เอนไซม์และยืนโพลีคีไทด์ซินเทสในเจตมูลเพลิงแดง



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POLYKETIDE SYNTHASE ENZYMES AND GENES IN PLUMBAGO INDICA

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การศึกษาในครั้งนี้เกี่ยวกับการค้นหาเคนไซม์ และยีนของเอนไซม์โพลีคีไทด์ซินเทสใน เจตมูลเพลิงแดงเพื่อความเข้าใจเกี่ยวกับชีวสังเคราะห์ระดับเอนไซม์ของสารพลัมบาจิน การเพาะเลี้ยง เนื้อยื่อของเจตมูลเพลิงแดงในครั้งนี้สามารถชักนำให้เกิดเป็นเนื้อเยื่อแคลลัส รากเพาะเลี้ยง และต้นพืช ขนาดเล็กในหลอดทดลอง โดยแคลลัสที่ได้เกิดจากการใช้ส่วนต้นอ่อนของพืช ภายใต้สภาวะของอาหาร ลูตร MS ที่ประกอบด้วย 2,4-D 1 มิลลิกรัมต่อลิตร และ BA 0.1 มิลลิกรัมต่อลิตร ส่วนรากเพาะเลี้ยง เกิดจากการใช้ใบอ่อน ภายใต้สภาวะของอาหารสูตร B5 ที่ประกอบด้วย NAA 1 มิลลิกรัมต่อลิตร และ kinetin 0.1 มิลลิกรัมต่อลิตร จากนั้นทำการเพิ่มปริมาณรากเพาะเลี้ยง ภายใต้สภาวะของอาหารเหลว สูตร MS ในขณะที่ต้นพืชขนาดเล็กในหลอดทดลองเกิดจากการใช้ส่วนข้อของพืช ภายใต้สภาวะของ อาหารสูตร LS เมื่อทำการวิเคราะห์หาปริมาณสารพลัมบาจินด้วยเทคนิค HPLC พบว่าส่วนต้นและราก ของต้นพืชที่เกิดจากการเพาะเลี้ยงเนื้อเยื่อพืช มีการสะสมของสารมากกว่าแคลลัสและรากเพาะเลี้ยง อย่างไรก็ มื่อใช้สารกัมมันตรังสี malonvl-CoA เพื่อตรวจหากิจกรรมของเอนไซม์โพลีคีไทด์ซินเทสใน สารสกัดเอนไซม์จากเนื้อเยื่อเพาะเลี้ยงต่างๆ ไม่พบว่ามีกิจกรรมของเอนไซม์โพลีคีไทด์ซินเทสในการ สร้างพลัมบาจิน ดังนั้นจึงมีการนำเทคนิคชีววิทยาโมเลกุล มาใช้ในการหายีนของโพลีคีไทด์ซินเทส โดยการทำ cDNA library ที่ได้จากการใช้อาร์เอ็นเอ (RNA) จากชิ้นส่วนราก วิธีการนี้ทำให้ค้นพบ cDNA จำนวน 1 ชิ้นที่ลมบูรณ์ ซึ่งเมื่อนำยืนนี้ไปแสดงออกและชักนำให้สร้างโปรตีนใน Escherichia coli แล้ว พบว่าเอนไซม์บริสุทธิ์ที่ได้สามารถใช้ acetyl-CoA ทำปฏิกิริยากับ malonyl-CoA ผลิตภัณฑ์ ที่เกิดจากปฏิกิริยานี้พบว่าเป็นสารกลุ่มไพโรน (pyrone) หลายชนิด ซึ่งได้แก่ triketide, tetraketide, pentaketide อย่างละ 1 ชนิด และ hexaketide pyrone อีก 3 ชนิด นอกจากนี้การใช้เอนไซม์บริสุทธิ์ ร่วมกับการเติมสารสกัดเอนไซม์จากเนื้อเยื่อเพาะเลี้ยงต่างๆ พบว่ากิจกรรมของเอนไซม์สามารถเปลี่ยน สาร hexaketide pyrone ตัวหนึ่งไปเป็นสารอีกชนิดหนึ่งที่ยังไม่รู้โครงสร้างทางเคมี อย่างไรก็ดีการ ทดลองที่ผ่านมายังไม่สามารถค้นพบการสร้างแนพโทควิโนนพลัมบาจินในสภาวะต่างๆที่มีการศึกษานี้ ปัจจัยต่างๆที่เกี่ยวข้องกับการสร้างสารพลัมบาจินในหลอดทดลองยังไม่ถูกค้นพบ และต้องดำเนินการ วิจัยต่อไป

