FLUORESCENCE SENSORS FROM PYRENO[4,5-d]IMIDAZOLE DERIVATIVES



A Dissertation Submitted in Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy in Chemistry Department of Chemistry FACULTY OF SCIENCE Chulalongkorn University Academic Year 2020 Copyright of Chulalongkorn University ฟลูออเรสเซนต์เซนเซอร์จากอนุพันธ์ของไพรีโน[4,5-d]อิมิดาโซล



วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรดุษฎีบัณฑิต สาขาวิชาเคมี คณะวิทยาศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย ปีการศึกษา 2563 ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

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ณิชาภา ชนะวังเมือง : ฟลูออเรสเซนต์เซนเซอร์จากอนุพันธ์ของไพริโน[4,5-d]อิมิดาโซล. (FLUORESCENCE SENSORS FROM PYRENO[4,5-d]IMIDAZOLE DERIVATIVES) อ.ที่ปรึกษาหลัก : ศ. ดร.ไพฑูรย์ รัชตะสาคร, อ.ที่ปรึกษาร่วม : ศ. ดร.มงคล สุขวัฒนาสินิทธิ์

งานวิจัยนี้มีการออกแบบและพัฒนาอนุพันธ์ของไพรีโนอิมิดาโซล (P1, P2, P3, P4 และ P5) ซึ่งสามารถสังเคราะห์ได้จากสารตั้งต้นไพรีนใน 2 ขั้นตอน ได้ผลิตภัณฑ์ร้อยละ 26 – 40% หลังจากนั้นทำการศึกษาคุณสมบัติทางแสงและการตรวจจับไอออนโลหะของโมเลกุลทั้งห้า ผลการ ทดลองพบว่าโมเลกุล P5 ไม่มีการการเลือกจับกับไอออนของโลหะใดๆ ในขณะที่โมเลกุล P1, P2, P3 และ P4 มีการเลือกจับกับไอออนของโลหะที่มีเวเลนซ์อิเล็กตรอน 3+ (บิทมัสไอออน, อะลูมิเนียมไอออน, โครเมียมไอออน, ไอออนเหล็ก) ในสารละลายอะซิโตไนไตรล์ (CH3CN) แต่ อย่างไรก็ตามมีเพียงอนุพันธ์ของไพรีโนอิมิดาโซลที่แทนที่บนวงอิมิดาโซลด้วยอนุพันธ์ของ 2-ไฮด รอกซิลฟีนิล (P2) ที่แสดงให้เห็นว่ามีความจำเพาะเจาจงในการตรวจจับไอออนของบิสมัทใน สารละลายผสม 20% ของไดเมทิลซัลฟอกไซด์ในอะซิโตไนไตรล์ แบบเพิ่มสัญญาณของฟลูออเรส เซนซ์ เนื่องมาจากกระบวนการเคลื่อนย้ายโปรตอน-อิเล็กตรอน ภายในโมเลกุลในสถานะกระตุ้น (excited state intramolecular proton transfer ,ESIPT) โดยการเกิดสารประกอบระหว่าง โมเลกุลและไอออนยับยั้งการเกิดพันธะไฮโดรเจนภายในโมเลกุล และโมเลกุล P2 มีการขยาย สัญญาณฟลูออเรสเซนซ์ที่ความยาวคลื่น 447 นาโนเมตร สำหรับกลไกการเกิดสารประกอบ ระหว่างโมเลกุลและไอออนบิสมัทสามารถตรวจสอบได้โดยวิธีทางสเปกโทรสโกปี ได้แก่ ¹H-NMR และ เทคนิคการวัดไทเทรซันด้วยยูวีและฟลูออเรสเซนซ์ ซึ่งบ่งชี้ให้เห็นว่าอัตราส่วนที่จับกันเป็น 1:1 ระหว่างสารประกอบ P2 และบิสมัทไอออน และให้ค่าต่ำสุดที่สามารถตรวจวัดได้ (LOD) ประมาณ 1.20 ไมโครโมลาร์ในสารละลายผสม 20% ของไดเมทิลซัลฟอกไซด์ในอะซิโตไนไตรล์ และ 3.40 ไม โครโมลาร์ใน สารละลาย 10% น้ำในไดเมทิลซัลฟอกไซด์ในอะซิโตไนไตรล์ที่สภาวะพีเอซ 5 ้นอกจากนี้ได้ทำการประยุกต์ใช้เซนเซอร์นี้ทดลองหาปริมาณบิสมัทในน้ำตัวอย่างสองชนิด พบว่ามี ค่าร้อยละการกลับคืน 91-97%.

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Nichapa Chanawungmuang : FLUORESCENCE SENSORS FROM PYRENO[4,5d]IMIDAZOLE DERIVATIVES. Advisor: Prof. PAITOON RASHATASAKHON, Ph.D. Co-advisor: Prof. MONGKOL SUKWATTANASINITT, Ph.D.

Herein, The five fluorescent probes (P1, P2, P3, P4 and P5) based on pyreno[4,5-d]imidazole were successfully prepared in two steps from pyrene in 26 - 40% overall yields. After that, all of the compounds were studied the photophysical properties and selectivity for sensing metal ions. The results showed compound P5 was not selective to metal ions while the others (P1, P2, P3 and P4) were selective to detection of trivalent metal ions such as Bi³⁺, Al³⁺, Cr³⁺ and Fe³⁺ in CH₃CN. However, only compound P2 with 2-hydroxyl phenyl substituent on imidazole ring formed complexation reaction selectively towards bismuth (III) ions and provided fluorescence enhancement signal attributing to the excited state intramolecular proton transfer (ESIPT) process. Compound P2 exhibited selective fluorescence turn-on at 447 nm. The sensing mechanism was determined by ¹H-NMR, and UV-VIS and fluorescence titrations, and the Job's plot indicated a 1:1 binding between P2 and bismuth (III) ions. The detection limit was 1.20 µM in CH₃CN-DMSO (8:2, v/v) mixture and 3.40 µM in 10% aqueous in the mixed 80% CH3CN-DMSO (pH = 5). Moreover, the sensor was applied to quantitative analysis of spiked Bi3+ in real water samples from different sources. The recovery of Bi³⁺ was completed at 91-97%.

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LIST OF SCHEMES



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LIST OF ABBREVIATIONS

Ar	aromatic
calcd	calculated
¹³ C NMR	carbon-13 nuclear magnetic resonance
CH ₃ CN	acetonitrile
DMSO	dimethyl sulfoxide
d	doublet (NMR)
dd	doublet of doublet (NMR)
equiv	equivalent (s)
ESI-MS	electrospray ionization mass spectrometry
EtOH	ethanol
FT-IR	fourier transform infrared spectroscopy
g	gram (s)
¹ H NMR	proton nuclear magnetic resonance
Hz	Hertz
h จหาะ	hour (s)
J Chula	coupling constant
K _{SP}	solubility product constant
MALDI-TOF	matrix assisted laser desorption/ionization-time of flight
mg	milligram (s)
mL	milliliter (s)
mmol	millimole (s)
m/z	mass per charge
m	multiplet (NMR)
M.W.	molecular weight
Μ	molar

MHz	megahertz
rt	room temperature
S	singlet (NMR)
THF	tetrahydrofuran
TLC	thin layer chromatography
UV	ultraviolet
δ	chemical shift
°C	degree Celsius
μL	microliter (s)
μΜ	micromolar (s)
Φ	quantum yield
% yield	percentage yield
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	จุฬาลงกรณ์มหาวิทยาลัย

CHAPTER I

1.1 Fluorescence

The phenomenon of fluorescence is one of radiative processes. Fluorescence is the emission of light that results from deactivation of excited molecules to lower energy level occurring in highly conjugated molecules or aromatic compounds. The absorption and emission of light in fluorescence process can be classically presented by the Jablonski diagram as shown in Figure 1.1 [1]. This diagram showed electronic and vibrational energy levels. When a molecule absorbed light energy, electron in the molecule were excited to the excitation state (S_1) to form excited molecule. The molecule is not stable and will rapidly relax to the lowest vibrational level of the first excited state (S_1) via releasing some energy as kinetic energy (rotation or vibration) or thermal energy (heat). This process is non-radiative transition known as geometric relaxation. Then, the molecule will deexcited and return to the ground state (S_0) via emission of a longer wavelength photon. The lifetime of fluorescence process takes in nano-second.



Figure 1.1 Jablonski diagram

1.2 Fluorescent sensor

Nowadays, fluorescent sensors play an important role as detection methods in chemical, biological, and environmental fields such as detection of metal ions, anions, and biological molecules in water samples or living cells. Fluorescence technique is a simple method that has several advantages such as high selectivity, high sensitivity, short response time, cost-effectiveness in instrumentation, and non-destructive technique of sample. In general, a fluorescent sensor consists of two components: one is a unit for selective binding with analyte of interest that is called "receptor" and the other is "reporter" that translates the signal changing. The reporter of fluorescent sensor is called "fluorophore". The response of fluorophore can be read-out from changing of fluorescence or wavelength shift (ratiometric sensor) [2] as shown in Figure 1.2.



