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APPENDICES

APPENDIX A

Instruments, materials, chemical reagents and glassware

1. Instruments and materials

- Analytical balance: Mettler Toledo model AG204, Switzerland.
- Autoclave: Tomy model SS-325, Japan.
- Centrifuges: Beckman model Avanti J25, U.S.A; Eppendorf model 5430, Germany; Sorvall model RC-5C Plus and Sorvall tabletop centrifuge model RC-5C Plus, USA.
- Circulating Water Bath: Techre model TE8 A, UK.
- Freeze Dryer: Savant model Super Modulya 233, USA.
- Hot plate and stirrer: Thermolyne model Crimarec2, USA.
- Incubator: Memmert model BE500(30°C, 37°C, 45°C, 50°C, and 55°C), Germany.
- Incubator shaker: New Brunswick Scientific model innova4300, U.S.A
- Magnetic stirrer: Ika model RO-10, Malaysia.
- Microwave: Sanyo model EM-815FW, Japan.
- Oven: Memmert UE 600, Germany.
- pH Meter: Mettler Toledo model CH-8603, Switzerland.
- Pipetteman: Gilson, Villiers-Le-Bel, France.
- Precision balance: Mettler Toledo model PB3002, Switzerland.
- Freezer : Sharp model FC27 (-20°C), Japan and Deep Freezer REVCO model ULT1790-7-V12 (-80°C), USA.
- Shaking Water Bath: Memmert, model WB22, Germany.
- Spectrophotometer: Sherwood Scientific model259, Cambridge, UK.
- Vortex mixer: Barnstead/Thermolyne model M37610-26, Iowa, USA.

2. Chemicals

Chemical	Company	Grade
Acetone	Merck	Analytical
Bovine serum albumin	Sigma	Analytical
Chloroform	Mallinckrodt	Analytical
Ethanol	Carlo Erba	Analytical
Ethylene diamine tetraacetic acid (EDTA)	Merck	Analytical
Ferric sulfate sevenhydrate	Carlo Erba	Analytical
Folin-Ciocalteu's phenol	Merck	Analytical
Hydrochloric acid	Merck	Analytical
Magnesium sulfate heptahydrate	Sigma	Analytical
Methanol	Merck	Analytical
Phenol	Carlo Erba	Analytical
Potassium hydrogen sulfate	Merck	Analytical
Di-potassium tartate	Carlo Erba	Analytical
Sodium chloride	Carlo Erba	Analytical
Tri-sodium citrate dihydrate	Merck	Analytical
Sodium dodecyl sulfate	Fluka	Analytical
Sodium hydroxide	Merck	Analytical
Sodium potassium tartate	Merck	Analytical
Trichloroacetic acid	Merck	Analytical
Trisma base	Merck	Analytical
Tyrosine	Sigma	Analytical
Xylose	Merck	Analytical

3. Glassware

- Culture tube 16x150 mm : Pyrex, U.S.A.
- Culture tube 25x250 mm : Pyrex, U.S.A.
- Petri-dish 90 mm: Millionant, SA.54, France.

APPENDIX B

Culture Media

1. Enrichment Media

1) Glucose-Ethanol medium

Glucose	1.5%
Ethanol	0.5%
Peptone	0.3%
Yeast Extract	0.3%
Acetic acid	0.35%

Adjust pH 3.5 with HCl

2) Sorbitol medium

Sorbitol	2.0%
Peptone	0.3%
Yeast extract	0.3%

Adjust pH 3.5 with HCl

3) Sucrose-Acetic acid medium

Sucrose	2.0%
Peptone	0.3%
Yeast extract	0.3%
Acetic acid	0.35%

Adjust pH 3.5 with HCl

4) Methanol-Peptone-Yeast extract (MPY) medium

Methanol	0.8% (added after autoclaving)
Peptone	0.3%
Yeast extract	0.3%

Adjust pH 4.0 with HCl

2. Glucose-Ethanol-Yeast extract-CaCO₃ (GEY- CaCO₃) agar plate

Glucose	2.0%
Ethanol	0.5%
Peptone	0.3%
Yeast Extract	0.3%
CaCO ₃	0.7%
Agar	1.5%

3. Cryoprotectant for preservation

Glucose	2.5%
Glycerol	20%
Peptone	0.5%
Yeast Extract	0.3%

4. Growth in test media (4 kinds)

4.1 Glucose-ethanol with 0.3% acetic acid medium

Glucose	1.5%
Ethanol	0.5%
Peptone	0.3%
Yeast Extract	0.3%
Acetic acid	0.3%

Adjust pH 3.5 with HCl

4.2 Glucose-ethanol without acetic acid medium

Glucose	1.5%
Ethanol	0.5%
Peptone	0.3%
Yeast Extract	0.3%

Adjust pH 3.5 with HCl

4.3 Sorbitol medium

Sorbitol	2.0%
Peptone	0.3%
Yeast extract	0.3%

Adjust pH 3.5 with HCl

4.4 Sucrose with 0.3% acetic acid medium

Sucrose	2.0%
Peptone	0.3%
Yeast extract	0.3%
Acetic acid	0.3%

Adjust pH 3.5 with HCl

5. Oxidation/fermentation medium

Glucose	10%
Peptone	2.0%
NaCl	5%
K ₂ HPO ₄	0.3%
Bromthymol blue	0.03%
Agar	3%

Adjust pH 7.1, autoclave at 110°C, 10 min

6. Acetate/Lactate medium

Peptone	0.2%
Yeast extract	0.2%
Sodium acetate/lactate	0.2%
Bromthymol blue	0.002%

7. Acid production medium

Yeast extract	2.5%
Bromocresol purple	0.2%
Carbon source	1.0%

8. Glucose-Glycerol-Yeast extract Potato (GGYP) medium

Potato	10%
Glucose	0.5%
Glycerol	1.0%
Yeast extract	1.0%
Peptone	1.0%

9. Glucose-Glycerol-Yeast extract (GGY) medium

Glucose	2.0%
Glycerol	1.0%
Yeast extract	0.5%
Agar	1.5%

APPENDIX C

Reagents and Buffers

1. Determination of protein

The protein content was measured by the method of Lowry *et al.* (1951) with bovine serum albumin as standard.

1.1 Reagents

A: 2% sodium carbonate in 0.1N NaOH

B: 0.5% CuSO₄.5H₂O in 1% sodium citrate

C: 1 N Folin-Ciocalteu's phenol reagent

(2N Folin Phenol was diluted with distilled water to the final concentration in 1N, the solution should be freshly prepared before use.)

D: 1 ml Reagent B + 50 ml Reagent A (or similar ratio), Make up immediately before use.

1.2 Procedure

1. Place 0.1 ml of proper dilution of culture broth (for protein determination) or clear supernatant of reaction mixture (for soluble peptide determination)
2. Add 1 ml of Reagent D into the tube and vortex immediately. Incubate at room temperature for 10 min.
3. After the 10 min incubation, add 0.1 ml of Reagent C to sample and vortex immediately. Incubate 30 min at room temperature.
4. Absorbance (OD) of samples was measured at 750 nm. Concentrations of the samples were compared to the standard curve for determination of values. Distilled water was used instead of sample as a blank.

1.3 Preparation of standard curve of protein

Standards of 0, 0.1, 0.2, 0.3, 0.5, 0.7 and 1.0 mg/ml were prepared from bovine serum albumin. The reactions were carried out with the same procedure as described previously. Absorbances were plotted against concentrations of standards.

