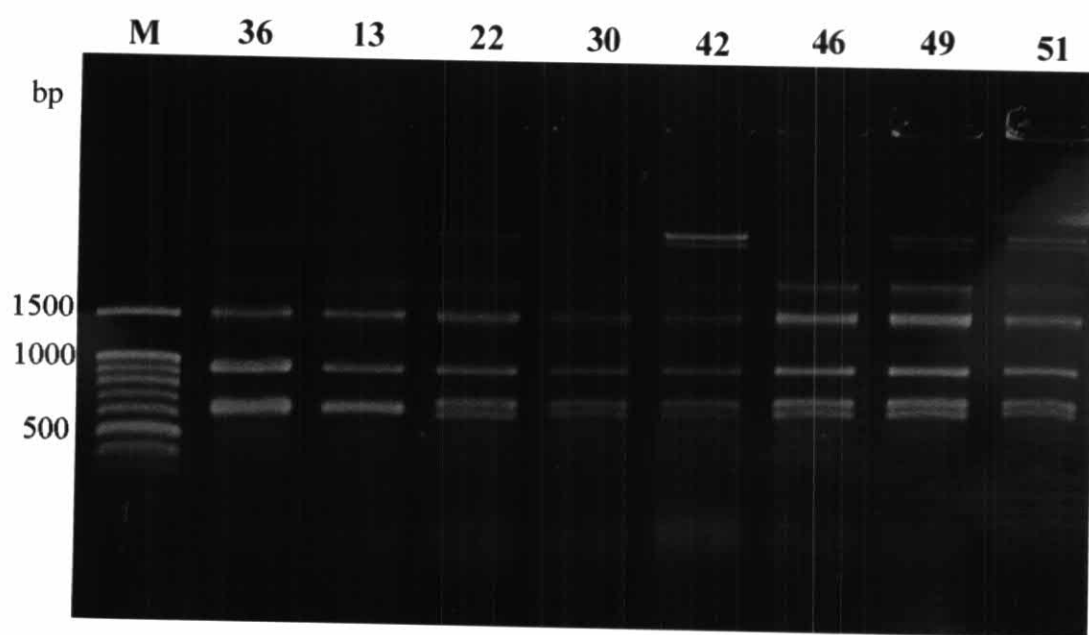


**Figure 4.4** 3' RACE PCR products of ER gene separated in 1.2% agarose gel. PCR products amplified from 3'RACE cDNA of liver (lane M1 is  $\lambda$ HindIII, lane M2 is 100 bp markers and lane 1 is 3' RACE PCR product of ER).

PCR product with a few different sizes was obtained. Band at the size of 1.76 kb which was close to an expected size was cloned. Several white colonies were checked for the insert size by colony PCR. Some examples of colony PCR products were shown in Fig. 4.5 A and B. Clone No.36 (insert size of 1.76 kb), 32 and 5 (insert size lower than 1.76 kb) were cultured for plasmid extraction and sequencing (see in appendix B; 3'ER/5, 32 and 36). After BLAST analysis (see in appendix C), it was found that 3'ER/5, 32 and 36 was homologous with estrogen receptor alpha of European sea bass (*Dicentrarchus labrax*), flathead mullet (*Mugil cephalus*) and thicklip grey mullet (*Chelon labrosus*) with E-value as  $7e-101$ , 0.0 and 0.0, respectively. Because of ER FR/59 sequence used for designing ER F-RACE primer was DNA binding domain which was very highly conserved region between ER $\alpha$  and ER $\beta$ . 3' RACE PCR product of ER gene could amplify 2 isoforms. Therefore, PCR product from 3'ER colonies with the insert size of 1.76 kb were digested with *Hind*III (Fig. 4.6).



**Figure 4.5 A and B;** Colony PCR product of 3'RACE *ER* separated in 1.2% agarose gel. Lane M1 is 100 bp DNA marker, lane M2 is  $\lambda$ HindIII, - is negative control, + is positive control and other lanes is 3'RACE *ER* colony PCR product; number of lane represent number of clone.



**Figure 4.6** 3'ER colony PCR product insert size = 1.76 kb digested with *Hind*III.  
Lane M is 100 bp DNA marker, other lanes is 3'ER colony PCR product  
insert size 1.76 digested with *Hind*III, No. of lane represent No. of clone.

Result in Fig. 4.6 showed 2 different patterns; group1 was No.13 and 36 and group2 was No. 22, 30, 42, 46, 49 and 51. No. 49 was sequenced (see in appendix B). BLAST analysis indicated it homologous with estrogen receptor alpha (*Chelon labrosus*) with E-value as  $3e-160$ . Three types of 3'end cDNA sequence of ER $\alpha$  gene (3'ER/5, 32 and 36) were aligned as shown in Fig. 4.7

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3'ER-5      CTTCAAGAGGAGCATCCAGGGTCACAA TGACTACATGTGCCCGGCGACCAATCAGTGCAC
3'ER-36     CTTCAAGAGGAGCATCCAGGGTCACAA TGACTACATGTGCCCGGCGACCAATCAGTGCAC
3'ER-32     CTTCAAGAGGAGCATCCAGGGTCACAA TGACTACATGTGCCCGGCGACCAATCAGTGCAC
*****

3'ER-5      TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGTCTTAGGAAGTGTATGAAGT
3'ER-36     TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGTCTTAGGAAGTGTATGAAGT
3'ER-32     TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGTCTTAGGAAGTGTATGAAGT
*****

3'ER-5      GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGTTCGCGTTTTGCGGCGCGACAAGCG
3'ER-36     GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGTTCGCGTTTTGCGGCGCGACAAGCG
3'ER-32     GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGTTCGCGTTTTGCGGCGCGACAAGCG
*****

3'ER-5      GCGGGCCGCGCAGAGACAAGGTTGCCAAAGATCCAGAGCACAAGCGTCGGCGCCCCC
3'ER-36     GCGGGCCGCGCAGAGACAAGGTTGCCAAAGATCCAGAGCACAAGCGTCGGCGCCCCC
3'ER-32     GCGGGCCGCGCAGAGACAAGGTTGCCAAAGATCCAGAGCACAAGCGTCGGCGCCCCC
*****

3'ER-5      TCAGGACGGGAGGAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAGGATC
3'ER-36     TCAGGACGGGAGGAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAGGAGGATC
3'ER-32     TCAGGACGGGAGGAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAG--GATC
*****

3'ER-5      TTCTGTGACTAACTTGCTCCAGACCAGGTGCTCCTCCTGCTCCAGGGAGCTGAGCCTCC
3'ER-36     TTCTGTGACTAACTTGCTCCAGACCAGGTGCTCCTCCTGCTCCAGGGAGCTGAGCCTCC
3'ER-32     TTCTGTGACTAACTTGCTCCAGACCAGGTGCTCCTCCTGCTCCAGGGAGCTGAGCCTCC
*****

3'ER-5      GATGCTTTGCTCCCGGCAAAAGCTGAGCCGACCTACACGGAGGTCACCATGATGACCTT
3'ER-36     GATGCTTTGCTCCCGGCAAAAGCTGAGCCGACCTACACGGAGGTCACCATGATGACCTT
3'ER-32     GATGCTTTGCTCCCGGCAAAAGCTGAGCCGACCTACACGGAGGTCACCATGATGACCTT
*****

3'ER-5      GCTCACCAGCATGGCCGACAAGGAGCTGGTGACATGATCGCCTGGGCCAAGAAGCTTCC
3'ER-36     GCTCACCAGCATGGCCGACAAGGAGCTGGTGACATGATCGCCTGGGCCAAGAAGCTTCC
3'ER-32     GCTCACCAGCATGGCCGACAAGGAGCTGGTGACATGATCGCCTGGGCCAAGAAGCTTCC
*****

3'ER-5      AGGTTTCCCTGCAGCTCTCCCTCCACGATCGGGTGCAGCTGCTGGAGAGCTCGTGGCTGGA
3'ER-36     AGGTTTCCCTGCAGCTCTCCCTCCACGATCAGGTGCAGCTGCTGGAGAGCTCGTGGCTGGA
3'ER-32     AGGTTTGGAGAAGCAC----CAACGA--GAGCATTTTTCTGCA-ACCTTGTGTTTGAA
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-5      GGTGCTGATGATCGGGCTCATATGGAGGTCACCCACTGTCCTGG-----
3'ER-36     GGTGCTGATGATCGGGCTCATATGGAGGTCACCCACTGTCCTGGAAAACCTATCTTCGC
3'ER-32     AAAACTG-CAGTGAACAATAATTATTGCTCGACACGCCGAGG-----
*** * * * * * * * * * * * * * * * * * * * * *

3'ER-5      -----GCAGTGGG-----
3'ER-36     CCAGGACCTCATACTGGACAGGAATGAAGGCGACTGCGTTGAGGGCATGGCTGAGATCTT
3'ER-32     -----ACAGAAGAGAATCTGCAGG-----
*

3'ER-5      -----
3'ER-36     TGACATGCTCCTGGCCACCGCTCCCGTTCCGCATGCTCAAGCTCAAACCCGAGGAGTT
3'ER-32     -----

3'ER-5      -----
3'ER-36     CGTGTGCTCAAAGCCATCATCTGCTCAACTCTGGTGCCTTTTCTTCTGCACCGGCAC
3'ER-32     -----

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3'ER-5 -----CCGAC--
3'ER-36 GATGGAGCCCCTCCACGACAACGTGGCCGTGCAGAACATGCTGGACACCATCACCGACGC
3'ER-32 -----CCGCTG-
***

3'ER-5 -----CAGTCCAGGCGGCAGGC
3'ER-36 GCTCATAATCACATCGCCAATCAGGGTTCTCGGCTCAGCAGCAGTCCAGGCGGCAGGC
3'ER-32 -----ATAGAC- GACGATTAGT
*** ** *

3'ER-5 CCAGCTGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCT
3'ER-36 CCAGCTGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCT
3'ER-32 TTAAGAGCGCAGCTTTGCCTGAGATTTGGAGCCGTTACAATCAAATATATGCTGCCTGT
* * * * *

3'ER-5 CTACAGCATGAAGTGCAAGAACAAGTGCCTCTGTACGACCTGCTGCTGGAGATGCTGGA
3'ER-36 CTACAGCATGAAGTGCAAGAACAAGTGCCTCTGTACGACCTGCTGCTGGAGATGCTGGA
3'ER-32 ATGTGGCTGGAAGT--TATTATGGTGTCTCCGCTCTGAATATTGTTGCTCCAGAAATAA
* ** * * * *

3'ER-5 CGCTCACCGCCTCCACCGCCCGGACAAACCGGGCCAGTTCGGGTTCCAGGTCGGCAAAGA
3'ER-36 CGCTCACCGCCTCCACCGCCCGGACAAACCGGGCCAGTTCGGGTTCCAGGTCGGCAAAGA
3'ER-32 GATTGATAGGTTTTAACTTCAAAAAAAAAAAAAAAAAAAAAA
* * * * *

3'ER-5 CCCTCCCCCACCACCAACAGAGACGGCGGGGTGCAGCCGGGGGAGGCGGTTCCGGGACC
3'ER-36 CCCTCCCCCACCACCAACAGAGACGGCGGGGTGCAGCCGGGGGAGGCGGTTCCGGGACC
3'ER-32 -----

3'ER-5 TCGAGGCAGCCACGAGAGCCCGAGCAGACCCCTCTGGTCCGGGCGTCTGCAGTACGG
3'ER-36 TCGAGGCAGCCACGAGAGCCCGAGCAGACCCCTCTGGTCCGGGCGTCTGCAGTACGG
3'ER-32 -----

3'ER-5 AGGCCCCAGATCTGACTGCACCCACATCTTATGAGGCAGGAAGGAAGGAGGAGCAGCAC
3'ER-36 AGGCCCCAGATCTGACTGCACCCACATCTTATGAGGCAGGAAGGAAGGAGGAGCAGCAC
3'ER-32 -----

3'ER-5 GCGAAGGTCAAAAAATCCTTTTCATTGATGTTGCGTTTACAGAATGAAAAGGTTTTGAG
3'ER-36 GCGAAGGTCAAAAAATCCTTTTCATTGATGTTGCGTTTACAGAATGAAAAGGTTTTGAG
3'ER-32 -----

3'ER-5 TTAATTTTCATGAGAGAATTATTTATAAATCTGTGATTTTAAAGCTGTTTAGGGAGAAGC
3'ER-36 TTAATTTTCATGAGAGAATTATTTATAAATCTGTGATTTTAAAGCTGTTTAGGGAGAAGC
3'ER-32 -----

3'ER-5 TTCCCTCGAACTGCTCCGACTCGCGTCAGTCTGAGCGTCGGTGCAGCTGATCTTACACCT
3'ER-36 TTCCCTCGAACTGTTCCGACTCGCGTCAGTCTGAGCGTCGGTGCAGCTGATCTTACACCT
3'ER-32 -----

3'ER-5 TTCATAATATCTGTGATTTCAGAGTGCCTCTTAACGGCTTTTTCGGTGTGTTTTATTACC
3'ER-36 TTCATAATATCTGTGATTTCAGAGTGCCTCTTAACGGCTTTTTCGGTGTGTTTTATTACC
3'ER-32 -----

3'ER-5 CGTGGCACTCTGTTGGTGATTTTGAATGACGAGCAGCTAATCTTTCCGTTTCTTTGCC
3'ER-36 CGTGGCACTCTGTTGGTGATTTTGAATGACGAGCAGCTAATCTTTCCGTTTCTTTGCC
3'ER-32 -----

3'ER-5 TCGACCAAAGTGCACTTCCCTCTCGCATTCAAGGGGTAAGGGCATTATTTTTACTTTTG
3'ER-36 TCGACCAAAGTGCACTTCCCTCTCGCATTCAAGGGGTAAGGGCATTATTTTTACTTTTG
3'ER-32 -----

3'ER-5 CATTTAAATAGGGTAAAAATAAATCGCGATAAACAGGTCAAAAAAAAAAAAAAAAAAAA
3'ER-36 CATTTAAATAGGGTAAAAATAAATCGCGATAAACAGGTTAAAAAAGAGTAAATGATTT
3'ER-32 -----

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```

3'ER-5      AAAAAAAAA-----
3'ER-36     AGTGTTCCTTCAACATCAGGTATTGTGTTACTCACAAATTTAAACTTGAGATATT
3'ER-32     -----

3'ER-5      -----
3'ER-36     GCAAAAAAAAAAAAAAAAAAAAAAAAAAAA
3'ER-32     -----

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**Figure 4.7** Nucleotide alignment of 3 types of 3'end of cDNA sequences of ER $\alpha$  gene (3'ER/5, 32 and 36) in *L. subviridis*. Yellow, blue, pink and green label are ER F-RACE primer, stop codon, polyadenylation signal and polyA tail, respectively.

Nucleotide sequences of 3 variant transcript of ER $\alpha$  gene were different. 3'ER/5 and 3'ER/32 lack some region of coding sequences but 3'ER/36 have complete coding sequence. Three variant of ER $\alpha$  gene contained stop codon, 1 polyadenylation signal (AATAAA), and polyA tail which different in length and polyadenylation site between variants. 3'untranslated region (3'UTR) were different in length between variants as shown in Fig. 4.7. Three types of 3'end cDNA sequence of ER $\alpha$  gene (3'ER/5, 32 and 36) were translated to amino acid sequences and aligned. DNA binding domain showed 100 % identical in 3 variant. DNA binding domain contained zinc finger region (red alphabet shown in Fig. 4.8) and D box (PATNQ) (underline in Fig. 4.8 which involved in receptor dimerization. D domain showed 100 % identical in variant 5 and 36, 99.08 % similarity between variant (5 and 36) with variant 32. Interestingly 1 amino acid residues is missed from variant 32. Ligand binding domain was complete in variant 36 but lack 66.67 % of this domain in variant 5 and 32. F domain was complete in variant 36 and 5 with 98.97 % similarity but lack almost portion of this domain in variant 32 as shown in Fig. 4.8.

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3'ER-36      FKRSIQGHNDYMCPATNQCTIDRNRKSCQACRLRKCYEVGMMKGGVVRKDRGRVLRDRDKR
3'ER-5      FKRSIQGHNDYMCPATNQCTIDRNRKSCQACRLRKCYEVGMMKGGVVRKDRGRVLRDRDKR
3'ER-32      FKRSIQGHNDYMCPATNQCTIDRNRKSCQACRLRKCYEVGMMKGGVVRKDRGRVLRDRDKR
*****

3'ER-36      RAGDRDKVAKDPEHKASAPPQDGRKRSSSCSSTGGGGGSSVTNLPDQVLLLLQGAEPP
3'ER-5      RAGDRDKVAKDPEHKASAPPQDGRKRSSSCSSTGGGGGSSVTNLPDQVLLLLQGAEPP
3'ER-32      RAGDRDKVAKDPEHKASAPPQDGRKRSSSCSSTGGGGG--SSVTNLPDQVLLLLQGAEPP
*****

3'ER-36      MLCSRQKLSRPYTEVTMMLTLLTSMADKELVHMIAWAKKLPGFQLSLHDQVQLLESSWLE
3'ER-5      MLCSRQKLSRPYTEVTMMLTLLTSMADKELVHMIAWAKKLPGFQLSLHDRVQLLESSWLE
3'ER-32      MLCSRQKLSRPYTEVTMMLTLLTSMADKELVHMIAWAKKLPG--LEKHQREHFFCNLVFE
*****
          * . * : : : . : *

3'ER-36      VLMIGLIWRSTHCPGKLIFAQDLILDRNEGDCVEGMAEIFDMLLATASRFRLKLPKPEEF
3'ER-5      VLMIGLIWRSTHCPG-----
3'ER-32      KTAVKQIYLRHAAG-----
          : * : . * . *

3'ER-36      VCLKAII LLNSGAFSFCGTMEPLHDNVAVQNMLDTITDALIHIGQSGFSAQQQSRQA
3'ER-5      -----QWADQSRQA
3'ER-32      -----QKR-----
          * . *

3'ER-36      QLLLLLSHIRHMSNKGMEHLYSMKCKNKVPLYGLLEMLDAHRLHRPDKPGQFGFQVGKD
3'ER-5      QLLLLLSHIRHMSNKGMEHLYSMKCKNKVPLYDILLEMLDAHRLHRPDKPGQFGFQVGKD
3'ER-32      -----ICR---PLID-----
          * : ** .

3'ER-36      PPPTTNSDGGGAAGGGGSGPRGSHEPSRPPSGPVLQYGGPRSDCTHIL
3'ER-5      PPPTTNRDGGGAAGGGGSGPRGSHEPSRPPSGPVLQYGGPRSDCTHIL
3'ER-32      -----

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**Figure 4.8** Deduced sequence of 3'ER/5, 32 and 36. Yellow label, red alphabet, underline, non label, blue label, and grey label is DNA binding domain, zinc finger region, D box, D domain, ligand binding domain, and F domain, respectively.

Nucleotide sequences of 3'ER/36 and 49 which were similar to *HindIII* digested pattern were aligned and showed in Fig. 4.9 A and B

3'ER-49-M13F -----  
3'ER-36 CTTCAAGAGGAGCATCCAGGGTCACAATGACTACATGTGCCCGGGACCAATCAGTGCAC

3'ER-49-M13F -----  
3'ER-36 TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGCTTAGGAAGTGTATGAAGT

3'ER-49-M13F -----  
3'ER-36 GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGGTGCGGTTTTGCGGGCGACAAGCG

3'ER-49-M13F -----  
3'ER-36 GCGGGCCGGGACAGAGACAAGGTTGCCAAAGATCCAGAGCACAAAGCGTCGGCGCCCC

3'ER-49-M13F -----  
3'ER-36 TCAGGACGGGAGGAAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAGGATC

3'ER-49-M13F -----  
3'ER-36 TTCTGTGACTAACTTGCCTCCAGACCAGGTGCTCCTCCTGCTCCAGGGAGCTGAGCCTCC

3'ER-49-M13F -----  
3'ER-36 GATGCTTTGCTCCCGGAAAAGCTGAGCCGACCCTACACGGAGGTCACCATGATGACCCT

3'ER-49-M13F -----  
3'ER-36 GCTCACCAGCATGGCCGACAAGGAGCTGGTGCACATGATCGCCTGGGCCAAGAAGCTTCC

3'ER-49-M13F -----  
3'ER-36 AGGTTTCCTGCAGCTCTCCCTCCACGATCAGGTGCAGCTGCTGGAGAGCTCGTGGCTGGA

3'ER-49-M13F -----  
3'ER-36 GGTGCTGATGATCGGGCTCATATGGAGGTCCACCCACTGTCTGGAAAACATCTTCGC

3'ER-49-M13F -----  
3'ER-36 CCAGGACCTCATACTGGACAGGAATGAAGGGGACTGCGTTGAGGGCATGGCTGAGATCTT

3'ER-49-M13F -----  
3'ER-36 TGACATGCTCCTGGCCACCGCGTCCCGTTTTCCGCATGCTCAAGCTCAAACCCGAGGAGTT

3'ER-49-M13F -----  
3'ER-36 CGTGTGTCTCAAAGCCATCATCTGCTCAACTCTGGTGCCTTTTCTTTCTGCACCGGCAC

3'ER-49-M13F -----  
3'ER-36 GATGGAGCCCCCTCCACGACAACGTGGCCGTGCAGAACATGCTGGACACCATCACCAGCG

3'ER-49-M13F -----  
3'ER-36 GCTCATACATCACATCGGCCAATCAGGTTCTCGGCTCAGCAGCAGTCCAGGGCGGACGG

3'ER-49-M13F -----  
3'ER-36 CCAGCTGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAGGCATGGAGCACCT

3'ER-49-M13F -----  
3'ER-36 CTACAGCATGAAGTGAAGAACAAGTGCCTCTGTACGGCCTGCTGCTGGAGATGCTGGA  
\*\*\*\*\*

3'ER-49-M13F -----  
3'ER-36 CGCTCACCGCCTCCACCGCCCGGACAAACCGGGCCAGTTCGGGTTCCAGGTCCGGCAAAGA  
CGCTCACCGCCTCCACCGCCCGGACAAACCGGGCCAGTTCGGGTTCCAGGTCCGGCAAAGA  
\*\*\*\*\*

3'ER-49-M13F -----  
3'ER-36 CCCTCCCCCACCACCAACAGCGACGGCGGGGTGCAGCCGGGGAGGCGGTTCCGGGAC  
CCCTCCCCCACC-ACCAACAGCGACGGCGGGGTGCAGCCGGGGAGGCGGTTCCGGGAC  
\*\*\*\*\*

3'ER-49-M13F CTCTGAGGCAGCCACGAGAGCCCGAGCAGACCCCTCTGGTCCGGGCGTCTGCAGTACG  
3'ER-36 CTCTGAGGCAGCCACGAGAGCCCGAGCAGACCCCTCTGGTCCGGGCGTCTGCAGTACG  
\*\*\*\*\*

3'ER-49-M13F GAGGCCCCAGATCTGACTGCACCCACATCTTATGAGGCAGGAAGGAAGGAGCAGCA  
3'ER-36 GAGGCCCCAGATCTGACTGCACCCACATCTTATGAGGCAGGAAGGAAGGAGCAGCA  
\*\*\*\*\*

3'ER-49-M13F CGCGAAGGTCAAAAAATCCTTTTCATTGATGTTGCGTTTACAGAATGAAAAGGGTTTTG  
3'ER-36 CGCGAAGGTCAAAAAA-TCCTTTTCATTGATGTTGCGTTTACAGAATGAAAAGGGTTTTG  
\*\*\*\*\*

3'ER-49-M13F AGTCAATTTTCATGAGAGAATTATTTATAAATCTGTGATTTTAAAGCTGTTTAGGGAGAA  
3'ER-36 AGTCAATTTTCATGAGAGAATTATTTATAAATCTGTGATTTTAAAGCTGTTTAGGGAGAA  
\*\*\*\*\*

3'ER-49-M13F GCTTCCTCGAACTGCTCCGACTCGCGTCAGTTTGGAGCGTCGGTGCAGCTGATCTTACAC  
3'ER-36 GCTTCCTCGAACTGCTCCGACTCGCGTCAGTTTGGAGCGTCGGTGCAGCTGATCTTACAC  
\*\*\*\*\*

3'ER-49-M13F CTTTCATAATATCTGTGATTCAGAGTGCCTCTAACGGCTTTTTCGGTGTGCTTTATTA  
3'ER-36 CTTTCATAATATCTGTGATTCAGAGTGCCTCTAACGGCTTTTTCGGTGTGCTTTATTA  
\*\*\*\*\*

3'ER-49-M13F CCCGTGGCACTCTGTTGGTGATTTTGAATGACGAGCAGCTAATCTTCCGTTTCTTTG  
3'ER-36 CCCGTGGCACTCTGTTGGTGATTTTGAATGACGAGCAGCTAATCTTCCGTTTCTTTG  
\*\*\*\*\*

3'ER-49-M13F CCTCGACCAAAGTGCACCTTCTCTCGCATTCAAGGGGTAAAGGGCATTATTTTACTTT  
3'ER-36 CCTCGACCAAAGTGCACCTTCTCTCGCATTCAAGGGGTAAAGGGCATTATTTTACTTT  
\*\*\*\*\*

3'ER-49-M13F TGCATTTAAAATAGGGTAAAAATAAAATCGCGATAAACAGGTAAAAAAGAGTAAATGAT  
3'ER-36 TGCATTTAAAATAGGGTAAAAATAAAATCGCGATAAACAGGTAAAAAAGAGTAAATGAT  
\*\*\*\*\*

3'ER-49-M13F TTAGTGTTTTCTTTCAAACATCAGGTATTGTGTTTACTCACAATTTAAACCTGAGATAT  
3'ER-36 TTAGTGTTTTCTTTCAAACATCAGGTATTGTGTTTACTCACAATTTAAACCTGAGATAT  
\*\*\*\*\*

3'ER-49-M13F TTGGAAAAAAAAAAAAAAAAAAAAAAAAA  
3'ER-36 TTGCAAAAAAAAAAAAAAAAAAAAAAAAA-  
\*\*\* \*\*\*\*\*

A

3'ER-36 CTTCAAGAGGAGCATCCAGGGTCACAATGACTACATGTGCCCGGCGACCAATCAGTGCAC  
3'ER-49-M13R CTTCAAGAGGAGCATCCAGGGTCACAATGACTACATGTGCCCGGCGACCAATCAGTGCAC  
\*\*\*\*\*

3'ER-36 TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGCTTTAGGAAGTGTATGAAGT  
3'ER-49-M13R TATTGACAGGAATCGGAGGAAGAGCTGCCAGGCTTGCCGCTTTAGGAAGTGTATGAAGT  
\*\*\*\*\*

3'ER-36 GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGTTCGCGTTTTCGGCGCGACAAGCG  
3'ER-49-M13R GGGCATGATGAAAGGAGGTGTGCGCAAAGACCGGGTTCGCGTTTTCGGCGCGACAAGCG  
\*\*\*\*\*

3'ER-36 GCGGGCCGGCGACAGAGACAAGGTGCCAAAGATCCAGAGCACAAGCGTCGGCGCCCC  
3'ER-49-M13R GCGGGCCGGCGACAGAGACAAGGTGCCAAAGATCCAGAGCACAAGCGTCGGCGCCCC  
\*\*\*\*\*

3'ER-36 TCAGGACGGGAGGAAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAGGA--  
3'ER-49-M13R TCAGGACGGGAGGAAGCGCAGCAGCAGCTGTAGCAGCACCGGAGGAGGAGGAGGAGGA  
\*\*\*\*\*

3'ER-36 -TCTTCTGTGACTAACTTGCCCTCAGACCAGGTGCTCCTCCTGCTCCAGGAGCTGAGCC  
3'ER-49-M13R ATCTTCTGTGACTAACTTGCCCTCAGACCAGGTGCTCCTCCTGCTCCAGGAGCTGAGCC  
\*\*\*\*\*

3'ER-36 TCCGATGCTTTGCTCCCGCAAAGCTGAGCCGACCCCTACACGGAGGTCACCATGATGAC  
3'ER-49-M13R TCCGATGCTTTGCTCCCGCAAAGCTGAGCCGACCCCTACACGGAGGTCACCATGATGAC  
\*\*\*\*\*