Figure 1.2 Modes of fluorescent sensors

1.3 Sensing Mechanisms

The photophysical signaling mechanisms between interaction or reaction of analyte and receptor which could be explained by the photoinduced electron transfer (PET) [3-7], fluorescence resonance energy transfer (FRET) [5,7-11], Intermolecular charge transfer (ICT) [5,12-15], excited-state intramolecular proton transfer (ESIPT) [12,13,16-20], structure isomerization, aggregation-induced enhancement fluorescence (AIE) [21-24], aggregation-caused quenching (ACQ) [22,23,25,26] and excimer formation [27]. Therefore, fluorescent sensors could be designed on one or more than one sensing mechanism which related to the responding of fluorophore. In this work, the sensing compounds will be designed based on ESIPT and PET process.

Photo-induced electron transfer (PET)

Photo-induced electron transfer (PET) is fluorescence quenching process that can occur when either the highest occupied molecular orbital (HOMO) or the lowest unoccupied molecular orbital (LUMO) level of receptor or analyte is located between the HOMO-LUMO gap of fluorophore. The first case, a fluorescent compound can act as an electron acceptor since the HOMO level of fluorophore is lower than the HOMO level of binding site (Figure 1.3A). After an electron of the fluorophore was excited, one of the electrons at the HOMO level of the donor (receptor or analyte) will be transferred to the singly occupied HOMO of the fluorophore. As the results of the proton transfer process, it precludes the releasing of energy from LUMO to HOMO for fluorophore, so the fluorescence process is inhibited. The other case, a fluorescent compound acts as an electron donor for the LUMO level of receptor or analyte and the HOMO-LUMO gap of fluorophore are shown in Figure 1.3B. In this case, an excited electron at the LUMO level of the fluorophore will be transferred to the empty LUMO of the receptor or analyte resulting in the return to the ground state but it is non-radiative process.



Figure 1.3 Principle of photo-induced electron transfer (PET) effect.

Excited state intramolecular proton transfer (ESIPT)

Excited state intramolecular proton transfer (ESIPT) is generally occurred for suitable organic compounds containing both a basic site or hydrogen bond acceptor such as carbonyl group, (-C=O) or imine nitrogen (=N-) and a protic group or hydrogen bond donor such as hydroxyl (-OH), amino (-NH2) or thiol group (-SH) that forms a fast process of an intramolecular hydrogen bond. The classical model of ESIPT photophysical process was demonstrated by 2- (2'-hydroxyphenyl) benzoxazole (HBO), -benzothiazole (HBT) and -benzimidazole (HBI) as illustrated in Figure 1.4 [16,28]. After photoexcitation, the excited state enol form (E_c^*) is converted to its excited state keto form (K^*) in the picosecond time scale. The significantly red shift emission and unusually large Stoke shift can be observed for ESIPT molecule. A large Stoke shift is useful in fluorescence sensing to keep away from the inner filter or the self-absorption effect.



Figure 1.4 Principle of Excited state intramolecular proton transfer (ESIPT) process

1.4 Fluorescent sensors based on Pyreno[4,5-d]imidazole

To design and synthesize new molecules based on pyrene as the reactant because of its photophysical properties. Pyrene is a polycyclic aromatic which has been wildly used for organic electronics, for examples, to make field-effect transistors and organic light-emitting diodes (OLEDs) [29-31]. It is also capable as fluorophores due to its emission wavelength in the visible region, long fluorescence lifetime and high fluorescence quantum yield [32-35]. Moreover, it could be connected and modified to different binding unit. The position of pyrene molecule could be easy to modify by electrophilic aromatic substitution reaction at position 1, 3, 6, and 8 and addition of steric hindrance to molecule could be oxidized to different site of steric hindrance to molecule could be oxidized to diones or tatraones depending on the reagents, equivalents of oxidizing reagent and time of reaction (Figure 1.5) [38-40].



Electrophilic Aromatic Substitution

Figure 1.5 The reactions on positions of pyrene

For the connecting unit is an imidazole fragment that contains N and H atom which play an essential role in an intramolecular and intermolecular interactions such as hydrogen bond, metal-organic coordination, excited state intramolecular proton transfer (ESIPT), or concerted proton-electron transfer (CPET) [41]. Thus, many researchers are interested to use imidazole part for design the fluorescent sensor.

In 2011, Ju and co-worker [42] synthesized 2-(1-phenyl-1*H*-benzo[d]imidazol-2yl)phenol (HBIZ) for sensing zinc (Zn²⁺). The results found that the fluorescence intensity of molecule was increased by addition of Zn²⁺ and Mg²⁺ in 120 and 15 folds, respectively (Figure 1.6a). The complexation of Zn²⁺ and HBIZ was 1:1 stoichiometry that was investigated by Job's plot. However, the coexist of other metal ions such as Cu²⁺, Fe²⁺, Ni²⁺ could be quenched the fluorescence intensities (Figure 1.6b).



Figure 1.6 (a) Structure of HBIZ; (b) The fluorescence ratio of HBIZ (50 μ M) in the presence of various metal ions (10 μ M) in *N*,*N*-Dimethylformamide, λ_{ex} = 330 nm.

In 2014, Zhang et al. developed derivative of 3-(4,5-diphenyl-1*H*-imidazol-2yl)naphthalen-2-ol (DPIN) as fluorescent sensor displaying the ESIPT process [19]. The hydroxyl group of DPIN was conjugated with acryloyl group to DRIN-A (Figure 1.7a) that can react with biothiols (glutathione (GHS), cysteine (Cys), homocysteine (Hcy)). The result showed high selectivity and sensitivity to detection of cys in water (Figure 1.7b).



Figure 1.7 (a) Synthesis of DPIN-A.; (b) Fluorescent spectra of DPIN-A (10 mM) in pure water (PBS buffer, pH =7.2) towards biothiols (10 eq of GSH, Cys, Hcy.) were obtained after 30 min and other amino acids (100 eq.) were recorded after 1 hour.), λ_{ex} =371 nm.

In 2015, Hu and co-worker designed and developed fluorescent sensor without sulfur atom (S) in the molecule to sensing mercury ions (Hg^{2+}) [43]. The sensor (L) was designed based on the benzimidazole group as recognition group and the quinoline group as a fluorophore group. The sensor was selectively quenched by Hg^{2+} in water- dimethyl sulfoxide (DMSO) (1:9, v/v) through concerted proton-electron transfer (CPEF) process (Figure 1.8b). The binding between sensor L and Hg^{2+} was presented to form a stable complex [L + Hg^{2+} + $2H_2O$] in Figure 1.8a that was investigated by ¹H NMR titration, IR spectra and mass spectrum.



Figure 1.8 (a) Possible sensing mechanism of sensor L and Hg²⁺.; (b) Fluorescence spectra of L (20 μ M) towards various metal ions (10 equiv.) in H₂O-DMSO (1:9, v/v) (λ_{ex} = 325 nm).

Based on previous research, small aromatic rings such as benzene or naphthalene compounds were acted to fluorophore. In 2016, phenanthrene, a large aromatic compound, were used as fluorophore connecting with imidazole moiety. For instance, a ratiometric fluorescent sensor based on ESIPT mechanism between phenolic (-OH) and imidazole nitrogen (C=N) was developed by Shinha and co-worker [44]. The sensor **1H** was selective with aluminum ions (Al³⁺) in CH₃CN-H₂O (1:9, v/v) about sixfold enhancement in fluorescence intensity and a blue-shift from 445 to 412 and 430 nm. The possible binding mechanism was shown in Figure 1.9a. However, the large concentration of chromium ions (Cr³⁺) and indium ions (In³⁺) was very small increased fluorescence intensity and copper ions (Cu²⁺) could affect to quench the fluorescence intensity of **1H** due to its paramagnetic property as shown in Figure 1.9b.



Figure 1.9 (a) Possible binding mechanism between 1H and Al³⁺; (b) Fluorescence spectra of 1H (2.5 μ M) towards various metal ions (10 equiv.) in CH₃CN/H₂O (1:9, v/v), λ_{ex} =375 nm.

Moreover, in 2017, Cheng et al. synthesized a novel fluorescent sensor (Probe 1) based on phenanthro-imidazole connecting with a N,N-bis(pyridin-2-ylmethyl) benzeneamine unit that was a binding site for detection of copper ions (Cu²⁺) (Figure 1.10a). Upon addition of Cu²⁺, the fluorescence sensing was quenched through the electron charge transfer (CT) process based on 1:1 stoichiometry binding. The sensing was also highly and quickly selective towards Cu²⁺ in ethanol (EtOH) without significant influence from other metal ions as presented in Figure 1.10b. The low detection limit was 1.77 ppb.