2. Reducing sugar

Standards of 0, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 and 1.0 $\mu\text{g}/\text{ml}$ were prepared from xylose. The reaction were carried out with the same procedure as described by Somogyi and Nelson method (1952).

3. 6 N HCl

Conc. HCl	60	ml
Distiller water	60	ml

Add conc. HCl into the distilled water.

4. 2 N H_2SO_4

Conc. H_2SO_4	2	ml
Distilled water	34	ml

Add conc. H_2SO_4 into the distilled water.

5. Phenol:Chloroform (1:1 v/v)

Crystalline phenol was liquidified in water bath at 65°C and mixed with chloroform in the ratio of 1:1 (v/v). The solution was stored in a light tight bottle.

6. 0.5M EDTA (pH 8.0)

800 ml of distilled water, 186.1 g of disodium ethylenediaminetetraacetate. $2\text{H}_2\text{O}$ was added and stirred vigorously on a magnetic stirrer. The pH was adjusted to 8.0 with NaOH (20 g of NaOH pellets). The volume was adjusted to 1 litre. The solution was dispensed into aliquots and sterilized by autoclaving for 15 minutes at 15 lb/in^2 .

7. 2xPBS

8 mM Na₂HPO₄

1.5 mM KH₂PO₄

137 mM NaCl

2.7 mM KCl

The 2xPBS was adjusted the pH to 7.0 with 1N NaOH or 1N HCL. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in².

8. 3 M Sodium acetate pH 5.2

To 800 ml of distilled water, 408.1 g of sodium acetate was added and adjusted the pH to 5.2 with glacial acetic acid. The volume was adjusted to 1 litre. The solution was sterilized by autoclaving for 15 minutes at 15 lb/in².

9. 10% Sodium dodecyl sulphate (SDS)

The stock solution of 10% SDS was prepared by dissolved 10 g of sodium dodecyl sulphate in 100 ml sterilized distilled water. Sterilization is not required for the preparation of this stock solution.

10. 1 M Tris-HCl pH 8.0

The 1M Tris was prepared by dissolving 121.1 g of Tris base in 800 ml of distilled water. The pH was adjusted to the desired value by adding conc. HCL (pH 8.0, 42 ml of HCl). The solution was cooled to room temperature before making final adjustment to the desired pH. The volume of the solution was adjusted to 1 litter with with distilled water and sterilized by autoclaving.

11. RNase A solution

RNase A	20	mg
0.15 M NaCl	10	ml

Dissolve 20 mg of RNase A in 10 ml 0.15 M NaCl and heat at 95° C for 5-10 minutes. Keep RNase A solution in -20°C.

12. Proteinase K

Proteinase K (Sigma)	4	mg
50 mM Tris-HCl (pH 7.5)	1	ml

Use freshly prepared solution.

13. Nuclease P₁ solution

Nuclease P1	0.1	mg
40 mM CH ₃ COONa+12 mM ZnSO ₄ (pH 5.3)	1	ml

Store at 4°C.

14. Alkaline phosphatase solution

Alkaline phosphatase	2.4	units
0.1 M Tris-HCl (pH 8.1)	1	ml

15. 0.1 M Tris-HCl buffer, pH 9

Tris	1.21	mg
Distilled water	100	ml

Adjust the pH to 9 with HCl.

16. TE buffer

10 mM Tris HCl (pH 8.0)

1 m M Na₂-EDTA (pH 8.0)**17. TE buffer + RNase A**

TE buffer 960 ml

RNase A (2 mg/ml) 100 µl

18. Saline-Na₂ EDTA

0.1 M NaCl

50 mM EDTA.2Na (pH 8.0)

19. Fehling's solution

Coppersulfate 34.64 g

Sodiumpotassiumtartate 173 g

Sodiumhydroxide 50 g

Solvent was composed of a mixture 500 ml of coppersulfate and 500 ml of mixture sodiumtatare and sodiumhydroxide.

20. Flagella staining

Basic fuchisin 0.5 g

Tannic acid 0.2 g

Aluminium sulfate 0.5 g

Solvent was composed of a mixture of 2.0 of 95% ethanol, 0.5 ml of glycerol, and 7.5 ml of Tris (hydroxymethyl) aminomethane (Tris) buffer.

APPENDIX D

16S rDNA nucleotide sequences

1. The 16S rDNA nucleotide sequence of strain PA3-3

CGCACGAAGGTTTCGGCCTTACTGGCGACGGGTGAGTAACCGTAGGTATCTATCCATGGGTGGGG
 GATAACACTGGGAAACTGGCTATACCCATGACACCTGAGGGTAAAGGCCTAAGGCCTAAGTCGCCTGTGGA
 GGAGCCTGCGTTGATTAGCTAGTTGGTGGGGTAAAGGCCTACCAGGGCATGATCAATAGCTGGTT
 GAGAGGATGATGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGG
 AATATTGGACAATGGGGCAACCTGATCCAGCAATGACGGTGTGAGAAGGCTTCGGGATTGT
 AAAGCACTTCGACGGGGACGATGATGACGGTACCCGTTAGAAGAAGCCCCGGCTAACCTCGTCCA
 GCAGCCCGGTAATACGAAGGGGCTAGCGTTGCTCGGAATGACTGGCGTAAAGGGCGTAGGGC
 GGTTTGACAGTCAGATGTGAAATCCCCGGCTTAACCTGGGAGCTGCATTGATACGTCAACTAG
 AGTGTGAGAGAGGGTTGTGAAATTCCCAGTGTAGAGGTGAAATTCTGAGATATTGGGAAAGAACACC
 GGTGGCGAAGGCGGCAACCTGGCTATTATCTGAGGCGCAAAGCGTGGGGAGCAAACAGGATTAG
 ATACCCGGTAGTCCACGCTGAAACGATGTGGCTAGATGTTGGGTGACTTAGTCATTAGTGTGCGA
 GTTAACCGCTAACGACCCGCCSGGGAGTACCAACCGCAAGGTTGAAACTCAAAGGAATTGACGGGG
 GCGCGCACAAAGCGTGGAGCATGTGGTTAACGCAAGCAACGCGAGAACCTTACCAAGGGCTTGAA
 TGTAGAGGCTGCAAGCAGAGATGTTGGGTTAACACACAGGTGCTGCATGGCTGTGCGT
 CAGCTCGTGTGAGATGTTGGGTTAACGCCCCAACGAGCGCAACCCCTATCTTAGTGTGCCATCA
 GGTTGGGCTGGGCACTCTAAAGAGACTGCCGTGACAAGCCCAGGAAAGGTGGGGATGACGTCAAG
 TCCCCCTGGCCCTTATGTTCTGGCTTCCCACGTCTACAAATGGCGGTGACAGTGGGAAGCTAGGTGG
 TGACCCCCCTGCTGATCTCTAAAGCCGTCTAGTCGGATTGCACTCTGCCAACCAAGTCATTAAAA
 GAATCCCCTAGTAATCCCCGGGCCACATCCCCGGTGAATACCTTTGGGCTTTACACACCCCGT
 CACACCATGGGAGTTGGTTGACCTTAAGCCGGTGAGTAAGGCAAG

