

REFERENCES

- Abe, S., Nagamine, Y., Omichi, K., and Ikenaka, T. 1991. Investigation of the active site of *Bacillus macerans* cyclodextrin glucanotransferase by use of modified maltooligosaccharides. *J. Biochem.* **110**: 756-761.
- Albeyan, V. A., Yamamoto, T., and Afrikyan, E. G. 1994. Isolation and characterization of cyclodextrin glucanotransferase using cyclodextrin polymers and their derivatives. *Biochemistry. (Moscow)* **59**: 573-579.
- Amaizo. 1993. *Cyclodextrin*. USA (Mimeographed).
- Bender, H. 1977a. Cyclodextrin-glucanotransferase von *Klebsiella pneumoniae* 1. Synthese, reingugung und eigenschchaften des enzymes von *K. pneumoniae* M5 a1. *Arch. Microbiol.* **111**: 271.
- Bender, H. 1977b. Cyclodextrin-glucanotransferase von *Klebsiella pneumoniae* 2. Bedeutung des enzyme fur den metabolismus der cyclodextrin bei *K. pneumoniae* M5 a1. *Arch. Microbiol.* **113**: 49.
- Bender, H. 1986. Production, characterization and application of cyclodextrins. *Adv. Biotech. Proc.* **6**: 31-71.
- Bender, H. 1988. Studies on the reaction mechanism of cyclodextrin glycosyltransferase: subsite analysis. In O. Huber and J. Szejtli (eds.), *Proceedings of the Fourth International Symposium on Cyclodextrins*, Munich, Kluwer, Academic Publisher: 19-26.
- Bender, H. 1991. On the role of histidine residues in cyclodextrin glycosyltransferase: chemical modification with diethylpyrocarbonate. *Carbohydr. Res.* **209**: 145-153. Bollage, D.M. and Eldstein, S.J. 1991. *Protein methods*. New York: John Wiley & Sons. Inc.

- Boonchai, J. 1995. *Determination of cyclodextrin glycosyltransferase gene from Bacillus sp. A11*, Master's Thesis, Faculty of science, Chulalongkorn University.
- Bovetto, L. J., Villette, J. R., Fontaine, I. F. Sicard, P. J., and Bouquet, S. J-L. 1992. Cyclomaltodextrin glucanotransferase from *Bacillus circulans* E192: II. Action pattern. *Biotech. Appl. Biochem.* 15: 59-68.
- Bradford, M. M. 1976. A rapid and sensitive method for the qualitatively of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72: 248-254.
- Casu, B. and Reggiani M. 1979. Methylated cycloamyloses and their inclusion properties. *Carbohydr. Res.* 76: 59-68.
- Depinto, J. A. and Campbell, L. L. 1964. Formation and degradation of cyclic dextrans by intracellular enzymes of *Bacillus macerans*. *Science.* 14-20.
- Depinto, J. A. and Campbell, L. L. 1968. Purification and properties of cyclodextrin glycosyltransferase from an alkalophilic bacteria forming primarily cyclodextrin. In D. Duchhene (ed.), *Proceedings of the Fifth International Symposium on Cyclodextrins*, Paris, Edition de Sante.: 25-31.
- Ellman, G. L. 1959. Tissue sulfhydryl groups. *Arch. Biochem. Biophys.* 82: 70-77.
- Englbrecht, A., Harrer, G., Lebert, M., and Schmid, G. 1990. Biochemical and genetic characterization of cyclodextrin glycosyltransferase from an alkalophilic bacteria forming primarily cyclodextrin. In D. Duchene (ed.), *Proceedings of the Fifth International Symposium on Cyclodextrins*, Paris, Edition de Sante.: 25-31.
- Ensuiko 1994. *Stabilization of natural colors by cyclodextrin*. Japan (Mimeographed).
- Fields, R. 1972. The rapid determination of amino groups with TNBS. *Methods Enzymol.* 25B: 464-468.
- French, D. and Rundle, R. E. 1942. The molecular weights of the schardinger alpha and beta dextrin. *J. Am. Chem. Soc.* 64: 165-172.

- Fujiwara, S., Kakihara, H., Woo, K. B., Lejeune, A., Kanemoto, M., Sakaguchi, K., and Imanaka, T. 1992. Cyclization characteristics of cyclodextrin glucanotransferase are conferred by the NH₂-terminal region of the enzyme. *J. Bacteriol.* 174: 7478-7481.
- Fuwa, H. 1954. A new method for microdetermination of amylase activity by the use of amylase as the substrate. *J. Biochem.* 41: 583-603.
- Hellman, J., Wahlberg, M., Karp, M., Kopela, T., and Mantsala, P. 1990. Effect of modifications at the C-terminus of cyclomaltodextrin glucanotransferase from *Bacillus circulans* var. *alkalophilus* on catalytic activity. *Biotech. Appl. Biochem.* 12: 387-396.
- Hoare, D. E. and Koshland, D. E. 1967. *J. Biol. Chem.* 242: 2447.
- Hofmann, B. E., Bender, H., and Schulz, G. E. 1989. Three-dimensional structure of cyclodextrin glycosyltransferase from *Bacillus circulans* at 3.4 Å^o resolution. *J. Mol. Biol.* 209: 793-800.
- Horikoshi, K. 1979. Production and industrial applications of beta-cyclodextrin. *Proc. Biochem.* 14: 26-30.
- Horikoshi, K., and Akiba, T. 1982. Alkalophilic microorganisms: A new microbial world. Japan Tokyu: *Scientific Societies Press*: 105-157.
- Horikoshi, K. 1988. Enzymology and molecular genetic of CD-forming enzyme. In O. Huber and J. Szejtli (eds.), *Proceedings of the Fourth International Symposium on Cyclodextrins*, Munich, Kluwer Academic Publisher: 7-17.
- Horikoshi, K., Ando, T., Yohida, K., Tokyu, J. P., Nakamura, N., Kunitachi, N. 1982. Enzeugung von cyclodextrin. *Ger Patent DE* 24, 53, 860.
- Janssen. 1992. *Encapsin HP13 Biotech N. V. Drug Delivery Systems*. Belgium (Mimeographed).
- Jantarama, J. 1997. *Mutation of Bacillus sp. A11 for increasing cyclodextrin glycosyltransferase activity*. Master's Thesis, Graduate School, Chulalongkorn University.