Figure 1.10 (a) Binding mechanism of Probe 1 with Cu^{2+} undergo CT process.; (b) Fluorescence intensity of Probe 1 (10 µM) without metal ions (**Green**), in the presence of various metal ions (1 equiv.) (**Blue**), and with 10 µM of Cu^{2+} in the presence of other metal ions (**Red**), $\lambda_{em} = 387$ nm.

In the same year, Chen et al. [45] designed a three-components fluorescent probe (PAN-Bn-PB), phenanthrol-imidazole, benzonitrile, and phenyl boronate, based on an intramolecular charge transfer (ICT) process for sensing hydrogen peroxide (H_2O_2) in vitro and in vivo. The turn-on fluorescence sensor was good selectivity and sensitivity to detecting H_2O_2 in several minutes. However, it could not be perfect probe for H_2O_2 in vivo because its emission wavelength was blue.



Figure 1.11 (a) Sensing mechanism of PAN-Bn-PB with H_2O_2 .; (b) Fluorescence intensity of PAN-Bn-PB (10 μ M) in various molecules that could exist in biological system (200 μ M).

Recently, Li and co-worker [12] synthesized and designed a pH fluorescence probe (PIF) from phenanthrol-imidazole connect with fluorescein by a hydroxyl phenyl imine group based on ESIPT and ICT process (Figure 1.12). The pH sensing of PIF is capable to respond pH from 6.5 to 9.5 without interference of cations and anions.



Figure 1.12 Proposed sensing mechanism of PIF in acid-base conditions.

For pyreno[4,5-*d*]imidazole derivatives, in 2013, Liu et al [46] reported the properties of a novel skeleton containing pyrene and imidazole units that was synthesized in one step. The results found that both PyDPI and PyDCzl were stable in high thermal and showed deep-blue emission in solution. Furthermore, both molecules tended to aggregate into thin films and crystallize as powders. These properties could be applied to use in material field, for examples, organic light emitting diodes (OLEDs) or dye-sensitized solar cells (DSSCs).

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Figure 1.13 The one-step to synthesis of pyrene–imidazole skeletons. Nevertheless, a few researches developed pyreno[4,5-*d*]imidazole derivatives to fluorescent sensor for sensing anions such as cyanide sensor, fluoride sensors. In 2015, 10-(1-*H*-imidazole-2-yl)-9*H*-pyreno[4,5-*d*]imidazole (Py-BiimzH₂) was synthesized by Mardanya et at. [47,48] that was studied to sensing anions. Upon addition of various anions to Py-BiimzH₂, it was selective to sensing cyanide ions (CN⁻) and fluoride ions (F⁻) in DMSO. The detection limits were calculated to 1.70×10^{-7} M. for CN⁻ and 1.54×10^{-7} M. for F⁻. However, in aqueous medium, only CN⁻ was enhanced fluorescence intensity at 475 nm. in H₂O–DMSO (9:1, v/v) as shown in Figure 1.14. It might be CN⁻ is a stronger base than F⁻ in aqueous solution. The detection limit was 1.04×10^{-6} M in aqueous solution.



Figure 1.14 Structure of Py-BiimzH₂ and luminescence spectra of Py-BiimzH₂ upon addition of various anions as their tetrabutylammonium (TBA) salts (300 equiv.) in H₂O-DMSO (9:1, v/v), $\lambda_{ec} = 350$ nm [47].

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In the same year, Mardanya and co-worker also developed high selectivity of fluorogenic sensors for cyanide ions in aqueous solution, the two complexation between Ru(II) or Os(II) with Py-BiimzH₂ and 2,2'-bipyridine was synthesized to form $[(bpy)_2Ru(Py-BiimzH_2)](ClO_4)\cdot H_2O$ (**A**) and $[(bpy)_2Os(Py-BiimzH_2)](ClO_4)\cdot H_2O$ (**B**). Quenching luminescence intensity was observed upon addition of CN⁻ for both sensors (Figure 1.15). Both complexes were high selectivity to detection of CN⁻ in aqueous media in the presence of various anions. The detection limit of **1** and **2** to CN⁻ in aqueous media was determined to be 1.03×10^{-8} M and 1.24×10^{-8} M, respectively.



Figure 1.15 UV-Vis spectra and luminescence spectra of complex A and B upon addition of various anions as their tetrabutylammonium (TBA) or sodium salts (300 equiv.) in pure water (HEPES buffer, pH = 7) [48].

In addition, Mardanya and co-worker [49] studied and synthesized the new complexation of Ru²⁺ (**C**) and Os²⁺ (**D**) with 10-pyridin-2-yl-9H-9,11diazacyclopenta[e]pyrene (HImzPPy) and 2,2'-bipyridine. The complexes could be enhanced intensity as turn-on luminescence for sensing dihydrogen phosphate ions $(H_2PO_4^{-})$ resulting from the interaction of hydrogen-bonding between NH proton of imidazole and $H_2PO_4^{-}$ and quenched intensity as turn-off luminescence for sensing fluoride ions (F⁻) and acetate ions (AcO⁻) resulting from the deprotonation of NH proton of imidazole. The range of detection limit of 1 and 2 to each anion was calculated about 10⁻⁹ M as shown in (Figure 1.16).



Figure 1.16 Luminescence spectra of complex **C** and **D** upon addition of various anions as their tetrabutylammonium (TBA) in CH₃CN.

In recently, Tabasi et al. [50] synthesized and studied the two isomers of (9*H*-pyreno[4,5-d]imidazol-10-yl)-benzaldehyde, para (4) and meta (5) isomers. The para isomer acts as ratiometric fluorescent sensor which could be reacted with fluoride ions (F⁻) in polar solvents (acetone and DMSO) undergo ICT process. (Figure 1.17)



Figure 1.17 Proposed mechanism and fluorescence titration of (9H-pyreno[4,5*d*]imidazol10-yl)-benzaldehyde in DMSO.

From the literatures, it could be support to design and synthesize new derivative of pyreno[4,5-*d*]imidazole in order to fluorescent sensor to detection of

metal ions, anions or other analytes because of a few research using this derivatives as fluorescence sensor.

1.5 Objective of this research

To this end, we designed about five pyreno[4,5-*d*]imidazole derivatives (**P1-P5**), (Figure 1.18) and aim to use them as specific fluorescent sensors for metal ions. The five compounds contain different type of aromatic ring that can provide information on mechanism of sensing such as the Excited State Intramolecular Proton Transfer (ESIPT) or photo-induced electron transfer (PET).



CHAPTER II EXPERIMENTAL

2.1 Materials and Chemicals

All reagents used in reactions were analytical grade purchased from Sigma-Aldrich (USA), Merck (Germany) or Fluka (Switzerland). Solvents for extraction and chromatography such as dichloromethane (CH_2Cl_2), hexanes, ethyl acetate (EtOAc) and methanol (CH_3OH) were commercial grade and used without further purification. All column chromatography was operated using Merck silica gel 60 (70–230 mesh) and Thin layer chromatography (TLC) was performed on silica gel plates (Merck F245). The stock solutions of the 3 compounds, 10-phenyl-9H-pyreno[4,5-d]imidazole (**P1**), 2-(9H-pyreno[4,5-d]imidazol-10-yl)phenol (**P2**) and 10-(2-methoxyphenyl)-9Hpyreno[4,5-d]imidazole (**P3**), were prepared at 1 mM in acetonitrile (CH₃CN) that were used as fluorescent sensors and at 1 mM in ethanol (EtOH) that were used for studies the photophysical properties.

2.2 Analytical instruments

The target molecules were characterized by MALDI-TOF mass spectrometer (Bruker Daltonics) using α -cyano-4-hydroxycinnamic acid (CCA) as a matrix. All ¹H-NMR spectra were acquired from sample solution in Deuterated NMR solvents such as CDCl₃, DMSO-D₆, D₂O and CD₃CN, on Varian Mercury 400 MHz and Jeol 500 MHz. ¹³C-NMR spectra were acquired using Bruker at 100 MHz. The UV-Vis absorption spectra were obtained from a Varian Cary 50 UV-vis spectrophotometer (Varian, USA) and the fluorescence emission spectra were recorded on a Varian Cary Eclipse spectrofluorometer (Varian, USA).

2.3 Synthesis and characterization

2.3.1 Pyrene-4,5-diones (1)



To the solution of pyrene (2.02g, 10 mmol) was dissolved in a 1:1 mixture of CH_2Cl_2 (40 mL) and CH_3CN (40 mL), then $RuCl_3.nH_2O$ (0.20 g, 0.96 mmol) was mixed to the solution. After that, $NalO_4$ (10.00 g, 46.8 mmol) and H_2O (50 mL) were added. The reaction was stirred overnight at room temperature. The mixture was poured into H_2O (500 mL). The organic phased was separated and the aqueous phase was extracted with CH_2Cl_2 (50 mL) 3 times. The combined organic phases were washed with H_2O (200 mL) 3 times and dried over anhydrous Na_2SO_4 to give an orange solution. The solvent was removed under reduced pressure to give a dark orange solid which was purified by column chromatography using CH_2Cl_2 . The product was obtained as bright orange crystal in 1.6302 g, 70% yield. ¹H-NMR (400 MHz, CDCl₃) δ 8.47 (dd, J = 7.4, 1.1 Hz, 2H), 8.16 (dd, J = 8.0, 1.2 Hz, 2H), 7.82 (s, 2H), 7.76 (t, J = 7.7 Hz, 2H).

