

## REFERENCES

- Akimenko, N. M., Tkhruni, F. N., Rusinov, A. V., Chernik, T. P. and Kriviskii, A. S. 1976. DNA compact form in solution. VII. Transforming activity of precompact forms of DNA. **Mol. Biol. (Mosk)**. 10: 1035-1041.
- Arps, P. J. and Winkler, M. E. 1987. Structural analysis of the *Escherichia coli* K-12. *hisT* operon by using a kanamycin resistance cassette. **J. Bacteriol.** 169: 1061-1070.
- Barnett, M. J. and Long, S. R. 1990. DNA sequence and translational product of a new nodulation regulatory locus: SyrM has sequence similarity to NodD proteins. **J. Bacteriol.** 172: 3695-3700.
- Bennett, D. C. 1974. Similarity in the sequence of *Escherichia coli* dihydrofolate reductase with other pyridine nucleotide-requiring enzymes. **Nature**. 248: 67-68.
- Binotto, J., Mac-Lachlan, P. R. and Sanderson, K. E. 1991. Electrotransformation in *Salmonella typhimurium* LT 2. **J. Microbiol.** 37: 474-477.
- Bryan, L. E. and Kwan, S. 1981. Aminoglycoside-resistant mutants of *Pseudomonas aeruginosa* deficient in cytochrome d, nitrite reductase and aerobic transport. **Antimicrob. Ag. Chemother.** 19: 958-965.
- Bryan, L. E., O Hara, K. and Wong, S. 1984. Lipopolysaccharide changes in impermeability-type aminoglycoside resistance in *Pseudomonas aeruginosa*. **Antimicrob. Ag. Chemother.** 26: 250-257.
- Chanama, S. and Crawford, R. L. 1997. Mutational analysis of *pcpA* and its role in pentachlorophenol degradation by *Sphingomonas (Flavobacterium) chlorophenolica* ATCC 39723. **Appl. Environ. Microbiol.** 63: 4833-4838.

- Chasseaud, L. F. 1979. The role of glutathione and glutathione *S*-transferase in the metabolism of chemical carcinogens and other electrophilic agents. **Adv. Cancer. Res.** 29: 175-274.
- Cohen, S. N., Chang, A. C. Y. and Hsu, L. 1972. Nonchromosomal antibiotic resistance in bacteria: genetic transformation of *Escherichia coli* by R-factor DNA. **Pro. Natl. Acad. Sci. USA.** 69: 2110-2114.
- Coleman, K., Athalye, M. and Clancey et al. 1994. Bacterial resistance mechanisms as therapeutic targets. **J. Antimicrob. Chemother.** 33: 1091-1095.
- Conley, E. C. and Saunders, J. R. 1984. Recombination-dependent recircularization of linearized pBR 322 plasmid DNA following transformation of *Escherichia coli*. **Mol. Gen. Genet.** 194: 211-218.
- Crosby, D. G., Beynon, K. I., Stoll, G. G., Vonk, J. W., Greve, P. A. and Korte, F. 1981. Environmental chemistry of pentachlorophenol. **Pure Appl. Chem.** 53: 1052-1080.
- Davies, J. E. 1983. Resistance to aminoglycosides: mechanisms and frequency. **Rev. Infect. Dis.** 5: 261-270.
- Deveraux, J., Haeverli, P. and Smithes, O. 1984. A comprehensive set of sequence analysis programs for the VAX. **Nucleic Acids Res.** 12: 387-395.
- Dower, W. J., Miller, J. F. and Ragsdale, C. W. 1988. High efficiency transformation in *E. Coli* By high voltage electroporation. **Nucleic Acid Res.** 16: 6127-6145.
- Ederer, M. M., Crawford, R. L., Herwig, R. P. and Orser, C. S. 1997. PCP degradation is mediated by closely related strains of the genus *Sphingomonas*. **Molecular Ecology.** 6: 39-49.
- Edgehill, R. U. and Finn, R. K. 1982. Isolation, characterization and growth kinetics of bacteria metabolizing pentachlorophenol. **Appl. Environ. Microbiol.** 45: 1122-1125.