- Jones, M. A., Hamilton, M. L., and Lash, T. D. 1997. Effect of covalent modification on coproporphyrinogen oxidase from chicken red blood cells. *Prep. Biochem. Biotech.* **27**(1): 47-57.
- Kaneko, T., Hamamoto, T., and Horikoshi, K. 1988. Molecular cloning and nucleotide sequence of the cyclomaltodextrin glucanotransferase gene from the Alkalophilic *Bacillus* sp. strain No. 38-2. *J. Gen. Micro. Biol.* **134**: 97-105.
- Kim, P. 1996. *Purification of cyclodextrin glycosyltransferase by immunoaffinity chromatography*, Master's Thesis, Graduate School, Chulalongkorn university.
- Kimura, K., Shinsuke, K., Yasumasa, I., Toshiya, T., and Yamane, K. 1987. Nucleotide sequence of the β -cyclodextrin glucanotransferase gene of alkalophilic *Bacillus* sp. Strain 1011 and similarity of its amino acid sequence of these of α -amylase. *J. Bacteriol.* **169**: 4399-4402.
- Kitahata, S., Tsuyama, N., and Okada, S. 1974. Purification and some properties of the cyclodextrin glycosyltransferase from a strain of *Bacillus* sp. *Agric. Biol. Chem.* **38**: 387-393.
- Kitahata, S. and Okada, S. 1975. Transfer action of cyclodextrin glycosyltransferase on starch. *Agric. Biol. Chem.* **28**: 2413-2417.
- Klein, C. and Schulz, G. E. 1991. Structure of cyclodextrin glycosyltransferase refined at 2.0 Å resolution. *J. Mol. Biol.* **217**: 737-750.
- Klein, C., Hollender, J., Bender, H., and Schulz, G. E. 1992. Catalytic center of cyclodextrin glycosyltransferase derived from X-ray structure analysis combined with site-directed mutagenesis. *Biochemistry.* **31**: 8740-8746.
- Kobayashi, S., Kainuma, K., and Suzuki, S. 1978. Purification and some properties of *Bacillus macerans* cycloamylose (cyclodextrin) glucanotransferase. *Carbohydr. Res.* **61**: 229-238.

- Kuttiarcheewa, W. 1994. *Immobilization of cyclodextrin glycosyltransferase on inorganic carriers*. Master's Thesis, Graduate School, Chulalongkorn University.
- Laloknam, S. 1997. *Detection of cyclodextrin glycosyltransferase gene by synthetic oligonucleotide probes*. Master's Thesis, Graduate School, Chulalongkorn University.
- Lawson, C. L., van Montford, R., Strokopytov, B., Rozeboom, H. J., Kalk, K. H., de Vries, G., Penninga, D., Dijkhuizen, L., and Dijkstra, B. W. 1994. Nucleotide sequence and X-ray structure of cyclodextrin glycosyltransferase from *Bacillus circulans* strain 251 in a maltose-dependent crystal form. *J. Mol. Biol.* **236**: 590-600.
- Lee, K. C. P. and Tao, B. Y. 1995. A kinetic study of cyclodextrin glycosyltransferase: substrate and product inhibitions. *Biotech. Appl. Biochem.* **21**: 111-121.
- Lundblad, R. L. 1991. *Chemical reagents for protein modification*. 2nd ed. CRC Press, Inc.
- Malai, T. 1994. *Cyclodextrin production from rice starch*. Master's Thesis, Graduate School, Chulalongkorn University.
- Mattsson, P., Susanna, M., and Timo, K. 1990. Analysis of cyclomaltodextrin glucanotransferase isozymes by isoelectric focusing in the immobilized pH gradients. *J. Biochem. Biophys.* **20**: 237-246.
- Mattsson, P., Battchikova, N., Sippola, K., and Korpela, T. 1995. The role of histidine residues in the catalytic activity of cyclomaltodextrin glucanotransferase from *Bacillus circulans* var. *alkalophilus*. *Biochem. Biophys. Acta.* **1247**: 97-103.
- Mattsson, P., Pohjalainen, T., and Korpela, T. 1992. Chemical modification of cyclomaltodextrin glucanotransferase from *Bacillus circulans* var. *alkalophilus*. *Biochem. Biophys. Acta.* **1122**: 33-40.