2. The 16S rDNA nucleotide sequence of strains MHM10-1

TTTGAGTTGATCCCTGGCTCAGAGCGAACGCTGGCGCATGCTAACACATGCAAGTCGCACGAAG
 GTTTCGGCCTTACTGGCGACGGGTGAGTAACCGTAGGAATCTATCCATGGGTGGGGATAACTCT
 GGGAAACTGGAGCTAACACCGCATGATACCTGAGGGTCAAGGCCTAACGCGAACGCTGTGGAGGAGCC
 TCGCTTGATTAGCTTGTGGGGTAATGGCTACCAAGGCATGATCAATAGCTGGCTGAGAG
 GATGATCACGCCCCACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATAT
 TGGACAATGGGGCAACCTGATCCAGCAATGCCGTGTGAGAAGAAGGTTTCGGATTGTAAGG
 ACTTCGACGGGACGATGATGACGGTACCCGTTAGAAGAAGCCCCGCTAACCTCGTGCCACGAGCC
 GCGTAATCGAACGGGCTAGCGTTGCTCGGAATGACTGGCGTAAAGGGCGTAGGGCTTGTG
 TACAGTCAGATGTGAAATCCCCGGCTAACCTGGGAGCTGATTGATACGTGAGACTAGAGTGT
 GAGAGAGGGTTGTGAAATTCCCAGTGTAGAGGTGAAATTCTGAGATATTGGGAAAGAACACGGTGG
 CGAAGGGCGAACCTGGCTAACACTGACGCTGAGGCGGAAAGCGTGGGGAGCAAACAGGATTAG
 ATACCCGGTAGTCCACGCTGAAACGATGTGTGCTAGATGTTGGTAACCTAGTTATTAGTGTGCG
 AGTTAACCGCTAACGACACCCGCTGGGAGTACGGCGCAAGGTTGAAACTCAAAGGAATTGACG
 GGGGCCCGAACAGCGTGGAGCATGTTGGTTATTCGAAGCAGCGCAGAACCTTACCAAGGGCTGT
 ATGGGTAGGCTGTGTCAGAGATGGGCAATTCCCGCAAGGGACCTGCCGACAGGTGCTGCATGGCT
 GTCGTCAGCTCGTGTGAGATGTTGGGTTAACGCCCCAACGAGCGCAACCCCTATCTTAGTGTG
 CAGCATTTGGGGCACTCTAGAGAGACTGCCGTGACAAGCCGGAGGAAGGTGGGGATGACG
 TCAAGTCCTCATGGCCCTTATGTCCTGGCTACACACGTGCTAACATGGCGGTGACAGTGGGAAGC
 TAAGAGGTGGAATCGCTAGTAATCGGGATCAGCATGCCGGTGAATACGTTCCGGCTTGTAC
 ACACCGCCCGTCACACCATGGGAGTTGGTTGACCTTAAGCCGGTGAGCGAACCCGCAAGGGG

3. The 16S rDNA nucleotide sequence of strain KLM13-1

TTTGAGTTGATCCCTGGCTCAGAGCGAACGCTGGCGCATGCTTAACACATGCAAGTCGCACGAAG
 GTTTCGCCCTAGTGGCGGACGGGTGAGTAACCGTAGGAATCTATCCATGGGTGGGGATAACTCT
 GGGAAACTGGAGCTAATACCGCATGATACCTGAGGGTCAAAGGCAGAAGTCGCCTGTGGAGGAGCC
 TCGTTTGATTAGCTTGTGGGGTAATGGCCTACCAAGGCAGTATCAATAGCTGGTCTGAGAG
 GATGATCAGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATAT
 TGGACAATGGGGCAACCTGATCCAGCAATGCCCGTGTGAAGAAGGTTTCGGATTGTAAGC
 ACTTTGCACGGGAGCATGACGGTACCGTAGAAGAAGCCCCGCTGACGATGATGACGGTAC
 CCGTAGAAGAAGCCCCGCTAACCTCGTGCACAGCCGGTAATACGAAGGGGGCTAGCGTTGCT
 CGGAATGACTGGCGTAAAGGGCGTAGGCGGTTGTACAGTCAGATGTGAAATCCCCGGCTAA
 CCTGGGAGCTGATTGATACGTACAGACTAGAGTGTGAGAGAGGGTTGGAATTCCCAGTGTAGAG
 GTGAAATTCTGAGATAATTGGAAGAACACCGGTGGCGAAGGCAGAACCTGGCTCATTATCTGAGGC
 GCGAAAGCGTGGGGAGCAACAGGATTAGATACCTGGTAGTCCACGCTGTAACAGATGTGGCTAG
 ATGTTGGGTGACTTAGTCATTAGTCAGTGTGCAAGTCAGCTTAAGCACCGCCTSGGAGTACCCG
 CAAGGTTGAAACTCAAAGGAATTGACGGGGGCCACAGCGTGGAGCATGTGGTTAATTGCA
 AGCAACCGCAGAACCTTACCAAGGGCTGAATGTAGAGGCTGCAAGCAGAGATTTGGTGGGGACTCTAGA
 ACCTCAGTCCCACAGGAGCGAACCCCTATCTTAGTGTGCAAGCATGTTGGTGGGGACTCTAGA
 GAGACTGCCGGTGACAAGCCGGAGGAAGGTGGGATGACGTCAAGTCCTCATGCCCTATGCTCTG
 GGCTACACACGTGCTACAATGGCGGTACAGTGGGAAGCTAGATGGGACATCGTGTGATCTCTAA
 AAACCGTCTCAGTCGGATTGACTCTGCAACTCGAGTACATGAAGGGGAATCGCTAGTAATCGCG
 GATCAGCATGCCCGGTGAATACGTTCCGGGCTTGTACACACCGCCGTACACCATGGGAGTTG
 GTTGACCTTAAGCCGGTAGCGAACCCGCAAGGGG

4. The 16S rDNA nucleotide sequence of strain BBM91-1

TGAGTTGATCCCTGGCTCAGAGCGAACGCTGGCGCATGCTTAACACATGCAAGTCGCACGAAGGT
 TTCCGCCCTAGTGGCGGACGGGTGAGTAACCGTAGGAATCTATCCATGGGTGGGGATAACTCTGG
 GAAACTGGAGCTAATACCGCATGATACCTGAGGGTCAAAGGCAGAAGTCGCCTGTGGAGGAGCC
 CGTTTGATTAGCTTGTGGGGTAATGGCCTACCAAGGCAGTATCAATAGCTGGTCTGAGAGGA
 TGATCAGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTG
 GACAATGGGGCAACCTGATCCAGCAATGCCCGTGTGTAAGAAGGTTTCGGATTGTAAGC
 TTTCGACGGGAGCATGATGACGGTACCGTAGAAGAACCCCCGGCTGACGATGATGACGGTACCC
 GTAGAAGAACCCCCGCTAACCTCGTGCACAGCCGGTAATACGAAGGGGGCTAGCGTTGCTC
 GGAATGACTGGCGTAAAGGGCGTAGGCGGTTGTACAGTCAGATGTGAAATCCCCGGCTTAAC
 CTGGGAGCTGCTATTGATACGTACAGACTAGAGTGTGAGAGAGGGTTGGAATTCCCAGTGTAGAGG
 TGAAATTCTGAGATAATTGGGAAGAACACCGGTGGCAAGGCAGAACCTGGCTCATTATCTGAGGCG
 CGAAAGCGTGGGGAGCAAACAGGATTAGATACCTGGTAGTCCACGCTGAAACGATGTGGCTAGA
 TGTTGGGTGACTTAGTCATTAGTCAGTGTGCCAGTTAACCGCTTAAGCACCGCCTSGGAGTACCC
 AAGGTTGAAACTCAAAGGAATTGACGGGGCCCGACAAGCGTGGAGCATGTGGTTAATTGAA
 GCAACCGCGAGAACCTTACCAAGGGCTTGAATGTAGAGGGTGAATTAGTCATTAGTCAGTGTG
 CGCTTAAGCACCCGCTSGGGAGTACCAACCGCAAGGTTGAAACTCAAAGGAATTGACGGGG
 GCACAAAGCGGTGGAGCATGTGGTTAATTGAAACCAACGCGCAGAACCTTACCAAGGGCTG
 GAGGCTGCAAGCAGAGATTTGTTCCGGGACCTCAGTCCCGCAACGAGCGAACCCCTATCTT
 TACACACCGCCCGTCACACCATGGGAGTTGGTTGACCTTAAGCCGGTAGCGAACCCGCAAGGGGG
 CTTGAATGTAGAGGCTGCAAGCAGAGATTTGTTCCGGGACCTCAGTCCCACGAGCGAAC
 CCCTATCTTGTGCAAGCAGAGATTTGTTGGGTGGGACTCTAGAG