- Eggink, G., Engle H., Vriend, G., Terpstra, R. and Witholt, B. 1990. Rubredoxin reductase of *Pseudomonas oleovorans*: structural relationship to other flavoprotein oxidoreductases based on one NAD and two FAD fingerprints. **J. Mol. Biol.** 212: 135-142.
- Foster, T. J. 1983. Plasmid-determined resistance to antimicrobial drugs and toxic metal ions in bacteria. **Microbiol. Rev.** 47: 369-375.
- Gasparich, G. E., Hackett, K. J., Stamburski, C., Renaudin, J. and Bove, J. M. 1993. Optimization of method for transferring *Spiroplasma citri* strain R8A2 HP with the spiroplasma virus SpV1 replicative form. **Plasmid.** 29: 193-205.
- Glumova, E. F. and Prozorov, A. A. 1981. Comparative study of chromosome and plasmid transformation in *Bacillus subtilis*: the effect of lysozyme and polyethylene glycol. **Genetika.** 17: 1581-1587.
- Gonzalez, J. F. and Hu, W. 1991. Effect of glutamate in the degradation of pentachlorophenol by *Flavobacterium* sp. **Appl. Microbiol. Biotechnol.** 35: 100-104.
- Grindley, N. D. F. and Joyce, C. M. 1980. Genetic and DNA sequence analysis of the kanamycin resistance transposon Tn 903. **Pro. Natl. Acad Sci USA.** 77: 7176-7180.
- Hamilton, C. M., Aldea, M., Washburn, B. K., Babitzke, P. and Kushner, S. R. 1989. New method for generating deletions and gene replacements in *Escherichia coli*. **J. Bacteriol.** 171: 4617-4622.
- Hardy, D. J., Legeai, R. J. and Callaghan, R. J. 1980. *Klebsiella neonatal* infections: mechanism of broadening aminoglycoside resistance. **Antimicrob. Ag. Chemother.** 18: 542-549.
- Jakoby, W. B. and Habig, W. H. 1980. Glutathione transferase, p. 63-94. In Jakoby, W. B. (ed). **Enzymatic basic of detoxication**, vol. 2. Academic Press, New York.

- Jasin, M. and Shimmel, P. 1984. Deletion of an essential gene in *Escherichia coli* by site-specific recombination with linear DNA fragments. **J. Bacteriol.** 159: 783-786.
- Jenson, s. and Renberg, L. 1972. Contaminats in pentachlorophenol: chlorination dioxin and perdioxin. **EMSO.** 1: 62-65.
- Khasanov, F. K., Zvingila, D. J., Zainullin, A. A., Prozorov, A. A. and Bashkirov, V. I. 1992. Homologous recombination between plasmid and chromosomal DNA in *Bacillus subtilis* requires approximately 70 bp of homology. **Mol. Gen. Genet.** 234: 494-497.
- King, S. R. and Richardson, J. P. 1986. Role of homology and pathway specificity for recombination between plasmid and bacteriophage. **Mol. Gen. Genet.** 204: 141-147.
- Klapath, T. R., Guerinot, M. L. and Lynd, L. R. 1996. Electrotransformation of *Clostridium thermosaccharolyticum*. **Ind. Microbiol.** 16: 342-347.
- Kucers, A., Crowe, S., Grayson, M. L. and Hog, J. 1997. **The use of antibiotics.** Bulterworth-Heinemann. England.
- Lange, C. C., Sshneider, B. L. and Orser, C. S. 1995. Verification of the role of PCP 4-monooxygenase in chlorine elimination from pentachlorophenol by *Flavobacterium* sp. strain ATCC 39723. **Biochem. Biophys. Res. Commun.** 219: 146-149.
- Lange, C.C. and Orser, C. S. 1994. Molecular analysis of pentachlorophenol degradation. **Biodegradation.** 5: 277-288.
- Lee, M. S., Seok, C. and Morrison, D. A. 1998. Insertion duplication mutagenesis in *Streptococcus pneumoniae*: targeting fragment length is a critical parameter in use as a random insertion tool. **Appl. Environ. Micro.** 64: 4796-4802.
- Levi-Meyrueis, C., Fodor, K. and Schaeffer, P. 1980. Polyethylene-induced transformation of *Bacillus subtilis* protoplasts by bacterial chromosomal DNA. **Mol. Gen. Genet.** 179: 589-594.