- Matzuzawa, M., Nakamura, N., and Horikoshi, K. 1975. An improve method for the preparation of schardinger beta-dextrin on an industrial scale by cyclodextrin glycosyltransferase of an alkaphilic *Bacillus* sp. ATCC 21783. *Strach/ Strake*. 27: 410-413.
- Mean, G. E. and Feeney, R. E. 1971. *Chemical modification of proteins*. Holden-Day, Inc.
- Mile, E.W. 1977. Modification of histidyl residues in proteins by diethylpyrocarbonate. *Methods Enzymol*. 47: 431-442.
- Miller, G. L. 1959. Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Anal. Chem*. 426-428.
- Mori, S., Hirose, S., Oya, T., and Kitahata, S. 1994. Purification and properties of cyclodertrin glucanotransferase from *Brevibacterium* sp. No. 9605. *Biosci. Biotech. Biochem*. 11: 1968-1972.
- Muramatsu, Y., Takahashi, K., and Nakamura, N. 1993. Characterization of a maltogenic amylase of *Thermomonospora viridis* and application in branched cyclodextrin production. *Strach/ Strake*. 3: 99-104.
- Nakamura, A., Haga, K., and Yamane, K. 1993. Three histidine residues in the active center of cyclodextrin glucanotransferase from *Bacillus* sp. 1011: Effect of the replacement on pH dependence and transition-state stabilization. *Biochemistry*. 32: 6624-6631.
- Nakamura, A., Haga, K., and Yamane, K. 1994. Four aromatic residues in the active center of cyclodextrin glucanotransferase from alkalophilic *Bacillus* sp. 1011: Effect of the replacement on substrate binding and cyclization characteristics. *Biochemistry*. 33: 9929-9936.
- Nagamoto, S., 1985. Cyclodextrins-expanding the development of their functions and application. *Chem. Econ. Eng. Rev*. 17: 28-34.

- Ohnishi, M., Abe, M., Azuma, T., Kubota, M., and Rokushika, S. 1994. Tryptophan residues of *Bacillus cycloamylose* glucanotransferase: Effect of modification with *N*-bromosuccinimide on the enzyme-catalysed synthesis of cyclomaltoheptaose from maltotriose. *Strach/ Strake*. **46**: 272-275.
- Ohnishi, M., Taniguchi, M., and Hiromi, K. 1983. Kinetic discrimination of tryptophan residues of glucoamylase from *Rhizopus niveus* by fast chemical modification with *N*-bromosuccinimide. *Biochem. Biophys. Acta*. **744**: 64-70.
- Penninga, D., Strokopytov, B., Rozeboom, J. H., Lawson, C. L., Dijkstra, B. W., Bergsma, J., and Dijkhuizen, L. 1995. Site-directed mutations in tyrosine 195 of cyclodextrin glycosyltransferase from *Bacillus circulans* strain 251 affect activity and product specificity. *Biochemistry*. **34**: 3368-3376.
- Pongsawasdi, P. and Yagisawa, M. 1987. Screening and identification of a cyclomalto-dextrin glucanotransferase producing bacteria. *J. Ferment. Technol.* **65**: 463-467.
- Pulley, O. A. and French, D. 1961. Studies on the schardinger dextrans. XI The isolation of new schardinger dextrans. *Biochem. Biophys. Res. Com.* **5**: 11-15.
- Ramachandran, L.K., and Witkop, B. 1967: *Method Enzymol.* **11**: 283
- Riodan, P. W., Wacker, W. E. C., Valke, B.L. 1965. *N*-acetylimidazole: A reagent for determination of "free" tyrosyl residues of proteins. *Biochemistry*. **4**:1758-1765.
- Rojtinnakorn, J. 1994. *Preparation of antibody against cyclodextrin glycosyltransferase from Bacillus All*. Master's Thesis, Graduate School, Chulalongkorn University.
- Rutchorn, U. 1993. *Production of cyclodextrin glucanotransferase in a fermenter and its immobilization on DEAE-cellulose*. Master's Thesis, Graduate School, Chulalongkorn University.

- Saenger, W. 1979. Circular hydrogen bonds in alpha- cyclodextrin hexahydrate
Nature. 279: 343.
- Saenger, W. 1982. Structure aspect of cyclodextrin inclusion compounds, In J. Szejtli (ed.), *Proceedings of the First International Symposium on Cyclodextrins*, Budapest, Akademiai Kiado: 141-145.
- Schardinger, F. 1903. Thermophile bakterien aus verschiedenen nahrungsmitteln und mch, und die gebildeten produkte, wenn diese bakterien innahrlosungen kultiviert werden, die kohlehydrate enthalten, *Z. Untersuch Nahr Genussm*. 6: 865-880.
- Schardinger, F. 1904. *Bacillus macerans* ein acetone bildender rottebacillus, *Zbl. Bakter. Parasiten Kunde II*. 14: 772-778.
- Schmid, G., Huber, O. S., and Eberle, H.J. 1989. Cyclodextrin glucanotransferase production yield enhancement by overproduction of cloned gene. *TIBTECH*. 17: 244-248.
- Sin, K-A., Nakamura, A., Masaki, H., Matsuura, Y., and Uozumi, T. 1994. Replacement of an amino acid residue of cyclodextrin glucanotransferase of *Bacillus obhensis* double the production of γ -cyclodextrin. *J. Biotech*. 32: 283-288.
- Strokopytov, B., Penninga, D., Rozeboom, H. J., Kalk, K. H., Dijkhuizen, I., and Dijkstra, B. W. 1995. X-ray structure of cyclodextrin glycosyltransferase complex with acarbose. Implications for the catalytic mechanism of glycosidase. *Biochemistry*. 34: 2234-2240.
- Strokopytov, B., Knegt, R. M. A., Penninga, D., Rozeboom, H. J., Kalk, K. H., Dijkhuizen, I., and Dijkstra, B. W. 1996. Structure of cyclodextrin glycosyltransferase complexed with a maltononaose inhibitor at 2.6 Å resolution. Implications for product specificity. *Biochemistry*. 35: 4241-4249.