3'ER-36 CCTGCTCACCAGCATGGCCGACAAGGAGCTGGTGCACATGATCGCCTGGGCCAAGAAGCT  
 3'ER-49-M13R CCTGCTCACCAGCATAGCCGACAAGGAGCTGGTGCACATGATCGCCTGGGCCAAGAAGCT  
 \*\*\*\*\*

3'ER-36 TCCAGGTTTCTGCAGCTCTCCCTCCACGATCAGGTGCAGCTGCTGGAGAGCTCGTGGCT  
 3'ER-49-M13R TCCAGGTTTCTGCAGCTCTCCCTCCACGATCAGGTGCAGCTGCTGGAGAGCTCGTGGCT  
 \*\*\*\*\*

3'ER-36 GGAGGTGCTGATGATCGGGCTCATATGGAGGTCCACCCACTGCTGGAAGAACTCATCTT  
 3'ER-49-M13R GGAGGTGCTGATGATCGGGCTCATATGGAGGTCCACCCACTGCTGGAAGAACTCATCTT  
 \*\*\*\*\*

3'ER-36 CGCCCAGGACCTCATACTGGACAGGAATGAAGGCGACTGCGTTGAGGGCATGGCTGAGAT  
 3'ER-49-M13R CGCCCAGGACCTCATACTGGACAGGAATGAAGGCGACTGCGTTGAGGGCATGGCTGAGAT  
 \*\*\*\*\*

3'ER-36 CTTTGACATGCTCCTGGCCACCGCTCCCGTTTCCGCATGCTCAAGCTCAAACCCGAGGA  
 3'ER-49-M13R CTTTGACATGCTCCTGGCCACCGCTCCCGTTTCCGCATGCTCAAGCTCAAACCCGAGGA  
 \*\*\*\*\*

3'ER-36 GTTCGTGTGCTCAAAGCCATCATCCTGCTCAACTCTGGTGCCTTTTCTTTCTGCACCGG  
 3'ER-49-M13R GTTCGTGTGCTC-----  
 \*\*\*\*\*

3'ER-36 CACGATGGAGCCCTCCACGACAACGTGGCCGTGCAGAACATGCTGGACACCATCACCGA  
 3'ER-49-M13R -----  
 -----

3'ER-36 CGCGCTCATAATCACATCGGCCAATCAGGGTTCTCGGCTCAGCAGCAGTCCAGCGGCA  
 3'ER-49-M13R -----  
 -----

3'ER-36 GGCCCAGCTGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAGGCATGGAGCA  
 3'ER-49-M13R -----  
 -----

3'ER-36 CCTCTACAGCATGAAGTGAAGAACAAGTGCCTCTGTACGGCCTGCTGCTGGAGATGCT  
 3'ER-49-M13R -----  
 -----

3'ER-36 GGACGCTCACCGCCTCCACCGCCCGGACAAACCGGGCCAGTTCGGGTTCCAGGTCGGCAA  
 3'ER-49-M13R -----  
 -----

3'ER-36 AGACCCTCCCCCACCACCAACAGCGACGGCGGGGTGCAGCCGGGGAGGCGGTTCCGGG  
 3'ER-49-M13R -----  
 -----

3'ER-36 ACCTCGAGGCAGCCACGAGAGCCCGAGCAGACCCCTCTGGTCCGGGCGTCTGCAGTA  
 3'ER-49-M13R -----  
 -----

3'ER-36 CGGAGGCCCCAGATCTGACTGCACCCACATCTTATGAGGCAGGAAGGAAGGAGGAGCAG  
 3'ER-49-M13R -----  
 -----

3'ER-36 CACGCGAAGGTCAAAAATCCTTTTCATTGATGTTGCGTTTACAGAATGAAAAGGGTTTT  
 3'ER-49-M13R -----  
 -----

3'ER-36 GAGTTAATTTTCATGAGAGAATTTTATAAATTCTGTGATTTTAAAGCTGTTTAGGGAGA  
 3'ER-49-M13R -----  
 -----

3'ER-36 AGCTTCCCTCGAACTGTTCCGACTCGCGTCAGTCTGAGCGTCCGGTGCAGCTGATCTTACA  
 3'ER-49-M13R -----  
 -----

3'ER-36 CCTTTCATAATATCTGTGATTCAGAGTGCCTCTTAACGGCTTTTCCGGTGCCTTTATT  
 3'ER-49-M13R -----  
 -----

3'ER-36 ACCCGTGGCACTCTGTTGGTGATTTTGAATGACGAGCAGCTAATTCTTTCCGTTTCTTT  
 3'ER-49-M13R -----  
 -----

```

3'ER-36      GCCTCGACCAAAGTGCACTTCCTCTCGCATTCAAGGGGGTAAAGGGCATTATTTTACTT
3'ER-49-M13R -----

3'ER-36      TTGCATTTAAAAATAGGGTAAATAATAATCGCGATAAACCCAGGTAAAAAAGAGTAAATGA
3'ER-49-M13R -----

3'ER-36      TTTAGTGTTTTCTTTCAAACATCAGGTATTGTGTTTACTCACAAATTTAAAACCTTGAGATA
3'ER-49-M13R -----

3'ER-36      TTTGC#####
3'ER-49-M13R -----

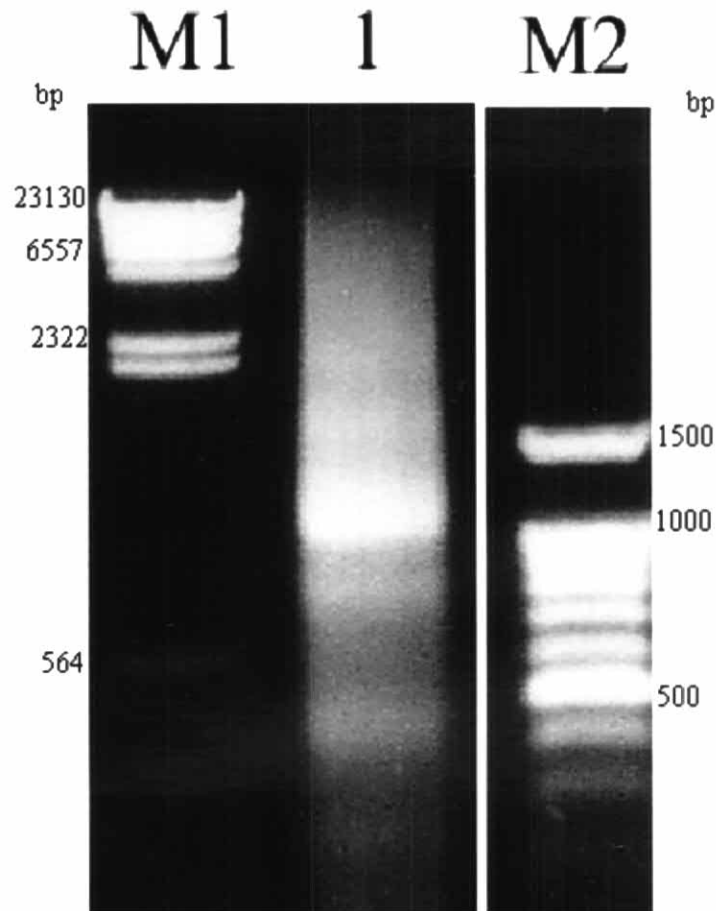
```

## B

**Figure 4.9** 3'ER/36 aligned with 3'ER/49 M13F sequence (A) and M13R (B). Blue, pink and green label were stop codon, polyadenylation signal and polyA tail, respectively.

cDNA sequences shown 98.57 % similarity between 3'ER/49 and 3'ER/36 as shown in Fig. 4.9 so the internal sequence of 3'ER/49 was not further sequenced.

ER $\alpha$  R RACE primer was designed from 3' cDNA sequence of ER $\alpha$  gene. A DNA fragment at the size of 1.2 kb was obtained from 5'RACE PCR amplification of ER $\alpha$  in liver (Fig.4.10). After sequencing (see in appendix B) and BLAST analysis (see in appendix C), this PCR product was homologous with estrogen receptor alpha of Nile tilapia (*Oreochromis niloticus niloticus*) with E-value of 0.0



**Figure 4.10** 5'RACE PCR product of *ER $\alpha$*  separated in 1.2% agarose gel. Lane M1 is  $\lambda$ HindIII, M2 is 100 bp DNA marker and lane 1 is 5'RACE PCR product of *ER $\alpha$* .





```

aggaaggaaaggaaggagcagcagcgaagggtcaaaaaatccttttcattgatgttgctt
tacagaatgaaaagggttttgagttatctcatgagagaattattataaattctgtgat
tttaaagctgtttagggagaagcttcctcgaactgttcgactcgcgtcagtcagcg
tcggtgcagctgatcttacacctttcataatatctgtgattcagagtgcgtctctaacgg
cttttcggtgctgtttattaccctggcactctgttggtgattttgaaatgacgagcag
ctaattctttccgtttctttgcctcgaccaaaagtgcacttcctcctcgattcaaggggt
aaagggcattattttacttttgcattttaaataagggtaaaataaaatcgcgataaaacca
ggttaaaaagagtaaatgatttagtgtttctttcaaacatcaggtattgtgtttactc
acaatttaaaacttgagatatttgcataaaaaaaaaaaaaaaaaaaaaaaaaaaaaa

```

**Figure 4.11** Full length cDNA sequence of *ER $\alpha$*  and deduced amino acid sequence of *ER $\alpha$* . Blue, red alphabet, blue alphabet, pink and green labels are *ER $\alpha$*  full length primer, start codon, stop codon, polyadenylation signal and polyA tail, respectively.

Result in Fig. 4.11 shows full length mRNA sequence of *L. subviridis ER $\alpha$*  at the size of 2,512 bp which contained 5'UTR, ORF and 3'UTR (contained 1 polyadenylation site) at the sizes of 114, 1,863 and 535 bp, respectively. ORF at the size of 1,863 bp encoded deduced amino acid sequence of *ER $\alpha$*  which contained 620 amino acid residues with the molecular weight at 67.55 kDa. *ER $\alpha$*  deduced sequence was analyzed with Prosite and SMART program. Domains and motifs found in the sequence showed in Fig. 4.12 and table 4.1 AF1-containing modulatory domain was found at the N-terminus in estrogen alpha-type receptors at amino acid residues position 7-183 (E-value = 1.10e-04), Nuclear hormone receptors DNA-binding domain found at amino acid residues position 184-243 (score = 19.144), zinc finger region (NR C4-type) found at amino acid residues position 187-207 and 223-242, P box directly interact with regulatory region of DNA found at amino acid residues position 205-209, D box involved in receptor dimerization found at amino acid residues position 224-228, and ligand binding domain of hormone receptors (E-value = 1.94e-33) were found at amino acid residues position 353-524.

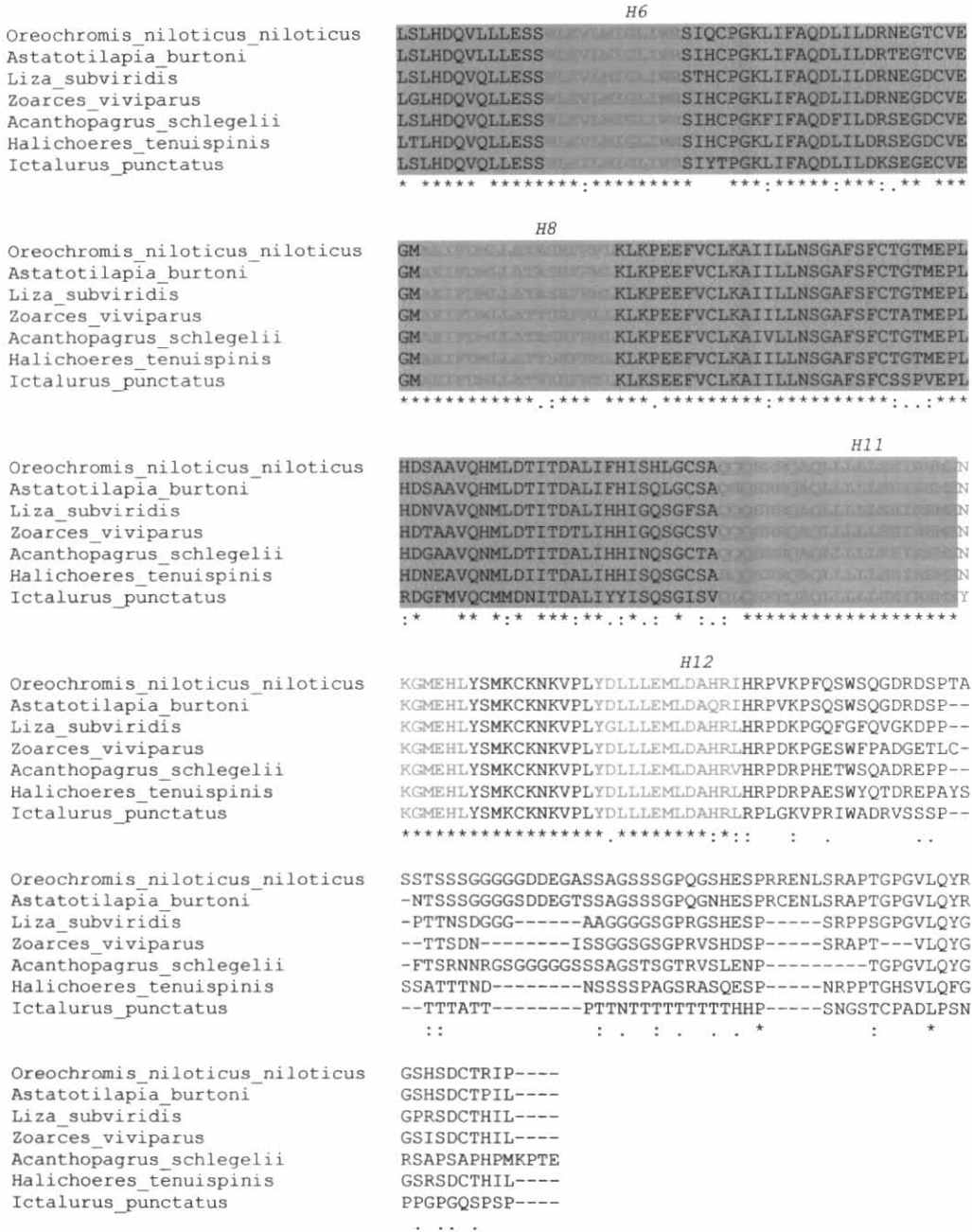
**Table 4.1** Domains, motifs, and consensus patterns of ER $\alpha$ 

Domain	Site	Consensus pattern
AF1-containing modulatory domain	7-183	Poorly conserved region which spans the first 180 residues and contains the activation function 1 (AF1) region ( <a href="http://pfam.janelia.org/cgi-bin/getdesc?name=Oest_recep">http://pfam.janelia.org/cgi-bin/getdesc?name=Oest_recep</a> )
DNA-binding domain	184-243	C - x(2) - C - x(1,2) - [DENAVSPHKQT] - x(5,6) - [HNY] - [FY] - x(4) - C - x(2) - C - x(2) - F(2) - x - R The 4 C's are zinc ligands ( <a href="http://br.expasy.org/prosite">http://br.expasy.org/prosite</a> )
P box	205-209	EGCKA (Attwood, Kroll, and Denslow, 2004)
D box	224-228	PATNQ (Attwood, Kroll, and Denslow, 2004)
zinc finger region (NR C4-type)	187-207, 223-242	C - x(2) - C - x(1,2) - [DENAVSPHKQT] - x(5,6) - [HNY] - [FY] - x(4) - C - x(2) - C - x(2) - F(2) - x - R ( <a href="http://br.expasy.org/prosite">http://br.expasy.org/prosite</a> )
ligand binding domain	353-523	Contained helices H3, H6, H8, and H11 surrounding the hydrophobic cavity in tertiary structure (Menuet et al, 2002)

MYKRQSPAQSRQPCGPV**LKPRNSPASSELETLSPSRPSPPRAPLGD**MYPEESRGSAGTATVDFLEGTYDYAAPSPAPTPLYSH  
**STTGYYSA**PLDAHGPPSDGSLQSLGSGPTSPLVFPSSPRLSPFMHPPSHHYLETTSTPVIYRSSHQPLSREEQCQCGARDEAYAS  
**GELGAGAGGFEMTKE**TRFCAVCSDYASGYHYGVWVSC**EGCKA**FFKRSIQGHNDYMC**PATNQ**CTIDRNRKSCQACRLRKCYEVGM  
MKGGRKDRGRVLRDRKRRAGDRDKVAKDPEHKASAPPQDGRKRSSSCSSTGGGGSSVTNLPPDQVLLLLQGAEPMLCSRQ  
KLSRPYTEVTMTLLT**SMADKELVHMIAWAKKLP**GFQLSLHDQVQLLESSWLEVLMIGLIWRSTHCPGKLI**FAQDL**LDRNEG  
DCVEGMAEIFDMLLATASRFRLKPEFVCLKAIILLNSGAFS**FCTGTME**PLHDNVAVQNMLDTITDALIHIGQSGFSAQ**Q**  
**QSRQAQ**LLLLL**SHIRHMS**NKGMEHLYSMCKNKVPLYGLLLEMLDAHRLHRPDKPGQFGFQVGKDPPTTNSDGGGAAGGGGS  
GPRGSHEPSRPPSGVQLQYGGPRSDCTHIL

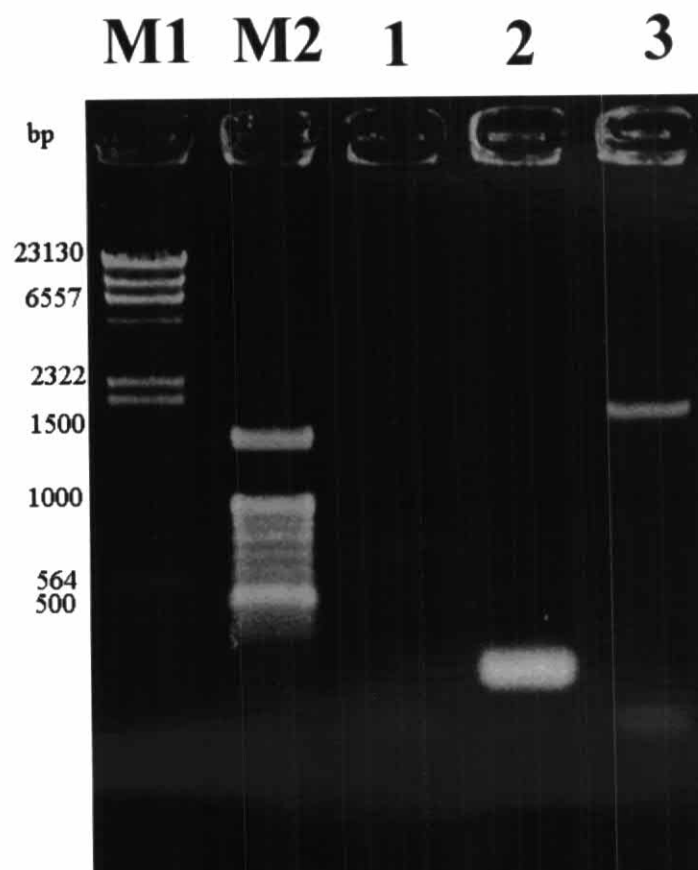
**Figure 4.12** ER $\alpha$  of *L. subviridis* showed important domains and motifs. Green label is AF1-containing modulatory domain, yellow label is nuclear hormone receptors DNA-binding domain, underline is zinc finger region (NR C4-type), blue alphabet is P box, red alphabet is D box and blue label is ligand binding domain of hormone receptors. ER $\alpha$  deduced sequence aligned with ER $\alpha$  amino acid sequences of several fish species was shown in Fig. 4.13





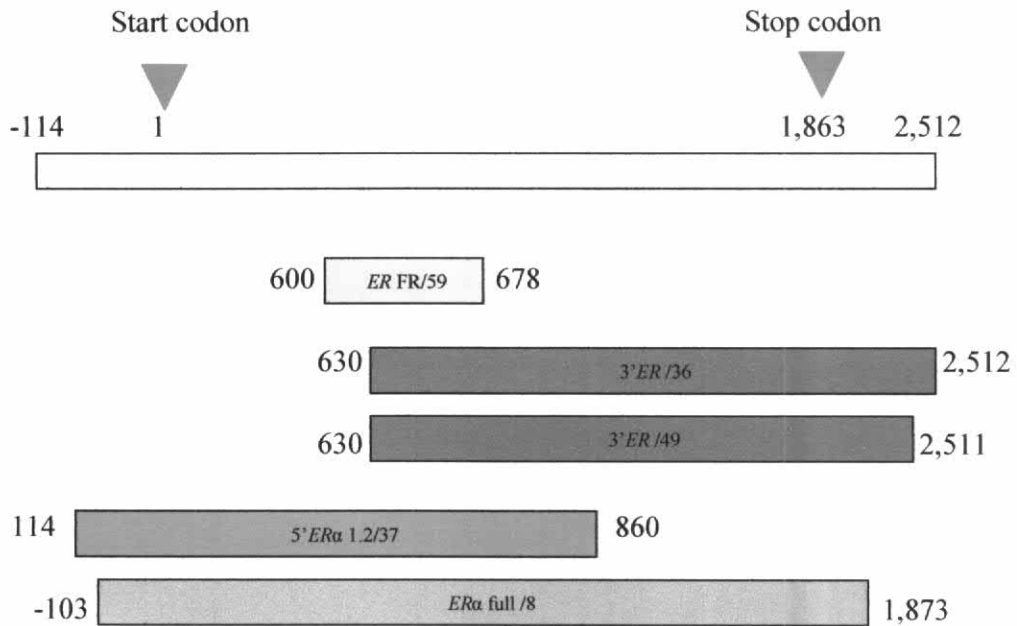
**Figure 4.13** Alignment of ERα amino acid sequences of *L. subviridis* and other fish species. DNA binding domain labeled with yellow containing 2 zinc finger regions (underline), red alphabet is P box (EGCKA), and D box (PATNQ) labeled with pink. Ligand binding domain is labeled with blue. Pink alphabet is Helices H3, H6, H8, and H11 surrounding the hydrophobic cavity in tertiary structure.

Results in Fig. 4.13 showed DNA binding domain contained P box (EGCKA) directly interact with regulatory regions of DNA, and D box (PATNQ) involved in receptor dimerization, and ligand binding domain of *L. subviridis* ER $\alpha$  which was highly conserved with these regions of other fish ER $\alpha$ . Helices H3, H6, H8, and H11 surrounding the hydrophobic cavity in tertiary structure of *L. subviridis* ER $\alpha$  which was highly conserved with these regions of other fish ER $\alpha$ . Open reading frame (ORF) of ER $\alpha$  was amplified using primers designed from start and stop codon sequences (Fig. 4.11). A DNA fragment at the size of 1.9 kb was obtained from the amplification in *L. subviridis* liver (Fig. 4.14)



**Figure 4.14** PCR product of *ERα* ORF separated in 1.2% agarose gel. Lane M1, M2, 1, 2, and 3 is  $\lambda$ *Hind*III, 100 bp DNA marker, negative control, and positive control and *ERα* ORF PCR product, respectively.

BLAST analysis of *ERα* ORF indicated that it was homologous with estrogen receptor alpha of largemouth bass (*Micropterus salmoides*) at the E-value of 4e-20. Structure of full length of *ERα* gene which shown position of cDNA sequence of *ERα* gene obtained from this study shown in Fig.4.15

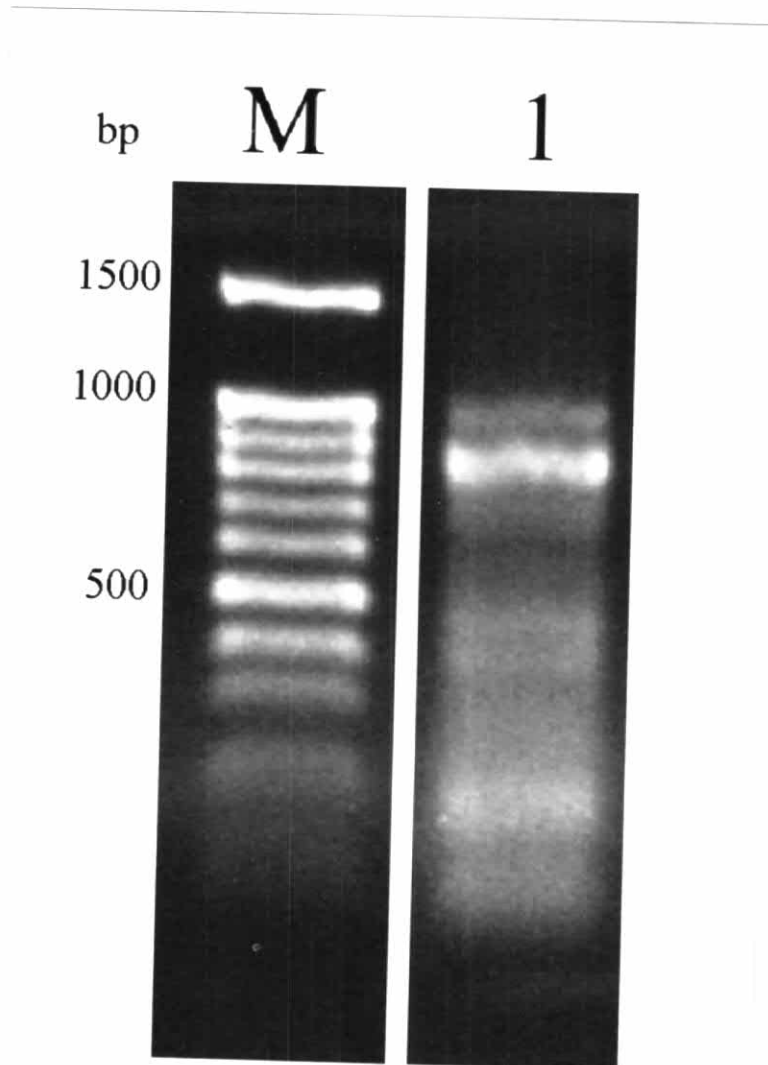


**Figure 4.15** Schematic representation of the full length of *ERα* gene. Yellow box is partial coding sequence of *ERα* amplified by degenerate PCR, green box is 3'end cDNA sequence of *ERα*, blue is 5'end cDNA sequence of *ERα*, and purple box is full length ORF of *ERα*.

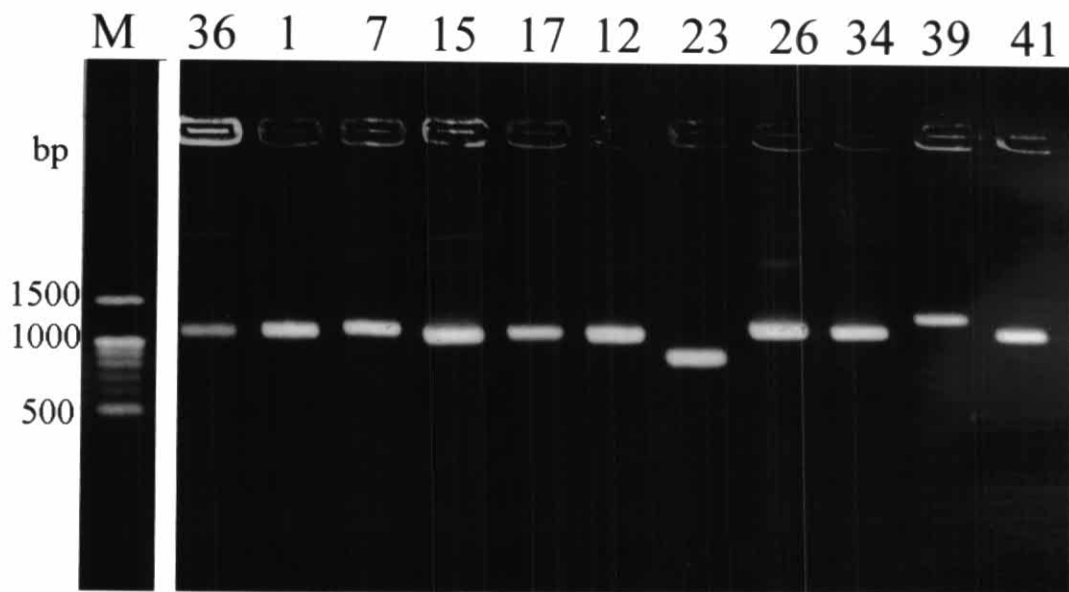


#### 4.3.4 Amplification of 5' and 3'cDNA end of *ERβ* by RACE-PCR

DNA fragment at the size of 850 bp was obtained from 5'RACE PCR amplification of *ERβ* in liver (Fig.4.16). The band was cloned. Several white colonies were checked for the insert size by colony PCR. Colony PCR products of 5'RACE *ER* were shown in Fig. 4.17. Clone No.36 (insert size of 850 bp), 23 (insert size lower than 850 bp) and 39 (insert size higher than 850 bp) were cultured for plasmid extraction, sequencing and BLAST analysis indicated that the sequences of 5'*ER*/23, 36 and 39 were homologous with *ERβ* of *Micropterus salmoides* with E-value of 7e-111, 3e-145 and 5e-141, respectively (see in appendix C).