- Lupski, J. R. 1987. Molecular mechanisms for transposition of drug-resistance genes and other movable genetic elements. **Rev. Infect. Dis.** 9: 357-365.
- Meletzus, D. and Eichenlaub R. 1991. Transformation of the phytopathogenic bacterium *Clavibacter michiganense* subsp. *michiganense* by eletroporation and development of a cloning vector. **J. Bacteriol.** 173: 184-190.
- Michelsen, B. K. 1995. Transformation of *E. coli* increase 260-fold upon inactivation of T<sub>4</sub> DNA Ligase. **Anal. Biochem.** 225: 172-174.
- Moellering, R. C. Jr. 1983. In vitro antibacterial activity of the aminoglycoside antibiotics. **Rev. Infect. Dis.** 5: 212-219.
- Oden, K. L., Deveaux, L. C., Vibat, C. R. T., Cronan Jr, J. E. and Gennis, R. B. 1990. Genomic replacement in *Escherichia coli* K-12. using covalently closed circular plasmid DNA. **Gene.** 96: 29-36.
- Okamoto, A., Kosugi, A., Koizumi, Y., Yanagida, F. and Udaka, S. 1997. High efficiency transformation of *Bacillus brevis* electroporation. **Biosci. Biotech. Biochem.** 6: 202-203.
- Orser, C. S., Topp, E. and Xun, L. 1992. Purification and characterization of a tetrachloro-*p*-hydroquinone reductive dehalogenase from a *Flavobacterium* sp. **J. Bacteriol.** 174: 8003-8007.
- Orser, C. S., Lange, C. C., Xun, L., Zahrt, T. C. and Schneider, B. J. 1993. Cloning, sequence, analysis, and expression of the *Flavobacterium* pentachlorophenol 4-monooxygenase gene in *Escherichia coli*. **J. Bacteriol.** 175: 411-416.
- Papadopoulou, B. and Dumas, C. 1997. Parameters controlling the rate of gene targeting frequency in the protozoan parasite *Leishmania*. **Nucleic Acids Res.** 25: 4278-7286.
- Patnaik, P. 1992. **A comprehensive guide to the hazardous properties of chemical substances.** Van Nostrand Reinhold., USA.