5. The 16S rDNA nucleotide sequence of strain FBM4-3

TGATCCCTGGCTCAGAGCGAACGCTGGCGGCATGCTAACACATGCAAGTCGCACGAAGGTTTCCGC
CTTAGTGGCGGACGGGTGAGTAACGCGTAGGAATCTATCCATGGGTGGGGGATAACTCTGGGAAACT
GGAGCTAATACCGCATGATACCTGAGGGTCAAAGGCGCAAGTCGCCTGTGGAGGAGCCTGCGTTGA
TTAGCTTGTGGGGTAATGGCCTACCAAGGCGATGATCAATAGCTGGTCTGAGAGGATGATCAG
CCACACTGGGACTGAGACACGGCCCAGACTCTACGGGAGGCAGCAGTGGGAATTGGACAATG
GGGGCAACCCTGATCCAGCAATGCCGGTGTGTAGAGAAGGTTTCGATTGAAAGCACTTCGAC
GGGGACGATGATGACGGTACCCGTAGAAGAAGCCCCGGCTGACGATGATGACGGTACCCGTAGAAG
AAGCCCCGGCTAACCTCGTGCCAGCAGCCCGGTAATACGAAGGGGCTAGCGTTGTCGGAATGAC
TGGCGTAAGGGCGTGTAGCGGTTTGACAGTCAGATGTGAAATCCCCGGCTAACCTGGGAGC
TGCATTGATACGTCAGACTAGAGTGTGAGAGAGGGTTGTGAAATTCCAGTGTAGAGGTGAAATTG
GTAGATATTGGGAAAGAACACCGGTGGGTACCCGTAGAAGAAGCCCCGGCTAACCTCGTGCAGCAG
CCGCGGTAATACGAAGGGGCTAGCGTTGTCGGAATGACTGGGCTAACGGGCGTGTAGCGGTT
TGTACAGTCAGATGTGAAATCCCCGGCTAACCTGGGAGCTGCATTGATACGTGCAGACTAGAGT
GTGAGAGAGGGTTGTGAAATTCCAGTGTAGAGGTGAAATTGTAGATATTGGGAAAGAACACCGGT
GGCGAAGGCGGCAACCTGGCTCATAACTGACGCTGAGGCCGAAAGCGTGGGAGCAAACAGGATT
AGATACCCGGTAGTCCACGCTGTAAACGATGTGTCTAGATGTGGTAACITAGTTATTCACTGTC
GCAGTTAACCGCTAACGACACCGCCTGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGA
CGGGGGCCCGACAAGCGGTGGAGCATGTGGTTATTCAAGCAGCGCAGAACCTTACCAAGGGCTT
GTATGGGTTAGAGAGACTGCCGTGACAAGCCGGAGGAAGGTGGGGATGACGTCAGTCCATGG
CCCTTATTCCCGGGACCTCAGTCCCGAACGAGCGAACCCCTATCTTAGTTGTCAGCAGTGG
GTGGGCACCTAGAG

6. The 16S rDNA nucleotide sequence of strain LBM3-1

AGCGAACGCTGGCGGCATGCTTAACACATGCAAGTCGACAGAACCTTCGGGGTTAGTGGCCGGACGG
GTGAGTAACCGTAGGAATCTGCCACGGGTGGGGATAACTCTGGAAACTGGAGCTAATACCGA
TGATACCTGAGGGCAAAGGCTGGAGCATGTGGTTAATTCCGCAAGTCGCCTGTTGAGGAGCCTCG
TTCGATTAGCTAGTTGGTGGGTAAGGCCTACCAAGGCATGATCGATAGCTGGTTGAGAGGATGA
TCAGCCACACTGGGACTGGACACGGCCCAGACTCCTACGGGCAACCTGATCAGCAATGCCGCGTG
TGTGAAGAAGGTCTCGGATTGTAAGCACTTTCGACGGGAGCATGATGACGTACCCGTAGAAGAA
GCCCGGCTAACCTCGTGCAGCAGCCCGGTAATACGAAGGGGGCTAGCGTGCCTCGGAATGACTGG
GCGTAAAGGGCGTGTAGGCCTTACACAGTCAGATGTAAATCCCCGGCTAACCTGGAGCTGCA
TTTGATACGTGTAGACTAGAGTGTGAGAGAGGGTTGGAATTCCCAAGTGTAAAGGTGAAATTCTGTAG
ATATTGGGAAAGAACACCGTGGCGAAGGCAGCAACCTGGCTCATGACTGACCTGAGGCGCGAAAGC
GTGGGGAGCAAACAGGATTAGATACCTGGTAGTCCACGCTGTAACAGATGGTCTAGATGTTGGGT
AACTTTGTTATTCACTGTGCGAGTTAACCGCTTAAGCACACCGCCTGGGGAGACGGCCGCAAGGTG
AAACTCAAAGGAATTGACGGGGGCCCGACAACCGGTGGAGCATGTGGTTAATTGAAAGCAACCGCG
CAGAACCTTACAGGGCTGAATGTAGAGGCTGTATTCAAGAGATGGATAATTCCCGCAAGGACCTCT
AACACAGGTGCTGCATGGCTGCTGTCAGCTGTCGTGAGATGTTGGGTAGTCCCGCAACGAGCG
CAACCCCTATCTTAGTTGCCAGCATGTTGGGTGGGACTCTAGAGAGACGCCGTGACAAGCCGG
AGGAAGGTGGGATGACGTCAGTCCTCATGGCCCTATGTCCTGGGCTACACGTGCTACAATGGCG
GTGACAGTGGGAAGCTAGATGGTGCACATCATGCTGATCTCTAAAAGCCGTCAGTCGGATTGCACTC
TGCAACTCGAGTGCATGAAGGTGGAATCGCTAGTAATCGCGGATCAGCATGGCGGTGAATACGTTCC
CGGGCCTTGTACACACCGCCGTCACACCATGGGAGTTGGTTGACCTTAACCGGTGAGCGAACCCG
CAAGGGCGCAGCCGACCACGGTCGGGTAGCGACTGGGGTGAAGTCGTAAGGTAGCCGTAGGG
GAACC