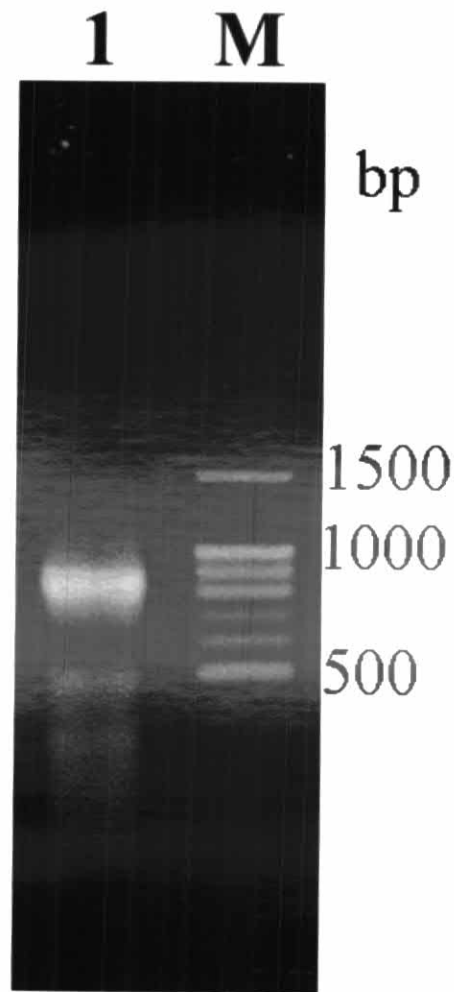


**Figure 4.16** 5'RACE PCR product of *ERβ* separated in 1.2% agarose gel. Lane M is 100 bp DNA marker, lane 1 is 5'RACE PCR product of *ERβ*.



**Figure 4.17** colony PCR product of 5' RACE *ER* separated in 1.2% agarose gel. Lane M is 100 bp DNA marker, number of lane represent number of clone.

Because of *ER* FR/59 sequence used for designing *ER* R-RACE primer was DNA binding domain which was very highly conserved region between *ER* $\alpha$  and *ER* $\beta$ . 5' RACE PCR product of *ER* could amplify 2 isoforms. Therefore, 5'RACE PCR amplification of *ER* in liver was again which used BD Advantage 2 Polymerase Mix (Clontech). DNA fragment at size of 850 bp was obtained from 5'RACE PCR amplification in liver (Fig. 4.18).



**Figure 4.18** 5'RACE PCR product of *ER* amplified by BD Advantage 2 Polymerase Mix (Clontech) separated in 1.2% agarose gel. Lane M is 100 bp DNA marker, lane 1 is 5'RACE PCR product of *ER* amplified by BD Advantage 2 Polymerase Mix (Clontech).



```

5'ER-36          TGGCCGTCCCACAGTCACGCCAGCATGCCTTCGCTCACGCTGCACTGCACCCAGCCGCTG
5'-ER-900-14    TGGCCGTCCCACAGTCACGCCAGCATGCCTTCGCTCACGCTGCACTGCACCCAGCCGCTG
5'-ER-23        TGGCCGTCCCACAGTCACGCCAGCATGCCTTCGCTCACGCTGCACTGCACCCAGCCGCTG
5'-ER-39        TGGCCGTCCCACAGTCACGCCAGCATGCCTTCGCTCACGCTGCACTGCACCCAGCCGCTG
*****

5'ER-36          GTCTACAACGAACCCAACCTCACACTCAGGCTGGCTGGAGCCCAAAGTCCACAGCATCAAC
5'-ER-900-14    GTCTACAACGAACCCAACCTCACACTCAGGCTGGCTGGAGCCCAAAGTCCACAGCATCAAC
5'-ER-23        GTCTACAACGAACCCAACCTCACACTCAGGCTGGCTGGAGCCCAAAGTCCACAGCATCAAC
5'-ER-39        GTCTACAACGAACCCAACCTCACACTCAGGCTGGCTGGAGCCCAAAGTCCACAGCATCAAC
*****

5'ER-36          CCCAGCAGCGCCGTCATCAGCTGTAACAAGCTGCTGGGGAAGAAATCGGAGGACGGAGTG
5'-ER-900-14    CCCAGCAGCGCCGTCATCAGCTGTAACAAGCTGCTGGGGAAGAAATCGGAGGACGGAGTG
5'-ER-23        CCCAGCAGCGCCGTCATCAGCTGTAACAAGCTGCTGGGGAAGAAATCGGAGGACGGAGTG
5'-ER-39        CCCAGCAGCGCCGTCATCAGCTGTAACAAGCTGCTGGGGAAGAAATCGGAGGACGGAGTG
*****

5'ER-36          GAGAGGGCGAAGGAGTCCCTCGTGTTCGTCGGCAGCAGGAAAGCCGACATGCACCTCTGC
5'-ER-900-14    GAGAGGGCGAAGGAGTCCCTCGTGTTCGTCGGCAGCAGGAAAGCCGACATGCACCTCTGC
5'-ER-23        GAGAGGGCGAAGGAGTCCCTCGTGTTCGTCGGCAGCAGGAAAGCCGACATGCACCTCTGC
5'-ER-39        GAGAGGGCGAAGGAGTCCCTCGTGTTCGTCGGCAGCAGGAAAGCCGACATGCACCTCTGC
*****

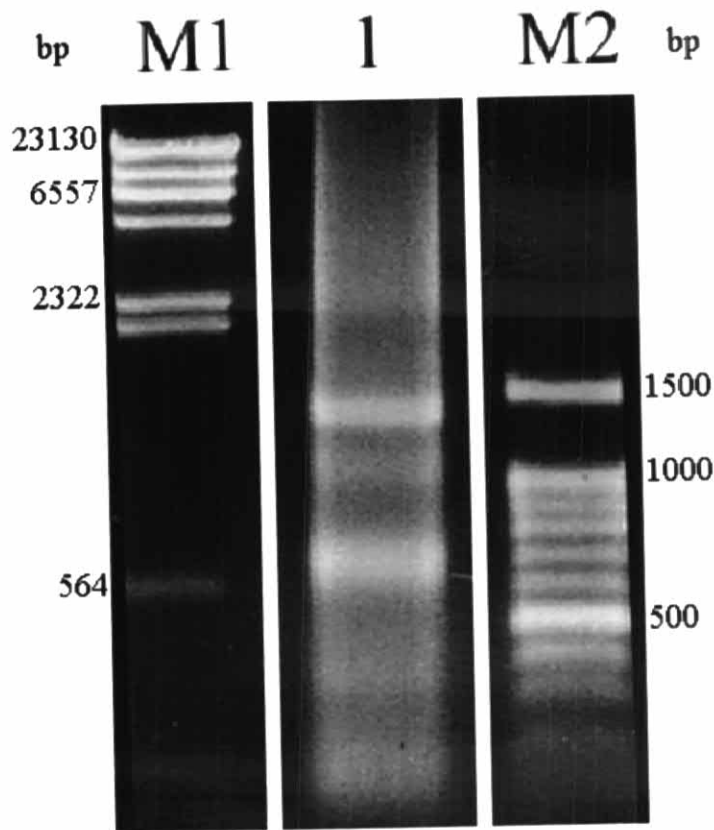
5'ER-36          GCCGTGTGCCACGACTACGCGTCCGGGTACCACTACGGCGTCTGGTCTGTGAGGGCTGC
5'-ER-900-14    GCCGTGTGCCACGACTACGCGTCCGGGTACCACTACGGCGTCTGGTCTGTGAGGGCTGC
5'-ER-23        GCCGTGTGCCACGACTACGCGTCCGGGTACCACTACGGCGTCTGGTCTGTGAGGGCTGC
5'-ER-39        GCCGTGTGCCACGACTACGCGTCCGGGTACCACTACGGCGTCTGGTCTGTGAGGGCTGC
*****

5'ER-36          AAGGCCTTCTTCAAGAGGAGCATCCAGGGTCACAATG
5'-ER-900-14    AAGGCCTTCTTCAAGAGGAGCATCCAGGGTCACAATG
5'-ER-23        AAGGCCTTCTTCAAGAGGAGCATCCAGGGTCACAATG
5'-ER-39        AAGGCCTTCTTCAAGAGGAGCATCCAGGGTCACAATG
*****

```

**Figure 4.19** Alignments of 4 types of 5'cDNA end of *ERβ*. Red is start codon

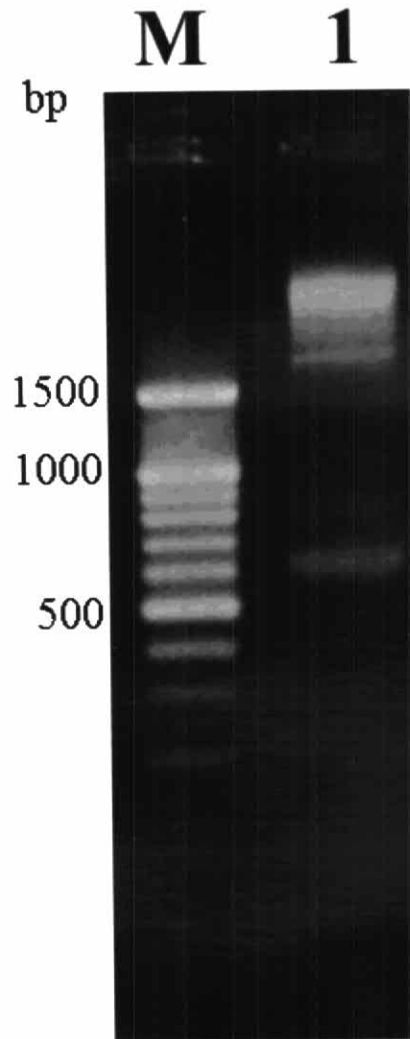
Results in Fig. 4.19 showed almost identical in coding sequences but differences in 5'UTR sequences of 5'cDNA ends between 4 types of *ERβ*. Further amplification of was conducted by RACE PCR using *ERβ* F RACE primer designed from the obtained 5'cDNA sequence of *ERβ*. DNA fragments at the size of 1.3 kb and 600 bp were obtained (Fig. 4.20). After sequencing (see in appendix B) and BLAST analysis (see in appendix C), these PCR products were homologous with *ERβ* of Atlantic croaker (*Micropogonias undulatus*) and largemouth bass (*Micropterus salmoides*) with E-value of 0.0 and  $3e-32$ , respectively.



**Figure 4.20** 3' RACE PCR product of *ERβ* separated in 1.2 % agarose gel. Lane M1, M2 and 1 are  $\lambda$ *Hind*III, 100 bp DNA marker and 3' RACE PCR product of *ERβ*, respectively.

*ERβ* F1-RACE primer designed from 3'*ERβ* 1.3/33 sequence for used in 3' RACE PCR amplification. DNA fragments at size 600 bp and large than 1.5 kb were obtained from 3' RACE PCR amplification (Fig. 4.21).





**Figure 4.21** 3' RACE PCR product of *ERβ* separated in 1.2 % agarose gel. Lane M and 1 are 100 bp DNA marker and 3'RACE PCR product of *ERβ*, respectively.

A DNA fragment at size 600 bp was cloned. After sequencing (see in appendix B) and BLAST analysis (see in appendix C), it was homologous with estrogen receptor beta b of *Astatotilapia burtoni* with E-value of  $1e-26$ . 3' cDNA end sequences of *ERβ* were aligned as shown in Fig. 4.22 and 4.23

```

3'ERbeta600-38      TGACTACATTTGCCCGCCACAAATCAGTGCACATATCGACAAGAATCGGCGTAAAAGCTG
3'ERbeta1.3-33      TGGCTACATTTGCCCGCCACAAATCAGTACACTATCGGCAAGAATCGGCGTAAAAGCTG
** *****

3'ERbeta600-38      CCAAGCCTGCCGCTGCGGAAATGCTACGAAGTGGGCGTGATGAAGGGCTAGCTGCCAA
3'ERbeta1.3-33      CCAAGCCTGCCGCTGCGGAAATGCTACGAAGTGGGCGTGATGAAGTG--TGGCG--TGA
***** * * * *

3'ERbeta600-38      ACCTTAAATTGTCTGTCCAGGGAGGCACTAGCAAAGTATTGTTGCTTGGGGTTGATG
3'ERbeta1.3-33      GCGGTGAACGCTGCAGCTATCGAGGAGCTCG-ACACCG-TCGTGGT---GGAGTCCAGT
* * * * * * * * * * * * * * * * * * * * * * * * *

3'ERbeta600-38      GACACACCTTCACACACCTG---TCTCTGAGTATGAACACATGGTTTGAGACAAAAAAA
3'ERbeta1.3-33      CTCGGGACGCCACGGCCGGGGCTTGGTGAAGTTCGGCCCGGTTCTCGGGCCAGCGGC
* * * * * * * * * * * * * * * * * * * * * * * * *

3'ERbeta600-38      ATATCAAAGAAAGGCAGGAAAAATATAAATAAAATC--TGCTTAATGTTCAAAAAACA
3'ERbeta1.3-33      ATCTCGACCTGGGACCGCCCTG-TCCCGCTGGCCTCCCTGCCCCAGGCCAACCCCTG
** * * * * * * * * * * * * * * * * * * * * * * * * *

3'ERbeta600-38      CATT--TCACCATTGAGTTGATACAACT-GTCTTCTGTCTACGCAA-TAAACACACCA
3'ERbeta1.3-33      CACCCTCAGCCATGAGCCCGGAGGAGTTCATCTCCCGCATCATGGAGGCCGAGCCTCCA
** * * * * * * * * * * * * * * * * * * * * * * * * *

3'ERbeta600-38      CGGAACAATCCTTAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA-----
3'ERbeta1.3-33      GAGATCTACCTGATGGAGGACCTGAAGAAGCCGTTACCCGAGGCCAGCATGATGATGTCC
* * * * * * * * * * * * * * * * * * * * * * * * *

3'ERbeta600-38      -----
3'ERbeta1.3-33      CTCACCAACCTGGCAGACAAGGAGCTGGTCTCATGATCAGCTGGGCCAAAAAGATCCCT

3'ERbeta600-38      -----
3'ERbeta1.3-33      GGCTTCGTAGAGCTGAGTCTAGCAGATCAGATCCACCTGCTGAAGTGTCTGGCTGGAG

3'ERbeta600-38      -----
3'ERbeta1.3-33      ATCCTCATGCTGGGTCTGATGTGGAGTCTGTGGATCATCTGGAAACTCATCTTCTCT

3'ERbeta600-38      -----
3'ERbeta1.3-33      CCAGACTTCAAACCTCAACAGGGAGGAGGCCAGTGTGTGGAGGCATCATGGAGATCTTT

3'ERbeta600-38      -----
3'ERbeta1.3-33      GACATGCTGCTGGCGGCCAGTCTCGCTTCAGAGAGCTGATGCTGCAGAGGGAGGATAC

3'ERbeta600-38      -----
3'ERbeta1.3-33      GTCTGCCTGAAGCCATGATCCTCCTCAACTCCAGTGAAGTTAACACCTCAACTCCAGTG

3'ERbeta600-38      -----
3'ERbeta1.3-33      AGTTTAAACACGGACGATCAGCTGCTGCTACTTCACTGAGAAACTGACAAACAGCAGCA

3'ERbeta600-38      -----
3'ERbeta1.3-33      AACGTTCTGATTTAAATGAGCCACCACAGGCCACGCCACGCCAGAAGAGGGCGTGGCC

3'ERbeta600-38      -----
3'ERbeta1.3-33      TGTGGATGTGAGGACAAAAAACAAGGCCCGCCACATGTGAAATTATCTCTCATTGT

3'ERbeta600-38      -----
3'ERbeta1.3-33      CCATGTCTGTGGAGGACAGTTGTCTTAGTGGAGTCCCAGGCTCTGGTTGCCATGG

```

```

3'ERbeta600-38 -----
3'ERbeta1.3-33 TAACCCCATATAAACATGTCTAACTCTTCTTCTGGGTTTCAGCCTCCAAGCTGCTGTGTG

3'ERbeta600-38 -----
3'ERbeta1.3-33 CACGGTGGGATGGACGGTGTCTCCACCTTAAATAAACTTATAGTCCACCTTAAGTGTCAA

3'ERbeta600-38 -----
3'ERbeta1.3-33 TCAAAATGGATCGGTGGCTCCCATGGTCCACTTCAGCTTGGTACCGCCACTCTGTGTCT

3'ERbeta600-38 -----
3'ERbeta1.3-33 GGTCTCTCCCTTTAAAAGTCACTTTTAAATTGGTGAAAAAAGTTAAAGAAATAAGGGAA

3'ERbeta600-38 -----
3'ERbeta1.3-33 GGATATAAAATAAAAAAAAAAAAAAAAAAAAAA

```

**Figure 4.22** Alignment of 3'*ER* $\beta$  1.3-33 and 600/38. Blue and red alphabet shown nucleotides which different, green, blue and yellow label is stop codon, polyadenylation signal and polyA tail, respectively.

```

3'ER-beta-600-38 -----
3'ER-beta-600-1  GCGGCCACCTCTCGCTTCAGAGAGCTGAAGCTGCAGAGGGAGGAGTACGTCTGCCTGAAG

3'ER-beta-600-38 -----
3'ER-beta-600-1  GCCATGATCCTCCTCAACTCCAGTGAGTTAACACGGACGATCAGCTGATGCTGCTTCAC

3'ER-beta-600-38 -----CTTCAAGAGGAGCATCCAGGGTCACAATGACTAC-ATTTGCCCCG
3'ER-beta-600-1  TGAGAAACACTGACAACACGACGCAACGTTCTGATTTAAATGAGCCACCACAGGCCACG
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 CCA----CAAATCAGTGCACTATCGACAAGAATCGGCGTAAAAAGCTGCCAAGCCTGCC-
3'ER-beta-600-1  CCCACGGCCAGAAGAGGGCGTGGTCTGTGGATGTGAGGACAAAAACAAGGCCCCGCC
          **          * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 GCCTGCGGAAATGCTA-CGAAGTGGGCGTG----ATGAAGGC-----CTAGCTG
3'ER-beta-600-1  ACATGTGAAATTATCTCTCATGTGCCATGTCTCTGTGGAGGACAGTGTGTTCTCTAGTGG
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 ----CCAAACCTTAATGTCTGTCCAGGGAGGCACTAGCAAA-CTGATTGTTGTCTT
3'ER-beta-600-1  AGTCCCAGGCTCTGGTTGCCATGGTAACCCCATATAAACATGTCTAACCTTCTTCTG
          *** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 GG-----GGTTGATGGACACACCTTCACACAC-CTGTCTCTGAGTATGAACACA
3'ER-beta-600-1  GGTTCAGCCTCCAAGCTGTGTGTGTCACGGTGGGATGGACGGTGTCTCCACCTTAA-ATA
          **          * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 TGTTTTGAGACAAAAAAATATCAAAGAAAGGCAGGAAAAAAT---ATAAAATAAAATC
3'ER-beta-600-1  AACCTATAGTCCACCTTAAGTGTCAAACAATGGATCGGTGGCTCCCATGGTCCAGTTC
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 TGCTTAATGTT-CAAAAAACACATTTACCATTGAGTTGATACAACTGTCTTCTGTCTT
3'ER-beta-600-1  AGCTTGGTACCGCCACTCTGTGTCTGGTCTCTTCCCTT-TAAAAGTCACTTTAAATTG
          **** * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 ACGCAA-----TAAACACACCAGGAACAATCCTAAAAA
3'ER-beta-600-1  ATGGAAAAAAGTTAAAGAAATAAGGGAAGGATAATAAATAAAAAA
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

3'ER-beta-600-38 AAAAAAAAAA
3'ER-beta-600-1  AAAAAAAAAA
          *****

```

**Figure 4.23** Alignment of 3'ERβ 600/1 and 600/38

Results in Fig. 4.22 and 4.23 showed different in nucleotide sequences of 3'cDNA end of ERβ, but we not determined additionally sequence (see in Fig. 4.23).

#### 4.3.5 Full length cDNA sequence determination of ERβ

5'ER/36 and 3'ERβ 1.3/33 cDNA sequence were connected *in silico* and obtained full length mRNA of ERβ at size 2,098 bp which contained 5'UTR, ORF, and 3'UTR at size 177, 1,431, and 490 bp, respectively. Domains and motifs found in the sequence showed in Fig. 4.24 and table 4.2. ERβ ORF encoded ERβ contained 476 amino acid residues with the molecular weight at 52.36 kDa which contained A/B domain, DNA binding domain, D domain and ligand binding domain but lack F domain. DNA binding domain was found at 178-253 with score as 17.312. Two zinc finger regions were found at 181-201 and 217-236. P box was

found at 199-204. D box was found at 218-222. Ligand binding domain was found at 352-476 with E-value as  $6.80e-15$  as shown in Fig. 4.24.

Table 4.2 Domains, motifs, and consensus patterns of ER $\beta$

Domain	Site	Consensus pattern
DNA-binding domain	178-253	C - x(2) - C - x(1,2) - [DENAVSPHKQT] - x(5,6) - [HNY] - [FY] - x(4) - C - x(2) - C - x(2) - F(2) - x - R The 4 C's are zinc ligands ( <a href="http://br.expasy.org/prosite">http://br.expasy.org/prosite</a> )
P box	199-203	EGCKA (Attwood, Kroll, and Denslow, 2004)
D box	218-222	PATNQ (Attwood, Kroll, and Denslow, 2004)
zinc finger region (NR C4-type)	181-201, 217-236	C - x(2) - C - x(1,2) - [DENAVSPHKQT] - x(5,6) - [HNY] - [FY] - x(4) - C - x(2) - C - x(2) - F(2) - x - R ( <a href="http://br.expasy.org/prosite">http://br.expasy.org/prosite</a> )
ligand binding domain	352-476	Contained helices H3, H6, H8, and H11 surrounding the hydrophobic cavity in tertiary structure (Menuet et al, 2002)

```

acgcgggggaaagcggttgagccacagagcctggacaagaagtcttctctctgttcccg
atgctcgtgaggtcttgaagtcggcctctagtcggacgtgtccacagaagacgatgcagt
agacgaaccaggtgcatcgtcttattgtccatcacctccaagtttctctgtccctgacatg
gctcgtcccctgggtggatgcggaaccctgttaccctgcttcagctacaggaggtggac
A S S P G L D A D P L P L L Q L Q E V D
tccagtaaagccaccgagagggcggagctccccaggactcctgccggtcatgtacagcct
S S K A T E R P S S P G L L P V M Y S P
cctgtggcggtggacgccacacgctctgcatcccttctcgttacacggacagtagccac
P V G V D S H T V C I P S P Y T D S S H
gactacagccacggacatggacctctgaccttctacagtcctgcatgctcagctacacc
D Y S H G H G P L T F Y S P S M L S Y T
cgccgccatcactgacagcccatcctctgtgtccccctctcageccctctgccttt
R P P I T D S P S S L C P P L S P S A F
tggcgtcccacagtcagccagcatgccttcgctcacgctgcactgcacccagccgctg
W P S H S H A S M P S L T L H C T Q P L
gttacaacgaacccaactcactcaggtggctggagccaaagtccacagcatcaac
V Y N E P N S H S G W L E P K V H S I N
cccagcagcgcgtcatcagctgtaacaagctgctgggaagaaatcggaggacggagtg
P S S A V I S C N K L L G K K S E D G V
gagagggcgaaggagtctcgtgtcgtcggcagcaggaaagccgacatgcacttctgc
E R A K E S S C S S A A G K A D M H F C
gccgtgtgccacgactacgctcgggtaccactacggcgtctggtcctgtgagggctgc
A V C H D Y A S G Y H Y G V W S C E G C
aaggccttctcaagaggagcatccagggtcacaatggctacattgccccgccacaat
K A F F K R S I Q G H N G Y I C P A T N
cagtacactatcggcaagaatcggcgtaaaagctgccaagcctgcccctgcggaatgc
Q Y T I G K N R R K S C Q A C R L R K C
tacgaagtggcatgatgaagtgtggcgtgagcgtgaacgctgcagctatcagaggact
Y E V G M M K C G V R R E R C S Y R G A
cgacaccgctcgtggtggagtccagtctcgggacgccacgggcccgggcttgggaagtc
R H R R G G V Q S R D A T G R G L V K V
ggccccggttctcgggcccagcggcatctcgacctgggaccgccctgtccccgctggcc
G P G S R A Q R H L D L G P P L S P L A
tccctgccccaggccaaccactgcaccactcagccatgagccggaggagttcatctcc
S L P Q A N H L H H S A M S P E E F I S
cgcatcaggagccgagcctccagagatctacctgatggaggacctgaagaagccttc
R I M E A E P P E I Y L M E D L K K P F
accgaggcagcatgatgtccctcaccacactggcagacaaggagctggtgctcatg
T E A S M M S L T N L A D K E L V L M
atcagctgggcaaaaagatccctgcttagagctgagcttagcagatcagatccac
I S W A K K I P G F V E L S L A D Q I H

```

```

ctgctgaagtgctgctggctggagatcctcatgctgggtctgatgctggaggtctgtggat
L L K C C W L E I L M L G L M W R S V D
catcctggaaaactcatctctcctcagacttcaaaactcaacagggaggaggccaggtg
H P G K L I F S P D F K L N R E E G Q C
gtggaggcatcatggagatctttgacatgctgctggcgccacgtctcgtctcagagag
V E G I M E I F D M L L A A T S R F R E
ctgatgctgcagaggaggagtacgtctgctgaaggccatgatcctcctcaactccagt
L M L Q R E E Y V C L K A M I L L N S S
gagtttaaacactcaactccagtgagtttaacacggacgatcagctgagtgtacttcaact
E F N T S T P V S L T R T I S
gagaaactgacaacacagcagcaaacgttctgatttaaatgagccaccacagccacgc
ccacggccagaagagggtggtcctgtggatgtgaggacaaaaaacaaggccccgccca
catgtgaaattattctctcattgtcctgtctctgtggaggacagttgttctctagtga
cgtcccaggtctggttgccatggttaaccccatataaacatgtctaactcttctctgg
gttcagcctccaagctgctgtgtcaggtgggatggacggtgtctccacctaaataaa
cctatagtccaccttaagtgtcaatcaaatggatcgggtgctcccattggtccactcag
cttggtaaccgccactctgtctgtgctctcctcctttaaagtcacttttaaatgggtg
aaaaagttaaagaataagggaaggataataataaaaaaaaaaaaaaaaaaaaaaaaaa
  
```

**Figure 4.24** Full length mRNA and deduced sequence of *ERβ*. Pink label, yellow label, blue alphabet, red alphabet, blue label, and green label is start codon, DNA binding domain, P box, D box, ligand binding domain, and stop codon, respectively.