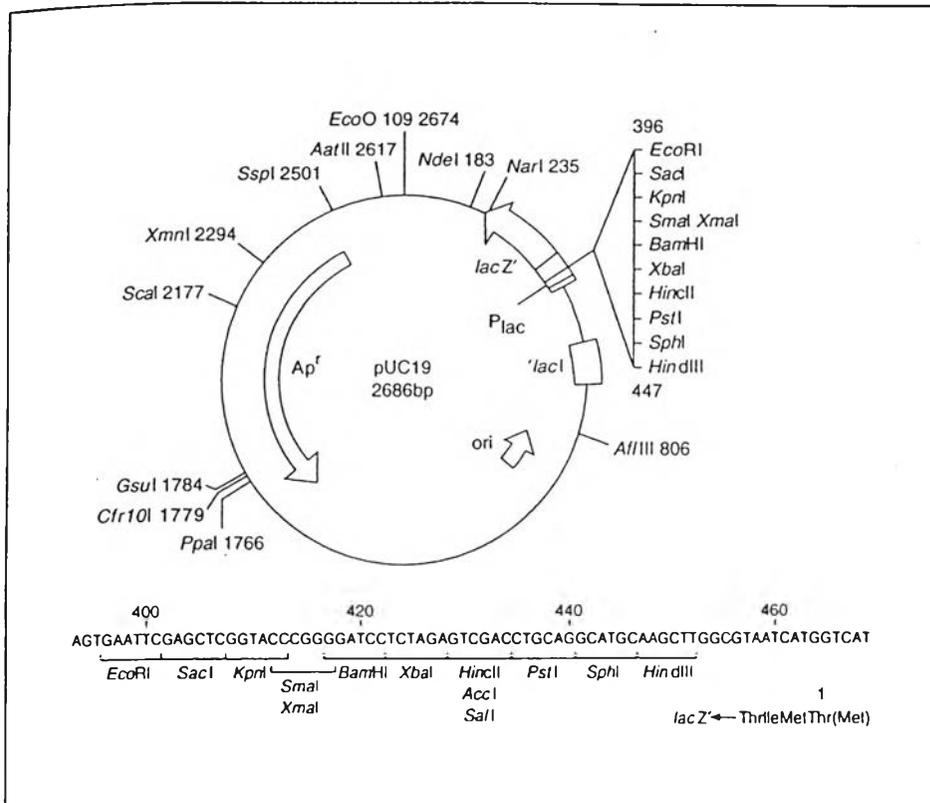
- Pignatello, J. J., Martinson, M. M., Steiert, J. G., Carlson, R. E. and Crawford, R. I. 1983. Biodegradation and photolysis of pentachlorophenol in artificial freshwater streams. **Appl. Environ. Microbiol.** 46: 1024-1031.
- Radding, C. M., Flory, J. Wu. A., Kahn, R., DasGupta, C., Gonda, G., Bianchi, M. and Tsang, S. S. 1982. Three phases in homologous pairing: polymerization of recA protein on single-stranded DNA, synapsis, and polar strand exchange. **Cold Spring Harbor Symp Quant Biol.** 47: 821-828.
- Rossmann, M. G., Moras, D. and Olsen, K. W. 1974. Chemical and biological evolution of nucleotide-binding protein. **Nature.** 250: 194-199.
- Russell, C.B., Thaler, D. S. and Dahlquist, F. W. 1989. Chromosomal transformation of *Escherichia coli recD* strains with linearized plasmid. **J. Bacteriol.** 171: 2609-2613.
- Saber, D. L. and Crawford, R. L. 1985. Isolation and characterization of *Flavobacterium* strains that degrade pentachlorophenol. **Appl. Environ. Microbiol.** 50: 1512-1518.
- Saunders, J. R. and Saunders, V. A. 1988. Bacterial transformation with plasmid DNA. In Grinsted, J. and Bennett, P. M., (ed). **Plasmid technology** (p. 87-101). Academic Press, London.
- Schell, M. A. and Sukordhman, M. 1989. Evidence that the transcription activator encoded by the *Pseudomonas plitida nahR* gene is evolutionarily related to the transcription activators encoded by the *Rhizobium nodD* genes. **J. Bacteriol.** 171: 1952-1959.
- Shannon, K. P. and Phillips, I. 1982. Mechanism of resistance to aminoglycoside antibiotics moderately impair granulocyte function. **Antimicrob. Ag. Chemother.** 138: 552.
- Shannon, K. P., Phillips, I. and King, B. A. 1978. Aminoglycoside-resistant among *Enterobacteriaceae* and *Acinetobacter* species. **Antimicrob. Ag. Chemother.** 4: 131-140.