7. The 16S rDNA nucleotide sequence of strain JR70-1

GAGTTTGATTCTGGCTCAGATTGAGGGGACGATGATGACGGTGCACGAAGGTTGCCCTAGTGGCGG
 ACGGGTGAGTAACCGTAGGGATCTATCCACGGGTGGGGACAACCTCGGAACTGGAGCTAATAC
 CGCATGATACTGAGGGTCAAAGCGCAAGTCGCCTGTGGAGGAACCTCGCTGATTAGCTAGTTGG
 TGGGGTAAAGGCCTACCAAGGCATGATCGATAGCTGGTTGAGAGGGATGATAGCCACACTGGGAC
 TGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGACAAGGGCGAACGCTGA
 TCCAGCAATGCCCGTGTGAAGAAGGTCTTCGGATTGTAAGCAGTGTGAGGGGACGATGATGA
 CGGTACCGTAGAAGAACGCCCCGGCTAACCTCGGCCAGCAGCCGGTAATCGAAGGGGCTAGC
 GTTGTCTCGGAATGACTGGCGTAAGGGCGCTAGGCGGTGTTACAGTCAGTGTGAAATCCCCGGG
 CTTAACCTGGGAACTGCATTGATACGTGACGACTAGAGTCAGAGAGGGTGTGGAATTCCAGTG
 TAGAGGTGAAATTCTGAGATATTGGGAGAACACCGGTGGCAAGGGCACCTGGCTGATACTG
 ACGCTGAGGCGCGAACGCTGGGAGCAAACAGGATTAGATACCCTGGTAGCCACGCTGTAACGA
 TGTGTCTGGATGTTGGGAAACTTAGTTCTCAGTCAGCTAACCGCTAGCACACCCTGGG
 GAGTACGGCCGCAAGGTTGAAACTCAAAGGATTGACGGGGCCCGACAACGGTGGAGCATGTGG
 TTTAATTGAAAGCAACCGCAGAACCTTACCAAGGGCTTGCATGGGGAGGACGGTCAGAGATGGACC
 TTTCTCGGACCTCCCGCACAGGTGCTGCATGGCTGTCAGCTCGTGTGGAGATGTTGGGTAAG
 TCCCGCAACGAGCGAACCCCTGTCTTAGTTGCCAGCACTTCAGGTGGGACTCTAGAGAGACTGC
 CGGTGACAAGCCGGAGGAAGGTGGGATGACGTCAAGTCCTCATGGCCCTATGTCCTGGGCTACACA
 CGTGTACAATGGCGGTGACAGTGGGAGCTACATGGTGACATGGTCTGCTCTAAAGCCGCTCA
 GTTCGGATTGACTCTGCAACTCGAGTACATGAAGGTGGAATCGCTAGTATCGCGGATCAGCATGCC
 GCGGTGAATACGTTCCCGGGCTTGTACACACCGCCGTACACCATGGGTTGTTGACCTTAAG
 CCGGTGAGCGAACCGCAAGGACGCAGCCGACCAACGGACGGGTGAGCGACGGGTGAAGTCGTAACA
 AGGTAGCCGTAGGGGAAACCTCGGGCTGGATCACCTCCTT

8. The 16S rDNA nucleotide sequence of strain AK32-2

TGGCTCAGAGCGAACGCTGGCGGATGCTTAACACATGCAAGTCGACGGATCTTCGGGATCAGTG
 CGGACGGGTCTATACCGAAAGCGTGGGAGCAACCGCATGATACCTGAGGGTCAAAGGGCGAAGT
 CGCCTGTGGAGGAACCTCGCTTCGATTAGCTATGGTGGGTAAGGCGTACCAAGGCATGATCGAT
 AGCTGGTTGAGAGGATGATGCCACACTGGACTGAGACACGGCCAGACTCTACGGGAGGCAG
 CAGTGGGAATATTGACAATGGCGAAAGCGATCCAGCAATGCCCGTGTGTAAGAAGGTCTTC
 GGATTGTAAGACATTGACGGGACGATGAGACGGTACCCGTAGAAGAAGCCCGCTAACCTTC
 GTGCCAGCAGCCCGGTAATACGAAGGGGCTCGTTCGGAATGACTGGCGTAAGGGCG
 TAGGCCTTATGCACTGAGATGTGAAATCCCCGTTAACCTGGGAACTGCAATTGAGACGCATAG
 ACTAGAGGTGAGAGAGGGTTGTGGAATTCCCATGTAAGAGGTGAAATTGTAAGATATTGGGAAAGAA
 CACCGTGGCAAGGGCGAACCTGGCTGATATGACGCTGAGGCCGAAAGCGTGGGAGCAAAC
 AGGATTAGATACCTGGTACTCCACGCTGTAAGATGTTGCTGGATTTGGTAACCTAGTTACTC
 AGTGTGAACTAACCGCCTAACGACACCCGCTGGAGTACGGCCGAAAGGTGAAACTCAAAGGA
 ATTGACGGGGGCGCACAAGCGGTGGAGCATGGTTAACCGAAGCAACCGCAGAACCTTACCA
 GGACTTGCATGGGAGGACGTACTCAGAGATGGATTCTCGGACCTCCCGCACAGGTGCTGCATGG
 CTGTCGTCAGCTCGTGTGAGATGTTGGTAATCCCGAACGAGCGAACCCCTGTCTTGTG
 CCAGCACTTCAGGTGGCACTCTAGAGAGACTGGGTGACAAGCGGAGGAAGGTGGGATGACGT
 CAAGTCCTCATGGCCCTATGTCCTGGCTACACAGTGTACAATGGCGGTGACAGTGGGAGCTAC
 ATGGCGACATGGTGTGATCTCTAAAGCCGCTCTCGGATTGTACTCTGCAACTCGAGTACATGAA
 GGTGGAATCGCTAGTAATCGCGGATCAGCATGCCCGGTGAAATACGTTCCGGGCTTGTACACACCG
 CCCGTACACCATGGGAGTTGGTGCACCTTAAGCGGTGAGCGAACCGCAAGGACGCAGCCGACC

9. The 16S rDNA nucleotide sequence of strain LD51-1

GAGTTTGATTCTGGCTAACACATGCAAGTCGCACGGATCTTCGGGATTAGTGGCGGACGGGTGAGT
 AACCGTAGGGATCTATCCATGGGTGGGGACAACACTCCGGAAACTGGAGCTAATACCGCATGATA
 CCTGAGGGTCAAAGGCGCAAGTCGCCTGTGGAGGAACCTCGCCTGATTAGCTAGTTGGTGGGTAA
 AGGCCTACCAAGGCGATGATCGATAGCTGGTTGAGAGGATGATCAGCCACACTGGGACTGAGACA
 CGGCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGACAATGGCGAAAGCCTGATCCAGC
 AATGCCCGTGTGAAGAAGGCTTCGGATTGAAAGCACCTTCGACGGGACGATGATGACGGTA
 CCCGTAGAAGAACGCCCCGCTAACCTCGTGCACGCCGGTAATACGAAGGGGCTAGCGTTG
 CTCGGAATGACTGGCGTAAAGGGCGCTAGGCCTGATGAGCTCAGATGTGAAATCCCAGGCTT
 AACCTGGGAACGTGATTGAGACGATTGACTAGAGTTGAGAGAGGGTTGTGGAATTCCCAGTGT
 GAGGTGAAATTCTGAGATATTGGAAAGAACACCGGTGGCGAAGCGGCAACCTGGCTCGATGACTGA
 CGCTGAGGCGCGAACAGCTGGGAGCAAACAGGATTAGATACCCCTGTTAGTCCACGCTGAAACGA
 TGTGTGCTGGATGTTGGTAACCTAGTTACTCAGTGTGAAAGCTAACCGCTAACGACACCCGCTGG
 GGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGACGGGGGCCGACAAGCGGTGGAGCATGT
 GGTTTAATTGAAAGCAACGCGCAGAACCTTACCAAGGGCTTGATGGCTGTCAGCTGTGTCGTGAGATGTTGG
 TTAAGTCCCGCAACGAGCACAAACCTTGTCTTATGTTGCCAGCACTTCAGGTGGGACTCTAGAGA
 GACTGCGGTGACAAGCCGGAGGAAGGTGGGATGACGTCAAGTCCTATGGCCTTATGTCCTGG
 CTACACACGTGCTACAATGGCGGTGACAGTGGGAGCTATGTTGACACAGTGTGATCTCTAAAA
 GCCGTCAGTTGGATTGACTCTGCAACTCGAGTACATGAAGGTGGAATCGCTAGTAATCGCGGA
 TCAGCATGCCCGGTGAATACGTTCCCGGCTTGTACACACCGCCCGTACACCATGGAGTTGG
 TCGACCTTAAGCCGGTGGAGCGAACCGCAAGGACGCAGCCGACCACGAGGGTCAGCGACTGGGTT
 GAAGTCGTAACAAGGTAGCCGTAGGGAACCTGCGGCTGGATCACCTCCTT