*ERβ* sequence of *L. subviridis* was aligned with *ERβ* of other fish species. The result was shown in Fig. 4.25

<p>AAG16711 AAO39210 AAP72179 Liza subviridis ABC68616</p>	<p>MASSPGLDPHPLMLQLQEVGSSKVSERPRSPGLLPAVYSPPLGMSHTVCIPSPYTDSS MASSPGLDADPLPLLQLQEVDSKASQRPSSPGLLPAVYSPPLGMSHTVCIPSPYTDSS MASSPGLDTPDLLLRLQEVDSKASERSSPGLLPAVYSPRQGMDSHTVYIPSPYTDNN MASSPGLDADPLPLLQLQEVDSKATERPSSPGLLPMVYSPVGVSDHTVCIPSPYTDSS MASSPGLNAEPLMLQLQEAYSSKPSERPTSPGLLPTMYSPLGIDSHTVCIIPSPYTDSS *****:..***:*.***. *** ::* *****.:*** *:***** *****..</p>
<p>AAG16711 AAO39210 AAP72179 Liza subviridis ABC68616</p>	<p>HEYNHSHGPLTFYSPSVLSYRPPITNSPSSLCPLSPSAFWPSPHNHPTMPSLTLHCPES HEYNHSHGPLTFYSPSVLSYARPPITDSPSSLCPLSPSAFWPSPHSHNMPSLTLRCPQP QEYNHSGSVSFYSPSVLSYARPSATDSPSSLCGPLSPSAFWPSPHSHNMPSLTLRCPQP HDYSHGHGPLTFYSPSMLS YTRPPIITDSPSSLCPLSPSAFWPSPHSHNMPSLTLHCTQP HDYNHGHGPLTFYGPSVLSYRPPITDSPTS LCPSPSAFWSSHSHHNVPVSLTLHCTQP :*. *. * :*:***:***:***. *:***:*** . *****.:* :*****:*.:</p>
<p>AAG16711 AAO39210 AAP72179 Liza subviridis ABC68616</p>	<p>IVYNEPSPHAWLESKAHNSINASSSIIIGCNKSLVKRSEEGVEDMNSSLCSSAVGKADMH LVYNEPSPHAWPEPKAHSINPSSS-ILGCNKPLGKRLEEGVEGVNSSLCSSAVGKADMH LGYNESGLHAWLESKPHNISSSSS-IIGCNKPLGKRSEEGVNGVNPVSLCSSVVGKADMH LVYNEPNSHSGWLEPKVHSINPSSA-VISCNKLLGKKEDEGVERAKESSCSSAAGKADMH LVYSEPGHPAWLNPKAHSINPSSS-VISCNRLGKPKDEGVEGVKSSSCSSAAGKADMH :*. * . * :*. * * . * . * :*:***: * * :*:*** : * * *. *****</p>
<p>AAG16711 AAO39210 AAP72179 Liza subviridis ABC68616</p>	<p>FCVACHDYASGYHYGVWSC<b>EGCKA</b>FFKRSIQGHNDYIC<b>PATN</b>CTIDKNRRKSCQACRLR FCVACHDYASGYHYGVWSC<b>EGCKA</b>FFKRSIQGHNDYIC<b>PATN</b>CTIDKNRRKSCQACRLR FCVACHDYASGYHYGVWSC<b>EGCKA</b>FFKRSIQGHNDYIC<b>PATN</b>CTIDKNRRKSCQACRLR FCVACHDYASGYHYGVWSC<b>EGCKA</b>FFKRSIQGHNDYIC<b>PATN</b>CTIDKNRRKSCQACRLR FCVACHDYASGYHYGVWSC<b>EGCKA</b>FFKRSIQGHNDYIC<b>PATN</b>CTIDKNRRKSCQACRLR ** *****</p>
<p>AAG16711 AAO39210 AAP72179 Liza subviridis ABC68616</p>	<p>KCYEVGMMKCGVRRERCSYRGARHRRGGLOPRDPTGRGLVVRVGLGSRAQRHLHLEAPLTP KCYEVGMMKCGVRRERCSYRGARHRRGGLOPRDPTGRGLVVRVGLGSRAQRHLHLEAPLAP KCYEVGMMKCGVRRERCSYRTRHRRGGLOPRDPTGRGLVVRVGLGSRAQRHLHLEAPLTP KCYEVGMMKCGVRRERCSYRGARHRRGGVQSRDATGRGLVKGVPGSRAQRHLDLGPPPLSP KCYEVGMMKCGVRRERCSYRGARHRRGGLOPR----GLVRIIGISRAQRLLPIELPFSS *****:*****:*****.:* * ***** : * ::</p>

```

AAG16711          LAPILQAKHVHLSAMSPEEFISRIMDAEPPEIYLME DLK KPFTEASMMMSLT
AAO39210          LTSLPQANHVHPSAMSPEEFISRIMEAEPPEIYLMEDMKKPFTEASMMMSLT
AAP72179          VTPLPQMSHVHHAAMSPEEFIMRIMEAEPPEIYLMEEQK KPFTEASMMMSLT
Liza subviridis  LASLPQANHLHHSAMSPEEFISRIMEAEPPEIYLME DLK KPFTEASMMMSLT
ABC68616          LVPVTQPN--HQSTMSPEEFISRIMEAEPPEIYLMDDLK KPFTEASMMMSLT
:..: * . * :***** **:*:*****: *****

H3

AAG16711          KIPGFVELSLADQINLLKCC          SVDHPGKLIFSPDFKLNREEG
AAO39210          KIPGFVDLSLADQIHLLKCC          SVDYPGKLIFSPDFKLNREEG
AAP72179          KIPGFVELCLADQIHLLKCC          SVDHPGKLIFSPDFKLNREEG
Liza subviridis  KIPGFVELSLADQIHLLKCC          SVDHPGKLIFSPDFKLNREEG
ABC68616          KIPGFVELSLADQIHLLKCC          SVDHPGKLIFSPDFKLNREEG
:*****:*.*****:*****:*****:*****

H6

AAG16711          QCVEGI          KLQREEYVCLKAMILLNSNLCTSSPQTAEELSRNKI
AAO39210          QCVEGF          KLQREEYVCLKAMILLNSNLCTSSPQTAEELSRNKI
AAP72179          QCVEGI          KLQREEYVCLKAMILLNSNLCTSSPQTAEELSRNKI
Liza subviridis  QCVEGI          KLQREEYVCLKAMILLNS-----
ABC68616          QCVEGI          KLQREEYVCLKAMILLNSNLCTSSPQTAEELSRNKI
*****:*****.***** *****

H8

AAG16711          LRL LDSVIDALVWAI SKMGLTT          NKGMDHLSTMKRKNVVL
AAO39210          LRL LDSVIDALVWAI SKLGLST          NKGMDHLSTMKRKNVVL
AAP72179          LRL LDSVIDALVWAI SKLGLST          NKGMDHLSTMKRKNVVL
Liza subviridis  -----SEFNTSTP-----VSLTRTIS-----
ABC68616          LRL LDSVIDALVWAI SKLGLST          NKGMDHLSTMKRKNVVL
*..: *          : * * : *

H11

AAG16711          VYDLLEMLDANTSSGGS-QPSSS-PSSETYS DQH QYPQPPSHLHPGSEQTTADHAI VPP
AAO39210          VYDLLEMLDANTSSGGS-QPSSS-PSSDTYS DQH QYPQPPSHLQPSDQTDGADHTTVPP
AAP72179          VYDLLEMLDANTSSGSSQSSSS-PNSDSYS DLH QYPQHPSHLQP--HQTADHNNMPA
Liza subviridis  -----
ABC68616          VYNLLEMLDANTSSSSSQTASSSPSSDTYS DGL QYPPSFHLQPGSDIVTPHPSTNSF

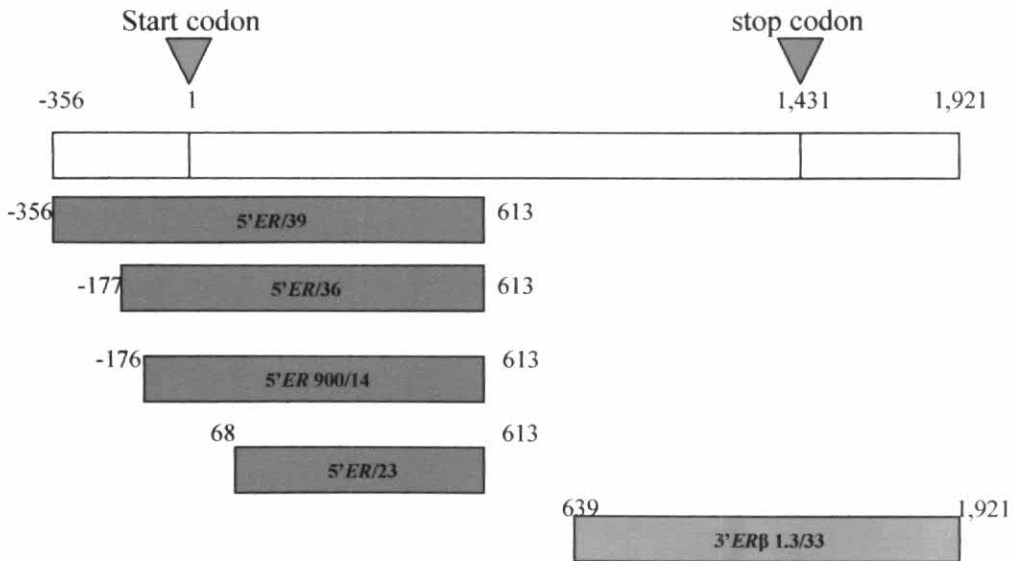
AAG16711          LGPTDDPILDGHL DAMPLQSSPPFQSLVVP HMDTNDYIHPEQ-WSLGTGDAAPSVEPTDY
AAO39210          HEPAEAPILDGHLQTL LHSSPPFQSLVVAHIDSNDYIHQEQ-WSLDTADAGPVEPTDY
AAP72179          HRQAEGQILEEELHTLPLQSSPPFHSQMATHMDRNEYVHPQHWSMDAEDAGVSVG--SY
Liza subviridis  -----
ABC68616          HEPPQEPAKDRTVGG-TFPSTPSPQTLVGSQIDSDDYI PAEH-WSLDAEDGGSSVEPVSY

AAG16711          ITTERVVMETALVTQP-
AAO39210          IVSDRVVMETALG---
AAP72179          MTSDRGVMEGALEVAGL
Liza subviridis  -----
ABC68616          IISDRVVMETTLEG---

```

**Figure 4.25** Alignment of ER $\beta$  amino acid sequence of *L. subviridis* and other species of fish. DNA binding domain is labeled with yellow. Two zinc finger regions indicate with underline, red alphabet is P box, D box is labeled with pink, ligand binding domain is labeled with blue, and helices H3, H6, H8, H11, and H12 surrounding the hydrophobic cavity in tertiary structure is pink alphabet.

Results in Fig. 4.25 showed highly conserved of amino acid sequences in DNA binding domain and ligand binding domain of fish ER $\beta$ . *L. subviridis* ER $\beta$  lack some region of ligand binding domain, helices H11 and H12 surrounding the hydrophobic cavity in tertiary structure and F domain. Structure of full length of ER $\beta$  gene which shown position of cDNA sequence of ER $\beta$  gene obtained from this study shown in Fig.4.26



**Figure 4.26** Schematic representation of the full length of ER $\beta$  gene. Green box is 5'end cDNA sequence of ER $\beta$ , blue box is 3'end cDNA sequence of ER $\beta$ .

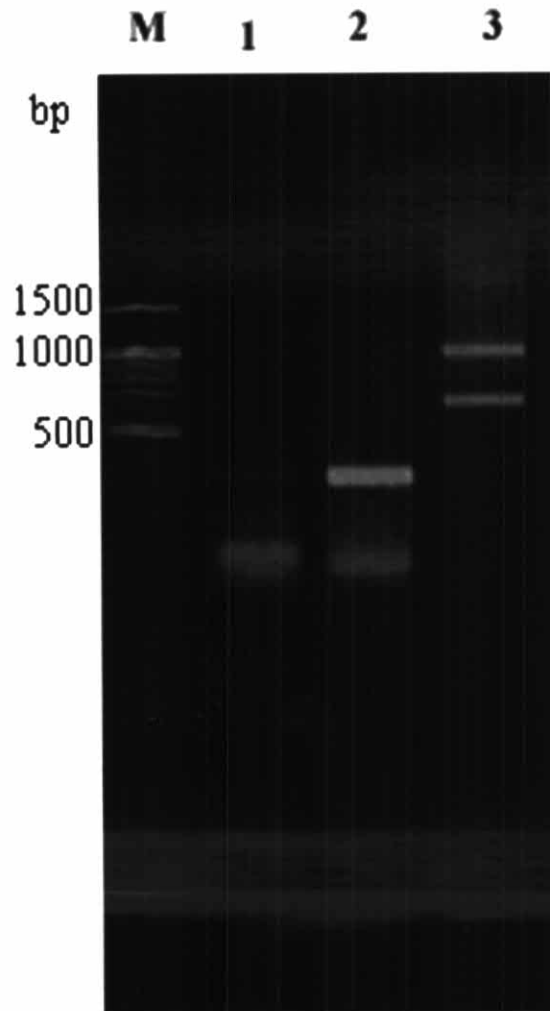
#### 4.4 Cloning and characterization of choriogenin (*chg*) genes

##### 4.4.1 choriogenin L (*chg*-L)

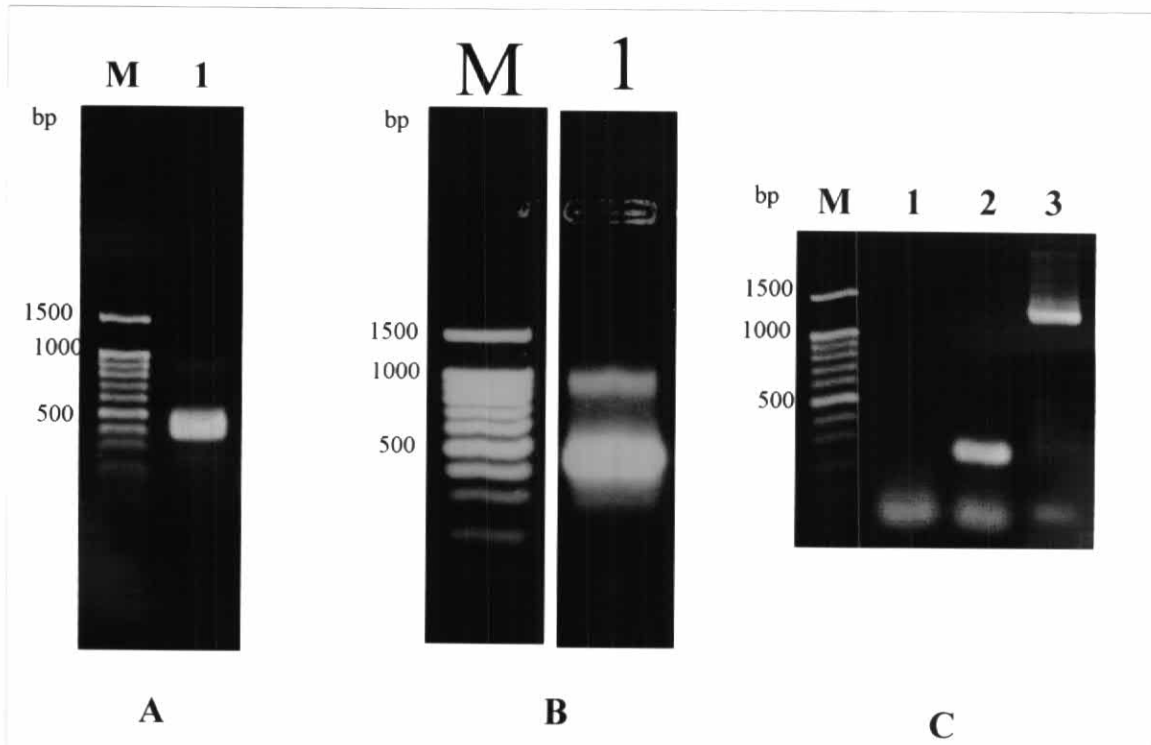
Partial cDNA sequence of *chg*-L gene was amplified from 1<sup>st</sup> strand cDNA templates of the fish liver. PCR products were determined by electrophoresis on 1.2% agarose (w/v) (Fig.4.27). DNA fragments at the sizes of 900 and 700 bp were obtained from PCR amplification. PCR product at the size of 900 bp which was close to the expected size of the product (921 bp) was eluted, reamplified, and determined by electrophoresis on 1.2% agarose (w/v). After sequencing (see in appendix B) and BLAST analysis (see in appendix C), the result indicated that this PCR product was homologous with chorion protein of gilthead seabream (*Sparus aurata*) with E-value of 2e-108. Further amplification was conducted by 5' and



3'RACE PCR using F RACE primer and GW R primer designed from the obtained partial sequence of *chg-L*. Two DNA fragments at the same size of 450 bp were obtained from 5' (Fig. 4.28 A) and 3' (Fig. 4.28 B) RACE PCR amplification. After sequencing of 2 different sizes of insert of 5' RACE PCR products of *chg-L* (5'L 500/12 and 18) and 3'RACE PCR product of *chg-L* (3'L 500/19 and 37) (see in appendix B) and BLAST analysis, these sequence were homologous with chorion protein of gilthead seabream (*Sparus aurata*), chorion protein of gilthead seabream (*Sparus aurata*), choriogenin L of Javanese rice fish (*Oryzias javanicus*), and choriogenin L of Javanese rice fish (*Oryzias javanicus*) with E-value as  $3e-14$ ,  $4e-22$ ,  $1e-04$ , and  $1e-04$  (see details in appendix C). Two clones of 5' and 3' cDNA ends of *chg-L* were aligned as shown in Fig. 4.29 and 4.30, respectively.



**Figure 4.27** *chg-L* PCR products separated in 1.2 % agarose gel. Lane M, 1, 2 and 3 are 100 bp DNA marker, negative control, positive control and *chg-L* PCR product, respectively.



**Figure 4.28** RACE PCR products of *chg-L*. 5' and 3' RACE PCR products of *chg-L* are shown in (A) and (B), full length (ORF) PCR product of *chg-L* is shown in (C). Lane M indicates 100 bp DNA markers, lane 1 of (A) and (B) is the products, lane 1, 2, and 3 of (C) is negative control, positive control and the products, respectively.

```

5'L500/12   ACGCGGGGACAGCACCTTGAGAATCTCTCAGATCGCTTGTCACTGTGGAGCCATGGTGAT
5'L500/18   ACGCGGGGACAGCACCTTGAGAATCTCTCAGATCGCTTGTCACTGTGGAGCCATGGTGAT
*****

5'L500/12   GAAGTGGACTGCTGCTTGCCTTGTGGCACTGGCTCTATTTGCCAGCGTCTGTGATGCTCA
5'L500/18   GAAGTGGACTGCTGCTTGCCTTGTGGCACTGGCTCTATTTGCCAGCGTCTGTGATGCTCA
*****

5'L500/12   GTGGGGAGAGTACACGCCTTCAAAATATCAGAAACCTGCACCTCCTGTGAAGCAAGAGCC
5'L500/18   GTGGGGAGAGTACACGCCTTCAAAATATCAGAAACCTGCACCTCCTGTGAAGCAAGAGCC
*****

5'L500/12   CAAACAAGTGCCTCAAGACACTCAACAGCATAAGCAGACATTGAAACACCACCTTCAATG
5'L500/18   CAAACAAGTGCCTCAAGACACTCAACAGCATAAGCAGACATTGAAACACCACCTTCAATG
*****

5'L500/12   GACATACCCTGAACCTCCCCGCCTGAACCTGCGCCTGAAATACCATTTGAACCCTACG
5'L500/18   GACATACCCTGAACCTCCCCGCCTGAACCTGCGCCTGAAATACCATTTGAACCCTACG
*****

5'L500/12   TCCTCAACCTGTTGCATCTGTGCTGTTGAGTGCAGAGAGAATGATGCTCATGTGGAAGT
5'L500/18   TCCTCAACCTGTTGCATCTGTGCTGTTGAGTGCAGAGAGAATGATGCTCATGTGGAAGT
*****

5'L500/12   CAGGAGGGATATGTTGGG-----
5'L500/18   CAGGAGGGATATGTTGGGACTGGCCAGTTGGTCAATCCGAATGACCTCACCTGGGGAA
*****

5'L500/12   ----
5'L500/18   CTG

```

**Figure 4.29** Alignments of 5'L500/12 and 18 sequences. Yellow label is start codon, blue label is *chg-L/D12* R2 primer, and green label is *chg-L/D12* R1 primer.

From result in Fig. 4.29 found 5'L500/12 and 18 is same sequence but amplified from different primer and shown length of 5'UTR of *chg-L* was 52 bp. Figure 4.30 shown in 100 % identical in coding sequences and some different in 3'UTR of 2 variant of 3'cDNA end cDNA sequences of *chg-L*, variant 19 have longer 3'UTR and polyA than variant 37, variant 19 contained polyadenylation signal but not found in variant 37.

```

3'L500-19      GGCTGGGGAGGTGGAGCTCATCCTAATACTCAAGAAAGGAAGATCCGTGATGTGTCTAAA
3'L500-37      GGCTGGGGAGGTGGAGCTCATCCTAATACTCAAGAAAGGAAGATCCGTGATGTGTCTAAA
*****

3'L500-19      TCTCAAACCTTTGGAATGGGAAGGTCAAATCACCGTGGGTCCCATTGACATCAAAGAGAGG
3'L500-37      TCTCAAACCTTTGGAATGGGAAGGTCAAATCACCGTGGGTCCCATTGACATCAAAGAGAGG
*****

3'L500-19      GTACTCGCTTAAACACGGTTGTCCATGACCATCACAGCAAACATTAGCCTTCATTCATTAC
3'L500-37      GTACTCGCTTAAACACGGTTGTCCATGACCATCACAGCAAACATTAGCCTTCATTCATTAC
*****

3'L500-19      TGTCTGTATGTTATTCTCTTGCTACTTGTGGCAGTTTGATCTACACAAGGAAGGAAAAA
3'L500-37      TGTCTGTATGTTATTCTCTTGCTACTTGTGGCAGTTTGATCTACACAAGGAAGGAAAAA
*****

3'L500-19      GAAAACATTTACTCCATTAAACACAATTGAAAAA-CTGGAAAAGGATGTCATAGCTATCA
3'L500-37      GAAAACATTTACTCCATTAAACACAATTGAAAAAAGCTGAAAAAAAAAAAAAAAAAAAAA
*****

3'L500-19      CCTTCCCCTCGGTGTAATAATAAATGTTACATGTGTACATGTAAAAAAAAAAAAAAAAAA
3'L500-37      AAAAAA-----

3'L500-19      AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
3'L500-37      -----

3'L500-19      AAAAAAAAAAA
3'L500-37      -----

```

**Figure 4.30** Alignments of 3'L500/19 and 37 sequences. Yellow label is *chg-L* F-RACE primer, blue label is stop codon, blue alphabet is polyadenylation signal and green label is polyA tail.

Full length (ORF) sequence of *chg-L* were amplified by PCR using primers designed from start and stop codon at 5' and 3' regions of *chg-L*. The product was obtained at the size of 1.3 kb (Fig. 4.27 C). Sequencing (see in appendix B) and BLAST analysis confirmed that this PCR product was *chg-L* which was the closest homologous with chorion protein of gilthead seabream (*Sparus aurata*) with E-value as  $7e-145$  (see detail in appendix C). *Chg-L* deduced sequence encoded from ORF was shown in Fig. 4.31. *L. subviridis* *Chg-L* which contained 419 amino acid residues with molecular weight at 46.09 kDa contained signal peptide at position 1-22, proline-rich region at position 35-86, zona pellucida (ZP) domain (ZP\_2) at position 93-353 (score = 35.841) which containing 261 amino acid residues, 8 conserved cysteines residues, and conserved N-glycosylation site at position 193-196 as shown in Fig. 4.31. Consensus pattern domain and motifs show in table 4.3.

```

gcttgcactgtggagccatgggtgatgaagtggactgctgcttgccttggcactggct
      M V M K W T A A C L V A L A
ctatttggcagcgtctgtgatgctcagtggggagagtacacgccttcaaatatcagaaa
L F A S V C D A Q W G E Y T P S K Y Q K
cctgcaactcctgtgaagcaagagcccaacaagtgcctcaagacactcaacagcataag
P A P F V K Q E P K Q V P Q D T Q Q H K
cagacatttgaaacaccacttcaatggacatacctgaacctccccgcctgaacctggc
Q T F E T P L Q W T Y P E P P P F E P A
cctgaaataccatttgaaccgctacgtcctcaacctgttgcatctgttgctgttgagtg
P E I P F F E P L R P Q P V A S V A V E C
agagagaatgatgctcatgtggaagtccaggaggatagtttgggactggccagttggc
R E N D A H V E V R R D M F G T G Q L V
aatccgaatgacctcaccctggggaactgtcctgctgctgcagaggatcctgcggctcaa
N P N D L T L G N C P A V A E D P A A Q
gtgttgattttgaagctgaactgcatgactgtttgagctcattgggttgacagaagat
V L I F E A E L H D C L S S L V L T E D
tcctgatctacatctcactctgaactacgatccccgacctctgggttctctccccgta
S L I Y I F T L N Y D P R P L G S S P V
gtaaggaccggcagtgagctgttattgtggaatgccactaccaagaagcacaatgtg
V R T G S A A V I V E C H Y P R K H N V
agcagccttctcttgaaccctgtggatcccatactctgcagttaagtgggcggaggaa
S S L P L E P L W I P Y S A V K V A E E
ttctgtacttcaacttaaaactcatgactgatgactggctgatgagaggccagtcacc
F L Y F T L K L M T D D W L Y E R P V N
cagtaactcctgggagacatcatttacatcgaggctatcgtcaagcagttctaccgctg
Q Y Y L G D I I Y I E A I V K Q F Y R V
ccctcctgtttacgtggacagttgtgtgggtactcttccctgacccaaactccagc
P L R V Y V D S C V G T L S P D P N S S
cccagatattccttcattgacaactatgggtgtctgattgatgctcgggtcacaggtct
P R Y S F I D N Y G C L I D A R V T G S
gcttcaaggttcttgcctcgcacagcagaaaacaagcttcagttcctgctggaggccttt
A S R F L P R T A E N K L Q F L L E A F
aggttcaaggggtgccgatagtgactgctctacattacatgccacttgaagcaacaact
R F K G A D S G L L Y I T C H L K A T T
actggccatcccatttgatggggaacaccgccttgttctacatcaacgggtggtctgg
T G H P I D G E H R A C S Y I N G W S E
gccagtgagtcgaatgctgcttgggatcctgtgattctggtttacctgatactggtgct
A S G V N A A C G S C D S G L P D T G A
ccagtggtggggaggtggagctcatcctaatactcaagaaaggaagatccgtgatgtg
P G G W G G G A H P N T Q E R K I R D V
tctaaatctcaaacttggaaatgggaaggtcaagtcaccgtgggtcccattgacatcaa
S K S Q T L E W E G Q V T V G P I D I K
gagagggactcgttcaacacggttgtccatga
E R V L A -

```

**Figure 4.31** Chg-L deduced sequence encoded from chg-L full length sequence.

Red and blue alphabet are start and stop codon, grey highlight indicates signal peptide, blue highlight indicates zona pellucida (ZP) domain (ZP<sub>2</sub>), green highlight indicates Proline-rich region, and pink highlight indicates conserved N-glycosylation site.

