

# **CHAPTER III**

# **MATERIALS AND METHODS**

## 1. Source of plant material

The plant materials were collected from various localities in Thailand and different periods of time as follows:

Number	Name	Place
ı	Aegle marmelos Corr.	Faculty of Pharmaceutical Science
		Chulalongkorn University,
		Bangkok, January 1996
2	Atalantia monophylla Correa	Rayong, June 1996
3	Citrus aurantifolia Swing.	Khamsakaesang
		Nakornratchasima, March 1996
4	Citrus hystrix DC.	Khamsakaesang
		Nakornratchasima, March 1996
5	Citrus maxima Merr.	Faculty of Pharmaceutical Science
		Chulalongkorn University,
		Bangkok, February 1996.
6	Citrus medica Linn	Rangsit, Prathumthani, March
		1996.
7	Citrus reticulata Blanco	Rangsit, Prathumthani, March
		1996.
8	Clausena anisata Hook.	Kanchanaburi, January 1996
9	Clausena excavata Burm.	Faculty of Pharmaceutical Science
		Chulalongkorn University,
	(1)	Bangkok, January 1996.

Number	Name	Place
10	Feronia limonia Swing.	Faculty of Pharmaceutical Science
		Chulalongkorn University,
		Bangkok, January 1996.
11	Glycosmis pentaphylla Corr	Saraburi , February 1996.
12	Hesperethusa crenulata Roem.	Rayong , June 1996
13	Micromelum minutum Wight&	Sakae-Raj Environmental Research
	Arn.	Station, Pak Thong Chai Nakorn
		Ratchasima, in April 1996
14	Murraya paniculata Jack.	Sakae-Raj Environmental Research
		Station, Pak Thong Chai Nakorn
		Ratchasima, in April 1996
15	Paramignya scandens Craib.	Sakae-Raj Environmental Research
		Station, Pak Thong Chai Nakorn
		Ratchasima, in April 1996
16	Toddalia asiatica Lamk.	Sakae-Raj Environmental Research
		Station, Pak Thong Chai Nakorn
		Ratchasima, in April 1996
17	Triphasia trifolia P. Wils.	Faculty of Pharmaceutical Science
		Chulalongkorn University,
		Bangkok, January 1996.
18	Zanthoxylum limonella Alston.	Sakae-Raj Environmental Research
		station, Pak Thong Chai, Nakorn
		Ratchasima , in April 1996

Authentication was achieved through comparison with herbarium specimen in the Botany Section, Technical Division, Department of Agriculture, Ministry of Agriculture and Coorperative, Thailand.

### 2. Volatile oil content and composition.

### 2.1 Volatile oil content determination.

Volatile oil was determined by the method described in the association of official analytical chemists (method 962.17,AoAc,1990). One hundred and fifty grams of each sample were put into a 500 ml round bottom flask. The tridistilled water were added into the flask to about half full and a few pieces of boiling chips. The flask was conneted to the apparatus for the determination of volatile oil (Fig. 1). The content of the flask was distilled until two consecutive reading taken at one hour interval showed no change in oil content (about four hours). After cooling, the oil volume was measured, calculated and expressed as millilitre of the oil per one hundred grams of sample. The volatile oil obtained was then collected and stored at 4 °C until being analysed for its chemical composition by GC-MS.

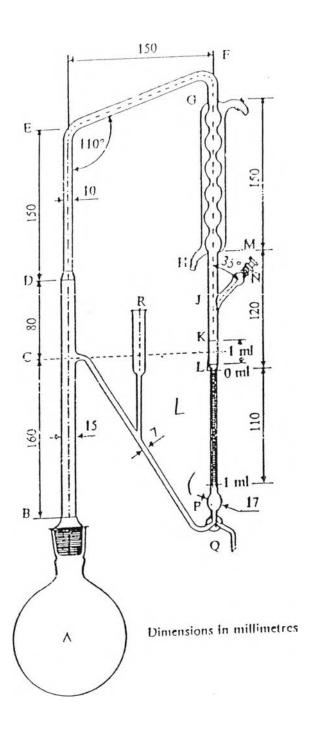


Figure 1 Apparatus for volatile oil content determination.

### 2.2 Gas chromatography - mass spectrometry

For identification of the composition of essential oil, a gas chromatographymass spectrometry (GC-MS) was used. The essential oil was diluted to 1:100 in methanol before being injected into GC-MS system. The condition of GC-MS was described below. The spectra were recorded and compared with the terpene library.

#### **GC-MS Condition**

#### Instrument model

Column fused silica capillary column

(30 mm. x 0.25 mm. i.d.) coated with

DB-5 (J&W) film thickness 0.25 µm

Column programming 60-180 °C rate 3 °C/min

Injector temperature 220 °C

Helium carrier gas 1 ml/min

Split ratio 100 : 1

Accelerating voltage 1700 volts

Sample size 1 μl

Solvent methanol were HPLC grade

#### Identification of the component

Identification of the components was based on GC retention times computer matching with of terpene library, comparison of the fragment pattern with those reported in the literature.