To a mixture of dione **1** (0.23 g, 1 mmol) and benzaldehyde (0.13 g, 1.2 mmol) in CH₃COOH (10 mL) was added ammonium acetate (0.54 g, 7.5 mmol). The reaction was refluxed for 4 h to form a yellow precipitate. The mixture was cooled and poured into ice-H₂O mixture (10 mL) to remove soluble salt and facilitate a complete precipitate. The solid crude product was filtrated, washed with water, and

dried by suction. The yellow solid was purified by column chromatography using 20-25% ethyl acetate in hexanes. 10-Phenyl-9H-pyreno[4,5-d]imidazole (**P1**) was obtained as a yellow solid in 75% yield. ¹H-NMR (400 MHz, DMSO-d6) δ ppm 13.84 (s, br, NH), 8.85 (d, J = 7.5 Hz, 2H), 8.40 (d, J = 7.5 Hz, 2H), 8.27 (d, J = 7.4 Hz, 2H), 8.21 (s, 2H), 8.17 (t, J = 7.6 Hz, 2H), 7.65 (t, J = 7.6 Hz, 2H), 7.54 (t, J = 7.3 Hz, 1H); ¹³C-NMR (100 MHz, DMSO-d6) δ 149.41, 133.5, 130.35, 129.32, 128.96, 126.22, 124.13, 121.77, 118.97; MS (MALDI-TOF) Calcd for C₂₃H₁₄N [M+]: 318.116; found: 318.027.





To a mixture of dione **1** (0.23 g, 1 mmol) and salicylaldehyde (0.15 g, 1.2 mmol) in CH₃COOH (10 mL) was added ammonium acetate (0.54 g, 7.5 mmol). The reaction was refluxed overnight to form a yellow-orange precipitate. The mixture was cooled and poured into ice-H₂O mixture (10 mL) to remove soluble salt and facilitate a complete precipitate. The solid crude product was filtrated, washed with water, and dried by suction. The orange solid was purified by column chromatography using dichloromethane. 2-(*9H*-Pyreno[4,5-*d*]imidazol-10-yl)phenol (**P2**) was obtained as bright orange solid in 55% yield. ¹H-NMR (400 MHz, DMSO-d6) δ ppm 13.99(s, OH), 13.21 (s, NH), 8.87 (d, *J* = 7.6 Hz, 2H), 8.78 (d, *J* = 7.6 Hz, 2H), 8.32 (d, *J* = 7.6 Hz, 1H), 8.23 (s, 2H), 8.19 (t, *J* = 6.9 Hz, 2H), 7.44 (t, *J* = 7.7 Hz, 1H), 7.16– 7.11 (m, 2H); ¹³C-NMR (100 MHz, DMSO-d6) δ 157.44, 149.56, 131.51, 131.18, 127.67, 126.46, 125.89, 124.62, 121.83, 119.13, 117.22, 112.98; MS (ESI) Calcd C₂₃H₁₄N₂O [M+H]: 335.12; found: 335.02.

2.3.2.3 10-(2-methoxyphenyl)-9H-pyreno[4,5-d]imidazole (P3)


To a mixture of dione 1 (0.23 g, 1 mmol) and 2-methoxybenzaldehyde (0.20 g, 1.5 mmol) in CH₃COOH (10 mL) was added ammonium acetate (0.54 g, 7.5 mmol). The reaction was refluxed for 12 h to form an orange precipitate. The mixture was cooled and poured into ice-H₂O mixture (10 mL) to remove soluble salt and facilitate a complete precipitate. The solid crude product was filtrated, washed with water, and dried by suction. The orange solid was purified by column chromatography using 20-25% ethyl acetate in hexanes. 10-(2-Methoxyphenyl)-*9H*-pyreno[4,5-*d*]imidazole (**P3**) was obtained as dark orange solid in 86 %yield. ¹H-NMR (400 MHz, DMSO-d6) δ ppm 13.07 (s, br, NH), 8.91 (d, *J* = 7.5 Hz, 2H), 8.31 (d, *J* = 6.8 Hz, 1H), 8.25 (d, *J* = 7.6 Hz, 2H), 8.21 (s, 2H), 8.13 (t, *J* = 7.6 Hz, 2H), 7.54 (t, *J* = 7.2 Hz, 1H), 7.31 (d, *J* = 8.3 Hz, 1H), 7.20 (t, *J* = 7.4 Hz, 1H), 4.08 (s, 3H); ¹³C-NMR (100 MHz, DMSO-d6) δ 156.79, 147.39, 131.65, 131.13, 130.31, 127.80, 126.33, 124.26, 121.92, 121.02, 119.41, 112.12, 56.11; MS (MALDI-TOF) Calcd C₂₄H₁₆N₂O [M+]: 348.126; found: 348.716.

2.3.2.4 2-methoxy-6-(9H-pyreno[4,5-d]imidazol-10-yl)phenol (P4)



To a mixture of dione 1 (0.23 g, 1 mmol) and *o*-vanillin or 2-hydroxy-3methoxybenzaldehyde (0.23 g, 1 mmol) in CH₃COOH (10 mL) was added ammonium acetate (0.54 g, 7.5 mmol). The reaction was refluxed overnight to form a precipitate. The mixture was cooled and poured into ice-H₂O mixture (10 mL) to remove soluble salt and facilitate a complete precipitate. The solid crude product was filtrated, washed with water, and dried by suction. The yellow solid was purified by column chromatography using 20-25% ethyl acetate in hexanes. 2-methoxy-6-(9*H*-pyreno[4,5-*d*]imidazol-10-yl)phenol (**P4**) was obtained as light yellow solid in 0.0463 g, 13% yield. ¹H-NMR (400 MHz, CDCl₃) δ ppm 8.74 (d, *J* = 6.1 Hz, 2H), 8.31 (d, *J* = 4.8 Hz, 1H), 8.25 (m, br, 2H), 8.09 (m, br, 3H), 7.87 (m, 2H), 7.01 (m, 2H), 4.02 (s, 3H); ¹³C-NMR (100 MHz, DMSO-d6) δ 157.19, 150.61,149.44, 149.22, 132.06, 128.27, 127.03, 125.09, 122.34, 119.63, 119.21, 118.01, 114.03, 113.72, 56.37 ppm; MS (MALDI-TOF) Calcd C₂₄H₁₆N₂O₂ [M+]: 364.121; found: 364.024.





To a mixture of dione 1 (0.23 g, 1 mmol) and picolinaldehyde (0.16 g, 1.5 mmol) in CH₃COOH (10 mL) was added ammonium acetate (0.54 g, 7.5 mmol). The reaction was refluxed overnight to form a precipitate. The reaction was followed by TLC. A green solution was obtained. After that, the mixture was cooled and poured into ice-H₂O mixture (10 mL) to remove soluble salt and facilitate a complete precipitate. The solid crude product was filtrated, washed with water, and dried by suction. The yellow solid was purified by column chromatography using 1-10% v/v of CH₂Cl₂ – MeOH. 10-(pyridin-2-yl)-9*H*-pyreno[4,5-*d*]imidazole was obtained as yellowed solid in only 0.0460 g, 15% yield. ¹H NMR (400 MHz, CDCl₃) **δ** 13.04 (br, NH), 8.92 (d, *J* = 3.1 Hz, 2H), 8.68 (d, *J* = 7.2 Hz, 1H), 8.52 (d, *J* = 7.5 Hz, 1H), 8.21 (m, 2H), 8.14 – 8.05 (m, 4H), 8.00 – 7.95 (m, 1H) 7.52 – 7.46 (m, 1H); ¹³C-NMR (100 MHz, DMSO-d6) **δ** 161.57, 150.81, 146.38, 145.82, 138.35, 135.99, 132.07, 129.22, 128.73, 128.21, 127.61, 126.73, 126.48, 126.36, 124.94, 123.80, 123.20, 122.98, 120.39, 119.92, 118.70 ppm; MS (MALDI-TOF) Calcd C₂₂H₁₃N₃ [M+]: 319.111; found: 319.445.

2.4 Studies of photophysical properties

2.4.1 UV-visible spectroscopy

The UV-Visible absorption spectra of the solutions of fluorophores were recorded at ambient temperature from 200-500 nm. The solution of each compound was prepared in EtOH. The concentration of them was diluted to 10 μ M.