9. The 16S rDNA nucleotide sequence of strain MG71-2

TTGAGTTGATCCTGGCTCGATGCTTAACACATGCAAGTCGCACGGATCTTCGGGATTAGTGGCGG
 ACGGGTAGTAACCGTAGGGATCTATCCACGGGTGGGGACAACACTCCGGAAACTGGAGCTAATA
 CCGCATGATAACCTGAGGGTCAAAGGCGCAAGTCGCCTGTGGAGGAACCTCGCCTGATTAGCTAGTT
 GGTGGGGTAAAGGCCTACCAAGGCGATGATCGATAGCTGGTTGAGAGGATGATCAGCCACACTGG
 GACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGACAATGGCGAAAGC
 CTGATCCAGCAATGCCGTGTGAAGAACGCTTCGGATTGAAAGCACCTTCGACGGGGACGAT
 GATGACGGTACCCCTAGAAGAACGCCCCGCTAACCTCGTGCACGCCGGTAATACGAAGGGG
 GCTAGCGTTGCTCGGAATGACTGGGCGTAAAGGGCGCTAGGCCTGATGAGCTCAGATGTGAAAT
 CCCGGGCTTAACCTCGGAACCTGCAATTGAGACGCATTGACTAGAGGTCGAGAGAGGGTTGTGGAAT
 TCCCAGTGTAGAGGTGAAATTCTGAGATATTGGAAAGAACACGGGTGGCGAACGGCAACCTGGC
 TCGATACTGACGCTGAGGCCGAAGCGTGGGAGCAAACAGGATTAGATACCCCTGGTAGTCCACG
 CTGTAACGATGTGCTGGATGTTGGTAACTTAGTTACTCAGTGTGAAAGCTAACCGCTAACGCA
 CACCGCCTGGGAGTACGGCCGAAGGTGAAACTCAAAGGAATTGACGGGGGCCGACAAGCGG
 TGGAGCATCTGGTTAATTGAAAGAACGGCAGAACCTTACCAAGGGCTTGATGGGAGGACGTC
 TCAGAGATGGGTATTTCTCGGACCTCCGCACAGGTGCTGATGGCTGTCAGCTGTGCTG
 GATGTTGGGTTAACGCGCAACGAGCGAACCCCTGTCTTATGTTGCCAGCAACTTCAGGTGGGAC
 TCTAGAGAGACTGCCGGTACAAGCCGGAGGAAGGTGGGATGACGTCAAGTCCTATGGCCTTAT
 GTCTGGGCTACACAGTGTACAAATGGCGTGACAGTGGGAAGCTATGTTGACACAGTGTGAT
 CTCTAAAAGCCGCTCAGTTGGATTGACTCTGCAACTCGAGTACATGAAGGTGGAATCGCTAGTA
 ATCGCGGATCAGCATGCCCGGTGAATACGTTCCCGGCTTGTACACACCGCCCGTACACCATGG
 GAGTTGGTTGACCTTAAGCCGGTGGAGCGAACCGCAAGGACGCAGCCGACCACGGACGGGTCAGCG
 AGCTGGGGTGAAGTCGTAACANG

10. The 16S rDNA nucleotide sequence of strain AN1-1

AGCGAACGCTGGCGGCATGCTAACACATGCAAGTCGCACGGATTTGGGATTAGTGGCGGACGG
 GTGAGTAACCGCTAGGGATCTATCCATGGGTGGGGACAACACTCCGGAAACTGGAGCTAATACCGC
 ATGATAACCTGAGGGTCAAAGCGTAAGTCGCCTGTGGAGGAACCTCGCTTCGATTAGCTAGTTGGTG
 GGGTAAAGGCCTACCAAGGCATGATCGATAGCTGGTTGAGAGGATGATCAGGCCACACTGGGACT
 GAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGGACAATGGCGAAAGCCTGA
 TCCAGCAATGCCCGTGTGTAAGAAGGTCTTCGGATTGTAAGACACTTCGACGGGACGATGATGATG
 ACGGTACCCGTAGAAGAAGCCCCGGCTAACCTCGTGCACAGCCGGTAATACGAAGGGGGCTA
 GCGTTGCTCGGAATGACTGGCGTAAAGGGCGCTAGGCGTTGATGCAGTCAGATGTGAAATCCCC
 GGGCTTAACCTGGGAACTGCATTGAGACGATTGACTAGAGTTGAGAGAGGGTTGTGGAATTCCC
 AGTGTAGAGGTGAAATTCTAGATATTGGGAGAACACCCGGTGGCGAAGGGCGGAAACCTGGCTCGA
 TACTGACGCTGAGCGCGAACAGCTGGGGAGCAAACAGGATTAGATACCTGGTAGTCCACGCTGT
 AAACGATGTGTGCTGGATGTTGGTAACCTAGTTACTCAGTGTGCAAGCTAACCGCCTAACGCACACC
 GCCTGGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGACGGGGGCCGACAAGCGGTGGA
 GCATGTGGTTAACCGCAACGCGCAGAACCTTACCAAGGGCTTGCATGGGAGGACGTACTCAG
 AGATGGGTATTCTCGGACCTCCCGCACAGGTGCTGCATGGCTGCTCAGCTCGTGTGAGATG
 TTGGGTTAACGCCCCGCAACGAGCGCAACCCCTGTCAGTTGCCCCACTTCAAGGTGGCACTCTA
 GAGAGACTGCCGTGACAAGCCGGAGGAAGGTGGGATGACGTCAAGTCCTCATGGCCCTATGTC
 CTGGGCTACACACGGTCTAACATGGCGGTGACAGTGGGAAGCTATGTTGACACAGTGTGATCTC
 TAAAAGCGTCTAGTCGGATTGTACTCTGCAACTCGAGTACATGAAGGTGGAAATCGCTAGTAATC
 CGGGATCAGCATGCCCGGGTGAATACGTTCCCGGGCTTGTACACACCGCCGTCACACCATTGGGAG
 TTGGTTGACCTTAAGCCGGTGAGCGAACCGCAAGGACGAGCCGAC