- Subhasitanont, P. 1993. *Chemical modification of Cassava linamarase*. Master's Thesis, Graduate School, Mahidol University.
- Svensson, B., Jespersen, H. M., Sierks, M. R., and Mac Gregor, E. A. 1989. Sequence homology between putative raw-starch binding domains from different starch-degrading enzymes. *Biochem. J.* 264: 309-311.
- Szejtli, J. 1988. Chapter I: *Cyclodextrin Technology*., Netherland: Kluwer Academic Publisher.
- Szejtli, J., and Paginton, J. 1991. Cholesterol removal with CDs, *Cyclodextrin News*. May 5 (9): 1-2.
- Techaiyakul, W. 1991. *Production and characterization of cyclodextrin glucanotrans-ferase from Bacillus spp.* Master's Thesis, Graduate School, Chulalongkorn University.
- Thoma, J. A., Dygert, S., and Hsue, K. 1965. A simple and specific assay for cyclodextrin transglucosidase. *Anal. Biochem.* 13: 91-99.
- Vikmon, M. 1981. Rapid and simple spectrophotometric method for determination of micro-amounts of cyclodextrins. *Proceedings of the First International Symposium on Cyclodextrins*, Hungary: 69-74.
- Villette, J. R., Helbecque, N., Albani, J. R., Sicard, P.J., and Bouquelet, S. J-L. 1993. Cyclodextrin glucanotransferase from *Bacillus circulans* E192: Nitration with tetranitromethane. *Biotech. Appl. Biochem.* 17: 205-216.
- Villette, J. R., Sicard, P.J., and Bouquelet, S. J-L. 1992. Cyclodextrin glucanotransferase from *Bacillus circulans* E192: III. Chemical modification by diethylpyrocarbonate: Evidence for an induce fit at the active site resulting from the binding of an acceptor. *Biotech. Appl. Biochem.* 15: 69-79.

- Villette, J. R., Krzewinski, F. S., Looten, P. J., Sicard, P.J., and Bouquelet, S. J-L. 1992. Cyclodextrin glucanotransferase from *Bacillus circulans* E192: IV. Evidence for a raw starch-binding site and its interaction with a β -cyclodextrin copolymer. *Biotech. Appl. Biochem.* 16: 57-63.
- Vittayakitsirikul, V. 1995. *Expression of cyclodextrin glycosyltransferase in Bacillus subtilis M1111 (RM 125) and Escherichia coli*. Master's Thesis, Graduate School, Chulalongkorn University.
- Voet, D. and Voet, J.G. 1990. *Biochemistry*. John Wiley & Sons, Inc.
- Wakayama, M., Tsutsumi, T., Yada, H., Sakai, K., and Morigushi, M. 1996. Chemical modification of histidine residues of *N*-Acyl-D-glutamate amidohydrolase from *Pseudomonas* sp. 5f-1. *Biosci. Biotech. Biochem.* 60 (4): 650-653.
- Walsh, C. 1979. Evidence that histidine plays a catalytic role at the active site. *Enz. Reac. Mech.* 84-93.
- Wanna, S. 1995. *Restriction map and localization of the presumed cyclodextrin glycosyltransferase gene cloned from Bacillus sp. A11*. Master's Thesis, Graduate School, Chulalongkorn University.
- Wind, H.D., Liebl, W., Buitelaar, R.M., Penninga, D., Spreinat, A., Dijkhuizen, L., and Bahl, H. 1995. Cyclodextrin formation by Thermostable α -amylase of *Thermoanaerobacterium thermosulfurigenes* EM1 and reclassification of the enzyme as a cyclodextrin glycosyltransferase. *Env. Micro.* 1257-1265.
- Yamamoto, M., Aritumi, H., Ilie, T., Hirayama, F., and Uekama, K. 1990. Pharmaceutical evaluation of branched β -cyclodextrins as parental drugs carriers. In D. Duchene (ed.), *Minutes of the Fifth International Symposium on Cyclodextrins*, Paris, Edition De Sante.: 541-544.

APPENDICES

APPENDIX 1: Preparation for polyacrylamide gel electrophoresis

1) Stock reagents

30% Acrylamide, 0.8% bis-acrylamide, 100ml

acrylamide 29.2 g

N, N'-methylene-bis-acrylamide 0.8 g

Adjusted volume to 100 ml with distilled water

1.5 M Tris-HCl pH 8.8

Tris (hydroxymethyl)-aminomethane 18.17 g

Adjusted pH to 8.8 with 1M HCl and adjusted volume to 100 ml
with distilled water

2 M Tris-HCl pH 8.8

Tris (hydroxymethyl)-aminomethane 24.2 g

Adjusted pH to 8.8 with 1M HCl and adjusted volume to 100 ml
with distilled water

0.5 M Tris-HCl pH 6.8

Tris (hydroxymethyl)-aminomethane 6.06 g

Adjusted pH to 6.8 with 1 M HCl and adjusted volume to 100 ml
with distilled water

1 M Tris-HCl pH 6.8

Tris (hydroxymethyl)-aminomethane 12.1 g

Adjusted pH to 6.8 with 1M HCl and adjusted volume to 100 ml
with distilled water