2.4.2 Molar extinction coefficient (ϵ)

The molar extinction coefficient (ϵ) of each fluorophore were calculated from the UV absorption spectra in EtOH at various concentrations. The absorption intensity of maximum wavelengths of each sample was plotted on Y-axis against the molar concentrations at the respective excitation wavelength on X-axis. Each graph should be a straight line and though origin-point. Then, the slope of graph was obtained to be the molar extinction coefficient of each fluorophore into the following equation:

 $A = \mathbf{E}bC$

Where A is the absorption intensity of sample.

 $\boldsymbol{\epsilon}$ is the molar extinction coefficient.

b is the pathlength in centimeter.

C is the molar concentration.

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2.4.3 Fluorescence spectroscopy

The fluorescence emission spectra of the solution of fluorophores were determined ambient temperature from 350-650 nm. The solution of each compound was prepared in EtOH. The concentration of them was diluted to 10 μ M.

2.4.4 Relative quantum yield

Fluorescence quantum yield (Φ_F) of P1, P2 and P3 were performed by using anthracene in EtOH (Φ_F = 0.27) as the standard reference. The UV-visible absorption spectra of reference samples and analytical samples were recorded under the condition that the maximum absorbance of all samples should never be above 0.1 at varied concentrations. The fluorescence emission spectra of the same samples at varied concentrations using appropriate excitation wavelengths selected were recorded based on the absorption maximum wavelength (λ_{max}) of each compound. The integrated fluorescence intensity on Y-axis were plotted against the absorbance at the respective excitation wavelengths on X-axis. Each plot should be straight line with 1 interception. Additionally, the fluorescence quantum yield (Φ_F) could be calculated into the following equation.

$$\Phi_{\rm X} = \Phi_{\rm ST} \left(\frac{{\rm Grad}_{\rm X}}{{\rm Grad}_{\rm ST}} \right) \left(\frac{\eta_{\rm X}^2}{\eta_{\rm ST}^2} \right)$$

Where Φ_{st} is the fluorescence quantum yield of standard reference.

 Φ_{χ} is the fluorescence quantum yield of sample.

 $Grad_{ST}$ is the gradient from the plot of integrated fluorescence intensity vs absorbance of standard reference.

 Grad_X is the gradient from the plot of integrated fluorescence intensity vs absorbance of sample.

 η_{ST} is the refractive index of standard reference.

 η_{X} is the refractive index of the solvent.

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2.5 Studies of metal ions sensing properties

The stock solutions of **P1**, **P2** and **P3** were prepared in CH₃CN and the stock solution of metal ions were prepared in Milli-Q water. They were pipetted and diluted to a desired concentration before spectrophotometric analysis

2.5.1 Effect of solvents

The stock solutions are diluted into 10 μ M by varied solvents such as CH₃CN, THF, EtOH, DMSO and water. The fluorescence intensities of fluorophore without and with analyte ware recorded using an excitation wavelength at room temperature.

2.5.2 Metal ion selectivity

The stock solutions of metal ions were prepared at 10 mM in Milli-Q water by dissolving commercial salts, LiNO₃, NaNO₃, KNO₃, AgNO₃, Ca(NO₃)₂, Mg(NO₃)₂, Ba(NO₃)₂, Co(NO₃)₂, Cd(NO₃)₂, Zn(NO₃)₂, Pb(NO₃)₂, Ni(NO₃)₂, Cu(NO₃)₂, Hg(OAc)₂, Fe(OAc)₂, Fe(NO₃)₃, Al(NO₃)₃, and Cr(NO₃)₃, except the solution of Bi(NO₃)₃ was prepared at 5 mM because it could not soluble to 10 mM.

The concentration of fluorophores and metal ions was diluted with solvent to 10 μ M and 100 μ M, respectively. The final volumes were adjusted to 1 mL.

2.5.3 Metal ions interference

The interfering effect of other metal ions on the detection of Bi^{3+} was studied by adding the other various competing metal ions into the solution of fluorophore in the presence of Bi^{3+} . The final concentration of fluorophore, Bi^{3+} and the other various competing metal ions were 10 μ M, 100 μ M and 100 μ M, respectively.

2.5.4 Fluorescence titration and Detection limit

Various concentration of Bi^{3+} (0-500 µM) was titrated into the solution of P2 (10 µM). The final volume was adjusted to 1 mL by adding CH₃CN/ DMSO (8:2 v/v). The fluorescent intensities were recorded from 300 nm to 700 nm at room temperature using an excitation wavelength. Moreover, the limit of detection (LOD) was obtained by plotting of the fluorescence intensity of fluorophore with concentrations of Bi^{3+} between 0-20 µM. The detection limit was calculated by the following equation:

Detection limit =
$$\frac{3\delta}{m}$$

Where δ is the standard deviation of the standard deviation (S.D) of ten independent measurements of a blank.

m is the slope between fluorescence intensity vs. concentrations of sample.

2.6 Studies of sensing mechanism

2.6.1 Time dependent

Fluorescence emission of **P2** was investigated after addition of Bi^{3+} in every 5 minutes for 30 minutes. The concentration of fluorophore and Bi^{3+} were 10 μ M and 100 μ M, respectively.

2.6.2 UV-visible and Fluorescence studies

To investigation of the interaction between **P2** and Bi^{3+} , concentration of Bi^{3+} from 1 µM to 500 µM was added to 10 µM of **P2**. The absorption wavelength and the fluorescence intensity were recorded. Moreover, the solution of sulfide ion (S²⁻) was added to the solution of **P2** in the presence of Bi^{3+} and adjusted with 20% DMSO/CH₃CN. The final concentration of **P2**, Bi^{3+} and S^{2-} were 10 µM, 100 µM and 100 µM, respectively.

2.6.3 ¹H NMR experiment

The fluorophore (**P2**) was dissolved in deuterated methanol (CD₃OD) and the $Bi(NO_3)_3$ was dissolved in deuterated water (D₂O). The ¹H-NMR data of **P2** with and without excess concentration of Bi^{3+} were investigated.

2.6.4 Job's plot จุฬาลงกรณ์มหาวิทยาลัย

The concentration of the fluorophore (**P2**) and Bi^{3+} were prepared at 10 μ M. The mole fraction (X) of **P2** from 0.1 to 1.0 was used in this experiment. The stoichiometry of metal-sensor complex was obtained by plotting of the fluorescence intensity of sensor's mole fraction, ((I-I₀)X_{sensor}), with the mole fraction of Bi³⁺ (X_{metal}).

2.7 Quantitative analysis for Bi³⁺

2.7.1 Effect of water content

The stock solutions of P2 are diluted into 10 μ M by varied water content from 10% to 90% water in CH₃CN. The fluorescence intensities of fluorophore ware recorded using an excitation wavelength at room temperature.

2.7.2 Effect of pH

The pH effect on fluorophores with and without analyte was investigated by data of fluorescence emission spectra in the series of buffer between pH 3.0 – 9.0, using acetate buffer for pH 3.0 and 5.0, HEPES buffer for pH 7.0, glycine-NaOH buffer for pH 9.0. The final concentrations of fluorophore, analyte and buffer solution were 10 uM, 100 uM and 20 uM, respectively. Moreover, the pH effect was studied using 0.1 M HCl and 0.1 M NaOH to adjust the solution to pH 3.0, 5.0, 7.0, 9.0 and 11.0. The final concentrations of fluorophore and analyte were 10 uM, and 100 uM, respectively.

2.7.3 Bi³⁺ in real water samples

To detection Bi³⁺ in water real samples, the water samples from tab and drinking water were studied by spiking the difference of Bi³⁺ to each water sample. The recovery of each sample was calculated from the calibration curve.



CHAPTER III RESULTS AND DISCUSSION

3.1 Synthesis and characterization of fluorophores (P1-P5)

The synthesis of pyreno[4,5-*d*]imidazole derivatives were started from the oxidation of pyrene using NalO₄ and RuCl₃ as a catalyst to give dione **1** as an orange solid in 47% yield. The spectrum of dione **1** was corresponding with a literature that was reported by Hu et al. in 2015 [51]. Then amination-condensation of dione **1** with aromatic aldehydes and ammonium acetated was performed at reflux temperature in acetic acid for 4-12 h depending on the types of the aromatic aldehydes. The target compound **P1**, **P2**, and **P3** were produced in moderate yield (55-86%) after purification by column chromatography. On the other hand, target compound **P4** was obtained in lower yield (15%) since the product was partially soluble in water even if it was worked-up with ice-water. The yellow aqueous solution was obtained after filtration. For the part of **P5**, many spots of product were shown in TLC so that the compound **P5** was obtained in low yield (15%) and not clean even after recrystallization by chloroform-ethanol. All the target compounds were characterized using ¹H NMR, ¹³C NMR and mass spectrometry.