- Shaw, K. J., Hare, R. S. and Sabatelli, F. J. 1991. Correlation between aminoglycoside resistance profiles and DNA hybridization of clinical isolates. **Antimicrob. Ag. Chemother.** 35: 2253-2260.
- Shaw, K. J., Rather, P. N., Hare, R. S. and Miller, G. H. 1993. Molecular genetics of aminoglycoside genes and familial relationships of the aminoglycoside modifying enzymes. **Microbiol. Rev.** 57: 138-142.
- Shen, P. and Huang, H. V. 1986. Homologous recombination in *Escherichia coli*: dependence on substrate length and homology. **Genetics.** 112: 441-457.
- Shevell, D. E., Abou-Zamzam, A. M., Demple, B. and Walker, G. C. 1988. Construction of an *Escherichia coli* K-12 *ada* deletion by gene replacement in a *recD* strain reveals a second methyltransferase that repairs alkylated DNA. **J. Bacteriol.** 170:3294-3296.
- Simon, J. R. and McEntee, K. 1989. A rapid and efficient procedure for transformation of intact *Saccharomyces cerevisiae* by electroporation. **Biochem Biophys. Res. Commun.** 164: 1157-1164.
- Singer, B. S., Gold, L., Gauss, P. and Doherty, D. H. 1982. Determination of the amount of homology required for recombination in bacteriophage T<sub>4</sub>. **Cell.** 31: 25-33.
- Taber, H. W., Mueller, J. D., Miller, P. F. and Arrow, A. S. 1987. Bacterial uptake of aminoglycoside antibiotics. **Microbiol. Rev.** 51: 439.
- Wierenga, R. K., Terpstra, P. and Hol, W. G. J. 1986. Prediction of the occurrence of the ADP-binding  $\beta\alpha\beta$ -fold in proteins using an amino acid sequence fingerprint. **J. Mol. Biol.** 187: 101-107.
- Winans, S. C., Elledge, S. J., Krueger, J. H. and Walker, G. C. 1985. Site-direct insertion and deletion mutagenesis with cloned fragments *Escherichia coli*. **J. Bacteriol.** 161: 1219-1221.

- Xun, L. and Orser, C. S. 1991a. Purification of a *Flavobacterium* pentachlorophenol-induced periplasmic protein (PcpA) and nucleotide sequence of the corresponding gene *pcpA*. **J. Bacteriol.** 173: 2920-2926.
- Xun, L. and Orser, C. S. 1991b. Purification and properties of pentachlorophenol hydroxylase, a flavoprotein from *Flavobacterium* sp. strain ATCC 39723. **J. Bacteriol.** 173: 4447-4453.
- Xun, L., Topp, E. and Orser, C. S. 1992a. Confirmation of oxidative dehalogenation of pentachlorophenol by a *Flavobacterium* pentachlorophenol hydroxylase and reaction stoichiometries. **J. Bacteriol.** 174: 5745-5747.
- Xun, L., Topp, E. and Orser, C. S. 1992b. Diverse substrate range of a *Flavobacterium* pentachlorophenol hydroxylase and reaction stoichiometries. **J. Bacteriol.** 174: 2898-2902.

## APPENDIX A

### Restriction map of pUC 19



pUC 19 DNA: Location of some restriction endonuclease sites in multicloning site

Enzyme	Location
<i>EcoR</i> I	396
<i>Sma</i> I	412
<i>BamH</i> I	417
<i>Pst</i> I	435
<i>Hind</i> III	447

## APPENDIX B

Nucleotide sequence of *pcpD* gene from *Sphingomonas chlorophenolica* ATCC 39723

NUCLEOTIDE SEQUENCE 28-999 = *pcpD* gene

PRODUCT: pentachlorophenol 4-monooxygenase reductase

ORGANISM: *Sphingomonas chlorophenolica* strain ATCC 39723

REFERENCE: Lange, C. C. and Orser, C. S.