**Table 4.3** Domains, motifs, and consensus patterns of Chg-L

Domain	Site	Consensus pattern
zona pellucida (ZP) domain (ZP <sub>2</sub> )	93-353	[LIVMFYW] - x(7) - [STAPDNLR] - x(3) - [LIVMFYW] - x - [LIVMFYW] - x - [LIVMFYW] - x(2) - C - [LIVMFYW] - x - [STA] - [PSLT] - x(2,4) - [DENSG] - x - [STADNQLFM] - x(6) - [LIVM](2) - x(3,4) - C The 2 C's may be involved in disulfide bonds

*L. subviridis* Chg-L was aligned with Chg-L of other fish species as shown in Fig. 4.32, Zona pellucida (ZP) domain of *L. subviridis* Chg-L was highly conserved with this domain of other fish species. ZP domain of Chg-L contained strictly conserved 8 cysteines residues which involved in disulfide bond formation of this protein and one N-glycosylation site. N and C terminus of Chg-L (non label) do not conserved between species of fish.

```

Oryzias_sinensis      -MMKFPAVCLVVLALLDGFCDAP--FYGKP---GPGSK-----TPQDP-----
Oryzias_latipes       -MMKFTAVCLVVLALLDGFCDAQH--NYGKPSYPPTGSK-----TPQDP-----
Liparis_atlanticus    MVMKYTAVCLLVLALFGTFCEAQRG--GFQKPYQKSPKQ---VPYEP-----
Liza_subviridis       MVMKWTAACLVALALFASVCDAQWG--EYTPSKYQKPAPP-----VKQEPKQV-----
Oncorhynchus_mykiss   MAMKWSVVLVAVAMLGCLCVAQNWPFFSKPVQQFFRPNRQPPQQPQQPPYQKPRIP
                        **:.:.**:.:.*: . * * * : . . . : *
Oryzias_sinensis      ---TQQ-KQLHEKEITWKYPADPQPEPKPVVPEQRFVPAATVAVECREDLAHVEAKKD
Oryzias_latipes       ---TQQ-KQLHEKELTWKYPADPQPEAKPVVPEQRYPVPAATVAVECREDLAHVEAKKD
Liparis_atlanticus    ---QQQAKQNFQKPLTWIFPEDPQPEAAVEVPFELRYPVAASVSVCRESAVHVEVKKD
Liza_subviridis       PQDTQQHKTFFETPLQWTYPEPPPPPEPAPEIPFEPLRQPVASVACRENDAHVEVRRD
Oncorhynchus_mykiss   PKDQTQAKQKFETPLDWTYPLDKPEPKIIGSSSEARTPVAANSVRACRENMVHVEAKHD
                        * * * . * . : * : * * * . . * * . . : * . * * * . * * * * : *
Oryzias_sinensis      LFGIGQFIDPADLTLGTCPPVAEDPAAQVLIFESELQNCGSVLTMTEDSLVYFTFLNYKP
Oryzias_latipes       LFGIGQFIDPADLTLGTCPPSAEDPAAQVLIFESEPLQNCGSVLTMTEDSLVYFTFLNYKP
Liparis_atlanticus    MFGIGQFINAADLTLGNCGAVAEDSAGQVLVFEAELQNCLSSLGMTEDSLIYFTFMNYIP
Liza_subviridis       MFGTGQLVNPNDLTLGNCPAVAEDPAAQVLIFEAELHDCSSLVLPEDSLIYFTFLNYDP
Oncorhynchus_mykiss   LLGIGQLIQLEDLTLGDCEMTGFDNINQVLIFESPLQSCGSQLRMTNSLIYIFFLYYKP
                        : * * * : : * * * * * * * * * * * * * * * * * * * * * * * * * * * *
Oryzias_sinensis      QPLGSAPVVRTSQAVVIVECHYPRKHNVSSLALDPLWVPFSAAKMAEEFLYFTLKLTTDD
Oryzias_latipes       KPLGSAPVVRTSQAVVIVECHYPRKHNVSSLALDPLWVPFSAAKMAEEFLYFTLKLTTDD
Liparis_atlanticus    QPLGGSPVVRTSQAAVIVECHYPRKHNVSSLALDPLWIPFSAVKMAEEFLYFTLKLMTDD
Liza_subviridis       RPLGSSPVVRTGSAAVIVECHYPRKHNVSSLPLEPLWIPYSAVKVAEEFLYFTLKLMTDD
Oncorhynchus_mykiss   KPLANTPLIRTNDAMINIECHYPRKHNVSSLALIPTWTPFSAAKYAEELLYFSMRLMTAD
                        : * * . : : * * . * : : * * * * * * * * * * * * * * * * * * * * * *
Oryzias_sinensis      FQFERPSYQYFLGDLIHIEATVKQYFHVPLRVYVDRCVATLSPDANSSPSYAFIDNYGCN
Oryzias_latipes       FQFERPSYQYFLGDLIHIEATVKQYFHVPLRVYVDRCVATLSPDANSSPSYAFIDNYGCL
Liparis_atlanticus    WQYERPSYQYFLGDTINIEAVVKQYFHVPLRIYVESCVATLEPDTSANPRYAFIDNNGCL
Liza_subviridis       WLYERPVNQYLLGDIYYIEAIVKQFYRVPLRVYVDSCVGTLSPDPNSSPRYSFIDNYGCL
Oncorhynchus_mykiss   WQYERAGNMYVLGDMVNIASVMQYFHVPLRIFVDSCVATLEPNINANPRYAFIENHGCL
                        : : * . * * * * : * * * * * * * * * * * * * * * * * * * * * *
Oryzias_sinensis      LDGRITGSNSKLVSRPAENKLDFQLEAFRFQG-----ADSGM
Oryzias_latipes       LDGRITGSNSKLVSRPAENKLDFQLEAFRFQG-----ADSGM
Liparis_atlanticus    LDARLTGSNSKLVSRPAENKLDFQLEAFRFQN-----AESGL
Liza_subviridis       IDARVTGSASRFLRPAENKLDFQLEAFRFQG-----ADSGL
Oncorhynchus_mykiss   IDAKMTGSHSQFMPRSADYKLYFQVEAFRFQIQKSGSDFINRQKTKIPFQAASDYPATLDM
                        : * : : * * * * : * . : * * * : * * * * : * * * * * : * : * : *
Oryzias_sinensis      IYITCHLKATSAAYPLDAEHRACSYIQGWHEVSGADPICASESGGFEVHAN---AVVS
Oryzias_latipes       IYITCHLKATSAAYPLDAEHRACSYIQGWHEVSGADPICASESGGFEVHAN---AVVS
Liparis_atlanticus    LYITCHLKATSASSSDNDHRACSWNNGWHEASKIDSVCGSCESAGLPPAANPRGANFVA
Liza_subviridis       LYITCHLKATTTCHPIDGEHRACSYINGWSEASGVNACGSCDSG-----
Oncorhynchus_mykiss   IFIITCHLKATTIAFPIDFEYKACSYINTWREAGGNDGVCGCCDST-----
                        : * * * * * * * * : . * : : * * * * : * * * * : * * * * *
Oryzias_sinensis      HGTSTLSGGGHGTGK-----PS-----
Oryzias_latipes       HGTSTLSGGGHGTGK-----PS-----
Liparis_atlanticus    SGPYTISGGGGGGGGSGGSGNYPSGSGNYPSGSGGLGGAGGGSGGYNGAHGGHGAHGINGG
Liza_subviridis       -LPDTGAPGGWGGGAH-----PN-----
Oncorhynchus_mykiss   -----CSNRKGR-----
                        . . . *

```

```

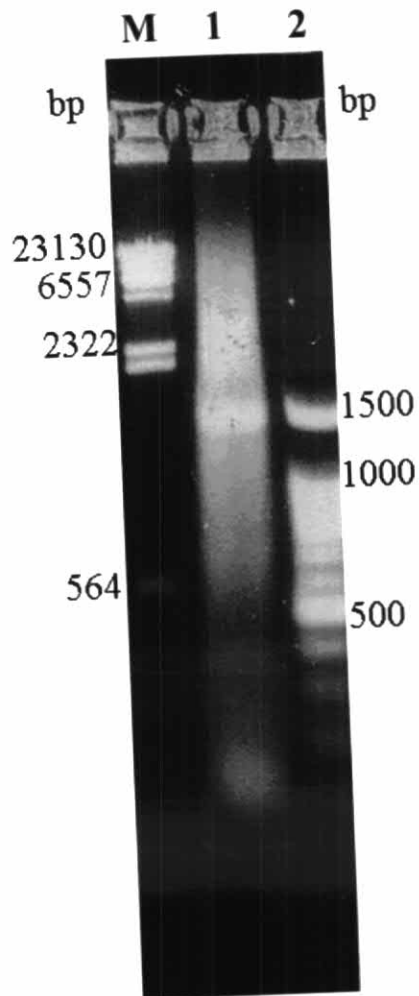
Oryzias_sinensis      -----DPSRKTREAAKTEVLEWEGDVTLGPIPIEERRV-
Oryzias_latipes       -----DPSRKTREAAKTEVLEWEGDVTLGPIPIEERRV-
Liparis_atlanticus    GGGGGGGGGGGGGYNGNSGGGIYNPSRKIRDVS--EVFEWEGDVTLGPIPIADKMVA
Liza_subviridis       -----TQERKIRDVSKSQTLEWEGQVTVGPIIDIKERVLA
Oncorhynchus_mykiss   -----DTTKHQKLVN-----IWEGDVQLGPIFISEKVAQ
                        :: : .      ***:* :*** * ::

```

**Figure 4.32** Alignments of Chg-L of *L. subviridis* and other species of fish. Zona pellucida (ZP) domain is labeled with green which contained 8 strictly conserved cysteines residues (label with yellow) and 1 conserved N-glycosylation site (label with pink).

An attempt to amplify 5'cDNA end sequence of *chg-H* was not successful. Non-specific bands and smear were obtained from 5'RACE PCR amplification. The result was shown in Fig. 4.34. After sequencing (see in appendix B) and BLAST analysis, this sequence was homologous with chorion protein (*Liparis atlanticus*) with E-value as 5e-62.





**Figure 4.33** 5'RACE PCR product of *chg-H* separated in 1.2 % agarose. Lane M1, M2 and 1 is  $\lambda$ *Hind*III, 100 bp DNA marker and 5'RACE PCR product of *chg-H*, respectively.

This sequence aligned with complete 5'cDNA end sequence of *chg-L* as shown in Fig. 4.34.

```

5'L500-12          ACGCGGGGACAGCACCTTGAGAATCTCTCAGATCGCTTGCTACTGTGGAGCCATGGTGAT
5'H1.5-10_chg-L_  -----

5'L500-12          GAAGTGGACTGCTGCTTGCCTTGTGGCACTGGCTCTATTTGCCAGCGTCTGTGATGCTCA
5'H1.5-10_chg-L_  -----

5'L500-12          GTGGGGAGAGTACACG-CCTTCAAATATCAGAAACCTGCACCTCCTGTGAAGCAAGAGC
5'H1.5-10_chg-L_  -----ACACGACCTTCAAATATCCTAAACCTGCACCTCCTGTGAAGCAAGAGC
                    *****

5'L500-12          CCAAACAAGTGCCTCAAGACACTCAACAGCATAAGCAGACATTTGAAACACCACTTCAAT
5'H1.5-10_chg-L_  CCAAACAAGTGCCTCAAGACACTCAACAGCATAAGCAGACATTTGAAACACCACTTCAAT
                    *****

5'L500-12          GGACATACCCTGAACCTCCCCCGCTGAACCTGCGCCTGAAATACCATTGAAACCGCTAC
5'H1.5-10_chg-L_  GGACATACCCTGAACCTCCCCCGCTGAACCTGCGCCTGAAATACCATTGAAACCGCTAC
                    *****

5'L500-12          GTCCTCAACCTGTTGCATCTGTTGCTGTGAGTGCAGAGAGAATGATGCTCATGTGGAAG
5'H1.5-10_chg-L_  GTCCTCAACCTGTTGCATCTGTTGCTGTGAGTGCAGAGAGAATGATGCTCATGTGGAAG
                    *****

5'L500-12          TCAGGAGGGATATGTTGGG-----
5'H1.5-10_chg-L_  TCAGGAGGGATATGTTGGGACTGGCCAGTTGGTCAATCCGAATGACCTCACCTGGGGA
                    *****

5'L500-12          -----
5'H1.5-10_chg-L_  ACTGTCTGCTGTCGCAAAGGATCTGCAGCTCAAGTGTGATTTTGAAGCTGAAGTGC

5'L500-12          -----
5'H1.5-10_chg-L_  ATGACTGTTGAGCTCATTGGTGTGACAGAAGATTCCCTGATCTGCATCTTCACTCTGA

5'L500-12          -----
5'H1.5-10_chg-L_  ACTACGATCCCCGACCTCTGGGTTCCCTCCCCGTAATAAGGACCGGCAGTGCAGCTGTTA

5'L500-12          -----
5'H1.5-10_chg-L_  TTGTGGAATGCCACTACCCAAGAAAGCACAATGTGAGCAGCCTTCCCTTGAACCCCTGT

5'L500-12          -----
5'H1.5-10_chg-L_  GGATCCCATACTCTGCAGTTAAAGTGGCGGAGGAATCTTGTACTTCACCTTAAACTCA

5'L500-12          -----
5'H1.5-10_chg-L_  TGACTGATGACTGGCTGTATGATAGGCCAGTCATACCCGCACTACCTGGGAGACATCATT

5'L500-12          -----
5'H1.5-10_chg-L_  TACATCAGGGCATTGTCCTCAAGCAATTTTCCACCTGCTCCCTCTGTGGTTACTCGTAAA

5'L500-12          -----
5'H1.5-10_chg-L_  GATGGGTGAGTAAT

```

**Figure 4.34** alignments of 5'L 500/12 and 5'H 1.5/10 (*chg-L*). Blue is start codon.

These two sequences were high similarity but 5'H 1.5/10 (*chg-L*) sequence lack overall 5'UTR and some portion of 5'end coding sequence as shown in Fig. 4.34.

```

5'L500-12      MVMKWTAACLVALALFASVCDAQWGEYTPSKYQKPAPPVKQEPKQVPQDTQQHKQTFETP
5'H1.5-10_chg-L_ -----TRPSKYPKPAPPVKQEPKQVPQDTQQHKQTFETP
                    *****

5'L500-12      LQWTYPEPPPEPAPPEIIPFEPLRPQPVASVAVECRENDAHVEVRRDMFG-----
5'H1.5-10_chg-L_ LQWTYPEPPPEPAPPEIIPFEPLRPQPVASVAVERKENDAHVEVRRDMFGTQQLVNPNDLT
                    *****

5'L500-12      -----
5'H1.5-10_chg-L_ LGNCPAVAKDPAAQVLIFEAEHLHDCLSSLVLTEDSLICIFTLNYDPRPLGSSPVIRTGSA

5'L500-12      -----
5'H1.5-10_chg-L_ AVIVECHYPRKHNVSLLPLEPLWIPYSAVKVAEEFLYFTLKLMTDDWLYDRPVIPALPGR

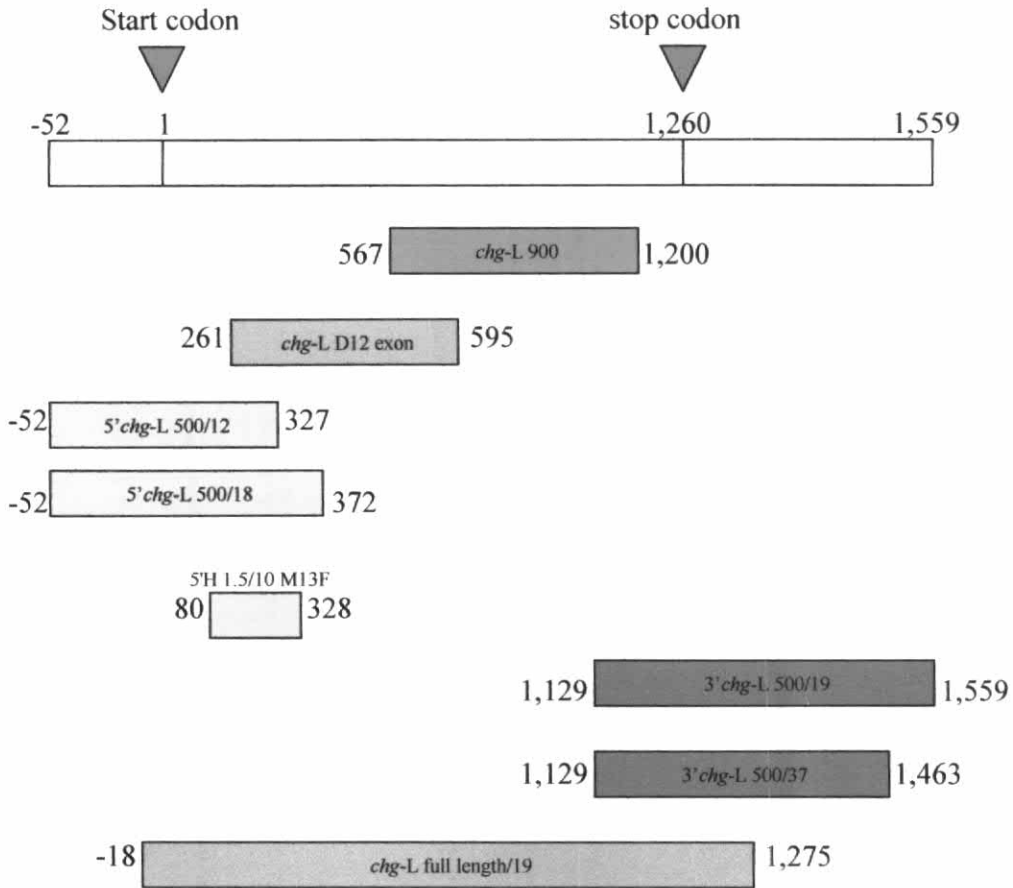
5'L500-12      -----
5'H1.5-10_chg-L_ HHLHQGIRPSNFSSTCSLCGYS-RWVSN

```

**Figure 4.35** Alignments of 5'L 500/12 and 5'H 1.5/10 deduced amino acid sequence.

Deduced amino acid sequence of 5'L 500/12 and 5'H 1.5/10 were high similarity as shown in Fig. 4.35.

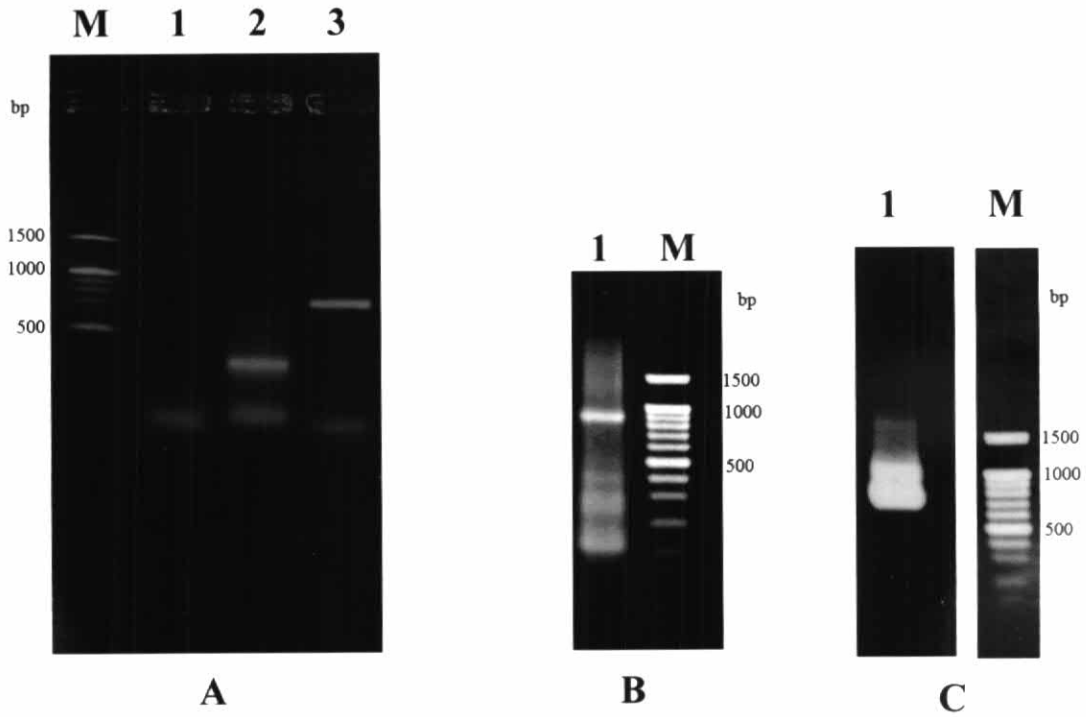
Structure of full length of *chg-L* gene which shown position of nucleotide sequence of *chg-L* gene obtained from this study shown in Fig. 4.36.



**Figure 4.36** Schematic representation of the full length of *chg-L* gene. Blue box is partial coding sequence of *chg-L*, orange box is exon obtained by GenomeWalk PCR from *DraI* GenomeWalk library, yellow box is 5'end cDNA sequence of *chg-L*, green box is 3'end cDNA sequence of *chg-L*, and purple box is *chg-L* full length (ORF).

#### 4.4.2 choriogenin H (*chg-H*)

*chg-H* PCR product at the size of 750 bp was amplified from 1<sup>st</sup> strand cDNA templates of livers as shown in Fig. 4.37 A. After sequencing (see in appendix B) and BLAST analysis, this PCR product was homologous with zona pellucida protein Bb of gilthead seabream (*Sparus aurata*) with E-value as 4e-104. 5'RACE PCR product of *chg-H* at the size of 900 bp and 3'RACE PCR products of *chg-H* at the sizes of 700 bp and 1 kb were amplified from livers (Fig. 4.37 B and C, respectively). 3'RACE PCR product of *chg-H* at size 700 bp and 1 kb were sequenced and BLAST analysis found it homologous with zona pellucida protein Bb of gilthead seabream (*Sparus aurata*) with E-value as 7e-62 and 2e-122, respectively (see detail in appendix B and C).



**Figure 4.37** *chg-H* PCR products separated in 1.2 % agarose gel. (A) *chg-H* PCR product separated in 1.2 % agarose gel Lane M, 1, 2, and 3 is 100 bp DNA marker, negative control, positive control and *chg-H* PCR product, respectively, (B) 5' RACE PCR product of *chg-H* separated in 1.2 % agarose gel (lane M and 1 is 100 bp DNA marker and 5' RACE PCR product of *chg-H*, respectively and (C) 3' RACE PCR product of *chg-H* separated in 1.2 % agarose gel (lane M and 1 is 100 bp DNA marker and 3'RACE PCR product of *chg-H*, respectively.

Two variant of 3'end cDNA sequence of chg-H were aligned as shown in Fig. 4.38.

```

3' chg-H800-51      CTACCCCTGTGGCCAAAGTATTGAGGGATCCTGTGTATGTGGAGGTTGAGCTCCTTGAAT
3' chg-H1.1-13     CTACCCCTGTGGCCAAAGTATTGAGGGATCCTGTGTATGTGGAGGTTGAGCTCCTTGAAT
*****

3' chg-H800-51      GACCGATCCAGCACTTGTCTGACTCTCGGCCGCTGTTGGACAACAACCCCCAATCC
3' chg-H1.1-13     GACAGATCCAGCACTTGTCTGACTCTCGGCCGCTGTTGGACAACAACCCCCAATCC
*** *****

3' chg-H800-51      TCACAGCCTGCCCCAGTGGGACATACTGGTGGACGGATGTCCATACAGGGATGATCGTTA
3' chg-H1.1-13     TCACAGCCTGCCCCAGTGGGACATACTGGTGGACGGATGTCCATACAGGGATGATCGTTA
*****

3' chg-H800-51      CTTGTCTGCATTGGTCCAGTCACTGGTGTGACCTCCAGGTCGTACAGACGTTTCCT
3' chg-H1.1-13     CTTGTCTGCATTGGTCCAGTCACTGGTGTGACCTCCAGGTCGTACAGACGTTTCCT
*****

3' chg-H800-51      TTTCAAATGTTACCTTTGTGGATCCTGCTTCATTGGAGCCCTGAGAGAATACGTGTA
3' chg-H1.1-13     TTTCAAATGTTACCTTTGTGGATCCTGCTTCATTGGAGCCCTGAGAGAATACGTGTA
*****

3' chg-H800-51      CATTCACTGTAGTACAGTGTGTGTGCTGCTGCACCAGGCCGTAACGTGTATCCATCATG
3' chg-H1.1-13     CATTCACTGTAGTACAGTGTGTGTGCTGCTGCACCAGGCCGTAACGTGTATCCATCATG
*****

3' chg-H800-51      CTACAGGAAAAAGAGATCTGTTGATGCCGTTGGTCCAGAGAAAGGATGAACCAAGGTTGT
3' chg-H1.1-13     CTACAGGAAAAAGAGATCTGTTGATGCCGTTGGTCCAGAGAAAGGATGAACCAAGGTTGT
*****

3' chg-H800-51      TGTTCCTTTTGGACCAGTGTATGCGCCGCCCTGAGGAGCAACAGGCTGAAGAAATAGAG
3' chg-H1.1-13     TGTTCCTTTTGGACCAGTGTATGCGCCGCCCTGAGGAGCAACAGGCTGAAGAAATAGAG
*****

3' chg-H800-51      CTGAGAACATTTGAAAAATATTTGGAACATAGTACAGTTGTTAGTTGTATGAAATTA
3' chg-H1.1-13     CTGAGAACATTTGAAAAATATTTGGAACATAGTACAGTTGTTAGTTGTATGAAATTA
*****

3' chg-H800-51      AATGTCAATGTACGTATTTGTACATTTTGCAGAAAGCCCGAGAGAACATGAAAAGTCA
3' chg-H1.1-13     AATGTCAATGTACGTATTTGTACATTTTGCAGAAAGCCCGAGAGAACATGAAAAGTCA
*****

3' chg-H800-51      TGTGTTGTA AAAACCAACTCGCGAAACGCTCATATTATAAAATTTAAATGAATAAATAC
3' chg-H1.1-13     TGTGTTGTA AAAACCAACTCGCGAAACGCTCATATTATAAAATTTAAATGAATAAATCC
*****

3' chg-H800-51      ATGAAAGCTC AAAAAAAAAAAAAAAAAAAAAAAAAAAAAA -----
3' chg-H1.1-13     ATGAAAGCTTACAAGGATTTGCTTCATGATATTATTCATAAGTCAGCAAGCTGCCATTT
***** * * * * *

3' chg-H800-51      -----
3' chg-H1.1-13     TAACTAGTAATGTTAGCCGGTGATATGGGAGTGGATTGCCTTGTAATGAGGGGTTCTG

3' chg-H800-51      -----
3' chg-H1.1-13     TTGTCAGTATTTGTTTATTCAAACACACACACATTTTTCAGTATAACTAAATTTAGCA

3' chg-H800-51      -----
3' chg-H1.1-13     TAATACTACAGCAAGTTACAGCACTGTATATGCCCCATGGATTACAGTGAACATTGCAA

```

```

3' chg-H800-51 -----
3' chg-H1.1-13 CAAAACGGGTCACTTAAATGTTTCATGCTTCTCAAATAAACTATATACGGGCAAAAAAA

3' chg-H800-51 -----
3' chg-H1.1-13 AAAAAAA

```

**Figure 4.38** Alignment of 3' end cDNA sequence of *chg-H*. Green label is stop codon, yellow label is polyadenylation signal and blue label is polyA tail.