Solution B (SDS PAGE)

2 M Tris-HCl pH 8.8 75 ml

10% SDS 4 ml

distilled water 21 ml

Solution C (SDS PAGE)

1 M Tris-HCl pH 6.8	50 ml
10% SDS	4 ml
distilled water	46 ml

2. Non-denaturing PAGE**7.5% Separating gel**

30% acrylamide solution	2.5 ml
1.5 M Tris-HCl pH 8.8	2.5 ml
distilled water	5.0 ml
10% $(\text{NH}_4)_2\text{S}_2\text{O}_8$	50 μl
TEMED	10 μl

5.0% stacking gel

30% acrylamide solution	0.67 ml
0.5 M Tris-HCl pH 6.8	1.0 ml
distilled water	2.3 ml
10% $(\text{NH}_4)_2\text{S}_2\text{O}_8$	30 μl
TEMED	5 μl

Sample buffer

1 M Tris-HCl pH 6.8	3.1 ml
glycerol	5.0 ml
1% bromophenol blue	0.5 ml
distilled water	1.4 ml

One part of sample buffer was added to four parts of sample.

Electrophoresis buffer, 1 litre

(25 mM Tris, 192 mM glycine)

Tris (hydroxymethyl)-aminomethane	3.0 g
Glycine	14.4 g

Dissolved in distilled water to 1 litre (final pH should be 8.8).

3. SDS-PAGE

7.5% separating gel

30% acrylamide solution	2.5 ml
solution B	2.5 ml
distilled water	5.0 ml
10% $(\text{NH}_4)_2\text{S}_2\text{O}_8$	50 μl
TEMED	10 μl

5.0% stacking gel

30% acrylamide solution	0.67 ml
solution C	1.0 ml
distilled water	2.3 ml
10% $(\text{NH}_4)_2\text{S}_2\text{O}_8$	30 μl
TEMED	5 μl

Sample buffer

1 M Tris-HCl pH 6.8	0.6 ml
50% glycerol	5.0 ml
10% SDS	2.0 ml
2-mercaptoethanol	0.5 ml
1% bromophenol blue	1.0 ml
distilled water	0.9 ml


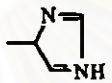
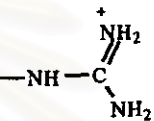
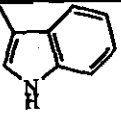
One part of sample buffer was added to four parts of sample. The mixture was heated 5 minutes in boiling water before loading to the gel.

Electrophoresis buffer, 1 litre

Tris (hydroxymethyl)-aminomethane	3.0 g
Glycine	14.4 g
SDS	1.0 g

Adjusted volume to 1 litre with distilled water
(pH should be approximately 8.3).

Appendix 2 Reactivities of amino acid side chains (continued)


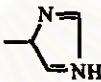
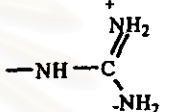
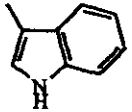
Reagent	—NH_2	—SH				—COOH		—S—S—	—S—CH_3
Phenylglyoxal	++	-	-	-	+++	-	-	-	-
Photooxidation	-	+++	±±	+++	-	-	+++	±	+++
Sodium borohydride	-	+++ ^b	++ ^b	++ ^b	-	-	-	-	-
Succinic anhydride	+++	+++	-	-	-	-	+++	-	-
Sulfite	-	+++	+++	+++	-	-	-	-	-
Sulfonyl halides	+++	+++	+++	-	-	-	+	-	+
Tetranitromethane	-	+++	+++	-	-	-	+	-	+
Tetrathionate	-	+++	-	-	-	-	-	-	-
Thiols	-	-	-	-	-	-	-	+++	-
Trinitrobenzenesulfonic acid	+++	++ ^b	-	-	-	-	-	-	-
Water-soluble carbodiimide and nucleophile	±	±	±	-	-	+++	-	-	-

^a -, +, ++, and +++ indicate relative reactivities; ±, ±±, and ±±± likewise indicate relative reactivities which may or may not be attained depending on the condition used.

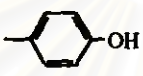
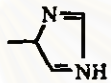
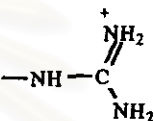
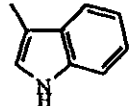
^b Spontaneously reversible under the reaction conditions or upon dilution, regenerating original group.