11. The 16S rDNA nucleotide sequence of strain MG71-1

AGCGAACGCTACGGACCCCTCGGGGTGAGTGGCGGACGGGTGAGTATCGCTAGGGATCTATCCATG
 GGTGGGGATAACATCGGAAACTGGTCTAACCTCGCATGATAACCTGAGGGTCAAAGGCCTAACAGT
 CGCCTGTGGAGGAGCCTGCGTTGATTAGCTTGGTGGTAAAGGCCTACCAAGGCATGATCG
 ATAGCTGGTCTGAGAGGATGATCAGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAGG
 CAGCAGTGGGAATATTGGACAATGGCGAAAGGCTGATCCAGCAATGCCCGTGTGAGAAGAAG
 TCTCGGATTGAAAGCACTTCGACGGGACGATGATGACGGTACCCGTAGAAGAAGCCCCGGCTA
 ACTTCGTCGACAGCAGCCGGTAATACGAAGGGGCTAGCGTCTCGGAAATGACTGGGCTAAAG
 GGCCTGAGCGGTTACACAGTCAGATGTGAAATTCCAGGGCTAACCTGGGCTGCATTGATA
 CGTGTAGACTAGAGTGTGAGAGAGGGTTGGAATTCCAGGTGAGAGGTGAAATTCTGTAGATATTG
 GGAAGAACACCGGTGGCGAAGGGCGAACCTGGCTCATTACTGACGCTGAGGCCGAAAGCGTGG
 GAGCAAACAGGATTAGATACCCCTGGTAGTCCACGCTGTAACGATGTGCTGGATGTTGGTAAC
 TAGTTACTCAGTGTGAAAGCTAACCGCTAACGCGCTAACACGCCCTGGGAGTACGGCGCAAGGTTGAA
 CTCAAAGGAATTGACGGGGCCCGACAAGCGGTGGAGCATGTTGTTAACCGAAGCAACCGC
 GAACCTTACCGGGCTTGACATGGGAGGCTGATCCAGAGATGGGTATTCCCGCAAGGGACCTCC
 TGCACAGGTGCTGCATGGCTGCTCAGCTCGTGTGAGATGTTGGGTTAACGCCCACCGAGC
 GCAACCTCGCTTAGTTGCCAGCACGTTGGGAGCTCTAGAGGAACCTGGGCTGACAAGGCC
 GGAGGAAGGTGGGATGACGTCAAGTCCTCATGGCCCTATGTCCTGGCTACACACGTGCTACAAT
 GCGGTGACAGTGGGAAGCTAGATGGTACATCGCTAACAGCGTCTAGTTGAACTGGGATT
 GTACTCTGCAACTCGAGTGCATGAAGGTGGAATCGCTAGTAATCGCGGATCAGCATGCCCGGTGAA
 TACGTTCCCGGGCCTTGTACACACCGCCGTCACACCGTCTAGTTGAGTGGGTTGACCCGAAGCCGGTGA
 GCGAACCGCAAGGACGAGCCGACCGACGGTGGTACGGCAACTGGGTG

12. The 16S rDNA nucleotide sequence of strain SIS32-2

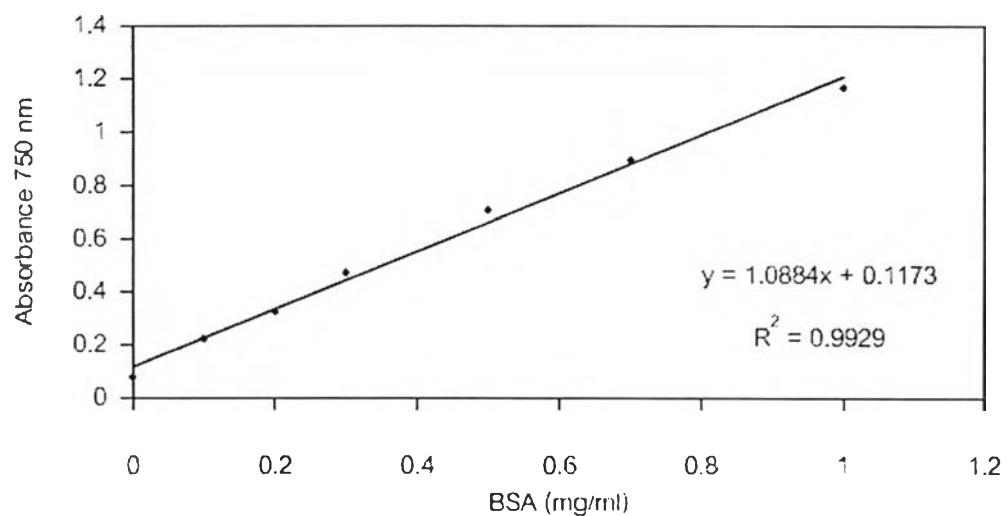
CGCACGGACGAGTAACGCGTAGGTATCTATCCATGGGTGGGGACAACACTCTGGGAAACTGGAGCTA
 ATACCGCATGACACCTGAGGGTCAAAGGCAGTCAGTCGCTGGAGGAGCCTCGCTCGATTAGCTTG
 TTGGTGGGTAATGGCTACCAAGGCATGATCGATAGCTGGCTGAGAGGATGATCAGCCACACTG
 GGACTGAGACACGCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGGACAATGGCGCAAG
 CTGATCCAGCAATGCCCGTGTGAAGAACGGTTTGGATTGTAAAGCACTITCGACGGGAGCAT
 GATGACGGTATCGATAGATATTGGAAAGAACACCGGTGGCAAGGCAGCACCTGGCTCATAACTGA
 CGCTGAGGCGCAGAGCTGGGAGCAAACAGGATTAGATACCCTGGTAGTCACGCTGTAAACGA
 TGTGTGCTGGATGTTGGTAGCTTAGTCATTCACTGTCAGTTAACGCGATAAGCACACGCCCTGG
 GGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGACGGGGCCCGACAAGCGGTGGAGCATGT
 GGTTTAATTGAAAGCAACGCCAGAACCTTACCAAGGCTTGCACATGGGAGGTATCCAGAGATG
 GGTATTCCCGAAGGGACCTCCTGCACAGGTGCTGCATGGCTGTCAGCTCGTGTGAGATGT
 TGGTTAAGTCATGACGGTATCGATAGATATTGGAAAGAACACCGGTGGCGAAGGCAGCACCT
 GGCTCATAACTGACGCTGAGGCCGAAAGCGTGGGAGCAAACAGGATTAGATACCCCTGGTAGTC
 ACGCTGAAACGATGTTGCTCGCAACGAGCGAACCCCTCGCTTAGTTGCGCATGATTGGGT
 GGGCACTCTAAAGGAACTGCCGTGACAAGCCGGAGGAAGGTGGGATGACGCTAAGTCCTCATGG
 CCCTTATGTCCTGGCTACACACGTCTACAATGGCGGTGACAGTGGGAAGCCAGGCAGCGATGCC
 AGCTGATCTCAAAAGCGCTCTCAGTCGGATTGCACTCTGCAACTCGAGTGCATGAAGGTGGAATC
 GCTAGTAATCGCGATCAGCATGCCGGTGAATACGTTCCGGCCTGTACACGGCGATGATCGA
 TAGCTGGTCTGAGAGGATGATCAGCCACACTGGGACTGAGACACGCCAGACTCCTACGGGAGGC
 AGCAGTGGGAATATTGGACAATGGCGCAAGCTGATCCAGCAATGCCCGTGTGAAGAAGGTC
 TTCGGATTGTAAGCACTITCGACGGGAGCATGACGGTA