Fig. 4.38 has shown different in 2 nucleotides position of coding sequence between 2 variant and high similarity in 3' UTR of 2 variant but 3'*chg-H* 1.1-13 have 3' UTR longer than 3'*chg-H* 800-51. Coding sequence of 2 variant was translated to amino acid sequence, 2 variant were 100 % identical in amino acid sequence as shown in Fig. 4.39.

```

3' Chg-H800-51 YPVAKVL RDPVYVEVELLEMTDPALVLT LGRCWTTTTPNPHSLPQWDILVDGCPYRDDRY
3' Chg-H1.1-13 YPVAKVL RDPVYVEVELLEMTDPALVLT LGRCWTTTTPNPHSLPQWDILVDGCPYRDDRY
*****

3' Chg-H800-51 LSALVPVTGVDLPGRYRRFLFKMFTFVDPASLEPLREYVYIHCSTAVCAAAPGRNCDPSC
3' Chg-H1.1-13 LSALVPVTGVDLPGRYRRFLFKMFTFVDPASLEPLREYVYIHCSTAVCAAAPGRNCDPSC
*****

3' Chg-H800-51 YRKKRSVDAVVQRKDEPKVVVSFGPVIMAAPEEQQAEE
3' Chg-H1.1-13 YRKKRSVDAVVQRKDEPKVVVSFGPVIMAAPEEQQAEE
*****

```

**Figure 4.39** Alignment of 3'Chg-H 800/51 and 1.1/13 deduced sequence.

Partial and 3' end cDNA sequence were ligated *in silico* and translated as shown in Fig. 4.40. It contained 2 polyadenylation signals.



```

ccagttcattgtgttagtagtagatgtcactctaccagcattgatctagagtcate
Q F I V V V A R D V T L P S I D L E S I
acactctgggacaagggtgaaggctgtacacctgtgactccaattcaaactatgccate
T L L G Q G E G C T P A D S N S N Y A I
taccagtttctgttaccgctgtggtagcatcgtaacggaggagcctgggttatagtc
Y Q F P V T A C G T I V T E E P G V I V
tatgaaacaggatgtcctcctcatatgaagtgtggttgaccattaggagtcattacc
Y E N R M S S S Y E V L V G P L G V I T
agagacagccactatgatgtgctcttccagtgacgtaccctggtaccactgttgaact
R D S H Y D V L F Q C R Y I G T T V E T
ctggttatagaagttagccccattagataatcctcctaccagttgctgctctgggacct
L V I E V A P L D N P P L P V A A L G P
attgcagtagcaatgagggtggccaatggtgaatgcactacaaaggttgcaatgaagtg
I A V R M R L A N G E C T T K G C N E V
gaagcagcatttaccctcctctatcagggatagactaccctgtggccaagtattgagg
E A A F T S F Y T E I D Y P V A K V L R
gatcctgtgtatggagggttgagctcctgaaatgacagatccagcactgtcctgact
D P V Y V E V E L L E M T D P A L V L T
ctcggccgctgttggacaacaacaaccccccaatcctcacagcctgccccagtgaggacata
L G R C W T T T T P N P H S L P Q W D I
ctgggtgagcaggtgtccatacagggatgatcgttacttctgctgcaactggtccagtcact
L V D G C P Y R D D R Y L S A L V P V T
gggtgcagcctcccaggtgcgtacagagctttccttttcaaatgttcaccttgtggat
G V D L P G R Y R R F L F K M F T F V D
cctgcttcattggagcccctgagagaatacgtgtacattcactgtagtagcagctgtgtgt
P A S L E P L R E Y V Y I H C S T A V C
gctgctgaccagggcctgaactgtgatccatcatgctacaggaagagagatctgttgat
A A A P G R N C D P S C Y R K K R S V D
gccgtggtccagagaaaggatgaaccaaggtgtgtttcttttgaccagtgatcatg
A V V Q R K D E P K V V V S F G P V I M
gccgccccctgaggagcaacagggctgaagaatagagctgagaacattttgagaataattt
A A P E E Q Q A E E -
ggaacatagtagcattgtttagttgtatgaaatataaatgtcaatgtacgtatttgatcat
tttgagaaagccccgagagaacatgaaaactgcatgtgttgtaaaaacccaactcgca
aacctcatattataaaatttaaatgataaaatccattgaaagcttacaaggatttgctt
catgatattattcataagtcagcaagctgccattttaaactagtaattgttagccggtgata
tggagtggtattgccttgaattgaggggtttctgtgtcagttattgtttattcaaaaca
cacacacatttttcagttataactaaatttagcataataactacagcaagttacagcact
gtatatgccccatggattcacgtgaactattgcaacaaaacgggtcacttaaaatgttca
tgctctcaataaaactatatacgggcaaaaaaaaaaaaaaaaaa

```

**Figure 4.40** Deduce sequence of Chg-H. Green label is ZP domain, yellow label is stop codon, blue label is polyadenylation signal, and pink label is polyA tail.

Partial Chg-H contained ZP domain at position 1-275 was aligned with other fish Chg-H as shown in Fig. 4.41.

```

Liza_subviridis -----
Oryzias_latipes MARHWSITVFSALALLCSFLGTEVDAQKGNPQDPKVPYPPYYPQPKPQDPQHVSPPYYPG
Liparis_atlanticus -----
Oncorhynchus_mykiss --MKWSAVCLVAVATLGWLCDQAIYLEK--PGWPPIQTPPSWPAQPPQ-----

Liza_subviridis -----
Oryzias_latipes KPQNPQKPSNPQYPSYPTPQNPQVPQNPQVPQNPQYPSYQNPSPYQNPSPYQYPSNP
Liparis_atlanticus -----
Oncorhynchus_mykiss KPIQPPQRPAQP-----PQWPVQP

Liza_subviridis -----
Oryzias_latipes PTSQNPSPQNPKLFQDGKPSNPQQPQVPQYPSKPPQNPQVPQYPSKPPQNPQVPQ
Liparis_atlanticus -----
Oncorhynchus_mykiss P-----QRPAQPPQRP-----

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Liza_subviridis -----
Oryzias_latipes YPSKPQPQPQPQVQYPSKPQPQPQPQVQYPSKPQPQPQPQVQYPSKPQPQPQPQVQV
Liparis_atlanticus -----MAAAAATAAGPAAAA
Oncorhynchus_mykiss -----AQPQRPAQPQQWPAQPQRPAQPQQ

Liza_subviridis -----
Oryzias_latipes YPSKPQPQPQPQVQYPSKPQPQPQYPSKPQYQAPQQPQPQVQYPSKPQDPGKNPNTPPIG
Liparis_atlanticus AAATAAMAASSAAAATVEKAETPSPATSPAGVAIVPMFGLIRSETPKTHRIKSDTFCQI
Oncorhynchus_mykiss WPAQPQRPAQPQPQRPAQPQRPAQPQRPAQPQVWPVHPQPWPVQPGTPLQRPK-FPSD

Liza_subviridis -----QFIVVVA
Oryzias_latipes PPPKSCVPRDVRVPCGVDPDISPSACDAIDCCHDQSCYFGTGATVQCTKDGHFIVVVA
Liparis_atlanticus ILLPRVVTQKRNTRVTCGLPDISTKHCEEINCCSDGQHCYFKAVTVQCTKDGQFIVVVA
Oncorhynchus_mykiss TGSKQSCDVGQHKVQCGLPDITAAHCDAINCCFDGRMCFYKSVTVQCTKDGQFVVVVA
: * : * * *

Liza_subviridis RDVTLPSIDLESITLLGQ-GEGCTPADSNSNYAIYQFPVTACCTIVTEEPGVIVYENRMS
Oryzias_latipes KDVTLPHIDLETISLLGQ-GQDCGPADSNSAFAYFPVTCYCTVVMEEPGVIVYENRMT
Liparis_atlanticus RDATLPNIDLESISLLRS-GPGCTHVDSNSGFAYNFGVTACCTVSEEPGVIIYENRMI
Oncorhynchus_mykiss RDATLPSLELDSISLLGTNGAHCHPIGTTTSVFAIYQFKVTECGTVMTEETDTIIYENRMS
: * . * * : * : * : * * * * * * * * * * : * * * * : * * * * * * * * * * * * * * * *

Liza_subviridis SSYEVLVGPLGVI TRDSHYDVLFCRYIGTTVETLVIEVAPLNDPPLPVAALGPVAVRMR
Oryzias_latipes SSYEVGVGVLGAI TRDSSFELLFCRYRATSVETLVVEVQPPDSP-LSIAELGPNLVYLQ
Liparis_atlanticus SSYEVDTGVPVGVISRDSQYELLFCRYIGTTVQTVVVEVSPLLDPPISVAAVGPIRVELR
Oncorhynchus_mykiss SSYQVGVGPFSGITRDSQYDLTFCRYKGSTIVAVVIDVKVPPPPNSDIAP-GPLIVELR
* * * : * . * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Liza_subviridis LANGECTTKGCNEVEAAFTSFYTEIDYPVAKVLRDLVYVEVELLEMTDPALVLTGRCWT
Oryzias_latipes IANGCQTKGCDEAAAAYTSFYTDADYPVTKVLRDPVYVDVQILGRTPNLVLTGRCWA
Liparis_atlanticus LGNGQCISKGCVEEDVAYASYTADYPVSKVLRDPVYVEVRLLEKTDNLVLTGRCWA
Oncorhynchus_mykiss LGSGGCLTKGCNEEEVAYTSYTEADYPVTKVLRDPVYTEVRILARTDPNLVLTGRCWA
: . * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Liza_subviridis TTPNPHSLPQWDILVDGCPYRDDRYLSAPVPVT---GVDLPSRYRFLFKMFTFVDPAS
Oryzias_latipes TTSPNPFSLPQWDILIDGCPYADDRYLSALVPI DHSSGLFPFTHHSRFLFKMFTFVDPHS
Liparis_atlanticus TTSPNPHSLPQWDILTDGCPNRRNDKYLSSLIPIGPSSGLFYPHSHYRFLFKMFTFVQTS
Oncorhynchus_mykiss TTPNPLSLPQWDLIDGCPYQDDRYLTPITVGPSSGLSYPTHYRFLFKMFTFVDPMS
* * : * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Liza_subviridis -----LEPLREYVYIHCSTAVCAAAPGRNCDPSCYRKK-RSDAVVQRKDEPK-VVVSFG
Oryzias_latipes -----MEPLREKVYIHCSTAACVPGQVSCPEPSCRRKGRDTEAVAIRTDERR-VVVSFG
Liparis_atlanticus SKSIKSQPLREQVYIHCSTAVCTPVKGYSCPEPVCYRKK-RDVMVDVQTSQPK-VVASVG
Oncorhynchus_mykiss -----MAPLRETVFIHCNTAVCLPSLGDSCPEPVCYRKR-RDIPA AVQKTARIKSNLVSSG
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Liza_subviridis PVIMAAPEEQAAE
Oryzias_latipes EVLMLAAADEPSEQ
Liparis_atlanticus PVDMGASWE-----
Oncorhynchus_mykiss ELIILTDPRELTN--
: : :

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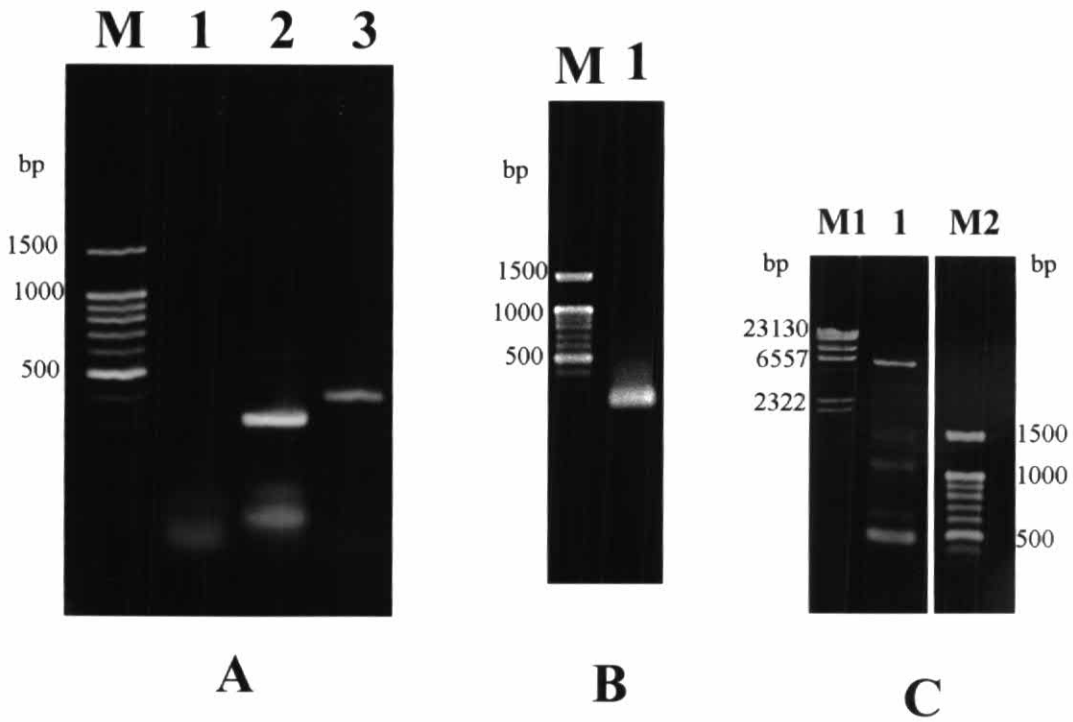
**Figure 4.41** *L. subviridis* Chg-H aligned with other fish Chg-H. Blue label is zona pellucida (ZP) domain, conserved cysteines residues label with yellow.

ZP domain of Chg-H was conserved and contained 12 conserved cysteines residues as shown in Fig. 4.41.

## 4.5 Cloning and characterization of vitellogenin (*vtg*) genes

### 4.5.1 *vtg-1*

Partial cDNA sequence of *vtg-1* at the size of 400 bp was amplified from 1<sup>st</sup> strand cDNA templates of liver (Fig. 4.42 A) The results of sequencing (see in appendix B) and BLAST analysis indicated its close similarity with *vtg1* of white mullet (*Mugil curema*) with E-value as 9e-114. 5' RACE PCR product at the size of 250 bp (Fig. 4.42 B) and 3' RACE PCR product at the sizes of 4.3, 1.5, 1.1, 0.6 and 0.5 kb (Fig. 4.42 C) were obtained. Only 250 bp product from 5' RACE PCR and 4.3 kb product from 3' RACE PCR were cloned and sequenced (see detail in appendix B). The result of sequencing and BLAST analysis indicated that both sequences were homologous with *vtgA* of charr (*Salvelinus alpinus alpinus*) and *vtg* of red seabream (*Pagrus major*) with E-value as 1e-04 and 0.0, respectively. Because 3'*vtg-1* sequence was very long, only 75% of the 4.3/51 fragment was sequenced. Nucleotide sequence of *vtg-1* (*in silico* ligated 5', 3' end cDNA sequence and partial coding sequence) shown in Fig. 4.43. Because internal sequence at size 950 bp not sequenced so sequence divided to 2 parts and translated to amino acid sequence of N and C-terminus of Vtg-1 as shown in Fig. 4.44 (A) and (B), respectively. *L. subviridis* Vtg-1 which contained 1,550 amino acid residues with molecular weight at 170.5 kDa which contained lipoprotein N-terminal domain (LPD\_N) at position 24-589, lipovitellin-phosvitin complex; beta-sheet shell regions at position 616-888 and von Willebrand factor (vWF) type D domain (VWD) at position 1,280-1,435 (Fig. 4.44). LPD\_N, beta-sheet shell regions, and VWD of *L. subviridis* Vtg-1 were aligned with these domains of other fish species (Fig. 4.45 (A), (B), and (C), respectively). LPD\_N, beta-sheet shell regions, and VWD of *L. subviridis* Vtg-1 were highly conserved with these domains of other fish species.



**Figure 4.42** *vtg-1* PCR products separated in 1.2% agarose gel. (A) *vtg-1* PCR product separated in 1.2% agarose gel, lane M, 1, 2 and 3 is 100 bp DNA marker, negative control, positive control and *vtg-1* PCR product, respectively, (B) 5' RACE PCR of *vtg-1* product separated in 1.2% agarose gel. Lane M and 1 is 100 bp DNA marker and 5' RACE PCR of *vtg-1*, respectively and (C) 3' RACE PCR of *vtg-1* separated in 1.2% agarose gel. Lane M1, M2 and 1 is  $\lambda$ HindIII, 100 bp DNA marker and 3' RACE PCR of *vtg-1*, respectively.

AC GCGGGAAGCAGTGTATCAACGCACAGTACGCGGGGACAACACCATCGTCCATGACAGCACTTGTACTTGCCCGGACACTGG  
 CCTTCATGGCTGGCCAGCGCAACATTTTGGCCCTGAATTTGCCGCCAGTAAACATTTGTCTACCACCTAGGAGGCATTGCTC  
 CCGGGGGTCTGTACAGGAAGGTTTGGCGAGAGCTGGACTCAAAGTCAGCAGCAAAGTTCTCATCAGCGCTGCAGTGAAAAC  
 AAATACCTGCTGAAGCTTGTGGAACCTGAGCTCTATGAATACAGTGGTGTGGCCCAAGGATCCTTTAATCCAGCAGCCAAG  
 CTGACTTACGCCCTGGCAGCTCAGCTTGTGACTCCCATCAAGTTTGAATATGCTAATGGTGTGGTAGGGAACCTGCTCGTCCCT  
 GAAGGAGTCTCAACAATGGTGTGAACATCCACAGAGGCATCCTGAATCTCCTTCAGCTCAACATCAAGAACTCAGAATGTA  
 TACGAGCTGCAGGAGGCTGGAACCTCAGGGAGTGTGCAAGACCTTTATGCCATCAGTGAAGATGAAAGGCTGAACTATCCTT  
 CTGACAAAGACCAGGGACATGAACCAATGTCAGGAAAAGATCATCAAGGATATGGGGTTGGCGTATACAGAGAATGTGCCAAG  
 TGTACGACAGGATCCAAAAACCTGAGAGGTGCAACATCATCAATATATCTTGAAGCCAGTTGCTAGTGGCGTCTTATCCTG  
 GAGGTAGCTGTGAATGAGGTGATCCAGTTCTCACCATTGCTGAGTTGAAAGGAGCTGCTCAGATGGAAACCAAGCAATCATG  
 GTCTTCTTGTGAGATTCAGAGAGCCCCATTGACCCATTGAGGCTCAGTATATTCATCAAGGATCTCTTAAGTACGAGTTCTCC  
 ACTGAGCTTCTTACAGACCCATTACAGTATAAAGATCAACAATGCACAGGCCAGATCGTGGATGTCTGAATCACCTGGTT  
 ATCCACAATGCGGAGAGAGTCCATGAGGATGCCCTCTGAAGTTTTTGGAAATTAATTACAGTCTCTGCCCGGCTGAGCTATAAAG  
 GACCTAGAAATGACCTGGAAACATACAAAAGGATCCCTGCCACAGACAGTGGATCCTGGATGCTGTCCAGCCATTGGAAAGTGC  
 TGCTGCTCTACGATTCATCATGGAGAAATTCCTGTGAGTACATAACTGTTGCTGAAGCAGCTCAGGCTTGATGTCATCTGTT  
 CACATGGTGACAGCAGACAGAGGCCATTAAGCTGATTAAGTCCCTGGCAGTCAATAGCAAAGTAAATCGACAACCCAGTTCTG  
 CGTGAGATTGCTCCTTGGGTATGGCACCATGATTTCCAAACACTGCGTTGAGTTGGCTGTCTGCCCGGCTGAGCTATAAAG  
 CCTATCCAGGACCTTCTTGTGAGGCTGTCAACAGGACGAGACAGAGGACATCATTCTGCTTCTGAAGGTTATGGAAATGCT  
 GGACATCTTGTAGCCTCAAGCAATCACAAAGATCCTGCCATACATGGTACTGCAGCTGCAGTTCTGCCAGTGAAGTTCAT  
 GTTCAAGCAATCATGGCCCTGAGGAACATTGCAAGAAGGACGCCAGAATGGTCCAGGAACCTGCTTCAACTCTATATGGAT  
 AAGGTTCTTACCAGAGCTTCGTATGATGCATGCATTGTGCTGTTGAGACAATGCCTCCAATGGGTTTGGTGACAACCTGTT  
 GCCAACATTTAAAGACTGAAGAGAATCTGCAGTGGCAAGCTTCACTTACTCTCATATGAATCCCTCACCAGGAGCAGTGA  
 GCTATTCATGCTTCAAGTGTGTCAGCTTGAACATTTGCCGTTAAACTCTTGAAGCAAGGCTGGACAGACTCAGCTTACGTTTC  
 AGCAAAGCCATCCATGTGGACGCTTACAACAATCCCTTGATGCTTGGTGTGCTGCTGGTGTCTTCTACATCAATGATGAGTGC  
 ACCAAGCTTCTGGAGGTTGGAGTGAAGACTGAGGAGCTACAGGAGGCTTCTTGAAGAACCCTGTGCTCATGACAATGCTGAC  
 AGGATCACCAAGATGAAGCGTGTCAATTAAGGCTCTCTCTCACTGGAGGCTCTTCCCTCCACCCCAACCGCTGGCCCTCGTCTAC  
 ATCAAGTTCCTTGGACAAGAAATGGCTTCGCTAACATTGACAAAGCCTTAATTGACCAAGCAATGGCGCTTGTACTGGACCC  
 TCTGTTCAAGCATTAGGCAGAAATGCTATCAAGACTCTGCTGTCTGGTGTCTTCTTACACTTTGCTAAGCCTCTGCTCATTACT  
 GAGATGCGTGCATCTGCCAAGTGTGCTGCTTCCAATGGAGCTCAGTCTGTACACTGCTGCTGTGGCTGCAGCAGGTTGCT  
 AAAGTCCAAGCAAAAACAGTACCAGCTCTGCCAGAAAACCTCCATTTCCGCTCAGCTCCTAAAGACAGACATACAGCTTGAGACC  
 GAGATCAGACCAAGTGTGCTATGAACACATTTGCCGTTATGGGTGTAACACTGCTGTACTGCAAGCTGCTGTGCTATCAAAA  
 GCTAAACTCAACTCCCTTGTGCCAGTAAAATTTGCTGCAAGACTTGACATCAATGAGGACACTTTAAGATTGAAGCTCTTCTCT  
 GGAACGTGCTGAAAACACTGCAGTTGTCCATGTTGAGACACCTGCTGTTGCAAGA (N) 55  
 TTACAGGTTTCAAAAAGAAATCAAGAACAATGGCTCGTACTTCTCGGGTGCCTCTGCAGTGGTTTCCAGAAGCAGGAGCAGTG  
 CCTCAAGCTTTGAGGCCATCTACAGAAGAATAAATTCCTAGGGAATGACGCTGCTCCTACCTTTGCCATCATCTTCCGTGCCA  
 TTAGAGCTGACAACATGATGCAGGGATACCAACTTGTGCTTATTTGGACAACCCAGTGCCAGATCCAGATCATTCTCGCTG  
 CCTTAGCTGTGACCTTGTCTTGAATCAGATTCTGCACAGGATGACAGATGAGATGAAATTCATGGTTCTCCTGAAGAAA  
 GATCACATCAAGCAGAACCACATCAATGTGAAAATTTGCTGACATAGATATTGACCTGTACCCAAAGAACAGTGACGTGGTTGTG  
 AAGGTTAATGAAATGAAATGCCCATCAGCAACCTCCATACCCAGCACCCACAGCCAAATATCCAGATCAGATCAAGGGTGAA  
 GGCATTGCTATGTACGCATCTAGCCATGGTCTTCATGAAGTCTACTTTGACAAGAACTCATGGAAGATTAAGTTGTGGACTGG  
 ATGAAGGGGAAGACATGTGGACTGTGTGAAAGGCTGATGGGAGATCAGACAGGAATACCCACACCCACAGGACGCTGACTGACT  
 AAGAATGCAGTCAGCTTTGCTCATTCTGGATTCTGCCAGCTGAGAGCTGCAGGGACACCACCTGGGTGCCGTGTGAAGCTTGA  
 TCTGTGACGCTAGAGAACAGGTGAACGTGCATGGTCTAGAGTCCAGATGCTACTCTGTTGAGCCTGTCTTCCCTGTTGGCT  
 GGCTGCTTCCCTTAAAGACCACAGTGTCACTGTTGGCTTCCACTGTCTACCTGTGATTCCATCCAGAGTCGCCCTGAGAGT  
 ATAAGCAACATTTATGAAAACGGCGTGGACTGAAGGAAAACAGCAGAGGCTCACCTTGCCTGCAGCTGCACCCGCTCAGTGGCT  
 TAACATGTCCCTGTATGTTATTTTAAATAGACTGCAACTGAAGCTGAAAGTCAAGCAAATGGGTACGAGCCTCCTATTGGAT  
 GATTACTGCACGGGTGATTATCCTTCAGCTGGCAACAAGTCAATTAACCCCACTGATGCAATGAGTAGCTTCTCATTTCTTAA  
 ACATCCATGTTGAAAGACACATCCTGTGTTTCATGAAAAGATGTTAGTTTGAAGAAGTCAAAATCATTGGCATGCATCAAGCAGAG  
 AAAACATTTAAGGAGATTGAAGAACTACCAAACTGGGTAACGAACCTGCTAACAATTAATAAACTGGAAGGATAGTGGAGA  
 ACCATTGCTTTGAGGAAGAAATGTGGTCAAAAAAAAATCTGATCGTGATAGGCGATCACTTAAATGTTAGGTGAAATCATA  
 TTGAAGAAAACAAGTTTTTTATTTTTTTTGGAGGTGACTATTTTTTGGCCAGGCAGTGTATAAAGGACTTTCATATGCATTTG  
 ATGTAAGGGGAAGCCAGTGTATGACCATCTGGATAATAGTATTTGAATGGTCAGATCACAGAAGAAAACCTTGAATTCAGAA  
 GCACCTGAATTCAGCAATACTACACAGTGTAAATAAGAGTCTAGAGTCTAAAAAGGTTTTCTGTCAATTAATATTTGTGAA  
 TACTTGATTGTTGACAACAAGAAAATTAACACCATCTTTGTGCAAAAAAAAAAAAAAAAAAAAAA

**Figure 4.43** Full length mRNA sequence of *vtg-1*. Yellow label is start codon, blue label is non-sequenced sequence in 3'v1 4.3/51 insert, green label is stop codon, and pink label is polyA tail.