Scheme 3.1 Synthesis of pyreno[4,5-d]imidazole derivatives (P1-P5)

For the NMR characterization, the ¹H NMR spectrum of dione **1** is shown in Figure 3.1 with 4 signals corresponding to all four aromatic protons of the symmetric molecule. The signal for aromatic proton d appears as a singlet peak at 7.84 ppm., the proton b appears as a triplet peak at 7.74 ppm., and proton a and c appear as doublet peaks at 8.47 and 8.16 ppm., respectively.



¹H NMR spectroscopy of the target compounds **P1**, **P2** and **P3** which obtained from condensation of dione **1** with aromatic aldehydes are shown in Figure 3.2. The presence of the imidazole ring was confirmed by the amine proton (NH) which appears at different positions (13.87, 13.21 and 13.07 ppm for **P1**, **P2** and **P3**, respectively). Moreover, the proton signal at 13.99 ppm confirmed the presence of the hydroxyl group (OH) in **P2**. The singlet proton signal at 4.08 ppm of **P3** confirmed the presence of the methoxy group (OCH₃).



Figure 3.2 ¹H NMR spectra of P1, P2 and P3 in DMSO-d₆

The ¹H-¹H correlation spectroscopy (COSY) was used to identify the coupled proton on phenolic ring as shown in Figure 3.3. The proton k (H_k) has been assigned at 8.40 ppm because the peak of proton at 8.40 ppm is triplet and coupled protons at 8.71 ppm (1H) and 9.49 ppm (1H). While the peak of proton at 8.7.1 ppm is triplet and coupled the proton at 8.43 ppm that indicated this peak is the proton l (H_l). Therefore, the proton m and j have been assigned at 8.43 and 9.49 ppm, respectively.



Figure 3.3 The ¹H-¹H COSY spectrum of P2 in DMSO-d₆.

Furthermore, FT-IR spectrometer could be confirmed absorption peak of hydroxyl group (-OH) on P2 at 3269.68 cm⁻¹ that is shown in Figure 3.4. The molecular weights of P1, P2 and P3 were confirmed by mass spectrometer (m/z = 318.027, 335.02 and 348.716) as seen in appendix (Figure A.4, A.8 and A.11).



Figure 3.4 FT-IR spectra of P1 and P2

Comparison of ¹H NMR of compound **P4** with compound **P2** in CD₃OD, proton of the phenyl ring that near the methoxy group (H₁) is shifted to upfield from 7.35 to 7.08 ppm and multiplicity changes from triplet to doublet. Moreover, the presence of the methoxy group was confirmed at 3.95 ppm (Figure 3.5). The molecular weight of **P4** was confirmed by mass spectrometer (m/z = 364.027) as seen in appendix (Figure A.14).



The ¹H NMR compound **P5** as shown in Figure 3.6 was corresponding with a literature that was reported in 2015 by Mardanya et al. [52].



Figure 3.6 Comparison of ¹H NMR spectra of each fractions of **P5** and picolinaldehyde in CDCl₃.

After that, mass spectroscopy was confirmed the product P5, the result was obtained at m/z = 319 in Figure 3.7.



3.2 Photophysical properties

The absorption and emission of fluorophores (P1-P5) were studied in EtOH. The normalized UV-Vis and fluorescence spectra are shown in Figure 3.8. The photophysical properties are summarized in Table 1. All of fluorophores exhibited two maximum absorption peaks which correspond to the characteristic peaks of pyrene derivatives[52]. One peak displayed around 278-290 nm with molar extinction coefficients around 9900-28500 M⁻¹ cm⁻¹ and the other showed around 331-335 nm with molar extinction coefficients 8500-21900 M⁻¹ cm⁻¹. The longer maximum absorption wavelengths (331-335 nm) are selected for excitation of each fluorophores to study the emission property. The maximum emission wavelengths for compound P1-P5 are observed at 407, 448, 389, 388 and 458 nm, respectively. The longest maximum emission in P2 and P5 may results from the Excited State Intramolecular Proton Transfer (ESIPT) which was initiated by the intramolecular hydrogen bonding between N atom on imidazole core and hydroxyl group on phenyl ring. Moreover, the ESIPT process has an effected on the fluorescent quantum yields $(\Phi_{\rm F})$ of P2 that is lower than two compounds. The relative quantum yields are found at 0.75, 0.41 and 0.78 for P1, P2 and P3, respectively.



Figure 3.8 Normalized absorption and emission spectra of P1-P3 in EtOH.

	Absorption ^a		Emission ^a		
compound	$\lambda_{\scriptscriptstyle max}$ (nm)	€ (M ⁻¹ cm ⁻¹)	λ_{ex} (nm)	$\lambda_{ ext{max}}$ (nm)	$\Phi^{\scriptscriptstyle ho}$
P1	290 331	9900 8500	331	407	0.75
P2	289 335	13000 11300	335	448	0.41
P3	290 333	11300 10400	333	389	0.78
P4	287 334	28500 21900	334	388	-
P5	278 335	27900 21100	335	458	0.24[52]

Table 3.1 Photophysical properties of molecules

^asolutions in absolute ethanol

^bAnthracene in ethanol (quantum yield 0.27) was used as the reference compound.

3.3 Metal ions sensing properties

3.1.1 Solvent effect

The absorption spectra of **P1-P5** in various solvents such as CH_3CN , THF, EtOH, DMSO and milli-q water are shown in Figure 3.9. All of target molecule are insoluble in water because of the hydrophobic structure of compounds and could be slightly soluble in DMSO. Therefore, these compounds are dissolved in CH_3CN , THF and EtOH to study fluorescent properties.



Figure 3.9 Absorption spectra of P1-P5 in various solvents.

The fluorescence spectra of **P1-P5** in CH₃CN, THF, and EtOH are shown in Figure 3.10. The same florescence spectra of **P1** and **P3** in each solvent could be indicated the types of solvent; polar solvent, non-polar solvent, and protic solvent, had no effect on **P1** and **P3**. The fluorescence intensities of **P4** are decreased in polar solvent. It may result from the stabilization of polar solvent. On the contrary, the fluorescence spectra of **P2** are different in each solvent and the intensity around 450 – 500 nm were expressed that might be the effect of hydroxyl group in

compound. In addition, the fluorescence intensity of P2 in CH_3CN is lower than THF and EtOH that might be the ground-state stabilization effect of the polar compound (P2) by the high polar solvent. While the fluorescence spectra of P5 show the different maximum emission wavelength around 400-470 nm in each solvent. As the results, CH_3CN is interesting to use as solvent for the selectivity screening as the turn-on fluorescence sensor.



Figure 3.10 Fluorescence spectra of P1-P5 in CH₃CN, THF and EtOH

3.3.2 Preliminary screening on metal ion selectivity

The selectivity of fluorophore **P1-P5** with various metal ions such as Li⁺, Na⁺, K⁺, Ag⁺, Mg²⁺, Ca²⁺, Ba²⁺, Co²⁺, Cd²⁺, Zn²⁺, Pb²⁺, Ni²⁺, Cu²⁺, Hg²⁺, Fe²⁺, Fe³⁺, Al³⁺, Bi³⁺, and Cr³⁺ were determined. After addition of 10 equiv. of metal ions to the solution of **P1-P3** in CH₃CN, the results show no selectivity of all sensor but the maximum emission wavelengths of some metal ions such as Fe³⁺, Al³⁺, Bi³⁺, and Cr³⁺ were red-shifted to around 450-460 nm. as shown in Figure 3.11. The result of **P1** is like **P3**, the fluorescence spectra show wavelengths shift from 380 -410 nm. to 460 nm. while fluorescence spectra of **P2** show fluorescence enhancement in the presence of trivalent metal ions. The result of **P4** is similar to **P2** but it was slightly enhanced by Zn^{2+} , Pb²⁺ and Ni²⁺. The different fluorescence mode of compounds may result from the difference of the complexation between fluorophores and metal ions. However, the fluorescence spectra of **P5** that contains hetero aromatic ring show fluorescence quenching by Cu²⁺, Ni²⁺ and Fe³⁺ due to an intra-ligand charge transfer transition (ILCT) from the pyrenyl-imidazole group to the pyridine group [52].





Figure 3.11 Fluorescence spectra and fluorescence enhancement ratio of P1-P5 (10 μ M) in CH₃CN upon addition of 10 equiv. of various metal ions

Interestingly, the fluorescence enhancement ratio (I/I₀) of **P2** is around 30-40 folds for four metal ions that is higher than 3-4 folds of **P4** and 8-10 folds of **P1** and **P3** (Figure 3.12). From this result, **P2** is focused on as the sensor to detection of metal ions.



Figure 3.12 Comparison of fluorescence enhancement ratio of P1-P5 (10 μ M) in CH₃CN upon addition of 10 equiv. of various metal ions

To improve in the selectivity of **P2** under the identical condition, anion effect was studied such as F^- , Cl^-Br^- , Γ , SCN^- , NO_3^- , CH_3COO^- , $S_2O_3^{2-}$, SO_3^{2-} , SO_4^{2-} , CO_3^{2-} , and PO_4^{3-} . Upon addition of 10 equiv. of anions, the fluorescence intensity was reduced by phosphate anion (PO_4^{3-}), is shown in Figure 3.13. It might be strong basicity of phosphate ion inhibited ESIPT process of **P2**.