13. The 16S rDNA nucleotide sequence of strain SI15-1

AGCGAACGCTGGCGCATGCTAACACATGCAAGTCGACGGACCTTCGGGGTAGTGGCGGACG
 GGTGAGTAACGCGTAGGGATCTATCCACGGGTGGGGATAACACCGGAAACTGGTCTAATACCG
 CATGATACCTGAGGGTCAAAGGCAGTCAGTCGCTGTGGAGGAGCCTCGCTCGATTAGCTGGT
 GGGGTAAAGGCCTACCAAGGCATGATCGATAGCTGGCTGAGAGGATGATCAGCCACACTGGGAC
 TGAGACACGCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGACAATGGCGCAAGCGT
 ATCCAGCAATGCCGTGTGAAGAACGGTTTGGATTGAAAGCACTTTAACGGGGACGATGAT
 GACGGTACCCAGAAGAACGCCGGCTACTTCGTGCCAGCAGCGCGTAATACGAAGGGGGCTAGC
 GTTGCTCGGAATGACTGGCGTAAGGGCGTGTAGGCGTTGACAGTCAGATGTGAAATTCCAGGG
 CTTAACCTGGGCTGCAATTGATACGTGTAGACTAGAGTGTGAGAGAGGGTTGGAATTCCAGGT
 GTAGAGGTGAAATTCTGAGATATTGGAAAGAACACCGGTGGCAAGGCCAACCTGGCTCATGAC
 TGACGCTGAGGCCGAAAGCGTGGGAGCAAACAGGATTAGATACCCCTGGTAGTCCACGCTGTAAA
 CGATGTTGCTGGATGTTGGTAACCTGGTTACTCAGTGTGCAAGCTAACCGCTAACGCC
 TGGGAGTACGCCGCAAGGTTGAAACTCAAAGGAATTGACGGGGCCCGACAAGCGGTGGAGCA
 TGTGGTTAACCGCAAGAACGCCAGAACCTAACAGGCTTGCACATGGGAGGCTGTATCCAGAG
 ATGGGTATTCCCGAACGGACCTCTGCACAGGTGCTGCATGGCTGTCAGCTCGTGTGAG
 ATGTTGGGTTAACCGCAACGAGGCCAACCTCGCTTGTAGTTGCCAGCACGTTGGTGGGCAC
 TCTAGAGGAACCTGGGTGACAAGCCGGAGGAAGGTGGGATGACGCTAACGTCCTCATGGCCCTAT
 GTCCTGGGCTACACACGTGCTACAATGGCGGTGACAGTGGGAAGCTAGTGGTACATCGTGGCGAT
 CTCTAAAAGCCGCTCAGTTGGATTGACTCTGCAACTCGAGTACATGAAGGTGGAATCGCTAGTA
 ATCGCGGATCAGCATGCCCGGTGAATACGTTCCGGCCTGTACACACCGCCGTCACACCATGG
 GAGTTGGTTGACCGAACGCCGAGCGAACCGCAAGGACGCG

14. The 16S rDNA nucleotide sequence of strain CT8-1

AGCGAACGCTGGCGCATGCTAACACATGCAAGTCGCACGGACCTTCGGGTCAAGTGGCGGACG
GGTAGAGTAACCGCTAGGGATCTATCCATGGGTGGGGATAACACTGGGAAACTGGTCTAATACCG
CATGATGCCCTGAGGGCCAAGGCAGTCGCCTGTGGAGGAGCCTGCGTTGATTAGCTTGGT
GGGGTAAAGGCCTACCAAGGCATGATCGATAGCTGGTCTGAGAGGATGATGCCACWCTGGAC
TGAGTCACGGCCCAGACTCCTACGGGAGGCAGCAGTGGGAATATTGGACAATGGCGCAAGCCTG
ATCCAGCAATGCCGTGTGAAGAAGGCATGATGCCCTGAGGGCCAAGGCAGTCGCCTG
TGGAGGAGCCTGCCGTTGATTAGCTTGGTGGTAAAGGCCTACCAAGGCATGATCGATAGCT
GGTCTGAGAGGATGATGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAG
TGGGAATATTGGACAATGGCGCAAGCCTGATCCAGCAATGCCGTGTGAAGAAGGTCTTCGG
ATTGTAAGCACTTCGACGGGACGATGATGACGGACCCCTAGAAGAAGCCCCGCTAACCTCGTGC
CCAGCAGCCGCGGTAATACGAAGGGGATGACGGACCCGTTAGAAGAAGCCCCGCTAACCTCGTGC
GCAGCCGCGGTAATACGAAGGGGCTAGCGTTGCTCGGAATAACTGGGCTAAAGGCAGCTAGGC
GGTTTGGACAGTCAGATGTGAAATTCTGGGCTTAACCTGGGCTGCTATTGATACGTACAGACTA
GAGTGTGAGAGAGGGTTGTGAAATTCCAGTGTAGAGGTGAAATTCTGAGATATTGGGAAAGAACAC
CGGTGGCGAAGGCTTCGAAAGTAAAGCACTTCGACGGGACGATGATGATCGGCCCTAGAA
GAAGCCCCGCTAACCTCGTCCAGCAGCCGCGTAATACGAAGGGGCTAGCGTTGCTCGGAATA
ACTGGGCGTAAGGGCGCTAGGCCTTGGACAGTCAGATGTGAAATTCTGGGCTAACCTGGG
GCTGCATTGATACGTACAGACTAGAGTGTGAGAGAGGGTTGTGAAATTCCAGTGTAGAGGTGAAA
TTCGTAGATATTGGGAAAGAACACCGGTGGCGAAGGCAGCAACCTGGCTCATGACTGACGCTGAGGC
GCGAAAGCGTGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCTGTAACCGATGTGCTG
GATGTTGGCAACTTAGTTGCTAGTGTCTAGCTAACCGCTAACCGAGTACGG
CCGC

APPENDIX E**Standard curve of Bovine serum albumin (BSA) and Total acid****1. Standard curve of Bovine serum albumin (BSA)****2. Total acid (Helrich, 1990)**

$$\text{Total acid (g/100ml)} = \frac{N \times V \times 60.1 \times 100}{1000 \times 10}$$

N = concentration of 0.1N NaOH

V = volume of 0.1N NaOH

APPENDIX F

Restriction size of *Acetobacter* and *Gluconobacter* type strains

Restriction of 16S-23S rDNA regions PCR product by digestion with five restriction endonucleases in *Acetobacter* and *Gluconobacter* type strains

strains	Molecular size of restriction fragments (bp) by digestion with				
	<i>Hpa</i> II	<i>Hae</i> III	<i>Bsp</i> 1286I	<i>Mbo</i> II	<i>Ava</i> II
<i>A. indonesiensis</i> NBRC 16471 ^T	340, 390	370,570	-	-	-
<i>A. cibinongensis</i> NBRC 16605 ^T	350, 400	370, 590	-	-	-
<i>A. lovaniensis</i> NBRC 13753 ^T	495	325, 570	-	-	-
<i>A. tropicalis</i> NBRC 16470 ^T	390	350, 570	-	-	-
<i>A. orientalis</i> NBRC 16606 ^T	400, 500	350, 580	-	-	-
<i>A. pasteurianus</i> TISTR 1056 ^T	350, 530	345, 570	-	-	-
<i>A. aceti</i> NBRC 14818 ^T	400, 550, 900	340, 575	-	-	-
<i>G. oxydans</i> NBRC 14819 ^T	-	-	94, 125, 440	610	315
<i>G. albidus</i> NBRC 3250 ^T	-	-	94, 125, 440	190, 360	315
<i>G. cerinus</i> NBRC 3267 ^T	-	-	94, 125, 180, 250	171, 356	714
<i>G. thailandicus</i> NBRC 100600 ^T	-	-	51, 92, 125, 244	360	714
<i>G. frateurii</i> NBRC 3264 ^T	-	-	51, 92, 125, 244	360	610

Biography

Miss Jintana Kommanee was born on November 8, 1981 in Khon Kaen Province, Thailand. She received her Bachelor Degree of Biotechnology 2004 from Department of Biotechnology, Faculty of Science, King Mongkut's Institute of Technology Ladkrabang, Thailand.

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