```

Pagrus_major      NKFLGNEAPPFAIIVRAVRADNKMMGYQLAVYLDKPSTRIQIILAALAA
Sillago_japonica NKFLGNAVAPTFAIIRAVKADKRVLGVELAVYLDKPTARLQIILANLAA
Verasper_moseri  NKYLKKEATPVFAIILRAIRADKMMGYELSVYLDKPTARLQIILAALAA
Liza_subviridis  NKFLGNDAAPTFAIIFRAIRADNMMQGYQLAAVYLDKPSARLQIILAALAL
Melanogrammus_aeglefinus NKYLGASAPPFAIILVRVVRANNQMMGYELSTYLDKPNKRVQLIILAALAK
**:* . .*.***: *.:*.: : **:.****. **:* **
Pagrus_major      DNNWKLKADGALLSKHKVTAKIIGWAECKQYDTMITAETGLVGPSPAARV
Sillago_japonica DNNWKICADGALLSKHKVTAKIIGWAECKQYDTIITAETGLVGPSPAARL
Verasper_moseri  DDNWKLCADGVLLSKFKVNKAVGWAECKQYDTMITAETGLVGPSPAARI
Liza_subviridis  T-----
Melanogrammus_aeglefinus GDNWKLKADGILLSKHKVTAKIIGWAECKEYSTMATAETGLVVKSPAARL

Pagrus_major      RVAWNLDPSAIKHYAKMYDLIPANMLPGLIKGDENSANQLSMTVIATS
Sillago_japonica RVAWNELPSAFKHYARKVYDIYPASSVPLIKGKDETSAKQLSLTVVATS
Verasper_moseri  RVAWNLPITALKRYAEKVYNAIPASMLAGLIQAKDEKSSNQLSATLIATS
Liza_subviridis  -----
Melanogrammus_aeglefinus TVSWETLPISFKTYAKLIYKIYPASILAGLVEGKEINIEKQVSLIVVATS

Pagrus_major      DRTIDFIWKSPTRTFFYKLALHLPYPLPLDGIKGLTPFDG-LADQVHYLFA
Sillago_japonica DKTEDFVWKIPTNTVYKLAHLPIALPLDEFKGLTPPDE-LADQIHYVFA
Verasper_moseri  DRELDLVWKTPIR-VYKLSLRLPIALPLDAIKGLTPPDD-AADRPHYWLA
Liza_subviridis  -----
Melanogrammus_aeglefinus NKDLDLIMKTPKHTIYKLALHLPALPLDGLTNLTPFDNIMEKVRYVLA

Pagrus_major      KAAAACECFNRDALTTFNGRKYNEMPLSCYQVLAQDCTNELKFMVLLKK
Sillago_japonica KAIAACECFRENTLTTFNRRYRNEMPLSCYQVLAQDCTDELKFMVLLKR
Verasper_moseri  KAGAAECFARDTLTTFNRRKYRTEFPLSCYQVLAQDCTDELKFMVLLKK
Liza_subviridis  -----LSCNQILAQDCTDEMFMVLLKK
Melanogrammus_aeglefinus QVYEAQCSYAKNTLTTFNRRYKNEEMPLSCNQVLAQDCKELKFMVLLKK
*** *.:****. :*:*****:

Pagrus_major      DHIEQNHINVKIADIDIDLYPKNTDVIIVKVNMEIPIINNLPHYQHTAKIQ
Sillago_japonica DHIEQSHINVKIADIDIDLYPKNTDVIIVKVNMEIPIINNLPHYQHTAKIQ
Verasper_moseri  DNIEQNHINVKIADIDIDLYPKNADVILKVNMEIPIISNLPHYQHTAKIQ
Liza_subviridis  DHIEQNHINVKIADIDIDLYPKNSDVVVKVNMEMPIISNLPHYQHTANIQ
Melanogrammus_aeglefinus DNIEQNHINVKIADIDIDLYPKNNDVIVKVNMQIPIISNLPHYQHTGTIQ
*.:* . **:.****.*****:* *.:****. :*.*****. .**
IRPKGEGISVYAPSLGLHEVYFDRNSWTVKVVDMWKGQTCGLCGKADGEV
IRPMGEGISVVFAPSHGLQEVNFDKNSWKVKVADWMMKGQTCGLCGKADGEV
IRPKEEGISVVFAPSLGLHEVYFDKNSWKVKVVDWMMKGQTCGLCGKADGEV
IRSKGEGIAMYASSHGLHEVYFDKNSWKVKVVDWMMKGQTCGLCGKADGEI
IKSNGEGISVYAAASHGLHEVYFDKNSWKVKLADWMMKGQTCGICGKADGEV
*.: **:.*: *.:** *.:*.:*.:****.***:*****:
RQEYRTPNGRVTKSAVSYAHSWVLPAESCRDTTECRMKLESVQLEKQVNI
KQEYSSPSGRVTKSAVSYAHSWVLPAESCRDTTECRMKLESVRLERQITV
QQEYHTPNGRRLTKNAVSYAHSWVLPAESCRDTTECRMKLESVQLQKQLNI
RQEYRTPNGRRLTKNAVSFAHSWVLPAESCRDTTGCRVKLESVQLEKQVNV
RQEFRTPNGLAKDAVSYAHSWVLPAESCRDASECRMREQESVKLERQIIII
*.:* .:*.***:****.***:***:***:***:***:***:***:***:

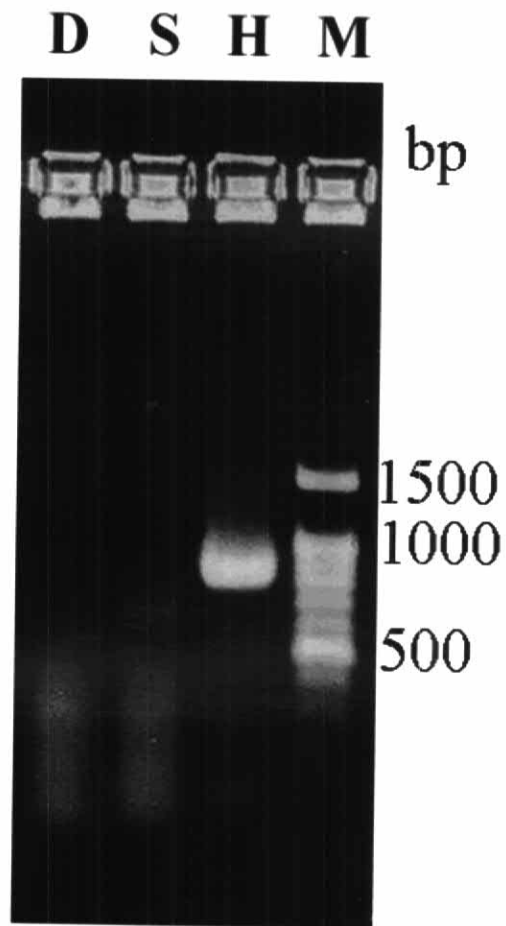
```

C

**Figure 4.45** Alignment of lipoprotein N-terminal domain (A), lipovitellin-phosvitin complex; beta-sheet shell regions (B), and von Willebrand factor (vWF) type D domain (C) of fish Vtg-1. Blue label is lipoprotein N-terminal domain, green label is lipovitellin-phosvitin complex; beta-sheet shell regions, and yellow label is von Willebrand factor (vWF) type D domain.

Additionally genomic sequences at 3'side of *vtg-1* were determined by GenomeWalk PCR. The results of PCR product was shown in Fig. 4.46.



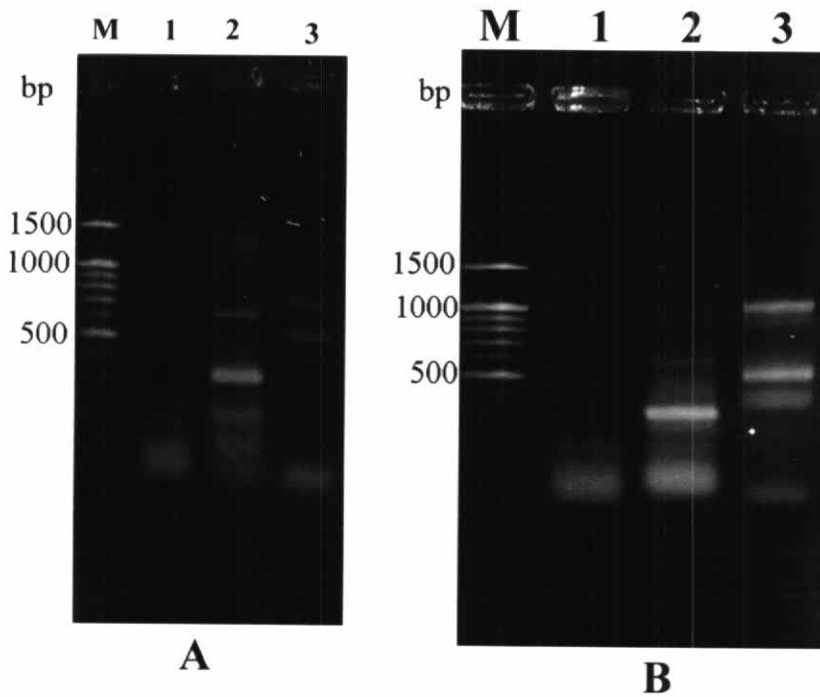


**Figure 4.46** 3' GenomeWalk PCR product of *vtg-1*. Lane M, D, S and H are 100 bp markers, 3' GenomeWalk PCR product of *vtg-1/DraI*, *SspI* and *HaeIII*, respectively.

3'GenomeWalk PCR product of *vtg-1/HaeIII* was cloned and sequenced (2 different size of insert) as shown in appendix B and BLAST analysis found it homologous with *vtg* of orange-spotted grouper (*Epinephelus coioides*) with E-value as  $5e-40$  and  $1e-64$ , respectively (see in appendix C).

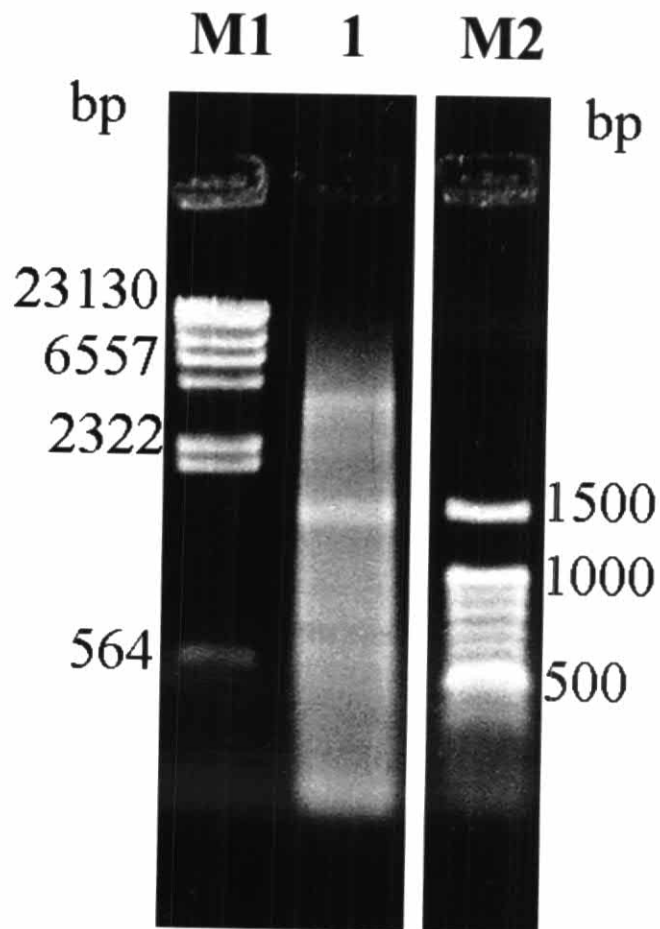
#### 4.5.2 *vtg-3*

Partial cDNA sequence of *vtg-3* at the sizes of 500 bp and 1 kb was amplified from 1<sup>st</sup> strand cDNA templates of liver (Fig. 4.47 A and B, respectively).

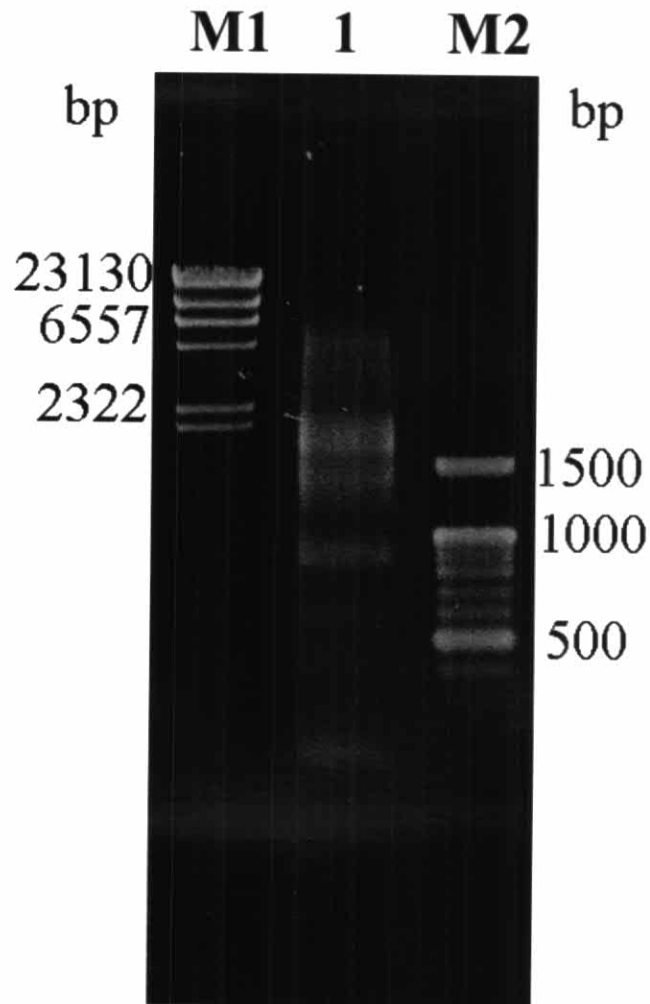


**Figure 4.47** PCR product of *vtg-3* at the sizes of 500 bp (A) and 1 kb (B). Lane M is 100 bp markers, lane 1, 2, 3A, 3B are negative control, positive control, PCR product of *vtg-3* at size 500 bp and 1 kb, respectively.

The results of sequencing and BLAST analysis indicated that both sequences were homologous with phosphovitinless vitellogenin of red seabream (*Pagrus major*) with E-value as  $9e-82$  and  $4e-120$ , respectively (see in appendix B and C). Resulting band from 5' RACE PCR amplification at the size of 1.5 kb (Fig. 4.48) was cloned and sequenced. BLAST analysis indicates that it was homologous with phosphovitinless vitellogenin of red seabream (*Pagrus major*) with E-value as  $5e-98$ . New RACE primer was designed from this sequence and used in 5' RACE PCR amplification. The result in Fig. 4.49 showed bands at the sizes of 1.8, 1.4 and 0.9 kb. The sequencing and BLAST analysis indicated that these bands were homologous with phosphovitinless vitellogenin of red seabream (*Pagrus major*) with E-value as 0.0,  $1e-178$ , and  $1e-92$ , respectively (see in appendix B and C).



**Figure 4.48** 5' RACE PCR product of *vtg-3* separated in 1.2 % agarose gel. Lane M1, M2 and 1 is  $\lambda$ *Hind*III, 100 bp DNA marker and 5' RACE PCR product of *vtg-3*, respectively.



**Figure 4.49** 5' RACE PCR product of *vtg-3* (continued) separated in 1.2 % agarose gel. Lane M1, M2 and 1 is  $\lambda$ *Hind*III, 100 bp DNA marker and 5' RACE PCR product of *vtg-3* (continued), respectively.





```

Liza_subviridis      TKGFLGFYGTNTELFPSGLQIE-KQNASLP-----
Pagrus_major        TKDFWVVFYGTNTELFQVGSSELKTKMPLAIPWKFTA KINVREKKFELDFPS
Gambusia_affinis    TKDFWVVFYGTNTELFQVGSSELKIKRPLAVPWKLSAKINVPERKFELDFPS
Acanthogobius_flavimanus TKSFWLFYGTNTELFQVGSSELKTKSPITIPWNFVVGKFNTRERKFELEFPR
Danio_erio          TKDHFLFHGINTDLFQCGETLKSVMGLPWAFDLKINPKQKQYEMNLT
**..  **.* **.* **.*  . : : *  : *

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B

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Acanthogobius_flavimanus RRMSKEVRT-----SSSTSREQQDRP----TRGME-GSPVPVLNFKALA
Danio_erio              QRVFKEKRD-----ENTSCERKTSSSLPVTQDLD-VTPDPVVTVKALS
Gambusia_affinis       ATQQNNPSS-----DSSES-DRDFNHRHHIILPEN-STTEAIFNVKAFA
Pagrus_major           RRLSKEATQGVRLSSDSASSAERSHSSHHDIVMETSNSTPEAVFSFQAF
Liza_subviridis        GRFPKKQPEGQSQSSDSASSDRGSHQSKRDMIMGGWESTLEDVFNKTFD
: : : : : : : : : : : : : : : : : : : : : : : : : : : : : :
Acanthogobius_flavimanus LSSSQRVEGYEASFYITPEGERQSTQLIVSHVGESSNWKMCVTT---AVA
Danio_erio              LSPQAKPLGYEGVAFYLPATAQKDDIEMIVSEVGEANWKMCANAHFDRTH
Gambusia_affinis       ICENQKPEGYNVVMYHSPEASIRNAKLIVSQVGASTNWRMCVES---SLD
Pagrus_major           MSGNQKPEGYDAAVYITPEANTQNAQLIVSQVGEDTNWKMCVDT-IVNER
Liza_subviridis        QVVNQKPEGYDASVYITPEANTQNAQLIVSQVGEDTNWKMCVDT-TVNAG
. : * : * : * : * : * : * : * : * : * : * : * : * : * : * :
Acanthogobius_flavimanus AQAKAHVAVWGEQCQPYSVS--VSSAYLSGNKPELKA VLRWDRVPEMTVLN
Danio_erio              TSAKAHLRWGAECQTYDVMRVSAAQCPEKSPISITKINWGTLPVFTTV
Gambusia_affinis       AKAKAHITWGDERQTYDMSMEATTAYINGSKPELKA VQWSRISEYMTL
Pagrus_major           THAKAHIRWGAECQSYEMSMRGATAHLPGSKPTL KAKVQWFRVPEMAEI
Liza_subviridis        SGAKAHIRWGAECQSYEISMRAATAYLPGSKPALKAKVHWTRVPEAMEDM
: * : * : * : * : * : * : * : * : * : * : * : * : * : * :
Acanthogobius_flavimanus GKRIAKYIPGMAFLLCFTQEEKNAMQEVSA SVIAASADS IDVKIKFPPEL
Danio_erio              GQIVQEQYVPGVSYIMGFYQKNEENPERQASVT VVASSPETFDLKVKIPER
Gambusia_affinis       GKRIERYIPGVAFLHGFSEKNERNPEREVSATV LAAWTDSVDVEIKLPEY
Pagrus_major           GKRIGSYVPGMAFFLGFSEQHERRNAKQEVSA SVVAAASADSVDVKIKFPEY
Liza_subviridis        GTRIESYIPGMAFLFGFYQQNERNAAQEVSA LVVAESADSDVVKIKFPEY
* : * : * : * : * : * : * : * : * : * : * : * : * : * :
Acanthogobius_flavimanus TLRREALPSPMELISSTETEIQ-----
Danio_erio              TIYKKKIPSPIELVGEAANLTMST-----
Gambusia_affinis       TVYKKAI PFLWNSQWFYENMTCSAEY-----
Pagrus_major           TVYRQAI PVLPPASFLFQPDIRNTTIDSFGQA
Liza_subviridis        TVYHQAI PFLLEPANFQEFHPSITNTTVDAGRA-

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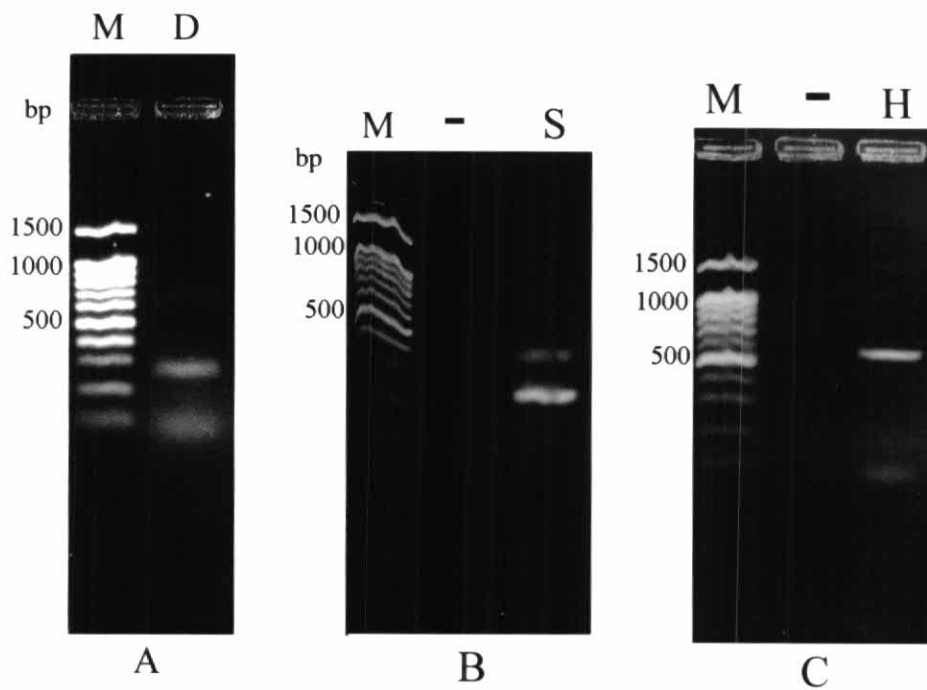
C

**Figure 4.51** Alignment of lipoprotein N-terminal domain (A), lipovitellin-phosvitin complex; beta-sheet shell regions on N-terminus (B), and C-terminus (C) of fish Vtg-3. Yellow label is lipoprotein N-terminal domain, blue label is lipovitellin-phosvitin complex; beta-sheet shell regions.

#### 4.6 ERE determination

ERE of *chg-L* gene was determined by amplifying 5' end of gene using genome walking technique. Genome Walk R primer designed from the DNA fragment of *chg-L* 900 and used in GenomeWalk PCR amplification with *DraI*, *SspI*, and *HaeIII* GenomeWalk library.





**Figure 4.52** Secondary 5' GenomeWalk PCR product of *chg-L* which amplified from *DraI* (A), *SspI* (B), and *HaeIII* (C) GenomeWalk library. The products were separated in 1.5 % agarose gel. Lane M, -, D, S, and H are 100 bp DNA marker, negative control, *chg-L/DraI*, *SspI*, and *HaeIII* GenomeWalk product, respectively.

The result of secondary PCR product was shown in Fig. 4.52. The primary PCR products were smear. After secondary PCR amplification, the products at the sizes of 650 (Fig. 4.52 A) and 350 bp (Fig. 4.52 B) were cloned, sequenced and BLAST analysis (see in appendix C) found it homologous with chorion protein (*Liparis atlanticus*) and choriogenin L (*Oryzias latipes*) with E-value as  $6e-33$  and  $3e-12$ , respectively. *chg-L/D12* sequence which shown exons and introns position shown in Fig. 4.53. Exons were join and translated to amino acid sequence as shown in Fig. 4.54.

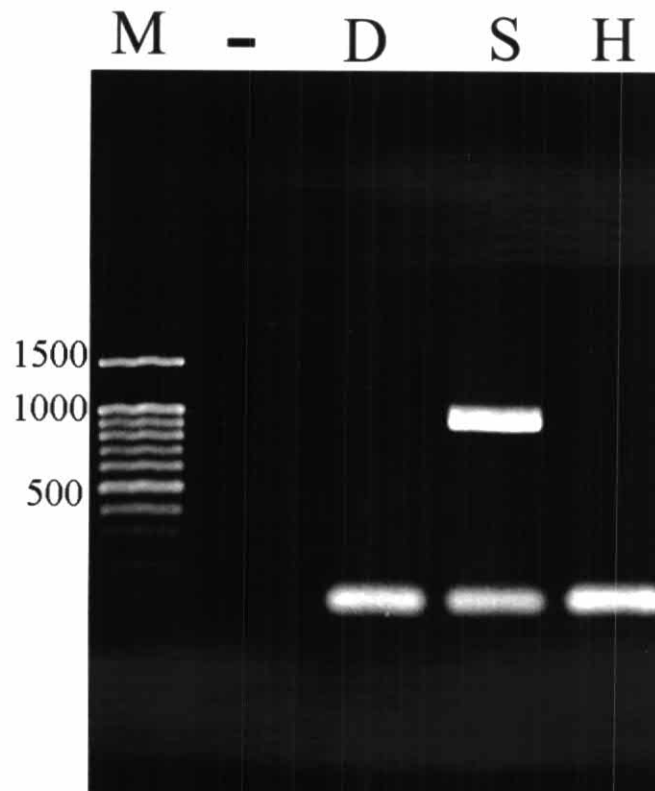
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CCTCACCTGGGGAAGTGTCTGCTGTCGAGAGGATCCTGCGGCTCAAGTGTGGA
TTTTTGAAGCTGAACTGCATGACTGTTTGAGCTCATTGGTGGTAAGTAAAATACAT
AATGCTGAAACTAAACAATATCTTATAATGAATCGTATATGAATATATCATCAAGG
TTTTTGGTCATCTTTGCTAGTTGACAGAAGATTCCCTGACCTACATCTTCACTCTG
AACTACGATCCCCGACCTCTGGGTTCCCTCCCCGTAAGGACCGGCAGTGCAGC
TGTTATTGTGGAATGCCACTACCCAAGGTGTGTTGACTGAAGATATGTGGCAATAA
ACAATCAAAACACAAGTAAAATAATCATCTTAATATGTAACGTAATGTGTAGCCTT
TAGCAGAAATGCACCGTGAAATCTAATCTGCACATGCCAGTGGTAGACGCATGTCA
CGCTGAGGATTTCTTTAGTCTTCTGTTTTAAGCGGGTATGTCTGTCTTGTCTCTTA
GAAAGCACAAATGTGAGCAGCCTTCCTC
```

**Figure 4.53** Nucleotide sequence of *chg-L/D12*. Blue label is exon and non label is introns.

```
gttgcatctgttgctgttgagtgagagagaatgatgctcatgtggaagtcaggagggat
V A S V A V E C R E N D A H V E V R R D
atgtttgggactggccagttggcaatccgaatgacctcaccctggggaactgtcctgct
M F G T G Q L V N P N D L T L G N C P A
gtcgcagaggatcctgcggtcaagtgttgatTTTTGAAGCTGAACTGCATGACTGTTG
V A E D P A A Q V L I F E A E L H D C L
agctcattggtggaagtttgacagaagattccctgacctacatcttcaactctgaactac
S S L V V S L T E D S L T Y I F T L N Y
gatccccgacctctgggttccctccccgtagtaaggaccggcagtgagctggtattgtg
D P R P L G S S P V V R T G S A A V I V
gaatgccactacccaaggaagcacaatgtgagcagccttcctc
E C H Y P R K H N V S S L P
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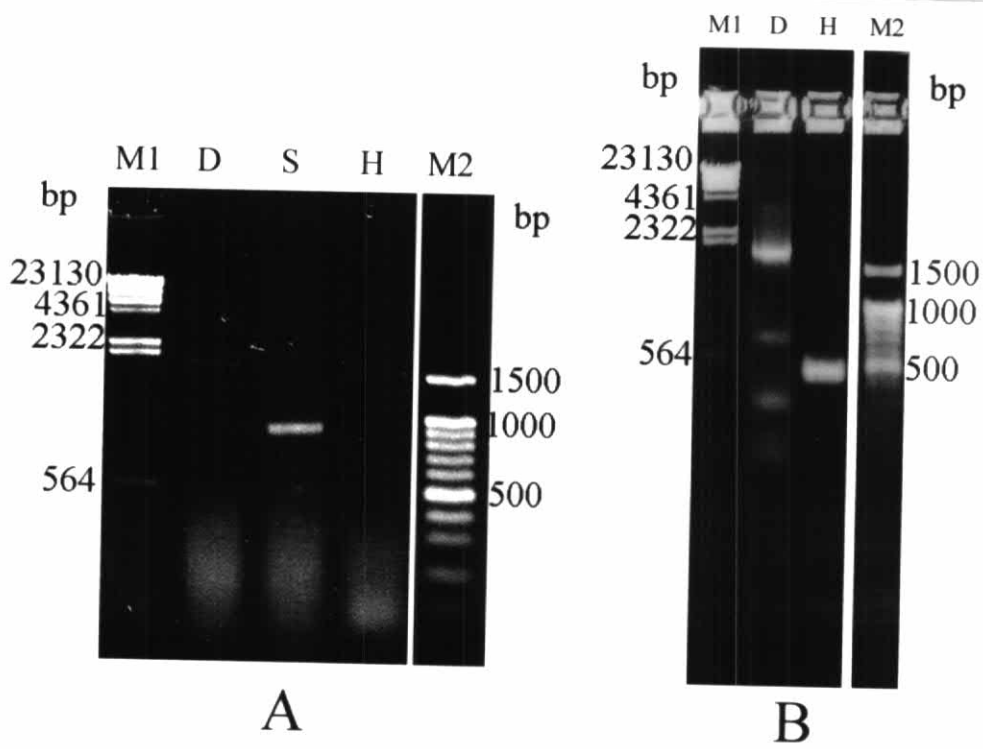
**Figure 4.54** Amino acid sequence translated from *chg-L/D12* sequence.

*chg-L/D12* GW primer were designed from this sequence and used for amplified further genomic sequence with GenomeWalk PCR amplification. *chg-L* GenomeWalk PCR product at size 900 bp was amplified from GenomeWalk *SspI* library (Fig. 4.55). After sequenced (see in appendix B) and BLAST analysis found it homologous with chorion protein (*Sparus aurata*) with E-value as  $8e-14$ . Additionally GenomeWalk PCR amplification of *chg-L* was do again (used EXT *Taq* in amplification), DNA fragment at size 1.76 kb and 900 bp were amplified from *DraI* and *SspI* GenomeWalk library, respectively (result shown in Fig. 4.56 A)



**Figure 4.55** Secondary 5' GenomeWalk PCR product of *chg-L* separated in 1.5 % agarose gel. Lane M, -, D, S, and H is 100 bp DNA marker, negative control, *chg-L/DraI*, *SspI*, and *HaeIII* GenomeWalk product, respectively.

Because *chg-L/DraI* primary PCR product was faint not enough for elution and *chg-L/SspI* may be same sequence as known sequence (*chg-L* D12-S4 see detail in appendix B and C), so primary *chg-L/DraI* and *HaeIII* were dilute 1:50 and used as template in secondary PCR amplification. DNA fragments at size 1.76 kb, 700 bp, and 300 bp were amplified from *chg-L/DraI* primary product as shown in lane D of Fig. 4.56 B. DNA fragment at size 500 bp was amplified from *chg-L/HaeIII* primary product as shown in lane H of Fig. 4.56 B.



**Figure 4.56** Primary (A) and secondary 5' GenomeWalk PCR product of *chg-L* (B) separated in 1.2 % agarose gel. Lane M1, M2, D, S, and H is  $\lambda$ HindIII, 100 bp DNA marker, *chg-L/DraI*, *SspI*, and *HaeIII* GenomeWalk product, respectively.