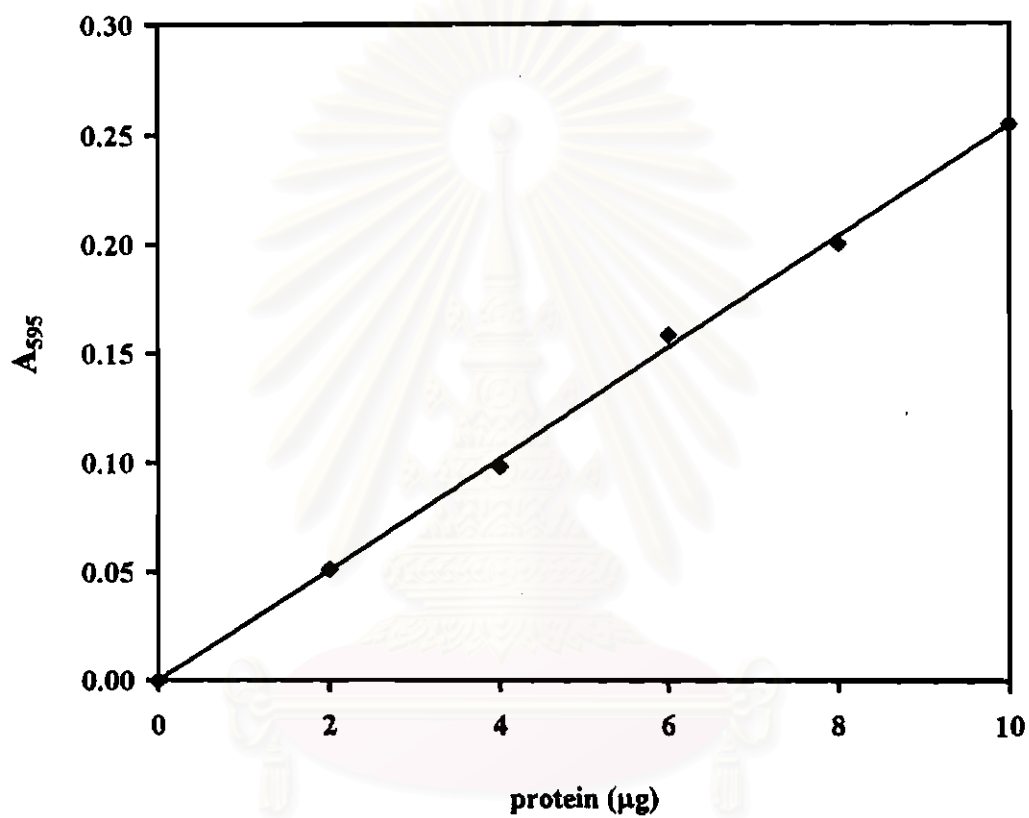
^c Easily reversible, regenerating original group.

Appendix 2 Reactivities of amino acid side chains (continued)

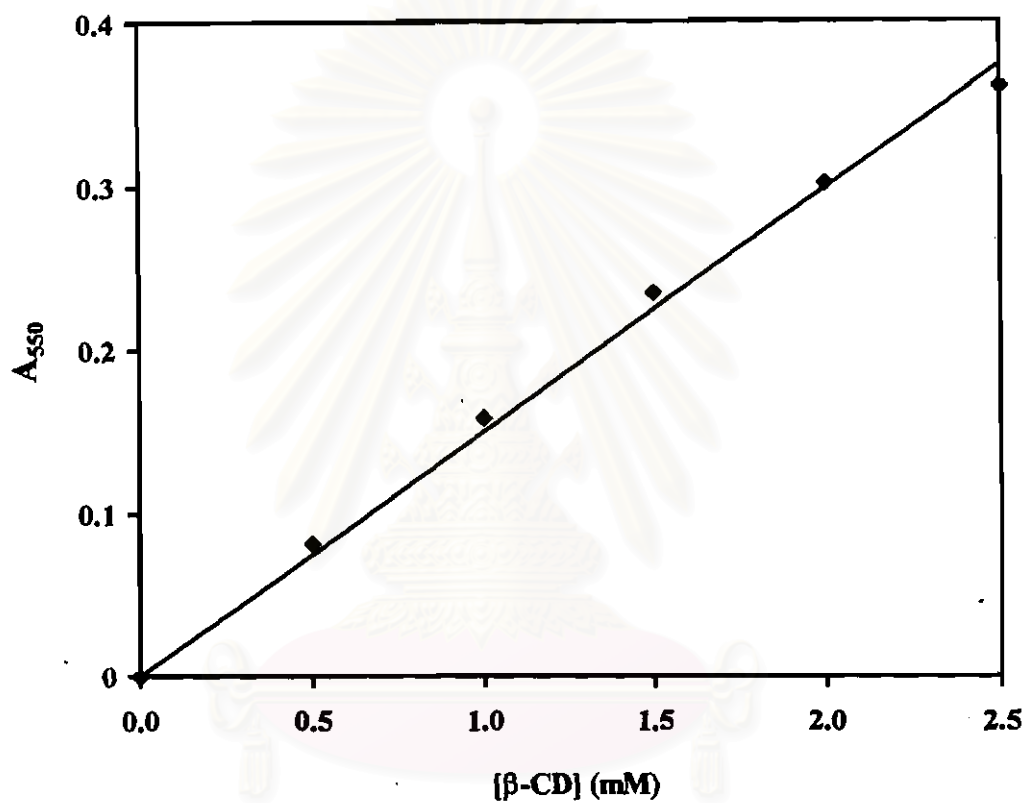
Reagent	—NH_2	—SH				—COOH		—S—S—	—S—CH_3
<i>N</i> -ethylmaleimide	++	+++	-	-	-	-	-	-	-
Ethyl thio-trifluoroacetate	+++ ^b	-	-	-	-	-	-	-	-
Formaldehyde	+++	+++	+++	+++	+	-	+	-	-
glyoxal	++	-	-	-	+++	-	-	-	-
Haloacetates	+	+++	-	+	-	-	-	-	+
Hydrogen peroxide	-	+++	-	-	-	-	+	+	+++
2-hydroxy-5-nitrobenzyl bromide	-	++	-	-	-	-	+++	-	-
Iodine	-	+++	+++	+++	-	-	-	-	-
<i>O</i> -iodosobenzoate	-	+++	-	-	-	-	-	-	-
Maleic anhydride	+++ ^e	++ ^e	++ ^b	++ ^b	-	-	-	-	-
<i>p</i> -mercuribenzoate	-	+++	-	-	-	-	-	-	-
Methanol/ HCl	-	-	-	-	-	+++	-	-	-
2-methoxy-5-nitropropone	+++ ^e	-	-	-	-	-	-	-	-
Methyl acetimidate	+++	-	-	-	-	-	-	-	-
<i>O</i> -methylisourea	+++	-	-	-	-	-	-	-	-
Nitrous acid	+++	+++	±	-	-	-	-	+	-
Performic acid	-	+++	-	-	-	-	++	+++	+++