Figure 3.13 Fluorescence spectra of P2 (10 μ M) in CH₃CN upon addition of 10 equiv. of various anions.

Besides, the titration of phosphate anions seem that more than 4 equiv. of phosphate anion did not affect to the fluorescence intensity of **P2** (Figure 3.14). After that, the new condition was observed using 10 equiv. of PO_4^{3-} in CH₃CN for screening of selectivity.



Figure 3.14 Fluorescence spectra of P2 (10 μ M) upon addition 1 to 5 equiv. of PO₄³⁻ in CH₃CN.

When the selectivity of **P2** towards 10 equiv. of various metal ions were determined, only Fe^{3+} could enhanced fluorescent intensity (Figure 3.15) but the enhancement fluorescence ratio is decreased from 25 folds to 5 folds. The selectivity of **P2** with Fe^{3+} was attributed to the solubility of phosphate salts that are shown in Table 3.2.



Figure 3.15 Fluorescence spectra of **P2** (10 μ M) in presence of PO₄³⁻ (10 equiv.) upon addition 10 equiv. of various metal ions in CH₃CN.

Compound	Formula	K _{sp} at 25 °C
Chromium phosphate	CrPO ₄	2.4×10^{-23}
Aluminium phosphate	AlPO ₄	9.84×10^{-21}
Iron(III) phosphate	FePO ₄ .	9.91 × 10 ⁻¹⁶
Bismuth phosphate	BiPO ₄	1.3×10^{-23}

Table 3.2 Solubility product constants (K_{sp}) [53]

The interaction between **P2** and Fe^{3+} was studied by UV-Vis titration spectra. After addition of Fe^{3+} from 0 to 18 equiv., the absorption band was slightly blue shift from 289 nm to 276 nm (Figure 3.16). The new absorption band might be ascribed to the complexation of **P2** and Fe^{3+} .



Figure 3.16 UV-Vis titration spectra of **P2** (10 μ M) in the presence of 10 equiv. of PO₄³⁻ upon addition of various concentrations of Fe³⁺ in CH₃CN.

However, the quantitative study of Fe³⁺ using fluorescence titration experiment showed the limitation of sensing as shown in Figure 3.17. The fluorescence signal of **P2** could not observed when the concentration of Fe³⁺ was less than 6 μ M because of the competitive binding between **P2**, Fe³⁺ and PO₄³⁻ at low concentration.



Figure 3.17 Fluorescence titration spectra of P2 (10 μ M) in the presence of 10 equiv. of PO₄³⁻ upon addition of various concentrations of Fe³⁺ in CH₃CN.

According to Zhange et al [54], the mixed organic solvent, EtOH-CH₃CN (1:4, v/v), was used for detection Bi³⁺. In order to improve selectivity of **P2**, another organic solvent was mixed with CH₃CN to alter the solubility of sensor. From the previous screening (Figure 3.7), the emission wavelength of **P2** with trivalent metal ions is approximate around 450 nm so that another organic solvent should not displayed the fluorescence intensity at 450 nm. The fluorescence spectra in Figure 3.18 showed that the mixed solvent between CH₃CN and DMSO could not increase the fluorescence intensity at 450 nm while the fluorescence intensities of the mixture of CH₃CN-EtOH appeared around 450 nm.



Figure 3.18 Fluorescence spectra of P2 (10 μ M) (a) in various ratio of CH₃CN-EtOH (b). in various ratio of CH₃CN-DMSO.

Then, addition 10 equiv. of four trivalent metal ions (Fe³⁺, Al³⁺, Bi³⁺ and Cr³⁺) to **P2** in the mixed solvents, the results showed the enhancement ratio at 80% CH₃CN-

DMSO in the presence of Bi^{3+} is higher than the others. Therefore, 80% CH₃CN-DMSO was optimized as the condition for study the selectivity of **P2** (Figure 3.19).



Figure 3.19 Fluorescence spectra and of fluorescence enhancement ratio of **P2** (10 μ M) in CH₃CN-DMSO 1:1 and 8:2, v/v upon addition of 10 equiv. of trivalent metal ions (Fe³⁺, Al³⁺, Bi³⁺ and Cr³⁺)

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The selectivity of **P2** toward various metal ions under the optimized condition were also studied. The fluorescence intensity at 450 nm was significant increased by addition 10 equiv. of Bi³⁺. The fluorescence enhancement ratio is shown about 7 folds although Fe³⁺ was slightly increased about 2 folds. On the other hand, the selectivity of **P1** and **P3** under this solvent system could not be improved that was shown in Figure 3.20.



Figure 3.20 Fluorescence spectra and fluorescence enchantment ratio of **P1-P3** (10 μ M) in CH₃CN-DMSO (8:2, v/v) before and after addition 10 equiv. of metal ions.

3.3.3 Interference studies

The possible interference of **P2** to Bi^{3+} was carried out in the presence of 10 equiv. of other metal ions under CH₃CN-DMSO (8:2, v/v). From Figure 3.21, it is observed that most metal ions cannot interfered to the detection of Bi^{3+} . However, only Fe³⁺ can be interfered for Bi^{3+} sensing that had small effect to increase the

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fluorescence signal. The increasing fluorescence intensity may come from the competition between coordination of P2 with Bi^{3+} and Fe^{3+} .



Figure 3.21 Interference of other metal ions (10 eq) on P2 (10 μ M) in the presence of Bi³⁺ (10 eq).

3.3.4 Fluorescence titration and detection limit

The qantitative analysis of **P2** with Bi^{3+} was examined by fluorescence titration experiment as shown in Figure 3.22. The emission intensity at 450 nm was gradually increased with Bi^{3+} concentration from 0 to 500 μ M.





Figure 3.22 Fluorescence spectra of P2 (10 μ M) in the different concentration of Bi³⁺. and relationship between concentration of Bi³⁺ and fluorescence enhancement ratio of complex.

A linear relationship between the fluorescence enhancement ratio and the concentration of Bi^{3+} was exhibited in the range from 0 to 20 μ M with the correlation coefficient of 0.9924 (Figure 3.23). The detection limit of **P2** with Bi^{3+} was calculated to be 1.20 μ M in CH₃CN-DMSO (8:2, v/v).at three-time noises.



Figure 3.23 Linear Plot between concentration of Bi^{3+} and fluorescence enhancement ratio of complex with P2 (10 μ M).

3.4 Sensing mechanism studies

3.4.1 Time dependence

The binding phenomenon of **P2** in presence of Bi^{3+} was determined to prove the complexation of **P2** and Bi^{3+} is stable by time effect studied at the period of 0-30 min. The result was showed in Figure 3.24. The fluorescence intensity ratio of **P2** upon addition of 10 equiv. of Bi^{3+} remained constantly ratio. It could be indicated that the process of complexation between **P2** and Bi^{3+} is stable.



Figure 3.24 Fluorescence intensity of P2 (10 μ M) upon addition 10 equiv. of Bi³⁺ in CH₃CN-DMSO (8:2, v/v) for 30 min.

3.4.2 UV-visible studies and decomplexation

The binding property of **P2** and Bi^{3+} was investigated by UV-Vis titration in CH3CN-DMSO (8:2, v/v) at room temperature. Upon addition of Bi^{3+} from 0.1 equiv. to 50 equiv., the absorption band was observed at 278 nm (Figure 3.25). The new band might be attributed to the complexation between **P2** and Bi^{3+} .



Figure 3.25 UV-Vis titration of P2 (10 μ M) upon addition of various concentrations of Bi³⁺.

Moreover, to confirm the complexation, sodium sulfide (Na₂S) was added to the solution of **P2** in the presence of Bi³⁺ in order to decomplexation. However, it is unsuccessful. After addition of Na₂S, although the UV-Vis spectrum shows the absorption band at 278 nm was disappeared, but the upper baseline shift was observed (Figure 3.22, left). It might be result from the formation of insoluble Bi₂S₃. Nonetheless, the fluorescence enhancement intensity at 450 nm was caused by the complexation of **P2** and Bi³⁺ which was reduced as same as the emission of **P2** without Bi³⁺ upon addition of Na₂S (Figure 3.22, right). The Na₂S could lead to the releasing Bi³⁺ and it was one of information for study the mechanism sensing.



Figure 3.26 UV-Vis and Fluorescence spectra of the restoration P2 in the presence of Bi^{3+} by Na_2S .

3.4.3 ¹H NMR experiment

The mechanistic experiment was also performed by ¹H NMR on **P2** in deuterated methanol (CD₃OD) before and after addition of Bi³⁺ that dissolved in deuterated water (D₂O) as exhibited in Figure 3.23. In the spectrum of **P2**, the peaks at 7.04, 7.08, 7.36 and 8.17 ppm were assigned to the proton signals of phenolic ring (H_k, H_m, H_l and H_j, respectively) and the others were the proton signals of pyrene. The downfield shift of most of the proton signals were observed except H_j which shifted slightly upfield. This result could be not only proved a coordination complex between **P2** and Bi³⁺ but also led to the deprotonation of phenolic photon. Consequently, the increasing fluorescence intensity might imply the inhibition of the excited-state intramolecular proton transfer (ESIPT) process.



Figure 3.27 ¹H NMR spectra of P2 in CD₃OD in the absence and the presence of Bi³⁺.

3.4.4 Job's plot

As can be seen in Figure 3.28, the Job's plot analysis was examined on the stoichiometry of the complex between **P2** and Bi³⁺. A plot of mole fraction of Bi³⁺ with fluorescence intensity of mole fraction of **P2** showed that the maximum point was obtained at 0.5 of mole fraction of Bi³⁺. The Job's plot confirmed the formation of 1:1 complex of **P2** with Bi³⁺.



Figure 3.28 Job's plot for determination of binding stoichiometry between P2 and Bi^{3+} .

3.5 Quantitative analysis for Bi³⁺

The application of this sensor was further studied for detection of Bi³⁺ in water samples so that the optimization of condition was determined.

3.5.1 Effect of water content

Due to the insolubility of **P2** in water, the content of water was controlled to avoid the precipitate. The UV-Vis spectra of 70-90% of water in CH₃CN disappeared original absorption bands and were broad peaks while 10 -60% of water were not remarkable changes (Figure 3.29). Although emission fluorescence intensity at 440-450 nm is not significant changes, however, 10% of water may be a mild condition to apply.



Figure 3.29 UV-Vis spectra and Fluorescence spectra of **P2** in the presence of water fraction in CH₃CN.

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3.5.2 Effect of pH

Then, the pH dependency of P2 was investigated by fluorescence emission spectra with and without Bi^{3+} . The first study, when 20 µM of pH buffers were used to control pH in 10 % water in the mixed organic solution (CH₃CN-DMSO; 8:2, v/v) of P2 as shown in Figure 3.30, fluorescent intensities were increased in the range between pH 3 and 5 for P2 in the presence of Bi^{3+} but no fluorescent intensity changes in the range between pH 7 and 9. It might be the binding between P2 and Bi^{3+} was disturbed by the solvation of buffer ions that were also observed by error value.



Figure 3.30 The variation of fluorescent intensity of **P2** (10 μ M) in the absence and presence of 10 equiv. of Bi³⁺ with difference pH levels (3-9) and fluorescence enhancement ratio in 10% pH-buffers (20 μ M) in mixed organic solvent CH₃CN-DMSO (8:2, v/v).

To prove this problem, the pH levels were adjusted by 1 mM HCl as acid and 1 mM NaOH as acid. In Figure 3.31, the enhancement fluorescent intensity of **P2** with Bi^{3+} was observed. However, the fluorescent intensity of **P2** in the presence and absence of Bi^{3+} at pH 3 is higher than the others due to the protonation of imidazole nitrogen atom while the hydroxide ion (OH) at the pH = 11 might compete for Bi^{3+} with **P2** that decreased fluorescent intensity. As the result, pH of real samples was adjusted by HCl and NaOH to 5 for quantitative application.



Figure 3.31 The variation of fluorescent intensity of **P2** (10 μ M) in the absence and presence of 10 equiv. of Bi³⁺ and fluorescence enhancement ratio in 10% aqueous solution at various pH (HCl-NaOH) in mixed organic solvent CH₃CN-DMSO (8:2, v/v)

3.5.3 Bi³⁺ in real water samples

To demonstrate using **P2** as sensor for detection of Bi^{3+} in real water samples. A calibration curve was investigated in 10% (v/v) of pH5 aqueous solution in mixed organic solvents, CH₃CN-DMSO (8:2, v/v) by fluorescence titration as shown in Figure 3.32. The detection limit in this system was calculated to be 3.40 μ M with three times noises.



Figure 3.32 Linear Plot between the concentration of Bi^{3+} and fluorescence enhancement ratio of **P2** in 10% (v/v) of pH 5 aqueous solution in mixed organic solvents (CH₃CN-DMSO (8:2, v/v)).

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The analysis of real water samples from tab water and drinking water were adjusted to pH at 5 using HCl-NaOH. And then were spiked the known amount of Bi^{3+} at 10 and 15 μ M. The results were illustrated in Table 3.3, showed that the recovery of Bi^{3+} in the spiked water samples at 91-97% was almost completed. The incomplete recover of Bi^{3+} might be caused by the anions such as phosphate, sulfate carbonate or sulfide in these water samples that formed or interacted to bismuth compounds so that it might decrease the Bi^{3+} ions.

Samples	Spike (µM)	found (µM)	Recovery
Tah water	10.00	9.72 ± 0.096	97.20% ± 0.96
Tab Water	15.00	13.77 ± 0.09	91.80% ± 0.60
Drinking water	10.00	9.61 ± 0.08	96.10% ± 0.80
	15.00	14.10 ± 0.01	94.00% ± 0.07

Table 3.3 Quantitative analysis for Bi(III) in real water samples (n=3)

In Table 3.4, the outcome of sensor (P2) is compared to others Bi^{3+} fluorescent sensors in terms of structure and detection limited. Even if the detection limited is not the lowest but it could be acceptable.



Sensors	Solvent(s)	Sensing mode	LOD µM	Ref.
Tiron HO S-ONA HO S-ONA	DI water	On-off	0.24	Taher et al., 2014 [55]
N-hydroxy 1,8- Naphthalimide	β-cyclodextrin, aqueous solution	Off-on	2.78	Kavitha and Slatin, 201 7 [56]
Rhodamine derivative on Zr-based MOFs	pH = 6, aqueous solution	Off-on	0.008	El-Sewify et al., 2018 [57]
Rhodamine derivative $\downarrow_{H} \rightarrow \downarrow_{H} $	pH 5.0–7.0, Ethanol- CH ₃ CN (1:4, v/v)	Off-on	2.69	Zhang et al., 2018 [54]
Rhodamine B derivative	CH ₃ CN-H ₂ O (99:1, v/v)	Off-on	0.0086	Guang et al., 2018 [58]
Pyren-1-ylmethylene- hydrazide	HEPES buffer pH 7.4, DMSO-H ₂ O (1:1, v/v)	Off-on	0.12	Saravanan et al. 2019 [59]
Pyreno[4,5-d]imidazole-	CH ₃ CN-DMSO (8:2, v/v)	Off-on	1.20	This work
HONG SETISON	pH =5, H ₂ O-mixed of CH ₃ CN/DMSO (1:9, v/v)	Off-on	3.40	This work

Table 3.4 Comparison of the Bi(III) fluorescent sensors

CHAPTER IV

In summary, five derivatives of pyreno[4,5-*d*]imidazole (**P1-P5**) were successfully synthesized from pyrene-4,5-dione and aromatic aldehydes in moderated to good yields. All of compounds except compound **P5** could be exhibit a fluorescent enhancement at 450 nm with trivalent cations such as Bi^{3+} , Fe^{3+} , Cr^{3+} and Al^{3+} in CH₃CN. However, the compound **P2** that consist of phenolic group could be used as sensor for selectively turn-on detection of Bi^{3+} in CH₃CN-DMSO (8:2 v/v). The detection limit of sensor (**P2**) was calculated at 1.20 µM. The sensing mechanism was investigated by UV-Vis titration, ¹H-NMR, Job's plot and using Na₂S to the decomplexation study, which suggests a 1 :1 binding to form complex between the sensor (**P2**) and Bi^{3+} . In the system that contain 10% aqueous in mixed organic solvents (CH₃CN-DMSO, (8:2 v/v)), the highest ratio of fluorescent enhancement was obtained by addition of Bi^{3+} at pH 5. The detection limit was estimated at 3.40 µM. For application, the quantitative detection of Bi^{3+} with **P2** in two real water sample could be recovered in acceptable percentage.

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APPENDIX



Figure A.1 ¹H NMR spectrum of dione in DMSO-d₆



Figure A.2 ¹H NMR spectrum of P1 in DMSO-d₆



Figure A.3 ¹³C NMR spectrum of P1 in DMSO-d₆



Figure A.4 MALDI-TOF-Mass spectrum of P1



Figure A.6 ¹H-¹H COSY NMR spectrum of P2 in DMSO-d₆



Figure A.8 ESI Mass spectrum of P2



Figure A.10 $^{\rm 13}{\rm C}$ NMR spectrum of P3 in DMSO-d_6



Figure A.12 ¹H NMR spectrum of P4 in CDCl₃



Figure A.13 ¹³C NMR spectrum of P4 in DMSO-d₆



Figure A.14 MALDI-TOF-Mass spectrum of P4



Figure A.16 $^{\rm 13}{\rm C}$ NMR spectrum of P5 in DMSO-d_



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