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Studies on the enzyme and gene of a polyketide synthase in Plumbagin indica have been performed in order to understand the enzymatic formation of plumbagin in plant. Tissue cultures of P. indica were successfully established in forms of callus, root culture and in vitro plantlets. Callus cultures derived from young stem explants were generated on MS medium supplemented with 1.0 g/l 2,4-dichlorophenoxyacetic acid (2,4-D) and 0.1 mg/l 6-benzylaminopurine (BA). Root cultures were established from young leaf segments on Gamborg's B5 (B5) medium supplemented with 1.0 mg/l α-napthaleneacetic acid (NAA) and 0.1 mg/l kinetin. Induced roots were cultured in Murashige and Skoog (MS) liquid medium without growth regulators for root proliferation. Plantlets were regenerated from nodal segments and maintained on Linsmaier and Skoog (LS) medium also without growth regulators. Plumbagin content present in these P. indica tissues were determined using high performance liquid chromatography (HPLC). The analysis revealed that the content of plumbagin in the aerial parts and roots of the micropropagated plantlets was significantly higher than that in callus and root cultures. By using a radiolabelled compound of malonyl-CoA as substrate, a standard enzyme assay was established to detect plant polyketide synthase activities in crude protein extracts prepared from the various tissue cultures of P. indica. The formation of plumbagin was, however, not detected in the established enzyme assay conditions. The technique of molecular cloning was, therefore, introduced to express and characterize the enzyme. A cDNA encoding a polyketide synthase, PinPKS, was isolated from a cDNA library prepared from the RNA of P. indica roots. The recombinant PinPKS was expressed in Escherichia coli and assayed for its polyketide synthase activity with acetyl-CoA as a starter molecule and malonyl-CoA as a co-substrate. Analysis of the resulting reaction mixture revealed that there was enzymatic formation of various sizes of pyrone products. These included the pyrones of triketide, tetraketide, pentaketide (one each) and three hexaketides. Addition of a crude protein extract of P. indica tissues into the reaction mixture could lead to an unknown compound which has not been identified. With many attempts, no structures of naphthoquinones formed from the hexaketide level have been found in the enzyme assay. Required factors for plumbagin formation in vitro remain to be discovered.

Program Pharmaceutical Chemistry and Natural Products

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| Figure 40 | Proposed steps of the formation triketide, tetraketide, pentaketide and | |
| | hexaketide pyrones from acetyl-CoA and malonyl-CoA in the standard | |
| | reaction mixture catalyzed by the PinPKS enzyme1 | 14 |
| Figure 41 | Proposed sequence taking place in the reaction mixture catalyzed | |
| | hy PinPKS 1 | 15 |

ABBREVIATIONS

Abs absorbance

ACS acridone synthase

ALS aloesone synthase

APS ammonium persulfate

B5 Gamborg medium

BA 6-benzylaminopurine

BAS benzalacetone synthase

BBS bibenzyl synthase

bp base pair

BPS benzophenone synthase

cDNA complementary DNA

CHS chalcone synthase

Ci Curie

cm centimeter (s)

CoA coenzyme-A

cpm count per minute

CTAS Coumaroyl triacetic acid lactone synthase

dNTP 2'-deoxynucleoside 5'-triphosphate

dpm disintegrations per minute

DNA deoxyribonucleic acid

DTT dithiothreitol

eV electron volt

h hour

HEDS homoeriodictyol/eriodictyol synthase

HPLC high performance liquid chromatography

IAA indole-3-acetic acid

IPTG isopropylthio-β-D-galactoside

g gram (s)

kinetin kinetin-6-furfurylaminopurine

kDa kilodalton (x10³ Da)

kg kilogram

L liter (s)

LC-ESI-MS liquid chromatography electrospray ionization mass spectrometry

LC-MS liquid chromatography mass spectrometry

LS Linsmaier and Skoog medium

m meter (s)

M molar

min minute (s)

mg milligram

ml milliliter

MMLV-RT moloney murine leukemia virus-reverse transcriptase

MS Murashige and Skoog medium

mRNA messenger RNA

M⁺ molecular ion

m/z mass to charge ratio

nm nanometer (s)

NAA α -naphthaleneacetic acid

NADPH nicotinamide adenine dinucleotide phosphate

OD optical density

OKS octaketide synthase

PCS polyketide chromone synthase

PCR polymerase chain reaction

Pfu plaque forming unit

PinPKS Plumbago indica polyketide synthase

PKS polyketide synthase

RNA ribonucleic acid

rpm rotation per minute

RT-PCR reverse transcriptase PCR

SDS sodium dodecyl sulfate

SDS-PAGE sodium dodecyl sulfate-polyacrylamide gel electrophoresis

sec second

STS stilbene synthase

TEMED N, N, N', N'-tetramethylethylenediamine

TLC thin layer chromatography

Tris tris (hydroxymethyl) aminomethane

U unit

V voltage

VPS phlorisovalerophenone synthase

v/v volume per volume (concentration)

w/v weight per volume (concentration)

w/w weight per weight (concentration)

X-gal 5-bromo-4-chloro-3-indolyl-β-D-galactoside

2,4-D 2,4-dichlorophenoxyacetic acid

2-PS 2-pyrone synthase

°C degree Celsius

 λ_{max} wavelength at maximum absorption

μg microgram (s) (10⁻⁶ g)

μl microliter (s) (10⁻⁶ l)

μM micromolar (s)