A DNA fragment at size 1.76 kb was sequenced (see in appendix B) and BLAST analysis (see in appendix C) found it homologous with chorion protein of *Sparus aurata* with E-value as  $3e-34$ . Nucleotide sequence of *chg-L* D12/D20 was shown in Fig. 4.57. The sequence contained 5' upstream region at size 1,424 bp which contained 3 half-site EREs (GGTCA at nucleotide position -326 and -956, and TGACC at nucleotide position -1189). The full site sequence of ERE was not found.

```

CGACGGCCCCGGGCAGGTAAATTAGATCCAGGGTGTGAGAAGTCTGAAAATCCCCATTTGAGC
TCAGAGCAGAATAAACACAGTCTGTTGGTGCCTGATGAAACAAAGGTTCAAAGGTTTTACTTGC
TTTTACAACCTGGATATCTTTTGGCTATTTGGTTATTTTTACAGTAATGATGTTCTTCATATATAT
GTATATATACAGTGATCAGGCATAACATTATGACCAGTTCCTAAATATTGTGTAAGTCTCCATTGTG
CCTCAGTTGTGGCTCATCAGAGAGTGAACATGGGCCCTCTGAGGGTGCCTGTGGTGTCTGTCAATA
TAATGTTGTTGGGGGGGAGGGGGCTTTGGCTCCTAAGGGCTGAAAGGAGGGGCCTCTGTGAATCAG
GCTTGTCCAGTCCATCCCAGAGACACTTGATCAGATTGGGAAGTGTGAATTTGGAGGCCAGGTCA
ACACCTAGTACTGTTGTGTCATGTTTTTTTAGTTGTTCTAAACTATTTTTGTGTGTTTGTCTGTG
TCAGGCTGCATCCTGCTGGGGATGGCTGCTGCTTGTATGGGGTGTGGTTGGGTTGTGGGGGGTAT
CTGGTCTATATATCAACCTTGCTTTGTGCACGTGAAGGATGGTTATGTAGAAAACATACCTACAATG
TGTTACTGTAGAGTCTACTAATGAAATACTGCAAAATATAAATAAATAAATAACAATCTATTATGAAG
ATTTATGTAATATAGGCTTAAACAAATCAAATAAAGCTACGATAGAAAACAAACATTTAATGGAATA
TCAGAAGAGGAGGAGAGGAAGGATCAGCCCAAATTAAGGTTTTGTGATGAGGTGTATGTGGCAGTA
ATATTATCAATAATAAAACAAGATCTTCAGTCCAGTGATTTTTCTCAGGTCTGAGCTGGTCTTA
AAGAATCGTAAATAAATAATATGGTAACTTACTGCAGGTGAAGCTGACAACCTGCCACCGTGGAG
CCATATCGACTGGCTGACATTCACAAAATGTACTTTGGCAAAACACTGAGTTCAGTCTGTGATGTC
AGGGCAGGTAAACCTAACCCAGGGTCAACATAACTTTTCTGCCTCTTTTATTCTGTCTTTCTATA
TATCTTCAAGCTATTTGTAAAGATTTTTTAATAATAATTTGTTTTTCAGAAAATGTGTCTGTATTTATC
TCATCCATAAGAAATAAATGTAATTTAGCTTTTCAGTCAACAGCTGCATTTTATTGGCTACAGCT
AGGGCTTACGAGTCTGCTGAGGGATGGCCAACTGAAGGGGCGGGGAAGTAATGAAAATAAACAGGTT
CACAGCGTTATAAAAGTAGAAGCAAGTCCCACCCTTTACAGCACCTTGAGAATCTCTCAGATCG
CTTGTCACTGTGGAGCCATGGTGTGTAAGTGGACTGCTGCTTGCTTGTGGCACTGGCTCTATTTGC
CAGCGTCTGTGATGCTCAGTGGGGAGAGTACACGCCTTCAAATATCAGAAACCTGCACCTCCTGTG
AAGCAAGAGCCCAAACAAGTGCCTCAAGACACTCAACAGCATAAGCAGACATTTGAAACACCCTTC
AATGGACATACCTGAACCTCCCCGCCTGAACCTGCGCCTGAAATACCATTTGAACCGCTACGTCC
TCAACCTGTTGCATCTGTTGCTGTTGAGTGCAGAGAGAATGATGCTCATGTGGAAGTCAGGAGGGAT
ATGTTTGGG

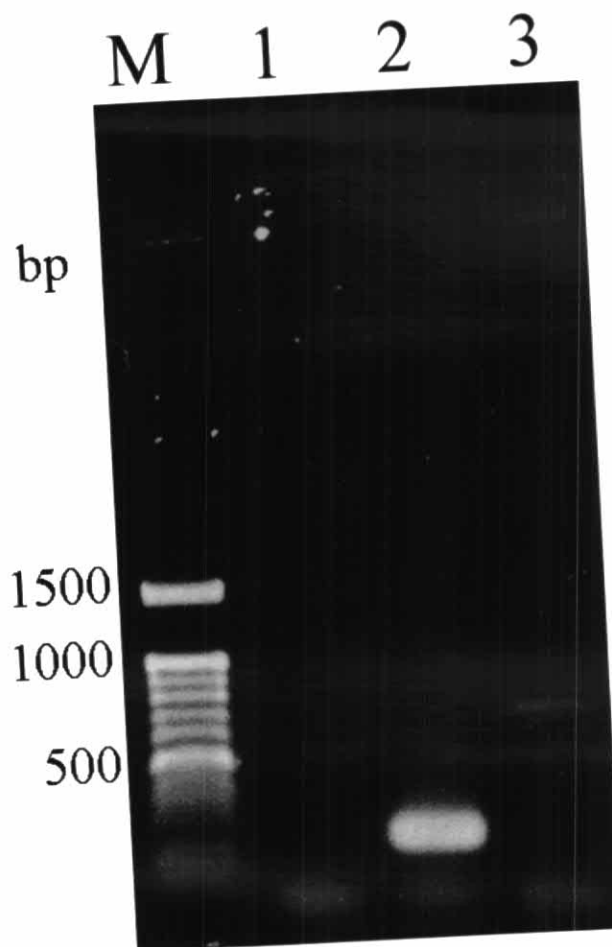
```

**Figure 4.57** Nucleotide sequence of *chg-L* D12/D20. Red alphabet is start codon, blue label is 5' upstream region, and yellow label is half-sites EREs.

#### 4.7 Cloning and characterization of transcriptional intermediary factor 2 (*tif2*) genes.

Partial sequence of *tif2* was amplified by PCR using *tif2* degenerate primer designed from conserved region in basic helix loop helix region, receptor interaction domain, and glutamine rich region of TIF2 from fish and other vertebrate species. PCR amplification with egg, testis, intestine, and liver 1<sup>st</sup> strand cDNA template of

*L. subviridis* but overall PCR product was amplified is other genes or transcript, so specific primer for amplification of *tif2* RID was designed from *tif2* cDNA sequence of zebrafish (*Danio rerio*) which reported in GenBank. DNA fragment at size 700 bp was amplified from adult head and visceral organ 1<sup>st</sup> strand cDNA template of normal *Danio rerio* as shown in Fig. 4.58.



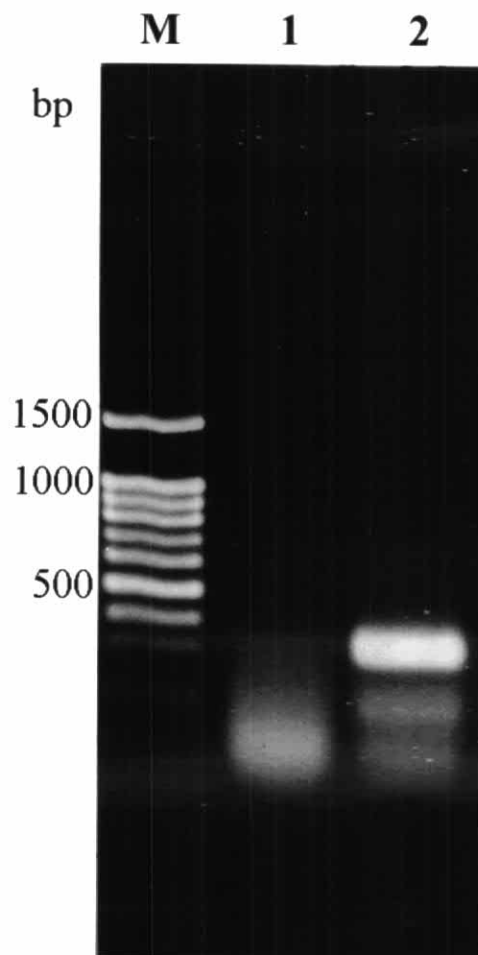
**Figure 4.58** *Danio rerio* *tif2* RID PCR product separated in 1.2 % agarose gel. Lane M, 1, 2, and 3 is 100 bp DNA marker negative control, positive control, and *Danio rerio* *tif2* RID PCR product, respectively



#### 4.8 Estrogen response of *chg-L*, *chg-H*, and *vtg-3* genes in liver of *L. subviridis* detected by semi-quantitative RT-PCR

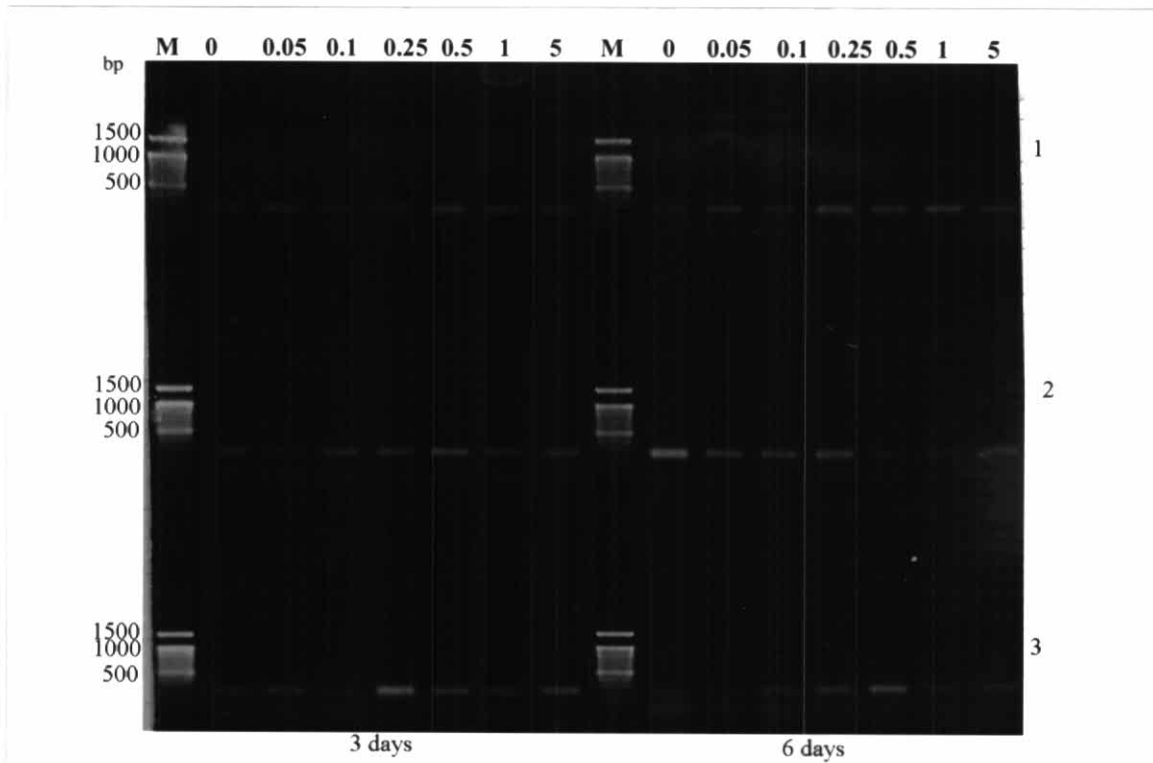
##### 4.8.1 $\beta$ -actin amplification

$\beta$ -actin PCR product of *L. subviridis* at size 300 bp was amplified from liver (used  $\beta$ -actin primer of *Macrobrachium rosenbergii* (designed by Preechaphol) in cross-amplified) result shown in Fig. 4.59, after sequencing and BLAST analysis (see in appendix B and C) it homologous with  $\beta$ -actin of swamp eel (*Monopterus albus*) with E-value as  $1e-117$ . Specific primer of *L. subviridis*  $\beta$ -actin was designed and used for internal control in semi-quantitative RT-PCR analysis.



**Figure 4.59**  $\beta$ -actin PCR product of *L. subviridis* separated in 1.2 % agarose gel. Lane M, 1, and 2 is 100 bp DNA marker, negative control, and  $\beta$ -actin PCR product, respectively.

$\beta$ -actin PCR product were amplified from liver of all sample for used as control as shown in Fig. 4.60

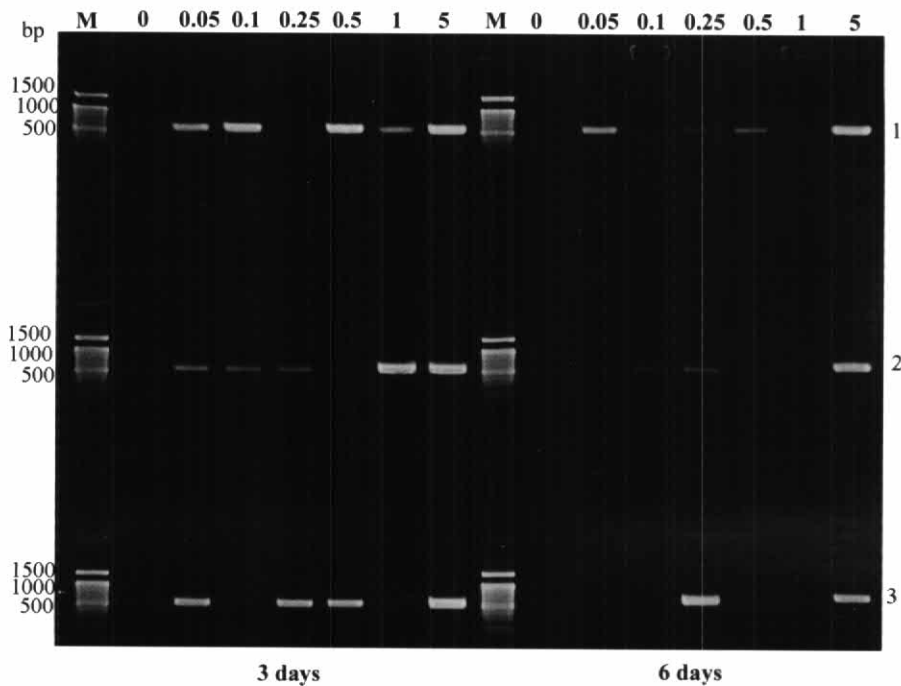


**Figure 4.60** Liver  $\beta$ -actin PCR products of *L. subviridis* exposed with E<sub>2</sub> at dose 0-5 mg/kg for 3 and 6 days separated in 1.6 % agarose gel. Lane M is 100 bp DNA marker, No. of lane indicated dose of E<sub>2</sub> (mg/kg), left and right panel is 3 and 6 days exposed, respectively, up, middle, and low row is replicate 1, 2, and 3, respectively.

Expression level of  $\beta$ -actin gene in liver is constitutive in all sample used in experiment which indicated quality of template were acceptable (Fig. 4.60).

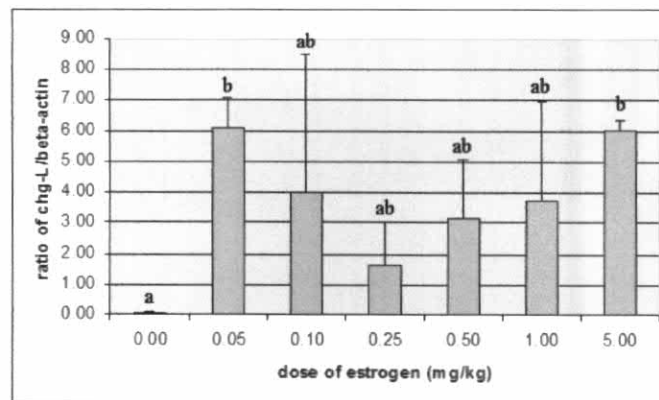
#### 4.8.2 *chg-L* expression level in liver of *L. subviridis* after exposed with $17\beta$ -estradiol (estrogen).

*chg-L* PCR product were amplified from liver of *L. subviridis* exposed with  $E_2$  at dose 0, 0.05, 0.1, 0.25, 0.5, 1, and 5 mg/kg for 3 and 6 days, results of amplification shown in Fig. 4.61.

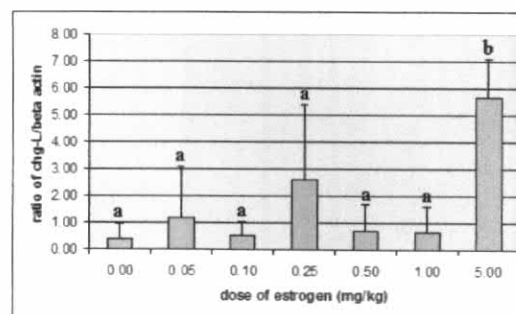


**Figure 4.61** Liver *chg-L* PCR products of *L. subviridis* exposed with  $E_2$  at dose 0-5 mg/kg for 3 and 6 days separated in 1.6 % agarose gel. Lane M is 100 bp DNA marker, No. of lane indicated dose of  $E_2$  (mg/kg), left and right panel is 3 and 6 days exposed, respectively, row 1, 2, and 3 is replicate 1, 2, and 3, respectively.

Intensity of *chg-L* band were measured and normalized with actin (ratio of *chg-L*/ $\beta$  actin). mRNA expression level of *chg-L* in liver of *L. subviridis* increased significantly ( $P < 0.05$ ) after exposed with estrogen at dose 0.05 and 5 mg/kg for 3 days but mRNA expression level of *chg-L* not changed when mullets exposed with  $E_2$  at dose 0, 0.1, 0.25, 0.5, and 1 mg/kg (Fig. 4.62 A). mRNA expression level of *chg-L* in liver of *L. subviridis* increased significantly ( $P < 0.05$ ) after exposed with estrogen at dose 5 mg/kg for 6 days but mRNA expression level of *chg-L* not changed when mullets exposed with  $E_2$  at dose 0, 0.05, 0.1, 0.25, 0.5, and 1 mg/kg (Fig. 4.62 B).



A

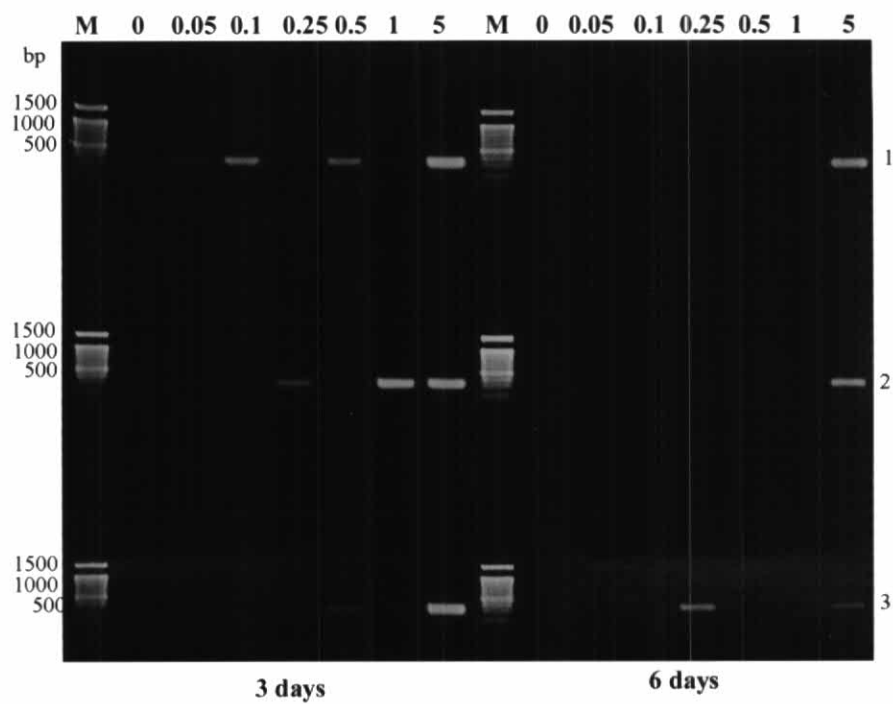


B

**Figure 4.62** mRNA expression level of *chg-L* gene in liver of *L. subviridis* which exposed estrogen by intraperitoneally injection at 3 days (A) at 6 days (B).

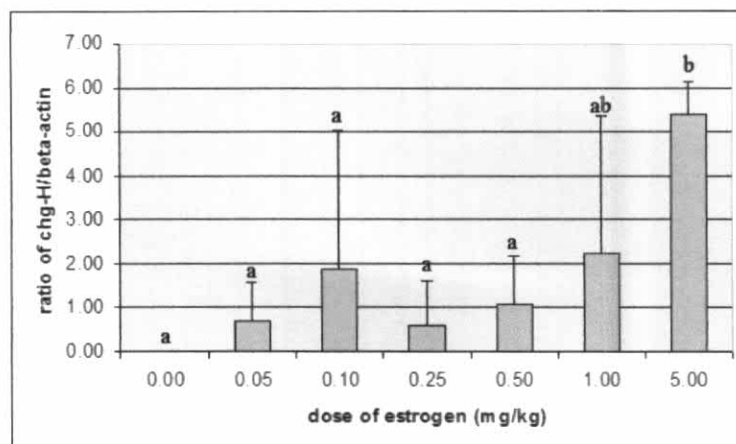
### 4.8.3 *chg-H* expression level in liver of *L. subviridis* after exposed with 17 $\beta$ -estradiol (estrogen).

*chg-H* PCR product were amplified from liver of *L. subviridis* exposed with E<sub>2</sub> at dose 0, 0.05, 0.1, 0.25, 0.5, 1, and 5 mg/kg for 3 and 6 days, results of amplification shown in Fig. 4.63.

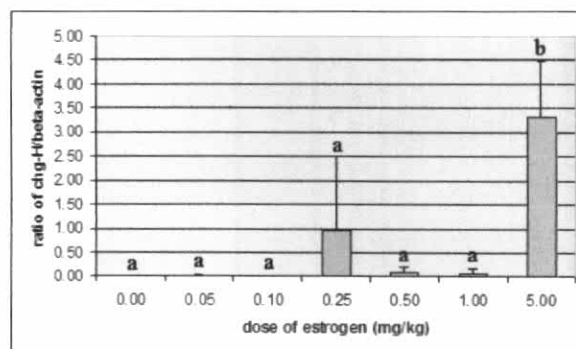


**Figure 4.63** Liver *chg-H* PCR products of *L. subviridis* exposed with E<sub>2</sub> at dose 0-5 mg/kg for 3 and 6 days separated in 1.6 % agarose gel. Lane M is 100 bp DNA marker, No. of lane indicated dose of E<sub>2</sub> (mg/kg), left and right panel is 3 and 6 days exposed, respectively, row 1, 2, and 3 is replicate 1, 2, and 3, respectively.

Intensity of *chg-H* band were measured and normalized with actin (ratio of *chg-H*/ $\beta$  actin). mRNA expression level of *chg-H* in liver of *L. subviridis* increased significantly ( $P < 0.05$ ) after exposed with estrogen at dose 5 mg/kg for 3 and 6 days but do not change when mullets exposed with  $E_2$  at dose 0, 0.05, 0.1, 0.25, 0.5, and 1 mg/kg (Fig. 4.64)



A

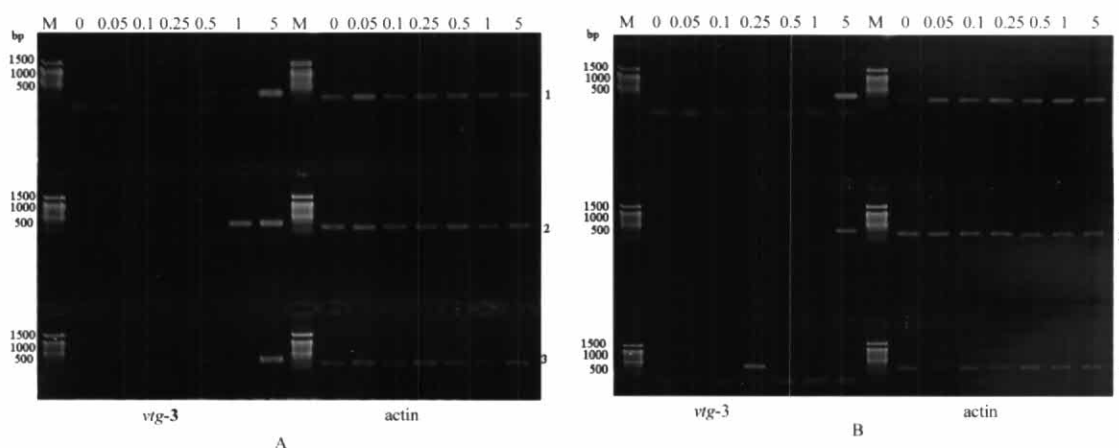


B

**Figure 4.64** mRNA expression level of *chg-H* gene in liver of *L. subviridis* which exposed estrogen by intraperitoneally injection at 3 days (A) at 6 days (B).

#### 4.8.4 *vtg-3* expression level in liver of *L. subviridis* after exposed with 17 $\beta$ -estradiol (estrogen).

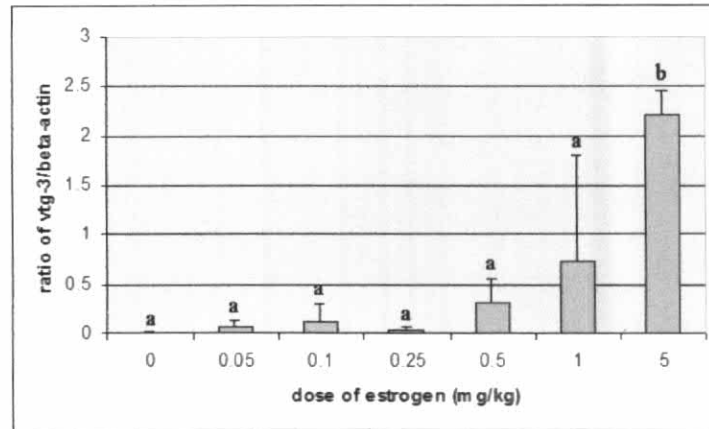
*vtg-3* PCR product were amplified from liver of *L. subviridis* exposed with E<sub>2</sub> at dose 0, 0.05, 0.1, 0.25, 0.5, 1, and 5 mg/kg for 3 and 6 days, results of amplification shown in Fig. 4.65 A and B, respectively.



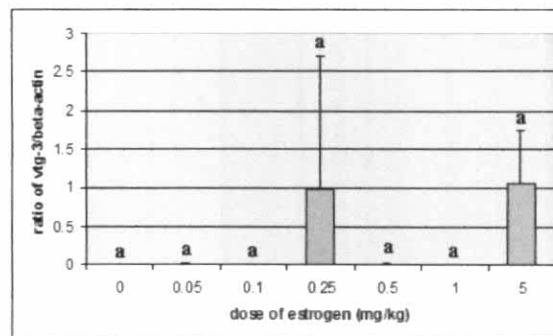
**Figure 4.65** Liver *vtg-3* and  $\beta$  actin PCR products of *L. subviridis* exposed with E<sub>2</sub> at dose 0-5 mg/kg for 3 days (A) and 6 days (B) separated in 1.6 % agarose gel. Lane M is 100 bp DNA marker, No. of lane indicated dose of E<sub>2</sub> (mg/kg), left and right panel is *vtg-3* and  $\beta$  actin PCR products respectively, row 1, 2, and 3 is replicate 1, 2, and 3, respectively.

Intensity of *vtg-3* band were measured and normalized with actin (ratio of *vtg-3*/ $\beta$  actin). mRNA expression level of *vtg-3* in liver of *L. subviridis* increased significantly ( $P < 0.05$ ) after exposed with estrogen at dose 5 mg/kg for 3 and 6 days but do not change when mullets exposed estrogen at dose 0, 0.05, 0.1, 0.25, 0.5, and 1 mg/kg. Results in Fig. 4.66 B shown mRNA expression level of *vtg-3* in

liver of *L. subviridis* not changes after exposed estrogen at dose 0, 0.05, 0.1, 0.25, 0.5, 1, and 5 mg/kg for 6 days.



A



B

**Figure 4.66** mRNA expression level of *vtg-3* gene in liver of *L. subviridis* which exposed estrogen by intraperitoneally injection at 3 days (A) at 6 days (B).