Appendix 2 Reactivities of amino acid side chains (Means and Feeney, 1971)

Reagent	—NH_2	—SH				—COOH		—S—S—	—S—CH_3
Acetic anhydride	+++	+++ ^b	+++ ^c	+++ ^b	-	-	-	-	-
N-acetylimidazole	±±	+++ ^b	+++ ^c	+++ ^b	-	-	-	-	-
acrylonitrile	±±	+++	-	-	-	-	-	-	-
Aldehyde/ NaBH ₄	+++	-	-	-	-	-	-	-	-
N-bromosuccinimide	-	+++	++	+	-	-	+++	-	-
N-carboxyanhydrides	+++	-	-	-	-	-	-	-	-
Cyanate	+++	+++ ^b	++ ^b	+ ^b	-	+ ^b	-	-	-
Cyanogen bromide	-	+	-	-	-	-	-	-	+++
1,2-cyclohexanedione	±	-	-	-	+++	-	-	-	-
Diacetyl trimer	+	-	-	-	+++	-	-	-	-
Diazoacetates	-	++	-	-	-	+++	-	-	-
Diazonium salts	+++	+	+++	+++	+	-	+	-	-
Diethylpyrocarbonat	+++	-	-	+++ ^c	-	-	-	-	-
Diketone	+++ ^c	-	+	-	-	-	-	-	-
Dinitrofluorobenzene	+++	+++	++	++	-	-	-	-	-
5,5'-dithiobis (2-nitrobenzoic acid)	-	+++ ^c	-	-	-	-	-	-	-
Ethyleneimine	-	+++	-	-	-	-	-	-	+

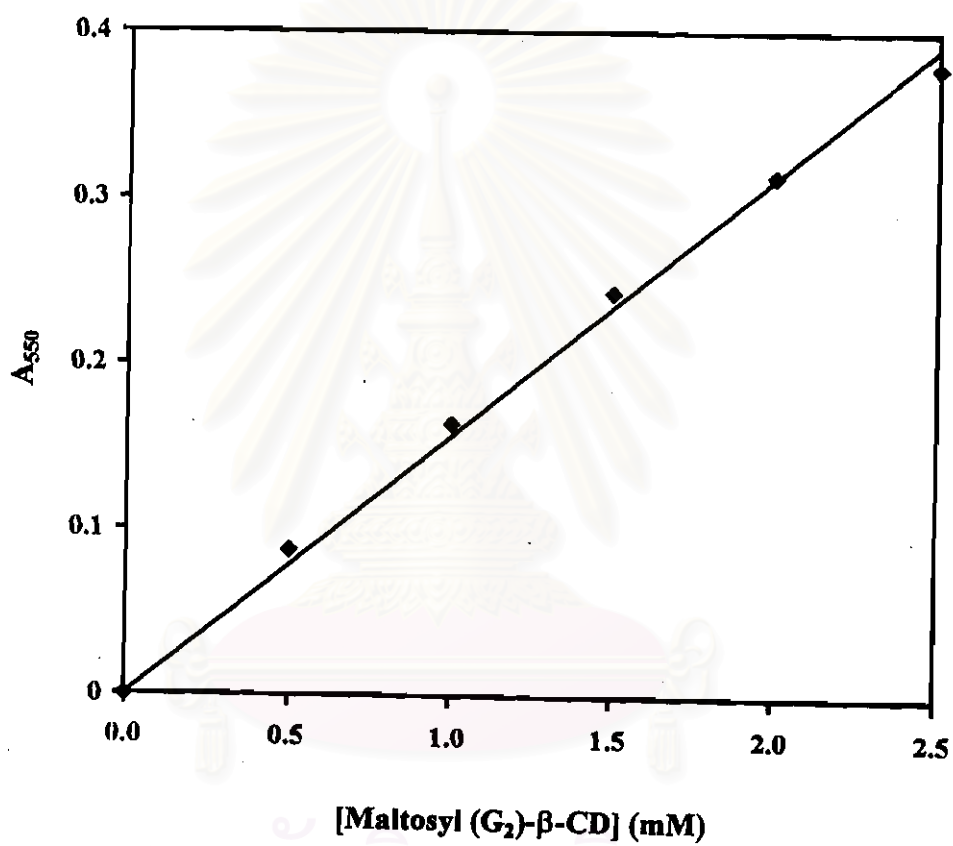
APPENDIX 3: Standard curve for protein determination by Bradford's method