JOURNAL: Thesis (1994) Microbiology, Molecular Biology and Biochemistry,  
University of Idaho, Moscow.

```

tgccgtgcg ctaggagac ccgtgcatg acaaaccgg ttcgacaat cgacatgacg 61
gtcacgcaga tcaccgcgt ggccaaggac atcggctctt acgaacttcg cccggaacc 121
ggcgtgatat tgccggagt caccgcggg gcgcatatcg gcgtttcget tccaacggg 181
atccagcgca gctattcgct cgtcaaccgg cagggcgaga gggaccgta cgtgatcacg 241
gtcaacctcg accgcaacag ccggggcggg tcgctacc tccacgagca gttgctgggc 301
gggcagcgcc tgtccatcgt accgcccgc aataatttcg ccctgggga gacagcccc 361
cactccgtcc tgttcgggg cggcatcggc atcacgccga tctggtcgat gatccaacgg 421
ttgctgggaa tcggttccac ctgggagctt caggacgct gtcgctggca ggatttcgtc 481
gcctaccgcc aggaactgga gcaggcggcg gcggaggctg gagcgagatt ccacctccac 541
ctcgatgaag aggcgacgg caaattcctg gacctggcg gccccgtggc gcaggcggc 601
caggacagca tcttctattg ctgctgccc gagcgatgc tccaggccta taaggcggcg 661
acggcacct gccgtccgaa cgggtgctgg tccaacattc tgctgctgct tgacggcgaa 721
ccggcggacg acgtgttcac ggtcgtcgtg gcgctggcgg ccggccagga attcacggtc 781
gaaccgggga tgacgatcct ggagacgctg ctccagaacg gcataagccg gaattactcc 841
tgcaccagg gcgtctgctg cacctgctg accaaggtgc tggaggcgca acccgatcat 901
cgcgactggg tcttgtccga cgagaagaag gcgtcaaatt cgacctggt gatctgctgt 961
tcgctgagca aatccccgcg gctggtgctg gacatctgaa tcgaccgctc gaaggacgac 1021

```

## APPENDIX C

Transposon Tn 903 fragment encodes for kanamycin resistance

DNA sequence 1-955: coordinates from transposon Tn 903 sequence are 1083-2038

NUCLEOTIDE SEQUENCE: 80-895 code for product neomycin phosphotransferase

SOURCE: *E. coli*.

1	ttacattgca	caagataaaa	atatacatc	atgaacaata	aaactgtctg	cttacataaa	cagtaataca	aggggtgta
81	tgagccalat	tcaacggga	acgtcttgct	cgaggccgg	atlaaatcc	aacatggatg	ctgattata	tgggtataaa
161	tgggctcgcg	ataatgtcgg	gcaatcaggt	gcgacaatct	actgattgta	tgggaagcc	gatgcgccg	agttgttct
241	gaaacatgc	aaagtagg	tgccaatga	tattacagat	gagatggca	gactaaactg	gctgacgga	ttatgcctc
321	ttccgaccat	caagcatttt	atccgtactc	ctgatgatga	atggttactc	accactgca	tccccggga	aacagcattc
401	caggtattag	aagaalatcc	tgattcaggt	gaaaatattg	ttgatgcgct	ggcagtgttc	ctgcgccggt	tgcattcgat
481	tcctgtttgt	aaltgtcctt	ttaacagca	tcgcglattt	cgctcgcctc	aggcgcaac	acgaatgaat	aacggtttgg
561	ttgatgcgag	tgattttgat	gacgagca	atggcaggc	agtgaacaa	gtctcgaaag	aaatgcataa	acttttgcca
641	ttctaccggg	attcagtcgt	cactcatggt	gatttctcac	ttgataacct	tattttgac	gaggggaat	taataggttg
721	tattgatggt	ggacgagtg	gaatcgcaa	ccgataccg	galcttgcca	tcctatggaa	ctgcctcggg	gagtttctc
801	ttcattaca	gaaacggcct	tttcaaaaat	atggtattga	taatcctgat	atgaataaat	tgcagttca	ttgatgctc
881	gatgagtttt	tctaatcaga	attggttaat	tggttglaac	actggcagg	cattacgctg	acttgacggg	acggc

## APPENDIX D

Transposon Tn 5 fragment encodes for neomycin and kanamycin resistance

NUCLEOTIDE SEQUENCE: 151-945 code for neomycin phosphotransferase

NUCLEOTIDE SEQUENCE: 13-62; function: aminoglycoside phosphotransferase

promotor.

SOURCE: *E. coli*

```

1 acagcaagcg aaccggaatt gccagctggg gcgcccctctg gtaaggttgg gaagccctgc
61 aaagtaaact ggatggcttt cttgccgcca aggatctgat ggcgcagggg atcaagatct
121 gatcaagaga caggatgagg atcgtttcgc atgattgaac aagatggatt gcacgcaggt
181 tctccggccg ctggggtgga gaggctattc ggctatgact gggcacaaca gacaatcggc
241 tgctctgatg ccgccgtggt ccggctgtca gcgcaggggc gcccggttct ttttgtcaag
301 accgacctgt ccggtgccct gaatgaactg caggacgagg cagcgcggct atcgtggctg
361 gccacgacgg gcgttccttg cgcagctgtg ctcgacgttg tactgaagc ggaaggac
421 tggctgctat tgggcgaagt gccggggcag gatctcctgt catctcacct tgctcctgcc
481 gagaaagtat ccatcatggc tgatgcaatg cggcggctgc atacgctga tccggctacc
541 tgcccattcg accaccaagc gaaacatcgc atcgagcgag cactgactcg gatggaagcc
601 ggtcttgtcg atcaggatga tctggacgaa gagcatcagg ggctcgcgcc agccgaactg
661 ttcgccaggc tcaaggcgcg catgcccgac ggcgaggatc tcgtcgtgac ccatggcgat
721 gcctgcttgc cgaatatcat ggtggaaaat ggccgctttt ctggattcat cgactgtggc
781 cggctgggtg tggcggaccg ctatcaggac atagcgttgg ctaccctgga tattgtgaa
841 gagcttggcg gcgaatgggc tgaccgcttc ctgctgcttt acggtatcgc cgctcccgat
901 tcgcagcgca tcgccttcta tcgccttctt gacgagttct tctgagcggg actctggggt
961 tcgaaatgac cgaccaagcg acgccaacc tgccatcac agatttcgat tccaccgccg
1021 cctctatga aaggttgggc ttcggaatcg tttccggga cgcggctgg atgatcctcc
1081 agcgcgggga tctcatgctg gagttcttcg cccaccccg gctcgatccc ctgcgagtt
1141 ggttcagctg ctgcctgagg ctggacgacc tcgaggagtt ctaccggcag tgcaaatccg
1201 tcggcatcca ggaaaccagc agcggctatc cgcgcatcca tgccccgaa ctgcaggagt
1261 ggggaggcac gatggccgct ttggtcgacc cggacgggac

```

//

## APPENDIX E

### Media composition for bacteria culture

#### Luria-Bertani (LB), per liter for *E. coli*

10 g Bacto-tryptone

5 g Bacto-yeast extract

10 g NaCl

#### Mineral salt medium, per liter for *S. chlorophenolica*

0.82 g  $K_2 HPO_4 \cdot 3H_2O$

0.19 g  $KH_2 PO_4$

0.5 g  $NaNO_3$

0.1 g  $MgSO_4 \cdot 7H_2O$

20  $\mu M$   $FeSO_4$

4 g L-glutamic acid sodium salt

#### SOC medium

0.5% (w/v) yeast extract

2% (w/v) tryptone

10 mM NaCl

2.5 mM KCl

10 mM  $MgCl_2$

20 mM  $MgSO_4$

20 mM glucose

## APPENDIX F

### **Solution and buffer for Southern blot hybridization**

#### **Hybridization**

10X SSC: 0.15 M NaCitrat, 1.5 M NaCl, pH 7.0

SDS, 10%

Standard hybridization buffer

5X SSC

N-lauroylsarcosine, 0.1% (w/w)

SDS, 0.02%

Blocking reagent, 1% (1/10 volumn of blocking solution, 10X conc.)

#### **Immunological detection**

Maleic acid buffer: 0.1 M maleic acid, 0.15 M NaCl; adjusted to pH 7.5

Blocking solution: diluting the 10X blocking solution 1:10 in maleic acid buffer

Washing buffer: Malic acid buffer plus 0.3% Tween 20 (v/V)

Detection buffer: 0.1 M Tris-HCl, 0.1 M NaCl, pH 9.5

## BIOGRAPHY

Miss Panarat Arunrattiyakorn was born on November 20, 1973 in Bangkok. She graduated with the degree of Bachelor of Science from the Department of Biotechnology at King Mongkut Institute of Technology, Ladkang (KMUTL) in 1995. She had studied for the degree of Master of Science at the Department of Biochemistry, Chulalongkorn University since 1996.