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

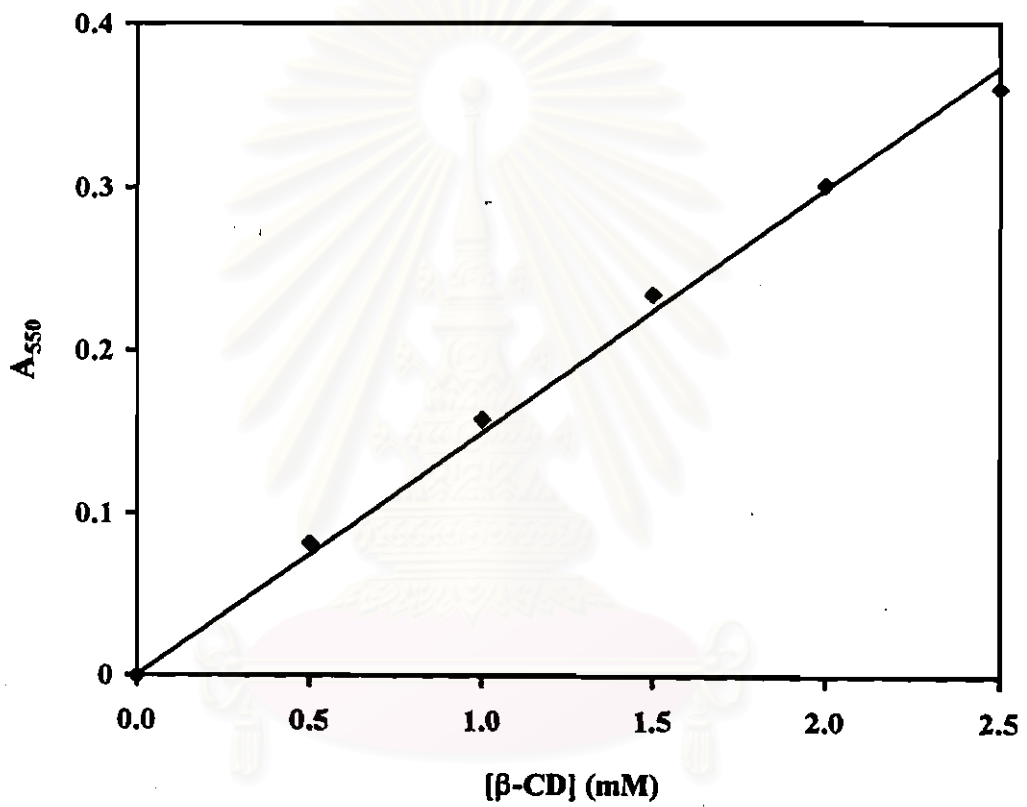
APPENDIX 4: Standard curve of β -cyclodextrin by phenolphthalein method

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

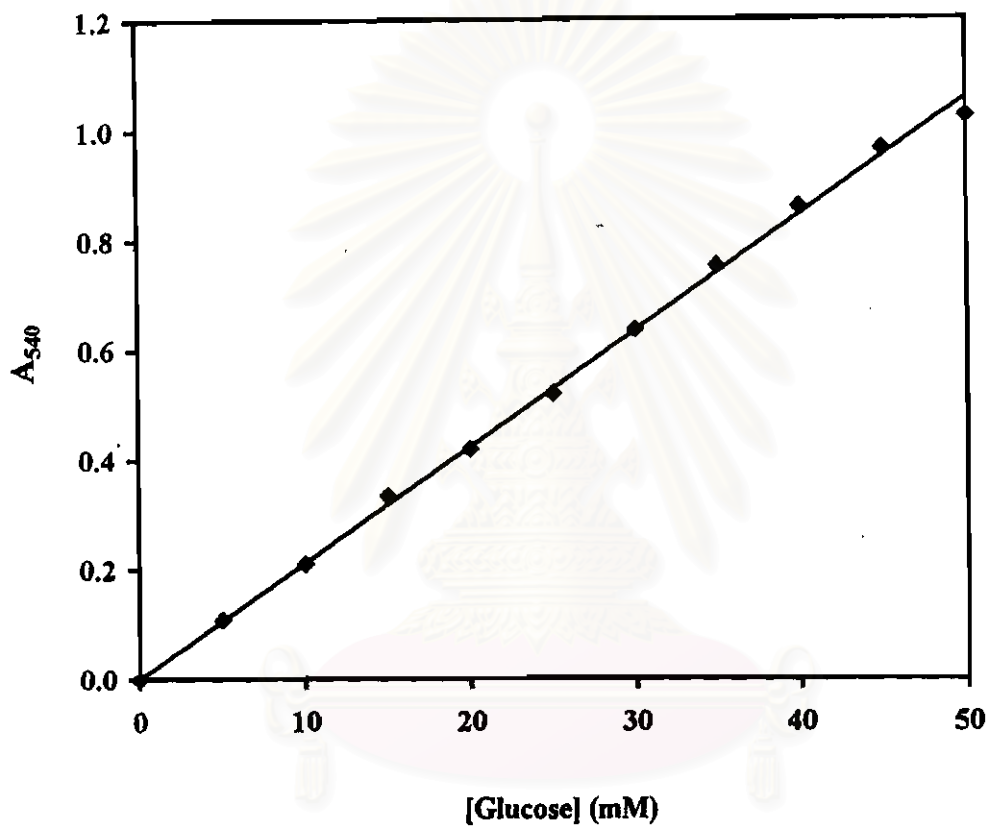
APPENDIX 5: Standard curve of maltosyl (G₂)- β -cyclodextrin by phenolphthalein method



สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

APPENDIX 6: Standard curve of methyl- β -cyclodextrin by phenolphthalein method

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

APPENDIX 7: Standard curve of glucose by dinitrosalicylic acid method

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

BIOGRAPHY

Miss Anchalee Tongsimma was born on May 3, 1973. She graduated with the Bachelor Degree of Science in Biochemistry from Chulalongkorn University in 1995 and continued studying for Master in Biochemistry Program.



สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย